## S2 Table. Representative function(s) and supporting published experimental data for HCV-human PPI network modules 1–7

Network module	Description (literature evidence)
Module 1:	(a) HCV E1/E2 can promote actin reorganization to induce internalization of the
Actin cytoskeleton	virion <sup>1</sup> .
organization and	(b) Adhesion between host cells can prevent invasion of pathogens <sup>2</sup> . However, during
cell adhesion	HCV entry, OCLN, a cell adhesion protein, interacts with HCV E2 <sup>3</sup> and relocalizes
	HCV to tight junctions where HCV internalization occurs <sup>1, 4, 5</sup> .
Module 2:	HCV infection has been associated with insulin resistance, a risk factor for
Peptide hormone	hepatocellular carcinoma (HCC) <sup>6</sup> .
processing	
Module 3:	HCV E1/E2 can trigger the unfolded protein response (UPR) <sup>7</sup> , a cellular homeostatic
Cellular	process triggered in response to stress induced by unfolded/misfolded proteins at the
homeostasis	endoplasmic reticulum <sup>8</sup> . HCV-triggered UPR can lead to autophagy, which is also a
	cellular homeostatic response <sup>9</sup> .
Module 4:	HCV infection enhances the growth of HCC <sup>10</sup> . For instance, in HCV-associated
Growth	cirrhosis tissues, upregulation of IGFBP3, an IGF-binding protein, can potentiate IGF
	signaling and contribute to tumor growth <sup>11, 12</sup> .
Module 5:	(a) During HCV infection, apoptosis in infected cells is triggered to decrease the
Apoptosis and cell	spread of the virus <sup>13, 14</sup> . However, HCV has evolved several anti-apoptotic
junction	strategies to increase its survival: 1) HCV E1 induces the production of reactive
organization	oxygen species (ROS) and the phosphorylation of STAT3 <sup>15</sup> , leading to cell
	survival; 2) HCV infection activates CHUK <sup>16</sup> , which can upregulate the activity
	of NF- $\kappa$ B and lead to enhancement of the expression of anti-apoptotic genes <sup>17</sup> ; 3)
	HCV E1/E2 can induce the phosphorylation of AKT proteins and activate PIK3-
	AKT signaling to enhance cell survival <sup>18</sup> .
	(b) Based on an <i>in vitro</i> study, HCV infection can regulate tight junction organization
	by downregulating the expression of CLDN1 and OCLN, which may lead to the
	observed morphological and functional alterations of HCV-infected hepatocytes <sup>19</sup> .
Module 6:	(a) Several studies have demonstrated that clathrin-dependent endocytosis is the main
Endocytosis and	route for HCV to enter human hepatocytes, mediated by HCV $E1/E2^{20-22}$ .
cell-cell signaling	(b) Upon HCV infection, cell-cell signaling in the host cell can be triggered to induce
	a systematic immune response against viral infection <sup>23</sup> .
Module 7:	(a) Receptor signaling, e.g., EGFR signaling and its downstream signaling by HRAS <sup>24</sup>
Receptor signaling	and PI3K-AKT <sup>18</sup> are pivotal to HCV entry <sup>24</sup> . These signaling events stimulate
and cytoskeleton	hepatocyte proliferation, which may contribute to hepatocellular carcinogenesis <sup>25,</sup>
organization	$^{26}$ .
	(c) During HCV entry, the virus induces cytoskeleton reorganization so that it is
	relocalized to the tight junction, where internalization and endocytosis occur <sup>1, 4, 5</sup> .
	PTPN11 upregulated by HCV infection <sup>27</sup> may reorganize the cytoskeleton <sup>28</sup> .

## S2 Table references

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