

**S3 Table. Published experimental evidence for relations between SLiMs and R6 proteins and between R6 proteins and module functions in Fig 3**

<b>R6 protein</b>	<b>Interacting HCV SLiM</b>	<b>Description</b>
<b>ABL1</b>	LIG_SH2_STAT5, LIG_SH3_3	Proteins of the ABL tyrosine kinase family, to which ABL1 belongs, regulate <i>cell-cell adhesion</i> (module 1) <sup>1</sup> . ABL1 also modulates the organization of <i>F-actin cytoskeleton</i> (module 1 and module 7) <sup>1,2</sup> .
<b>AKT1</b>	MOD family*	(a) Active AKTs regulate many cellular processes, including <i>apoptosis</i> (module 5) <sup>3</sup> . (b) HCV E2 induces the phosphorylation of AKT, which enhances HCV entry. This phosphorylation is mediated by the interactions between HCV E2 and two HCV entry host factors, CD81 and CLDN1 <sup>4</sup> .
<b>CHUK</b>	MOD family*	(a) CHUK is a subunit of the IKK complex, which when active can induce the NF- $\kappa$ B pathway to inhibit <i>apoptosis</i> (module 5) <sup>5</sup> . (b) HCV infection induces CHUK activity <sup>6</sup> .
<b>CSNK2A1, CSNK2A2</b>	MOD family*	CSNK2A1 and CSNK2A2 are subunits of CK2 protein kinase <sup>7</sup> . CK2 regulates <i>cell proliferation</i> (module 4) <sup>8</sup> and <i>ion channel</i> activity <sup>9</sup> , which is a function associated with module 8 although its enrichment in module 8 was not considered significant (P = 0.055).
<b>CTTN</b>	LIG_SH3_3	CTTN regulates <i>actin cytoskeleton organization</i> (module 1) by binding and regulating the Arp2/3 complex <sup>10</sup> .
<b>DLG1, DLG4</b>	LIG_SH3_3	DLG1 and DLG4 are PDZ domain-containing proteins that can regulate <i>cell-cell junctional signaling and organization</i> (module 6) <sup>11</sup> .
<b>GABARAP</b>	LIG_LIR_Gen_1, LIG_LIR_Nem_3	GABARAP, a functional homologue of LC3, is a key protein involved in the initial steps of autophagosome formation, which is a critical step of autophagy, a process that regulates <i>cellular homeostasis</i> (module 3) <sup>12</sup> .
<b>GRB2</b>	LIG_SH2_STAT5, LIG_SH3_3	(a) GRB2 plays a role in modulating <i>actin-based cytoskeleton functions</i> (module 1 and module 7) <sup>13</sup> . GRB2 is also an adaptor protein that mediates EGFR downstream <i>signaling</i> (module 7) and promotes <i>cell proliferation</i> (module 4) <sup>14</sup> . (b) Silencing of GRB2 expression inhibits HCV entry <sup>15</sup> .
<b>GRK5</b>	MOD family*	GRK5 is a G-protein-coupled receptor kinase that can enhance the <i>growth</i> (module 4) of cancer <sup>16, 17</sup> .
<b>MLLT4</b>	LIG_FHA_1, LIG_FHA_2	MLLT4 is a cell-cell junction-associated protein, which bridges <i>Ras signaling toward regulation of cell-cell contact</i> (module 6) <sup>18</sup> .
<b>NCK1</b>	LIG_SH2_STAT5, LIG_SH3_3	NCK1 regulates <i>actin remodeling</i> (module 1 and module 7) by activating the WASp/Arp2/3 pathway, linking this pathway to <i>receptor signaling</i> (module 7) <sup>19</sup> .

**S3 Table. Published experimental evidence for relations between SLiMs and R6 proteins and between R6 proteins and module functions in Fig 3 (continued)**

<b>R6 protein</b>	<b>Interacting HCV SLiM</b>	<b>Description</b>
<b>NEDD4</b>	DOC_WW_Pin1_4	NEDD4 is a ubiquitin ligase that can mediate ubiquitination of the <i>ion transporter</i> NHE1 <sup>20</sup> , a module 8 function although not found to be enriched to a statistical significant level (P = 0.055).
<b>NEDD4L</b>	DOC_WW_Pin1_4	NEDD4L is a ubiquitin ligase that can regulate <i>epithelial sodium transport</i> (module 8) <sup>21</sup> .
<b>PIK3R1</b>	LIG_SH2_STAT5	(a) PIK3R1, a regulatory subunit of PI3K, is a pivotal member of the PI3K-AKT <i>signaling pathway</i> (module 7) <sup>22</sup> and induces <i>actin skeleton reorganization</i> (module 7) in a Cdc42-mediated manner <sup>23</sup> . (b) The PI3K-AKT pathway transiently activated by HCV enhances viral entry <sup>4</sup> .
<b>PRKCA</b>	MOD family*	(a) Activation of PRKCA promotes tumor cell <i>proliferation</i> (module 4) and inhibits <i>apoptosis</i> (module 5) <sup>24</sup> . Furthermore, PRKCA can promote internalization of integrin $\beta$ 1, which is involved in <i>integrin signaling</i> (module 6) <sup>25</sup> , via <i>caveolae-mediated endocytosis</i> (module 6) <sup>25</sup> . (b) PRKCA is a conventional protein kinase C, which plays a role in alpha interferon-mediated HCV clearance <sup>26</sup> .
<b>PRKCD</b>	MOD family*	PRKCD, a member of the kinase C (PKC) family, can mediate cell <i>apoptosis</i> (module 5) <sup>27</sup> .
<b>SRC</b>	LIG_SH2_STAT5, LIG_SH3_3	(a) SRC modulates <i>actin cytoskeleton organization</i> (modules 1 and 7) <sup>28</sup> and the activity of <i>EGFR</i> (module 7) <sup>29</sup> . Activation of SRC is regulated by Na/K-ATPase, an <i>ion transporter</i> (module 8) <sup>30</sup> . (b) The HCV replicon activates SRC by inducing generations of reactive oxygen species <sup>31</sup> .
<b>SHC1</b>	LIG_SH2_STAT5	(a) SHC1 is an adaptor protein that links EGFR to its downstream <i>signaling</i> (module 7) <sup>32</sup> . (b) Silencing of SHC1 expression inhibits HCV entry <sup>15</sup> .
<b>TGFBR1</b>	MOD family*	(a) Formation of the ligand-receptor complex of TGFBR1 and TGF- $\beta$ activates TGF- $\beta$ signaling, which is involved in <i>apoptosis</i> (module 5) <sup>33</sup> . (b) Hepatocytic TGF- $\beta$ signaling, a tumor-suppression signaling, can be shifted to fibrogenesis by HCV infection and increase risk for HCC <sup>34</sup> .

\*MOD family: MOD\_CK1\_1, MOD\_CK2\_1, MOD\_GSK3\_1, MOD\_NEK2\_1, MOD\_NEK2\_2, and MOD\_ProDKin\_1.

### S3 Table references

1. Zandy NL, Playford M, Pendergast AM. Abl tyrosine kinases regulate cell–cell adhesion through Rho GTPases. *PNAS*. 2007;104(45):17686-91.
2. Woodring PJ, Litwack ED, O'Leary DDM, Lucero GR, Wang JYJ, Hunter T. Modulation of the F-actin cytoskeleton by c-Abl tyrosine kinase in cell spreading and neurite extension. *The Journal of Cell Biology*. 2002;156(5):879-92.
3. Song G, Ouyang G, Bao S. The activation of Akt/PKB signaling pathway and cell survival. *Journal of Cellular and Molecular Medicine*. 2005;9(1):59-71.
4. Liu Z, Tian Y, Machida K, Lai MMC, Luo G, Fong SKH, et al. Transient activation of the PI3K-AKT pathway by hepatitis C virus to enhance viral entry. *J Biol Chem*. 2012;287(50):41922-30.
5. Israël A. The IKK Complex, a Central Regulator of NF- $\kappa$ B Activation. *Cold Spring Harb Perspect Biol*. 2010;2(3).
6. Li Q, Pène V, Krishnamurthy S, Cha H, Liang TJ. Hepatitis C virus infection activates an innate pathway involving IKK- $\alpha$  in lipogenesis and viral assembly. *Nat Med*. 2013;19(6):722-9.
7. Litchfield DW, Bosc DG, Canton DA, Saulnier RB, Vilks G, Zhang CJ. Functional specialization of CK2 isoforms and characterization of isoform-specific binding partners. *Mol Cell Biochem*. 2001;227(1-2):21-9.
8. Lebrin F, Chambaz EM, Bianchini L. A role for protein kinase CK2 in cell proliferation: evidence using a kinase-inactive mutant of CK2 catalytic subunit alpha. *Oncogene*. 2001;20(16):2010-22.
9. Bachhuber T, Almacá J, Aldehni F, Mehta A, Amaral MD, Schreiber R, et al. Regulation of the Epithelial Na<sup>+</sup> Channel by the Protein Kinase CK2. *J Biol Chem*. 2008;283(19):13225-32.
10. Cosen-Binker LI, Kapus A. Cortactin: The Gray Eminence of the Cytoskeleton. *Physiology*. 2006;21(5):352-61.
11. Roberts S, Delury C, Marsh E. The PDZ protein discs-large (DLG): the 'Jekyll and Hyde' of the epithelial polarity proteins. *FEBS Journal*. 2012;279(19):3549-58.
12. Mariño G, Niso-Santano M, Baehrecke EH, Kroemer G. Self-consumption: the interplay of autophagy and apoptosis. *Nature Reviews Molecular Cell Biology*. 2014;15(2):81-94.
13. Carlier M-F, Nioche P, Broutin-L'Hermite I, Boujemaa R, Clainche CL, Egile C, et al. GRB2 links signaling to actin assembly by enhancing interaction of neural wiskott-aldrich syndrome protein (N-WASp) with actin-related protein (ARP2/3) complex. *J Biol Chem*. 2000;275(29):21946-52.
14. Downward J. The GRB2/Sem-5 adaptor protein. *FEBS Letters*. 1994;338(2):113-7.
15. Zona L, Lupberger J, Sidahmed-Adrar N, Thumann C, Harris HJ, Barnes A, et al. HRas

- signal transduction promotes hepatitis C virus cell entry by triggering assembly of the host tetraspanin receptor complex. *Cell Host Microbe*. 2013;13(3):302-13.
16. Kim JI, Chakraborty P, Wang Z, Daaka Y. G-Protein Coupled Receptor Kinase 5 Regulates Prostate Tumor Growth. *The Journal of Urology*. 2012;187(1):322-9.
  17. Kaur G, Kim J, Kaur R, Tan I, Bloch O, Sun MZ, et al. G-protein coupled receptor kinase (GRK)-5 regulates proliferation of glioblastoma-derived stem cells. *Journal of Clinical Neuroscience*. 2013;20(7):1014-8.
  18. Yamamoto T, Harada N, Kano K, Taya S-i, Canaani E, Matsuura Y, et al. The Ras Target AF-6 Interacts with ZO-1 and Serves as a Peripheral Component of Tight Junctions in Epithelial Cells. *The Journal of Cell Biology*. 1997;139(3):785-95.
  19. Chaki SP, Rivera GM. Integration of signaling and cytoskeletal remodeling by Nck in directional cell migration. *BioArchitecture*. 2013;3(3):57-63.
  20. Simonin A, Fuster D. Nedd4-1 and  $\beta$ -Arrestin-1 Are Key Regulators of Na<sup>+</sup>/H<sup>+</sup> Exchanger 1 Ubiquitylation, Endocytosis, and Function. *J Biol Chem*. 2010;285(49):38293-303.
  21. Rotin D, Staub O. Nedd4-2 and the regulation of epithelial sodium transport. *Front Physio*. 2012;3:212.
  22. Miled N, Yan Y, Hon W-C, Perisic O, Zvelebil M, Inbar Y, et al. Mechanism of two classes of cancer mutations in the phosphoinositide 3-kinase catalytic subunit. *Science*. 2007;317(5835):239-42.
  23. Jiménez C, Portela RA, Mellado M, Rodríguez-Frade JM, Collard J, Serrano A, et al. Role of the Pi3k Regulatory Subunit in the Control of Actin Organization and Cell Migration. *The Journal of Cell Biology*. 2000;151(2):249-62.
  24. Michie AM, Nakagawa R. The link between PKC alpha regulation and cellular transformation. *Immunol Lett*. 2005;96(2):155-62.
  25. Caswell PT, Vadrevu S, Norman JC. Integrins: masters and slaves of endocytic transport. *Nature Reviews Molecular Cell Biology*. 2009;10(12):843-53.
  26. Fimia GM, Evangelisti C, Alonzi T, Romani M, Fratini F, Paonessa G, et al. Conventional Protein Kinase C Inhibition Prevents Alpha Interferon-Mediated Hepatitis C Virus Replicon Clearance by Impairing STAT Activation. *J Virol*. 2004;78(23):12809-16.
  27. Reyland ME. Protein kinase C $\delta$  and apoptosis. *Biochemical Society Transactions*. 2007;35(5):1001-4.
  28. Yeatman TJ. A renaissance for SRC. *Nat Rev Cancer*. 2004;4(6):470-80.
  29. Biscardi JS, Maa M-C, Tice DA, Cox ME, Leu T-H, Parsons SJ. c-Src-mediated Phosphorylation of the Epidermal Growth Factor Receptor on Tyr845 and Tyr1101 Is Associated with Modulation of Receptor Function. *J Biol Chem*. 1999;274(12):8335-43.
  30. Weigand KM, Swarts HGP, Fedosova NU, Russel FGM, Koenderink JB. Na,K-ATPase activity modulates Src activation: A role for ATP/ADP ratio. *Biochimica et Biophysica Acta (BBA) - Biomembranes*. 2012;1818(5):1269-73.

31. Waris G, Turkson J, Hassanein T, Siddiqui A. Hepatitis C virus (HCV) constitutively activates STAT-3 via oxidative stress: Role of STAT-3 in HCV replication. *J Virol*. 2005;79(3):1569-80.
32. Ravichandran KS. Signaling via Shc family adapter proteins. *Oncogene*. 2001;20(44):6322-30.
33. Massague J. TGF beta signalling in context. *Nat Rev Mol Cell Bio*. 2012;13(10):616-30.
34. Matsuzaki K, Murata M, Yoshida K, Sekimoto G, Uemura Y, Sakaida N, et al. Chronic inflammation associated with hepatitis C virus infection perturbs hepatic transforming growth factor  $\beta$  signaling, promoting cirrhosis and hepatocellular carcinoma. *Hepatology*. 2007;46(1):48-57.