- 1 Mitogen-activated protein kinases (MAPKs) regulate IL-6 over-
- 2 production during concomitant influenza virus and Staphylococcus
- 3 aureus infection

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### Supplementary

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#### Methods

#### Additional cell lines and bacterial strain

The human hepatic cell line HepG2 (a kind gift from Ute Albrecht, University 29 30 of Duesseldorf, Germany) was cultivated in DMEM/F-12 medium 1:1. The human monocytic cell line THP-1 was cultivated in RPMI 1640 with 1% 31 glutamine. The murine lewis lung carcinoma (LLC) and mouse embryonic 32 33 fibroblast (MEF) cell lines were cultivated in DMEM. Each medium was 34 supplemented with 10% FBS. Primary human umbilical vein endothelial cells 35 (HUVEC) were cultivated in endothelial cell growth medium with supplement 36 mix (PromoCell, Heidelberg, Germany). 37 THP-1 cells (6-well: 2×10<sup>6</sup>, 12-well: 1×10<sup>6</sup>) were stimulated with 10 ng ml<sup>-1</sup> phorbol-12-myristate-13-acetate (PMA, AppliChem, Darmstadt, Germany) for 38 differentiation into macrophages 16 h prior to infection. LLC (6-well: 0.3×10<sup>6</sup>) 39 and MEF cells (6-well: 0.25×10<sup>6</sup>) were seeded 16 h and HUVECs (6-well: 40 0.5×10<sup>6</sup>) were seeded 48 h prior to infection. During infection culture medium 41 42 supplemented as indicated in the method section was used. 43 The mouse-adapted S. aureus strain LS1 was cultivated as described in the 44 method section.

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#### Plaque assay

47 Standard plague titration was performed as described earlier<sup>50</sup>.

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#### Transfer of conditioned medium

HepG2 cells (12-well:  $0.3\times10^6$ ) were seeded in 1 ml culture medium 40 h prior to stimulation. Calu-3 cells were infected as described in the method section. Supernatants of three 6-wells were pooled and sterile filtrated with a 0.2 µm filter to avoid transfer of living pathogens. HepG2 cells were washed and stimulated with 800 µl conditioned medium for 20 h at 37°C, 5% CO<sub>2</sub>.

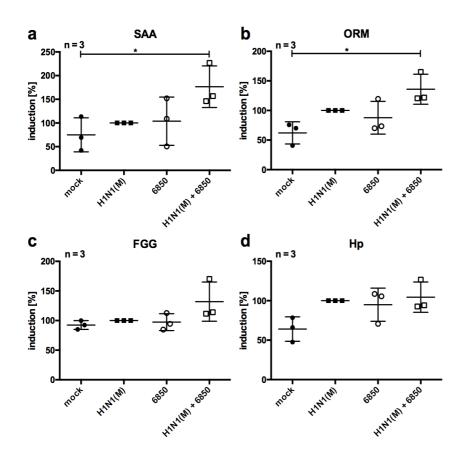
### **Actinomycin D treatment**

A549 cells were infected with IV H1N1(M) (MOI 5) and *S. aureus* 6850 (MOI 50) as described in the material and method section. At 6 h p.i. cells were lysed, or washed and incubated in 1 ml of DMEM/BA supplemented with 1 μg ml<sup>-1</sup> actinomycin D (Santa Cruz Biotechnology, Dallas, USA) for further 0.5, 1 or 2 h at 37°C, 5% CO<sub>2</sub>. Cells were lysed and total cellular RNA was isolated as described in the method section. For investigation of *IL*-6 mRNA stability, the amount of *IL*-6 mRNA was measured for each time point indicated by RT-qPCR and *IL*-6 levels were compared to untreated cells (0 h).

## Stimulation of cells with recombinant human IFN $\beta$

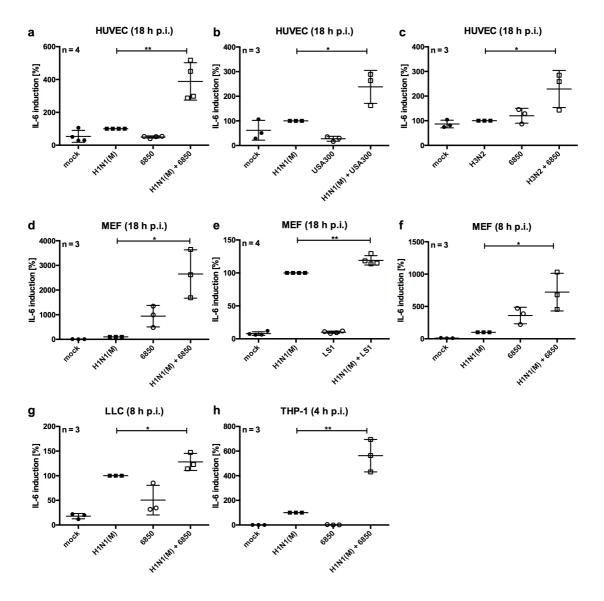
A549 cells were incubated in 1 ml of DMEM/INV with *S. aureus* 6850 (MOI 50) for 3 h at 37°C, 5% CO<sub>2</sub>. Extracellular bacteria were removed by gentamicin treatment for 0.5 h at 37°C, 5% CO<sub>2</sub>. Cells were incubated in 1 ml of DMEM/BA supplemented with 100 U recombinant human IFNβ (PBL Assay Science, Piscataway, USA) or same volume of PBS for 4.5 h at 37°C, 5% CO<sub>2</sub>.

### 73 Supplementary figures



Supplementary figure S1: Conditioned medium of IV and *S. aureus* super-infected Calu-3 cells induces expression of APPs in HepG2 cells.

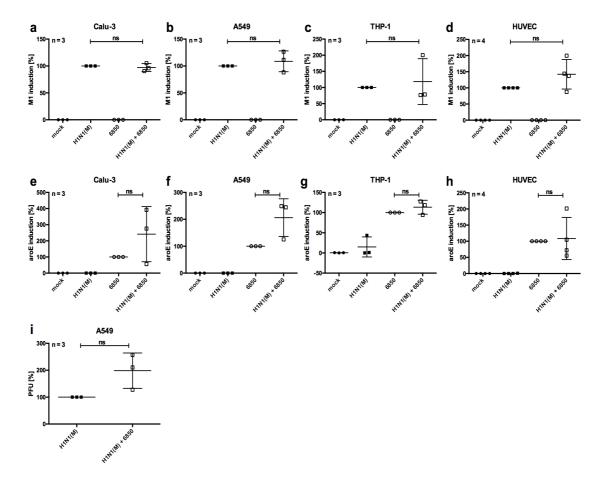
Calu-3 cells were infected with IV H1N1(M) (MOI 5) for 0.5 h and super-infected with *S. aureus* 6850 (MOI 50). Extracellular bacteria were removed by gentamicin treatment 3 h post bacterial infection. At 8 h p.i. cell supernatants were sterile filtrated and added to HepG2 cells for another incubation of 20 h. Levels of SAA (a), ORM (b), FGG (c) and Hp (d) were measured in duplicates. Means  $\pm$  SD of three independent experiments are shown. Samples, which were treated with conditioned medium of IV-infected Calu-3 cells, were arbitrarily set 100%. After normalisation, one-way ANOVA followed by Dunnett's multiple comparison tests were used to compare mock and IV and/or *S. aureus* infected samples (\* p < 0.05).



Supplementary figure S2: Increased *IL-6* expression during super-infection is cell line independent.

HUVEC (a – c), MEF (d – f), LLC (g) or THP-1 (h) cells were infected with H1N1(M) (MOI 0.5 (a, b, d, e), MOI 3 (f – h)) or H3N2 (MOI 0.5 (c)) for 0.5 h and super-infected with *S. aureus* 6850 (MOI 0.1 (a, c, d), MOI 10 (f, g), MOI 1 (h)), MRSA USA300 (b) or LS1 (e) (MOI 0.01). Extracellular bacteria were removed by antibiotic treatment 3 h post bacterial infection. *IL-6* mRNA levels were measured at 18 h (a – e), 8 h (f, g) or 4 h p.i. (h). Means  $\pm$  SD of at least three independent experiments are shown. Levels of IV-infected samples

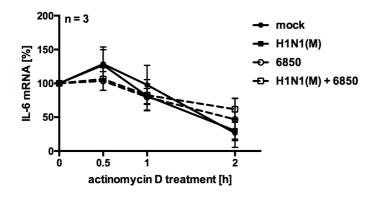
- 97 were arbitrarily set as 100%. After normalisation, two-tailed unpaired t-tests
- 98 were performed for comparison between IV-infected and IV/S. aureus super-
- 99 infected samples (\* p < 0.05, \*\* p < 0.01).



Supplementary figure S3: Levels of viral *M1* and bacterial *aroE* are not significantly changed upon super-infection with IV and *S. aureus*.

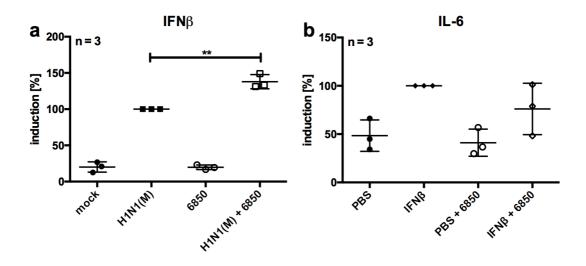
Calu-3 (a, e), A549 (b, f, i), THP-1 (c, g) or HUVEC (d, h) cells were infected with H1N1(M) (MOI 5 (a, b, e, f, i), MOI 3 (c, g), MOI 0.5 (d, h)) and superinfected with *S. aureus* 6850 (MOI 50 (a, b, e, f, i), MOI 1 (c, g), MOI 0.1 (d, h)). Extracellular bacteria were removed by antibiotic treatment 3 h post bacterial infection. M1 (a – d) and aroE (e – h) levels were measured at 8 h (a, b, e, f), 4 h (c, g) or 18 h p.i. (d, h). Infectious virus titres were determined by standard plaque assay at 8 h p.i. (i). Means  $\pm$  SD of at least three independent experiments are shown. Levels of IV (a – d, i) or *S. aureus* (e – h) infected samples were arbitrarily set as 100%. After normalisation, two-

- tailed unpaired t-tests were performed for comparison between single and
- super-infected samples (ns not significant).



Supplementary figure S4: *IL-6* mRNA stability is not altered by infection or super-infection with IV H1N1(M) and/or *S. aureus* 6850.

A549 cells were infected with IV H1N1(M) (MOI 5) for 0.5 h and super-infected with *S. aureus* 6850 (MOI 50). Extracellular bacteria were removed by gentamicin treatment 3 h post bacterial infection. At 6 h p.i. cells were lysed or treated with 1 μg ml<sup>-1</sup> actinomycin D for 0.5, 1 or 2 h. *IL-6* mRNA levels were measured in duplicates by RT-qPCR. Means ± SD of three independent experiments are shown. Untreated samples (0 h) were arbitrarily set as 100%.



Supplementary figure S5: Increased IFN $\beta$  synthesis after super-infection does not induce hyper-transcription of *IL-6*.

A549 cells were infected with IV H1N1(M) (MOI 5) for 0.5 h and super-infected with *S. aureus* 6850 (MOI 50) (a) or cells were directly incubated with *S. aureus* 6850 (MOI 50) (b). Extracellular bacteria were removed by gentamicin treatment 3 h post bacterial infection. Subsequently cells were left untreated (a) or were stimulated with 100 U of recombinant human IFNβ or PBS as control (b). *IFNβ* (A) and *IL*-6 (b) mRNA levels were measured in duplicates at 8 h p.i.. Means  $\pm$  SD of three independent experiments are shown. IV-infected (a) or IFNβ-stimulated (b) samples were arbitrarily set as 100%. After normalisation, two-tailed unpaired t-test was performed for comparison of IV H1N1(M)-infected and IV H1N1(M)/*S. aureus* 6850 super-infected samples (\*\* p < 0.01).

# 141 Supplemental table S6: Primer sequences for RT-qPCR

142 The tables list the sequences of used RT-qPCR primers.

## 143 human

Gene	Forward (5' – 3')	Reverse (5' – 3')			
CCL3	AGTTCTCTGCATCACTTGCTG	CGGCTTCGCTTGGTTAGGAA			
CCL5	CGGCACGCCTCGCTGTCATC	GCAAGCAGAAACAGGCAAAT			
CXCL8	ACTGAGAGTGATTGAGAGAGTGGAC	AACCCTCTGCACCCAGTTTTC			
FGG	GACGCTGCTACTTTGAAGTCC	TGGATTTGCACCGTGTCTTTG			
GAPDH	GCCAATTCCATGGCACCGT	GCCCCACTTGATTTTGGAGG			
Нр	CAGCACAGTCCCCGAAAAGAA	CAGTCGCATACCAGGTGTCC			
IL-1β	CAGCTACGAATCTCCGACCAC	GGCAGGGAACCAGCATCTTC			
IL-6	AACCTGAACCTTCCAAAGATGG	TCTGGCTTGTTCCTCACTAGT			
ΙΕΝβ	TCTGGCACAACAGGTAGTAGGC	GAGAAGCACAACAGGAGAGCAA			
SAA	CAAATACTTCCATGCTCGGGG	CGCAGCCTCTAACTTCTCCAC			
ORM	GCTGTTCCTTAGGGACACCAA	TGACATCTGACCTGGGAATGC			
TNFα	ATGAGCACTGAAAGCATGATC	GAGGGCTGATTAGAGAGAGGT			

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## 145 murine

Gene	Forward (5' – 3')	Reverse (5' – 3')			
GAPDH	TGACCACAGTCCATGCCATC	GACGGACACATTGGGGGTAG			
Нр	GCTATGTGGAGCACTTGGTTC	CACCCATTGCTTCTCGTCGTT			
IL-1β	GCAACTGTTCCTGAACTCAACT	ATCTTTTGGGGTCCGTCAACT			
IL-6	TAGTCCTTCCTACCCCAATTT	TTGGTCCTTAGCCACTCCTTC			
ORM	CGAGTACAGGCAGGCAATTCA	ACCTATTGTTTGAGACTCCCGA			
SAA	TGGCTGGAAAGATGGAGACAA	AAAGCTCTCTCTTGCATCACTG			
TNFα	CCCTCACACTCAGATCATCTTCT	GCTACGACGTGGGCTACAG			

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# 148 pathogenic

Gene	Forward (5' – 3')	Reverse (5' – 3')
aroE	CTATCCACTTGCCATCTTTTAT	ATGGCTTTAATATCACAATTCC
M1	TGCAAAAACATCTTCAAGTCTCTG	AGATGAGTCTTCTAACCGAGGTCG

# Supplemental table S7: Raw Ct values of *IL*-6 in different cell lines upon

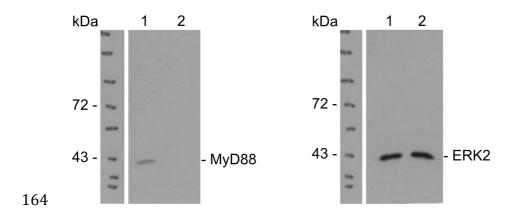
### infection

Analyses of *IL-6* mRNA were determined with Brilliant SYBR Green Mastermix (Agilent, Waldbronn, Germany) by RT-qPCR on a Stratagene Cycler (Agilent Technologies, Santa Clara, USA). Means and SD of at least three independent experiments with at least two biological replicates are shown. Ct values <35 were considered as specific.

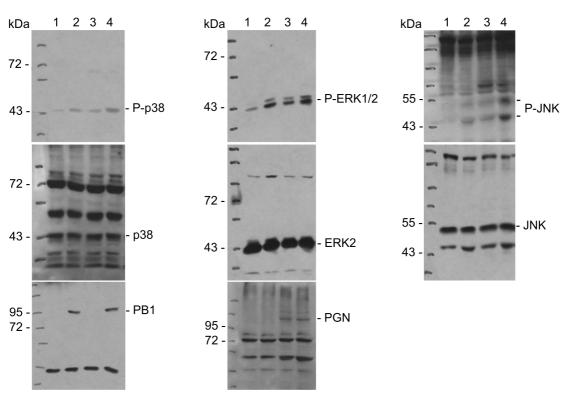
		sample							
		mock		H1N1(M)		6850		H1N1(M) + 6850	
cell line	n	mean	SD	mean	SD	mean	SD	mean	SD
Calu-3 (8 h p.i.)	16	22.34	0.73	17.44	0.69	19.98	0.93	15.86	1.20
A549 (8 h p.i.)	24	30.38	0.89	23.71	0.53	28.62	1.30	22.00	0.49
THP-1 (4 h p.i.)	12	32.93	1.52	25.08	0.81	30.87	1.75	22.52	0.87
HUVEC (18 h p.i.)	16	22.34	1.40	21.68	1.86	22.26	1.55	20.04	0.91

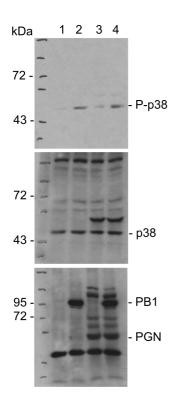
A 2-fold dilution series of cDNA from Calu-3 cells was used for PCR efficiency (E) calculation  $^{61}$ . Both primer pairs for IL-6 and GAPDH show high amplification efficiency, with an estimated E of 98.5% ( $\pm$  1.8%) and 106.2% ( $\pm$  6.1%), respectively.

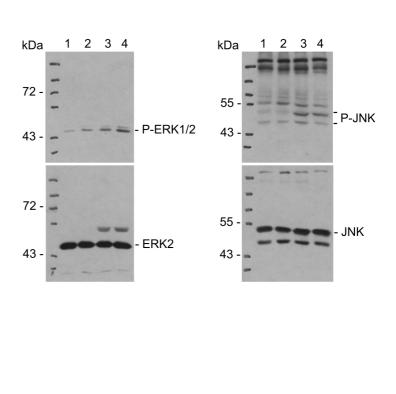


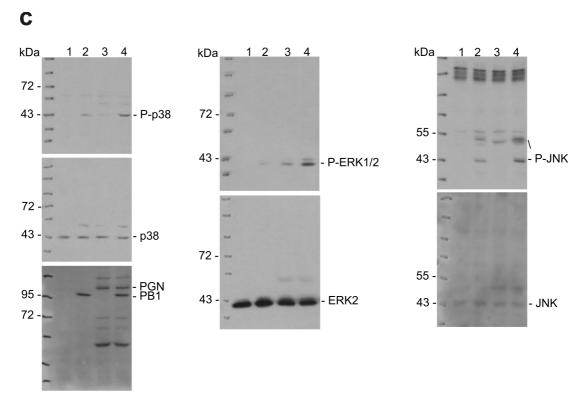


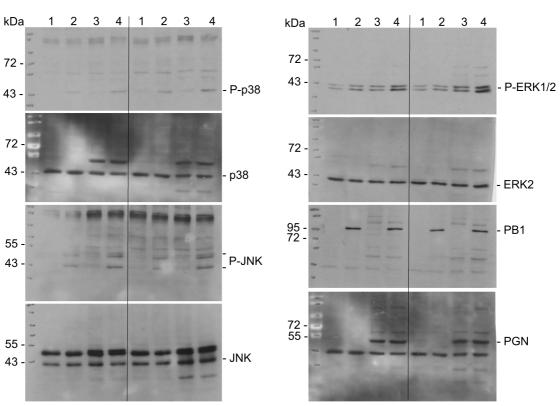




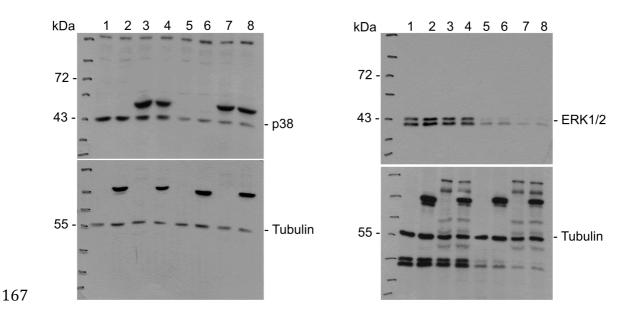








# d



Supplementary figure S8: Original blots of figures 4g, 5a, e, i and 6d, e.

Lane 1: mock-infected, lane 2: IV infection, lane 3: bacteria infection, lane 4: super-infection (a-d). In figure S8d lanes 5 - 8 represent the same infection scheme as described above, but in a siRNA scenario.

Figure S8a represents the original blots of figure 4g. Within figure S8b the upper panel shows the original blots of figure 5a, the lower panel represents the original blots of figure 5e. The upper panel of figure S8c corresponds to figure 5i (experiments performed with *S. aureus* 6850). On the left-hand side of the lower panel original blots of figure 5i (experiments performed with *S. aureus* SH1000) are shown. Blots on the right-hand side stand for figure 5i (experiments performed with *S. aureus* USA300).