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Supplemental Information

C/EBP-Induced Transdifferentiation Reveals Granulocyte-Macrophage

Precursor-like Plasticity of B Cells

Branko Cirovic, Jörg Schönheit, Elisabeth Kowenz-Leutz, Jelena Ivanovska, Christine Klement, Nina Pronina, Valérie Bégay, and Achim Leutz

Supplemental Figures



Ly-6G







F

G

| | upreg | ulated | | |
|---------|----------|--------------|--------|--|
| Ahr | Csf1r | ll18r1 | Notch2 | |
| Arhgdib | Csf2rb | ll1m | Nt5e | |
| Btnl1 | Ctsc | II28a | Pecam1 | |
| Ccl12 | Cybb | <i>I</i> I33 | Plaur | |
| Ccl24 | Ddx58 | Irak2 | Runx3 | |
| Ccl3 | Entpd1 | Irak3 | S100a9 | |
| Ccl6 | Ets1 | Irf7 | Sele | |
| Ccl9 | Fcgr1 | Itgal | Sell | |
| Ccr5 | Fcgr2b | Itgam | Ski | |
| Ccrl2 | Gfi1 | Kir3dl1 | Smad3 | |
| Cd4 | Gpi1 | Klra7 | Stat3 | |
| Cd44 | Gpr183 | Lair1 | Tagap | |
| Cd55 | H2-Ea-ps | Lilrb3 | Tgfbi | |
| Cd69 | Hcst | Lilrb4 | Tlr1 | |
| Cdkn1a | Hfe | Ltb4r1 | Trem1 | |
| Ceacam1 | Hlx | Mbl2 | Tyrobp | |
| Cfb | lfih1 | Muc1 | Vcam1 | |
| Csf1 | lfitm1 | Nfil3 | Vtn | |

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|---|----|-----|----|----|-----|----------|
| u | υw | 111 | eq | uı | aı | eu |

| Aicda | Ciita | Klrc3 | Tnfrsf13c |
|-------|--------------|----------|-----------|
| Bcl3 | Cmklr1 | Lta | Traf4 |
| Btla | Cxcr2 | Ltb | Xcr1 |
| C1qa | Cxcr6 | Masp1 | |
| C1s | Fas | Masp2 | |
| C8a | Fn1 | Pax5 | |
| Ccr6 | lcam2 | Pdcd1lg2 | |
| Ccr7 | lfna2 | Pla2g2a | |
| Ccr8 | lfng | Pou2f2 | |
| Cd160 | lkzf4 | Rag1 | |
| Cd163 | <i>l</i> /10 | Rag2 | |
| Cd2 | ll17b | Rorc | |
| Cd22 | <i>II20</i> | Slamf1 | |
| Cd244 | <i>I</i> /22 | Slamf7 | |
| Cd40 | ll7r | Tfrc | |
| Cd74 | Irf4 | Tgfb3 | |
| Cd79b | Irf8 | Tigit | |
| Cfd | Klrb1 | Tnfaip6 | |









Figure S1: Lympho-myeloid conversion potential of C/EBP family members,

Related to Figure 1

(**A**) Immunoblot analysis of C/EBP family members of total protein lysates from Plat-E cells 72 h after transduction with C/EBP-MIG constructs. Untransduced cells served as control and ACTB as housekeeping gene. (**B**) Flow cytrometric analysis of B cells transduced with individual C/EBPs focusing on macrophage (CD11b⁺CD115⁺), dendritic (CD11b⁺CD11c⁺) and granulocytic (CD11b⁺Ly-6G⁺) markers 4 days after transduction. (**C**) Semi-quantitative RT-PCR analysis of C/EBP expression in murine primary FACS-purified B cells (CD19⁺), granulocytes (CD11b⁺Ly-6G⁺) and macrophages (CD11b⁺CD115⁺). *Gapdh* served as housekeeping gene. (**D**) Flow cytometric analysis of C/EBP-transduced GFP⁺-gated HAFTL1 cells 4 days after induction.

Figure S2: The role of endogenous Cebpa and Cebpb on transdifferentiation,

Related to Figure 3

(A) Semi-quantitative RT-PCR expression analysis of endogenous C/EBP family members in C10 cells treated with OHT for 4 days and non-induced controls (Bussmann et al. 2009). RNA from total bone marrow or monocytic Raw264.7 cells served as controls. *Gapdh* was used as housekeeping gene. (B) Expression kinetics of endogenous C/EBP α , and β protein in C/EBP ϵ -transduced B cells based on immunoblot analysis. (C) Workflow for the generation of *Cebpa* and *Cebpb* double knockout v-Abl immortalized B cells and isogenic controls derived from a mouse with homozygous floxed *Cebpa* and *Cebpb* alleles. Deletion of *Cebpa* and *Cebpb* by incubating cells with recombinant Cre-recombinase. (D) *Cebpa* and *Cebpb* genotypes of established Cre-incubated cell clones was assessed by PCR. Pools of Cre-treated and untreated cells served as controls. Clone 4 was selected for further experiments.

(**E**) Quantification of transdifferentiated cells (CD11b⁺, related to Figure 3C) from four independent experiments. \pm SEM, **p<0.01. (**F**) Nanostring analysis showing upregulated genes 24 h after C/EBP ϵ transduction in GFP⁺-sorted cells compared to control B cells (MIG-transduced). Genes that were not upregulated in *Cebpa*^{Δ/Δ};*Cebpb*^{Δ/Δ} cells are marked in red. (**G**) Downregulated genes in GFP⁺ FACS-purified control cells transduced for one day with C/EBP ϵ compared to empty vector. Genes refractory to down-regulation in *Cebpa*^{Δ/Δ};*Cebpb*^{Δ/Δ} cells are marked in red. See also Table S1.

Figure S3: Effect of C/EBPε dosage on transdifferentiated cell type outcome, Related to Figure 4

(A) Sorting scheme of GFP⁺ Plat-E cells transduced with empty vector control (MIG) or C/EBP ϵ -MIG for 72 h. (B) Immunoblot detection of C/EBP ϵ , GFP and ACTB in protein lysates of 1 × 10⁶ FACS-purified cells from GFP-fractions in (A). Two exposure times (short and long) are shown. (C) B cell were transduced with pMSCV-based vectors either containing bi-cistronic C/EBP ϵ -IRES-EGFP (top panels) or C/EBP ϵ -EGFP fusion (bottom panels) transgenes. Transduced cells were classified according to GFP intensity and analysed for CD11b⁺Ly-6G⁺ and CD11b⁺CD115⁺ fractions.

Figure S4: Myeloid signalling-response of transdifferentiated bi-potential cells,

Related to Figure 5

(**A**) Bright field images of long-term C/EBPβ-transduced cells, untreated or treated with 10 ng/ml GM-CSF for 24 h. Empty vector (MIG) transduced B cells served as control. (**B**) Subsequent cytospin preparations and May-Grünwald-staining after 16 days of culture.

Figure S5: Persistence of the GMP-like phenotype after serial subcloning,

Related to Figure 5

Analysis of 17 subclones derived from a GMP-like parental clone presented in Figure 5G,H. Flow cytometric analysis of CD115 versus Ly-6G in the GFP+CD11b+-gated population indicates persistence of the GMP-like potential.

Supplemental Tables

Table S1: Gene expression signature in C/EBP-transduced B cells,

Related to Figure 2, 3

Nanostring analysis (gene set GXA-MIM) of GFP⁺ B cells ($Cebpa^{t/f}$; $Cebpb^{t/f}$) transduced with individual C/EBPs for 24 h. C/EBPɛ-infected $Cebpa^{\Delta/\Delta}$; $Cebpb^{\Delta/\Delta}$ B cells (B-DKO) were included for comparison. MIG-transduced cells served as control. Normalized gene expression values are shown.

Table S2: List of primer sequences,

| Oligonucleotide | Sequence 5'-3' |
|-------------------|---------------------------------|
| Cebpa FW | GCCAGTTGGGGCACTGGGTG |
| Cebpa RV | CCGCGGCTCCACCTCGTAGA |
| Cebpb FW | GCGTTCATGCACCGCCTGCT |
| Cebpb RV | TAGGCCAGGCAGTCGGGCTC |
| Cebpd FW | AGAACCCGCGGCCTTCTAC |
| Cebpd RV | ATGTAGGCGCTGAAGTCGAT |
| Cebpe FW | CACACTGCGGGCAGACAG |
| Cebpe RV | GTGCCTTGAGAAGGGGACT |
| Gapdh FW | AATGTGTCCGTCGTGGATCTGA |
| Gapdh RV | GATGCCTGCTTCACCACCTTCT |
| Cebpa flox FW | TGGCCTGGAGACGCAATGA |
| Cebpa flow RV | CGCAGAGATTGTGCGTCTTT |
| Cebpa ∆ FW | GCCTGGTAAGCCTAGCAATCCT |
| Cebpa Δ RV | TGGAAACTTGGGTTGGGTGT |
| Cebpb flox/Δ FW | GAGCCACCGCGTCCTCCAGC |
| Cebpb flox RV | GGTCGGTGCGCGTCATTGCC |
| Cebpb ΔRV | AGCAGAGCTGCCCCGGCAAA |
| DQ52 FW1 | ACGTCGACTTTTGYAAGGGATCTACTACTGT |
| DFS FW2 | ACGTCGACGCGGACGACCACAGTGCAACTG |
| JH4A RV | GGGTCTAGACTCTCAGCCGGCTCCCTCAGGG |

Related to Experimental Procedures, Figure 4, Figure S1, S2