Application of carbohydrate arrays coupled with mass spectrometry to detect activity of plantpolysaccharide degradative enzymes from the fungus *Aspergillus niger*

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Supplementary information

Supplementary Information:

Supplementary Figure S1: Thin layer chromatography of **a** oligosaccharides generated by acid hydrolysis of wheat hemicelluloses and **b** of oligosaccharides generated by enzymatic hydrolysis of wheat arabinoxylan, with relevant fractions labelled below the sample application site (numbers H1-H11 and AX5-AX18) and M indicating the H and AX mixture of oligosaccharides before size separation, **c** MALDI-ToF MS spectra with indicated the DP (DPx) and spacing of the main pentose oligosaccharide series and spacing of a minor secondary series consisting of 4Me-GLcA decorated xylose oligosaccharides for oligosaccharides generated by acid hydrolysis of wheat hemicelluloses and **d** for oligosaccharides generated by enzymatic hydrolysis of wheat arabinoxylan. All labelled peaks are sodium adducts.

Supplementary Figure S2. a MALDI-ToF MS spectra for fraction AX10 with (a1) unlabeled starting oligosaccharide, (a2) labelling of free oligosaccharide by reductive amination and (a3) immobilization of labelled carbohydrates on the gold plate via hydrophobic interactions **b** MALDI-ToF MS spectra for fraction H9 with (b1) unlabeled starting oligosaccharide, (b2) labelling of free oligosaccharide via reductive amination and (b3) immobilization of labelled carbohydrates on the gold plate via hydrophobic interactions.

Supplementary Figure S3. MALDI-ToF MS analysis of signals derived from background hydrolysis and/or impurities in carbohydrate arrays used with commercial enzyme activity. Spectra showing substrate incubated with buffer only under reaction conditions of **a** β -glucosidase on cellotetraose-containing arrays, with DP4 + Na 914.5 m/z, DP4 + H 892.5 m/z (100%), DP3 + Na 752.4 m/z, DP3 + H 730.5 m/z (2.2%) while DP3 peak resulting from enzyme activity (Fig. 4) was 48%. **b** endoxylanase on xylohexaose-containing arrays with DP6 + H 1036.9 m/z (100%), DP5 + H 904.9 m/z (1.2%), DP4 + H 772.9 m/z (0.6%), DP3 + H 640.8 m/z (2.1%) while peaks resulting from enzyme activity (Fig. 4) were 5.3%, 9.1% and 6.4% respectively.

Supplementary Table S1. qRT-PCR primers

Supplementary Table S2. Shotgun proteomics identification of proteins in the filtrate of *A. niger* cultures grown in the presence of wheat straw with Score: SEQUEST protein score, #PSMs: total number of identified peptide sequences.

Supplementary Table S3. a Chemical shifts of ¹H-¹H TOCSY NMR of fraction H8, **b** chemical shifts of ¹H and 2D ¹H-¹³C HSQC NMR and **c** ratios of integrated ¹H signals.

Supplementary Table S4: Ratio of the intensities of MALDI-ToF MS signal, calculated for DP5/DP8, DP6/DP8, DP5/DP6 of fraction AX10 after labelling via reductive amination using various reaction times and procedures. *Procedure a*: Excess of oligosaccharide, hexadecylamine (limiting reagent), excess of NaBH₃CN, sodium borate/sodium sulfate buffer. *Procedure b*: Oligosaccharide (limiting reagent), excess of hexadecylamine, excess of NaBH₃CN, sodium borate/sodium sulfate buffer. *Procedure c*: Excess of oligosaccharide, hexadecylamine (limiting reagent), excess of NaBH₃CN, sodium borate/sodium sulfate buffer.

Supplementary Table S5: Interday reproducibility of array generation, given as mean with standard deviation of the ratio of the intensities of MALDI-ToF MS signal, calculated for DP5/DP8, DP6/DP8, DP5/DP6 of fraction AX10 and H9 immobilised over multiple days.







AX5 AX6 AX7 AX8 AX9 AX10 AX11 AX12 AX13 AX14 AX15 AX16 AX17 AX18 M

FIG. 51C



rig. 51a



Intensity (a.u.)





Fig. S3



Supplementary Table S1, primers used for qRT-PCR

gene name	gene ID	primer name	sequence (5'-3')	reference
actA	An15g00560	An15g00560Fw	TCCTGGGTCTGGAGAGCGGTG	van Munster, J. M. et al. Fungal Genet Biol (2014), 72:34-47
		An15g00560Rv	CTGCATACGGTCGGAGATACCGGG	van Munster, J. M. et al. Fungal Genet Biol (2014), 72:34-47
cbhA	An07g09330	cbhAFw	CCAGCAAGCCGGAACGCTCA	9
		cbhARv	AACGCGCCGTTTAGCCCACA	9
cbhB	An01g11660	cbhBFw	CCAGCGATGGCAGCTGCAC	9
		cbhBRv	CTGCCGGACGTGGTCACACC	9
xynB	An01g00780	xynBFw	CACGACTCTGTCGCCCAGCG	9
		xynBRv	GGGGGTCAGTGGTCCAGCCA	9

Suppleme	lementary Table S2; Shotgun Proteomics of A. niger culture filtrates					Biological replicate B			Biological replicate A			
						# Uniqu					# Unique	
Accession	Description	# AAs M\	N [kDa] c	alc. pl	Score	Coverage	Peptides	# PSMs	Score	Coverage	Peptides	# PSMs
A2QFV7	Probable endo-1,4-beta-xylanase C OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=xinC PE=1 SV=1 - [XYNC_ASPNC]	327	35.46	6.65	3836.167	87.77	38	1293	4386.343	87.77	38	1437
AZQPGZ	Probable 1,4-beta-b-glucan celloblonydrolase A OS=Aspergillus niger (strain CES 513.88 / FGC A1513) GN=CDNA PE=3 SV=1 - [CBHA_ASPNC]	452	48.22	4.32	2510.741	52.43	12	694	3181.424	52.43	12	863
A2Q710	Probable endo-1,4-beta-xylanase B US-Aspergillus niger (strain CBS 513.88 / HSSC A1513) GN=XinB PE=3 SV=1 - [XYNB_ASPNC]	225	24.04	5.45	2237.921	36.44	5	/15	2421.212	30	4	/59
A2QFV9	Probable alpha-L-arabinoturanosidase axina US=Aspergillus niger (Strain US 5 L3.88 / Hosc AL5L3) GN=axina PE=3 SV=1 - [AXIA_ASPINC]	332	35.81	4.86	1629.407	61.45	13	502	2057.846	61.45	13	601
A2R511	Probable alpha-L-arabinoturanosidase & US=Aspergillus niger (strain US 513.88 / FGSC A1513) GN=abtB Pe=3 SV=1 - [AB+B_ASPNC]	499	52.48	4.44	1160.002	35.87	11	356	12/3./08	35.87	11	3/6
AZQAI7	Probable 1,4-beta-D-glucan celloblon/grolase B DS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=CDIB FE=3 SV=1 - [CBHB_ASPNC]	530	20.52	4.30	988.6004	34.51	10	328	1214.966	34.51	10	430
AZUSIS	Probable ferulogi esterabe A OS-Aspergilius inger (strain CBS 513.88 / FOSC A1513) GM=1deA PE=3 SV=1 - [FAEA_ASPNC]	281	30.53	4.55	591.2076	44.48	10	207	/35.0302	44.48	10	247
AZQAZ7	Probable exo-1,4-beta-xylosidase xind US=Asperginus niger (strain Cas 51.38) FGSC A1513) GN=Xind PE=3 SV=1 - [XIND_ASPNC]	804	87.16	4.89	410.8238	34.45	22	150	403.6227	34.58	22	149
AZQPC3	Probable endo-beta-1,4-glucanase B US=Aspergillus niger (strain USS 513.88 / FGSC A1513) GN=egils PE=3 SV=1 - [EGLE_ASPNC]	331	36.54	4.55	332.6013	28.4	10	169	332.4204	31.42	10	1/8
A2RAL4	Probable beta-glucosioase A OS-Aspergillus niger (strain CBS 513.88 / HOSC A1513) GN FordA PE=3 SV=1 - [BGLA_ASPNC]	860	93.17	4.78	307.6645	33.37	28	111	276.1129	36.16	29	101
AZQYU7	Probable refuloyi esterase C OS=Aspergillus niger (strain CBS 513.88 / FOSC A1513) GN=TaeC PE=3 SV=1 - [FAEC_ASPNC]	270	28.57	5.02	233.4921	36.67	6	65	284.6185	36.67	6	12
AZQASZ	Probable mannosyl-oligosaccharide alpha-1,2-mannosidase IB US=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=mnS1B PE=3 SV=1 - [MINS1E	513	55.89	5.05	213.7005	31.97	14	64	225.6573	36.26	15	65
AZRZL1	Probable mamnogalacturonate lyase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GB=rgiA PE=3 SV=2 - [RGLA_ASPNC]	531	57.10	4.60	208.3747	20.9	10	/1	270.6945	22.22	9	90
AZROEO	Extracellular exo-inulinase inue OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=inue PE=2 SV=1 - [INUE_ASPNC]	537	59.11	5.43	197.594	38.73	16	88	141.211	34.45	13	60
ASABES	Probable beta-glucosidase M US=Aspergillus niger (strain US 513.88 / Host A1513) G M=bgliM PE=3 SV=1 - [BGLM_ASPNC]	765	82.07	4.74	163.5841	21.96	13	54	162.2629	23.66	14	57
A2R3L3	Aspartic protease pep1 OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=pep1 PE=3 SV=1 - [PEPA_ASPNC]	394	41.25	4.67	131.3/12	25.38	8	56	119.9708	24.62	8	48
A2R2S8	Cell wall protein phiA OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=phiA PE=2 SV=1 - [PHIA_ASPNC]	184	19.17	5.03	120.3915	23.37	4	34	101.3156	25.54	5	32
A2QEJ9	Probable alpha-galactosidase B OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GR=agIB PE=3 SV=1 - [AGALB_ASPNC]	443	48.80	5.08	107.8469	23.25	9	49	114.5235	19.64	6	53
A2QK14	Probable mannan endo-1,4-beta-mannosidase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=manA PE=1 SV=1 - [MANA_ASPNC]	383	41.22	4.61	104.2/14	31.59	/	32	118.8116	30.29	6	32
A2QAN3	Probable beta-galactosidase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=lacA PE=3 SV=1 - [BGALA_ASPNC]	1007	109.64	5.21	//.29582	20.56	15	33	/1.6482/	21.75	15	32
A2QH21	Probable glucan endo-1,3-beta-glucosidase egIC OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=egIC PE=3 SV=2 - [EGLC_ASPNC]	460	46.32	4.49	74.7548	19.57	5	25	64.61966	19.57	5	24
A2QK83	Probable endo-xylogalacturonan hydrolase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=xghA PE=3 SV=1 - [XGHA_ASPNC]	406	42.10	4.51	72.05174	17.24	6	30	58.36289	17.24	6	23
A2RB93	Probable arabinogalactan endo-beta-1,4-galactanase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=galA PE=3 SV=1 - [GANA_ASPNC]	350	38.67	4.07	64.57549	24.29	7	20	88.52927	24.29	6	25
A2R2S6	Probable alpha-galactosidase D OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=aglD PE=3 SV=2 - [AGALD_ASPNC]	660	71.18	4.83	61.81282	16.97	10	24	50.46856	19.7	11	25
A2RAR6	Probable glucan 1,3-beta-glucosidase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=exgA PE=3 SV=1 - [EXGA_ASPNC]	416	45.50	5.19	58.95625	25.24	9	23	49.66892	27.88	8	19
A2R3X3	Probable alpha-glucuronidase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=aguA PE=3 SV=1 - [AGUA_ASPNC]	841	93.67	5.38	55.70176	20.57	15	36	80.58374	22.47	16	38
A2QEQ6	Beta-glucuronidase OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=An02g11890 PE=1 SV=1 - [GUS79_ASPNC]	541	58.82	5.47	49.53932	20.52	8	17	50.59749	16.82	7	15
A2R0Z6	Probable feruloyl esterase B OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=faeB PE=3 SV=1 - [FAEB_ASPNC]	521	57.17	5.20	48.00242	14.01	5	14	54.39299	15.55	6	16
A2QYR9	Probable 1,4-beta-D-glucan cellobiohydrolase C OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=cbhC PE=3 SV=1 - [CBHC_ASPNC]	459	48.10	4.53	45.85379	10.89	3	27	45.22075	18.52	4	23
A2Q7E0	Probable alpha-L-arabinofuranosidase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=abfA PE=3 SV=1 - [ABFA_ASPNC]	628	67.91	4.30	45.31327	13.22	6	15	29.4525	13.22	6	12
A2QT85	Probable arabinan endo-1,5-alpha-L-arabinosidase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=abnA PE=3 SV=1 - [ABNA_ASPNC]	321	34.46	4.22	42.86552	4.67	2	14	37.9504	4.67	2	11
A2QK82	Probable pectinesterase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=pmeA PE=3 SV=1 - [PMEA_ASPNC]	327	34.60	4.39	42.15743	19.57	4	18	59.39917	23.24	5	24
A2R060	Probable exopolygalacturonase X OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=pgaX PE=3 SV=1 - [PGLRX_ASPNC]	435	47.26	4.82	39.48248	15.63	5	13	22.17836	13.56	3	10
A2QHG0	Probable exopolygalacturonase B OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=pgxB PE=3 SV=1 - [PGXB_ASPNC]	438	48.35	4.86	39.31279	22.6	9	18	25.81601	18.95	7	14
A2R797	Probable alpha-fucosidase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=afcA PE=3 SV=1 - [AFCA_ASPNC]	793	86.51	4.91	24.53108	16.52	12	15	22.9456	11.6	7	10
A5AAG2	Probable arabinan endo-1,5-alpha-L-arabinosidase C OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=abnC PE=3 SV=1 - [ABNC_ASPNC]	318	34.03	4.87	24.49346	10.69	4	12	NA	NA	NA	NA
A2RAY7	Putative galacturan 1,4-alpha-galacturonidase C OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=rgxC PE=2 SV=1 - [RGXC_ASPNC]	423	45.87	5.20	22.18053	16.55	4	9	20.71233	11.58	3	6
A2Q877	Probable xyloglucan-specific endo-beta-1,4-glucanase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=xgeA PE=3 SV=1 - [XGEA_ASPNC]	241	25.47	3.88	21.76691	14.11	2	11	21.61399	14.11	2	12
A2QA64	Probable beta-galactosidase B OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=lacB PE=3 SV=2 - [BGALB_ASPNC]	1017	111.74	5.00	21.22608	6	6	9	15.32841	5.9	6	8
A2QV36	Probable pectate lyase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=plyA PE=3 SV=1 - [PLYA_ASPNC]	323	34.30	4.82	19.75821	13.62	4	14	28.6039	13.62	4	15
A2QAH3	Probable endopolygalacturonase I OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=pgal PE=3 SV=1 - [PGLR1_ASPNC]	368	38.09	4.48	19.3904	13.32	4	11	22.07549	11.96	3	9
A2QW66	Probable exopolygalacturonase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=pgxA PE=3 SV=1 - [PGXA_ASPNC]	434	46.94	4.79	18.59541	10.14	5	10	22.42316	10.14	5	10
A2QYE5	Probable rhamnogalacturonase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=rhgA PE=3 SV=1 - [RHGA_ASPNC]	446	46.93	4.23	16.74227	15.02	4	7	24.95915	5.83	2	8
A2QL72	Probable alpha-galactosidase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=aglA PE=3 SV=1 - [AGALA_ASPNC]	537	59.18	5.03	16.61905	11.92	5	10	18.05999	5.4	3	8
A2R3I1	Probable pectin lyase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=pelA PE=3 SV=1 - [PELA_ASPNC]	379	39.76	4.34	16.34141	16.09	5	11	14.46537	8.97	3	9
A2QTU5	Alpha-xylosidase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=axlA PE=1 SV=1 - [XYLA_ASPNC]	736	82.54	5.66	14.92676	13.04	7	10	22.78718	13.18	7	12
A2QEW2	Probable exopolygalacturonase C OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=pgxC PE=3 SV=2 - [PGXC_ASPNC]	440	47.82	5.20	14.81849	9.09	3	6	NA	NA	NA	NA
A2QWU9	Beta-mannosidase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=mndA PE=3 SV=1 - [MANBA_ASPNC]	931	103.96	5.29	14.74085	12.14	7	11	6.310669	5.91	4	4
P0C953	Histone H2A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=httA PE=3 SV=1 - [H2A_ASPNC]	134	14.14	10.55	8.224466	11.94	2	4	8.499234	11.94	2	4
A2R2W3	Probable cutinase 1 OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=An14g02170 PE=3 SV=1 - [CUTI1_ASPNC]	262	26.67	4.88	7.27599	9.92	2	3	10.04579	9.92	2	3
A2QCV8	Probable endopolygalacturonase B OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=pgaB PE=3 SV=1 - [PGLRB_ASPNC]	362	37.80	6.77	7.110866	9.67	3	4	4.506392	6.63	2	2
A2QUQ2	Endochitinase A OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=ctcA PE=2 SV=1 - [CHIA1_ASPNC]	1257	122.02	4.55	5.929324	0.88	2	3	4.313844	0.88	2	2
A2QY49	Histone H2B OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=htb1 PE=3 SV=1 - [H2B_ASPNC]	141	14.99	10.13	4.44716	18.44	3	4	4.437262	13.48	2	3
A2QE32	40S ribosomal protein S0 OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=rps0 PE=3 SV=1 - [RSSA_ASPNC]	298	31.93	4.91	4.07477	6.71	2	2	NA	NA	NA	NA
A5ABH4	Probable rhamnogalacturonate lyase B OS=Aspergillus niger (strain CBS 513.88 / FGSC A1513) GN=rglB PE=3 SV=1 - [RGLB_ASPNC]	706	78.37	5.11	2.444819	4.25	3	4	0	2.97	2	4

	Chemical shift (p.p.m.)							
Protons	Internal β-Xylp Non-reducing terminal β-Xylp							
¹ H	4.48	4.46						
² H	3.29	3.26						
³ Н	3.556	3.42						
⁴ H	3.79	3.62						
⁵H	4.10, 3.38	3.97, 3.31						

Table S3a, Upfield ¹H chemical shifts of xylose oligosaccharide as determined from the 2D TOCSY spectrum of fraction H8

	Chemical shift (p.p.m.)						
	Fractio	on AX14	Fract	ion H8			
Residue	¹ H (1D)	¹³ C (HSQC)	¹ H (1D)	¹³ C (HSQC)			
α-Xylp ¹	5.184	93.1	5.183	92.9			
β-Xylp ¹	4.584	97.6	4.584	97.6			
β-Xylp ^{2/3/4/5}	~4.47	~102.7	~4.48	~102.8			
α-Ara <i>f</i> -X ^N -O3s	5.395	108.7	NA	NA			
α-Ara <i>f</i> -X ^N - <i>O</i> -2	5.224	109.8	NA	NA			
α-Ara <i>f</i> -X ^N - <i>O</i> -3d	5.273	109.2	NA	NA			
β-Xyl <i>p</i> ^N -A <i>f-O</i> -2,3	4.640	100.9	NA	NA			
β-Xylp ^N -A <i>f-O</i> -3	4.43-4.45	NR	NA	NA			
4- <i>O</i> -Me-α-GlcA-X ^N -2	NA	NA	5.285	~98.7			
α-GlcA-X ^N -2	NA	NA	5.303	~98.7			
β-Xylp ^N -GlcA	NA	NA	4.641	~102.7			
β-Xylp ^N -MeGlcA	NA	NA	4.623	~102.7			

Table S3b, Assignment of chemical shifts of ¹H and ¹H-¹³C HSQC NMR anomeric resonances. Xylp¹ is reducing end xylopyranose; β -Xylp^{2/3/4/5} is the total of unsubstituted Xylp residues other than the reducing end; α -Araf-X^N-O- single (3s) or double (2 and 3d) arabinofuranosidase decoration on unidentified Xylp via α -1,3-, or α -1,2- and α -1,3- linkage respectively; β -Xylp^N-Af-O-3/2,3 Xylp with single/double Araf decoration respectively; (4-O-Me)- α -GlcA-X^N-C2 signal for GlcA and 4MeGlcA attached via α -1,2 to an unidentified Xylp, β -Xylp^N-(Me)GlcA unidentified Xylp decorated with GlcA or 4MeGlcA, NA not identified in this fraction, NR not resolvable.

	Fraction					
Signal integration ratio ¹ H (1D)	H6	H7	H8	AX14	AX15	
Xylp ¹	1	1	1	1	1	
β-Xyl <i>p</i> ^{2/3/4/5}	8.2	6.5	6.3	NR	NR	
(4- <i>O</i> -Me)-α-GlcA-X ^N -C2	1.1	0.7	0.6	NA	NA	
β-Xylp ^N -(Me)GlcA	1.2	0.8	0.6	NA	NA	
β-Xylp ^N -A <i>f</i> -O-2,3	NA	NA	NA	0.4	0.3	
β -Xylp ^N -Af-O-3 + β -Xylp ^{2/3/4/5}	NA	NA	NA	3.5	3.0	
α-Ara <i>f</i> -X ^N -O-3s	NA	NA	NA	0.9	0.7	
α-Ara <i>f</i> -X ^N - <i>O</i> -2	NA	NA	NA	0.4	0.3	
α-Ara <i>f-</i> X ^N - <i>O</i> -3d	NA	NA	NA	0.4	0.3	

Table S3c, Signal integration ratios ¹**H NMR.** With the sum of the signal for the α- and β-anomer of the reducing end xylopyranose (Xylp¹) set to 1, β-Xylp^N-Af-O-3 + β-Xylp^{2/3/4/5} combined ratio for β-Xylp^N-Af-O-3 and β-Xylp^{2/3/4/5}, other notations as in Table S3b.

Supplementary Table S4

	DP5	5/DP8		DP6	5/DP8		DP5		
Procedure	a	b	C	a	b	C	a	b	С
1 hour	0.32	0.40	0.35	0.70	0.79	0.74	0.46	0.52	0.47
2 hours	0.39	0.39	0.38	0.77	0.80	0.76	0.50	0.50	0.50
4 hours	0.54 ± 0.02	0.37	0.40	0.92 ± 0.02	0.81	0.85	0.59 ± 0.01	0.48	0.47
8 hours	0.55	0.36	0.47	0.92	0.77	0.89	0.60	0.47	0.53
1 day	0.56	0.35	0.52	0.93	0.75	0.99	0.60	0.46	0.52
3 days	0.56	0.33	0.55	0.92	0.74	0.95	0.60	0.45	0.53
Unlabelled 0.24 oligosaccharide			0.53 0.45				.45		

Supplementary Table S5

	DP5/DP8	DP6/DP8	DP5/DP6
AX10 day 1	1.71	1.82	0.94
AX10 day 2	1.77	1.81	0.99
AX10 day 3	1.75	1.81	0.99
AX10 variability	1.74 ± 0.03	1.81 ± 0.01	0.97 ± 0.03
H9 day 1	1.93	1.68	1.15
H9 day 2	1.93	1.62	1.19
H9 day 3	1.97	1.65	1.24
H9 variability	1.94 ± 0.02	1.65 ± 0.03	1.19 ± 0.05