

# Response of rhizosphere soil microbial to *Deyeuxia angustifolia* encroaching in two different vegetation communities in alpine tundra

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TableS1. Sampling sites, geographical location and dominant species that use plant coverage (%) to indicate.

Types	Altitude(m)	Latitude	Longitude	Dominant species & Coverage (%)
NH	2182	41°59'25.57"	128°1'0.23"	<i>Ligularia jamesii</i> (Hemsl.) Kom. 43%, <i>Sanguisorba sitchensis</i> 46%, <i>Veratrum nigrum</i> L. 7%
EH	2182	41°59'25.57"	128°1'0.23"	<i>D. angustifolia</i> 40%, <i>Ligularia jamesii</i> (Hemsl.) Kom. 22%, <i>Sanguisorba sitchensis</i> 26%
NS	2182	41°59'25.57"	128°1'0.23"	<i>Rhododendron aureum</i> Georgi 60%, <i>Vaccinium uliginosum</i> Linn. 28%
ES	2182	41°59'25.57"	128°1'0.23"	<i>D. angustifolia</i> 35%, <i>Rhododendron aureum</i> Georgi 35%, <i>Vaccinium uliginosum</i> Linn. 25%
DA	2182	41°59'25.57"	128°1'0.23"	<i>D. angustifolia</i> 92%
NS	2178	41°59'20.99"	128°0'52.65"	<i>Vaccinium uliginosum</i> Linn. 68%, <i>Phyllodoce caerulea</i> (L.) Bab. 24%
ES	2178	41°59'20.99"	128°0'52.65"	<i>D. angustifolia</i> 49%, <i>Vaccinium uliginosum</i> Linn. 28%, <i>Phyllodoce caerulea</i> (L.) Bab. 15%
NS	2178	41°59'20.99"	128°0'52.65"	<i>Rhododendron aureum</i> Georgi 83%, <i>Vaccinium uliginosum</i> Linn. 8%
ES	2178	41°59'20.99"	128°0'52.65"	<i>D. angustifolia</i> 49%, <i>Rhododendron aureum</i> Georgi 42%
DA	2178	41°59'20.99"	128°0'52.65"	<i>D. angustifolia</i> 90%
NS	2143	41°59'25.02"	128°0'45.68"	<i>Rhododendron aureum</i> Georgi 95%, <i>Vaccinium uliginosum</i> Linn. 3%
ES	2143	41°59'25.02"	128°0'45.68"	<i>D. angustifolia</i> 45%, <i>Rhododendron aureum</i> Georgi 51%
NS	2143	41°59'25.02"	128°0'45.68"	<i>Vaccinium uliginosum</i> Linn. 92%
ES	2143	41°59'25.02"	128°0'45.68"	<i>D. angustifolia</i> 55%, <i>Vaccinium uliginosum</i> Linn. 36%
NH	2142	41°59'25.02"	128°0'45.68"	<i>Ligularia jamesii</i> (Hemsl.) Kom. 82%, <i>Trollius chinensis</i> Bunge 7%, <i>Veratrum nigrum</i> L. 5%
EH	2143	41°59'25.02"	128°0'45.68"	<i>D. angustifolia</i> 45%, <i>Ligularia jamesii</i> (Hemsl.) Kom. 42%, <i>Trollius chinensis</i> Bunge 6%
DA	2143	41°59'25.02"	128°0'45.68"	<i>D. angustifolia</i> 93%
NS	2034	41°59'20.92"	128°0'9"	<i>Phyllodoce caerulea</i> (L.) Bab. 34%, <i>Rhododendron aureum</i> Georgi 52%, <i>Vaccinium uliginosum</i> Linn. 9%
ES	2034	41°59'20.92"	128°0'9"	<i>D. angustifolia</i> 47%, <i>Phyllodoce caerulea</i> (L.) Bab. 15%, <i>Rhododendron aureum</i> Georgi 23%, <i>Vaccinium uliginosum</i> Linn. 7%
NH	2034	41°59'20.92"	128°0'9"	<i>Sanguisorba sitchensis</i> 60%, <i>Trollius chinensis</i> Bunge 22%, <i>Aconitum monanthum</i> Nakai 8%, <i>Veratrum nigrum</i> L. 5%
EH	2034	41°59'20.92"	128°0'9"	<i>D. angustifolia</i> 55%, <i>Sanguisorba sitchensis</i> 30%, <i>Trollius chinensis</i> Bunge 8%, <i>Aconitum monanthum</i> Nakai 5%
NH	2034	41°59'20.92"	128°0'9"	<i>Sanguisorba sitchensis</i> 30%, <i>Aconitum monanthum</i> Nakai 21%, <i>Veratrum nigrum</i> L. 15%, <i>Trollius chinensis</i> Bunge 18%, <i>Ligularia jamesii</i> (Hemsl.) Kom 10%
EH	2034	41°59'20.92"	128°0'9"	<i>D. angustifolia</i> 42%, <i>Sanguisorba sitchensis</i> 18%, <i>Aconitum monanthum</i> Nakai 12%, <i>Trollius chinensis</i> Bunge 8%, <i>Veratrum nigrum</i> L. 5%
DA	2034	41°59'20.92"	128°0'9"	<i>D. angustifolia</i> 90%, <i>Veratrum nigrum</i> L. 5%
NH	2026	41°59'15.72"	128°0'6.29"	<i>Trollius chinensis</i> Bunge 35%, <i>Sanguisorba sitchensis</i> 30%, <i>Aconitum monanthum</i> Nakai 12%, <i>Veratrum nigrum</i> L. 8%, <i>Ligularia jamesii</i> (Hemsl.) Kom 12%
EH	2026	41°59'15.72"	128°0'6.29"	<i>Trollius chinensis</i> Bunge 25%, <i>Sanguisorba sitchensis</i> 9%, <i>Aconitum monanthum</i> Nakai 7%, <i>Veratrum nigrum</i> L. 4%
DA	2026	41°59'15.72"	128°0'6.29"	<i>D. angustifolia</i> 83%, <i>Trollius chinensis</i> Bunge 6%

TableS2. Primers and real-time PCR conditions used in this experiment.

function genes	primers	real-time PCR protocol
<i>nifH</i>	PolF 5'-TG CGAY CCS AAR GCB GAC TC-3'	94 °C for 5 min; 40 cycles of 94 °C for 30s, 57 °C for 45s(54 °C for <i>nifH</i> gene, 60 °C for AOB), 72 °C for 1 min.
	PolR 5'-ATS GCC ATC ATY TCR CCG GA-3'	
AOA	Arch-amoAF 5'-STA ATG GTC TGG CTT AGA CG-3'	
	Arch-amoAR 5'-GCG GCC ATC CAT CTG TAT GT-3'	
AOB	amoA-1F 5'-GGG GTT TCT ACT GGT GGT-3'	
	amoA-2R 5'-CCC CTC KGS AAA GCC TTC TTC-3'	
<i>nosZ</i>	nosZ-F 5'-CGY TGT TCM TCG ACA GCC AG-3'	
	nosZ 1622R 5'-CGS ACC TTS TTG CCS TYG CG-3'	

TableS3. Pearson correlation coefficient (*r*) between soil properties and enzymatic activities, Asterisk indicates significant differences \*indicate significant ( $P<0.05$ ) effects, \*\* indicate significant ( $P<0.01$ ) effects.

	TN	TOC	TP	TK	NO <sub>3</sub> <sup>-</sup>	NH <sub>4</sub> <sup>+</sup>	Moisture	pH	MBC	MBN	AN	AP	AK	C/N
Urease	0.616**	0.715**	0.329*	-0.288	-0.153	0.082	0.567**	-0.336	0.016	-0.076	0.515**	0.290	0.422*	0.606**
Sucrase	0.484**	0.714**	0.177	-0.260	-0.278	0.275	0.402*	-0.168	0.094	-0.318	0.317	0.084	0.324*	0.683**
ACP	0.062	0.242	0.490**	-0.086	-0.030	0.240	0.310	-0.409	0.037	-0.087	0.297	0.651**	0.543**	0.233
Catalase	0.748**	0.731**	0.387*	0.034	0.013	0.305	0.592**	-0.244	0.262	-0.100	0.440*	0.336*	0.413*	0.561**

TableS4. Pearson correlation coefficient ( $r$ ) between N cycling function genes abundance and soil properties, just list significant ( $P < 0.05$ ) correlation. Bolded values indicate significant ( $P < 0.05$ ) effects.

	AOB		AOA		<i>nosZ</i>		<i>nifH</i>	
	$r$	$P$	$r$	$P$	$r$	$P$	$r$	$P$
<b>Total</b>	<b>n=27</b>							
NO <sub>3</sub> <sup>-</sup>	<b>-0.404</b>	<b>0.036</b>	<b>0.451</b>	<b>0.018</b>	-0.073	0.719	-0.050	0.806
AP	<b>-0.410</b>	<b>0.040</b>	-0.061	0.764	0.291	0.141	-0.324	0.099
TP	-0.245	0.218	0.298	0.131	0.039	0.847	<b>0.461</b>	<b>0.016</b>
NH <sub>4</sub> <sup>+</sup>	-0.118	0.557	-0.132	0.512	<b>0.560</b>	<b>0.002</b>	-0.207	0.300
Moisture	0.020	0.922	-0.096	0.634	<b>0.490</b>	<b>0.009</b>	0.013	0.950
<b>Herbaceous</b>	<b>n=10</b>							
AP	<b>-0.636</b>	<b>0.048</b>	-0.240	0.505	-0.341	0.334	-0.463	0.178
TOC	0.249	0.489	<b>0.833</b>	<b>0.003</b>	0.207	0.506	<b>0.815</b>	<b>0.004</b>
TP	-0.232	0.518	<b>0.672</b>	<b>0.033</b>	-0.023	0.950	<b>0.668</b>	<b>0.035</b>
C/N	0.293	0.412	<b>0.701</b>	<b>0.024</b>	0.190	0.599	<b>0.698</b>	<b>0.025</b>
TK	-0.514	0.128	<b>-0.671</b>	<b>0.034</b>	0.193	0.592	<b>-0.632</b>	<b>0.048</b>
MBC	-0.425	0.221	-0.213	0.554	<b>-0.633</b>	<b>0.049</b>	-0.469	0.171
MBN	-0.452	0.190	-0.357	0.311	<b>-0.665</b>	<b>0.036</b>	-0.542	0.106
Moisture	0.475	0.165	<b>0.626</b>	<b>0.530</b>	0.456	0.185	<b>0.633</b>	<b>0.037</b>
<b>Shrub</b>	<b>n=12</b>							
AN	0.420	0.174	<b>-0.591</b>	<b>0.043</b>	-0.029	0.928	0.533	0.074
AK	-0.173	0.582	<b>-0.643</b>	<b>0.024</b>	<b>0.334</b>	<b>0.288</b>	0.431	0.161
NH <sub>4</sub> <sup>+</sup>	-0.231	0.471	-0.437	0.155	<b>0.620</b>	<b>0.031</b>	-0.332	0.291
MBC	-0.240	0.460	-0.143	0.658	<b>0.841</b>	<b>0.001</b>	0.134	0.679

TableS5. Alpha Diversity Metrics for Native sites (NH, NS), Encroaching sites (EH, ES) in two vegetation types and *D. angustifolia* (DA) bacteria.

	chao1	observed species	PD whole tree	shannon	simpson	goods coverage
NS	1100.42	857.5	54.29	7.86	0.99	0.99
ES	925.80	826.83	48.02	7.40	0.98	0.99
NH	1037.12	865.40	52.10	7.45	0.98	0.99
EH	998.05	834.80	51.02	7.36	0.98	0.99
DA	863.95	747.75	48.38	7.62	0.98	0.99

TableS6. A non-parametric test showed differences between encroaching and native sites in two treatments (NS/ES and NH/EH), compare the microbial diversity *D. angustifolia* to native samples (DA/NS/NH) and to encroaching samples (DA/ES/EH). Bolded values indicate significant ( $\alpha < 0.05$ ) effects.

Alpha name	NS/ES	NH/EH	DA/NS/NH	DA/ES/EH
chao1	<b>0.004</b>	0.55	<b>0.05</b>	0.09
observed species	0.618	0.31	0.26	0.17
PD whole tree	<b>0.025</b>	0.84	0.48	0.36
shannon	<b>0.037</b>	0.84	0.70	0.82
simpson	0.113	1.00	0.33	0.76
goods coverage	0.857	0.84	<b>0.03</b>	0.22

TableS7. Phylum with a greater relative abundance associated with native and encroaching sites under woody and herbaceous treatment to have a significant effect based on ANOSIM Analysis. Bolded values indicate significant ( $P < 0.05$ ) effects.

Phylum	NS/ES	NH/EH	DA/NS/NH	DA/ES/EH
Acidobacteria	0.07	0.43	0.17	0.17
Proteobacteria	<b>0.05</b>	<b>0.05</b>	0.12	0.66
Actinobacteria	0.11	0.43	0.43	0.15
Bacteroidetes	0.09	0.25	<b>0.04</b>	0.58
Chloroflexi	0.61	0.66	0.41	0.71
Candidatus Saccharibacteria	0.69	0.93	0.08	0.20
Planctomycetes	<b>0.04</b>	1.00	0.42	0.31
Firmicutes	0.61	0.93	0.35	0.45
Gemmatimonadetes	<b>0.02</b>	0.79	<b>0.04</b>	0.64
Verrucomicrobia	0.08	0.54	0.51	0.61
Nitrospirae	0.50	1.00	<b>0.04</b>	0.63
Latescibacteria	1.00	0.40	<b>0.05</b>	0.68
Cyanobacteria/Chloroplast	0.17	0.27	0.31	0.47
Chlamydiae	0.26	0.51	0.61	0.70

TableS8. The correlation between community composition and environmental variables for bacteria. Values in bold indicate significant correlation ( $P < 0.05$ ).

<b>Variable</b>	<b><i>r</i></b>	<b><i>Pr</i> (&gt;<i>r</i>)</b>
<b>TN</b>	<b>0.353</b>	<b>0.009 **</b>
<b>Moisture</b>	<b>0.262</b>	<b>0.030 *</b>
<b>pH</b>	<b>0.261</b>	<b>0.024 *</b>
<b>MBN</b>	<b>0.274</b>	<b>0.029 *</b>
MBN	0.204	0.068
AK	0.174	0.096
Elevation	0.153	0.128
TOC	0.157	0.129
NH <sub>4</sub> <sup>+</sup>	0.100	0.266
AP	0.055	0.520
AN	0.034	0.663
TK	0.026	0.722
NO <sub>3</sub> <sup>-</sup>	0.022	0.745
TP	0.020	0.794

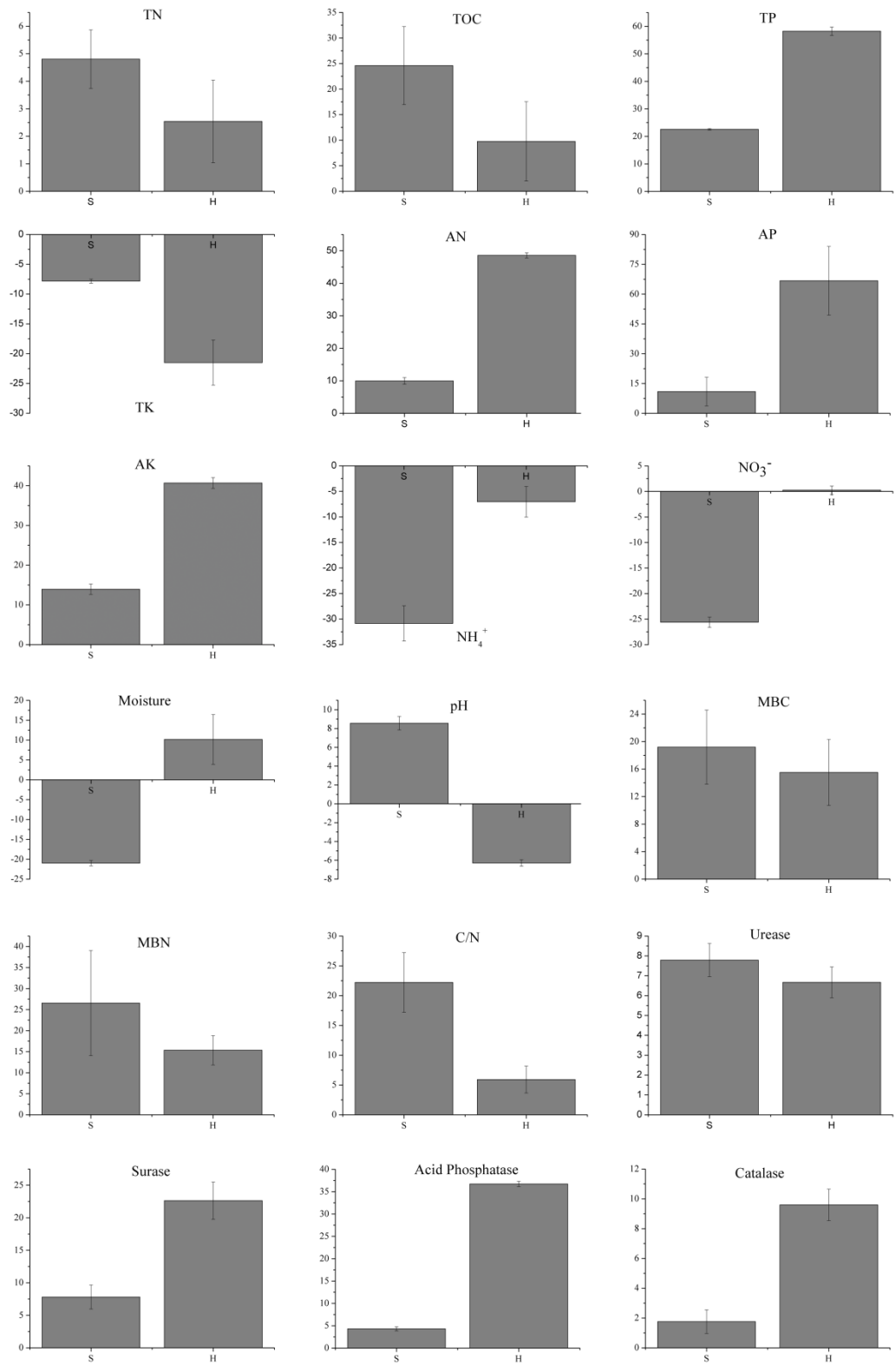


Fig.S1. The effect of *D. angustifolia* encroaching on soil nutrients are shown. Data are pooled over vegetation types and were generated by looking at the percentage loss or gain of a particular attribute *D. angustifolia* encroaching relative to the native control values. Bars are standard errors.

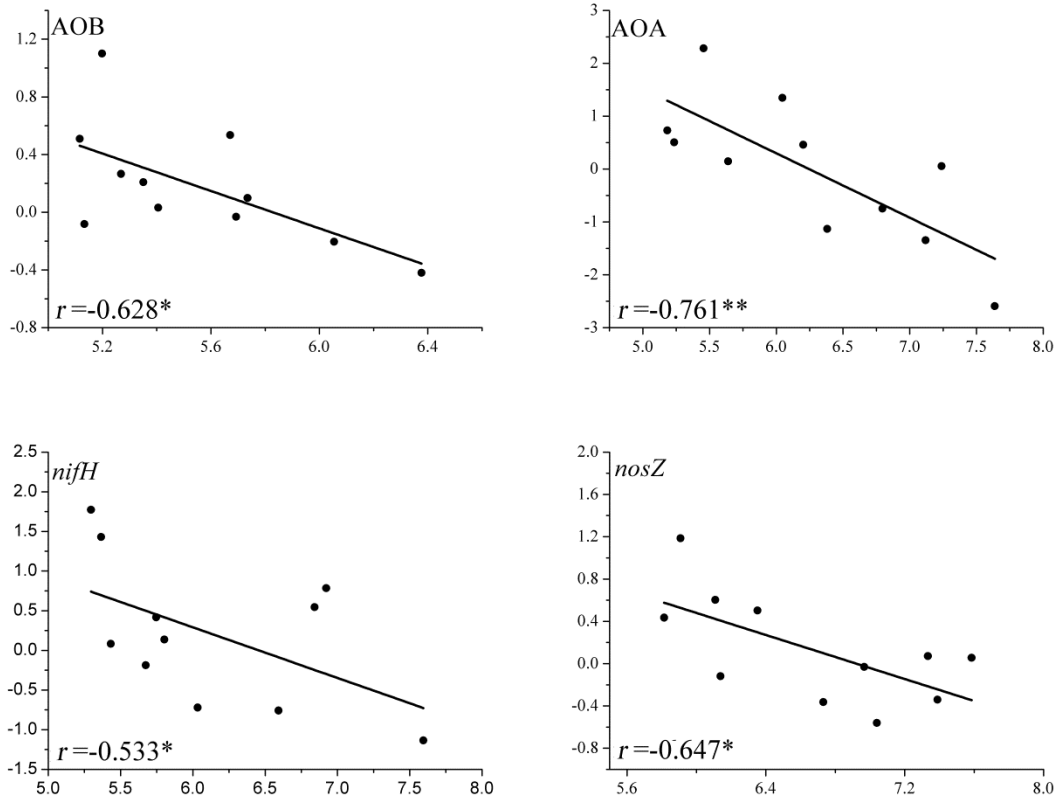


Fig.S2. Impact of invasive species on N-transforming function genes in native plots (*N*). Impact is calculated as the difference between the value of the parameter in encroaching plots (*E*) minus the value of the parameter in native plots (*N*). A positive value of *E-N* indicates an increased concentration of that element in the invaded soil Pearson correlation coefficient (*r*) between *N* and *E-N*. Significance levels(\* $P < 0.05$ , \*\* $P < 0.01$ ).

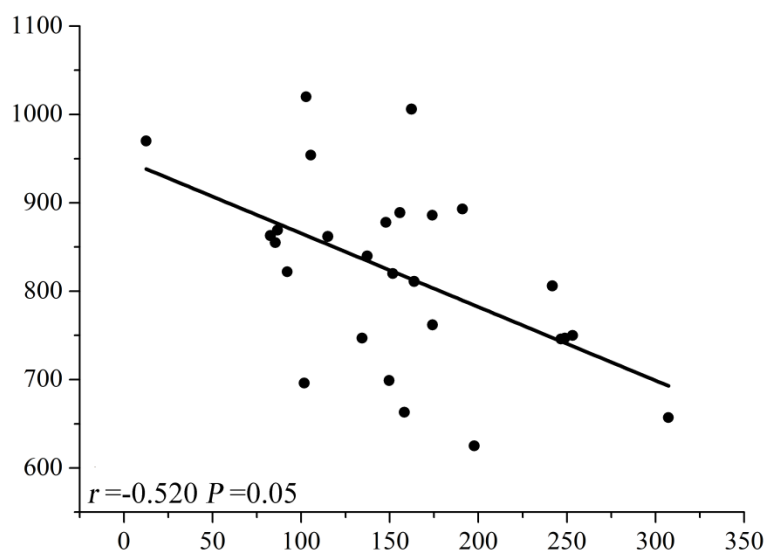


Fig.S3. Pearson correlation coefficient (*r*) between OTU richness and TOC.