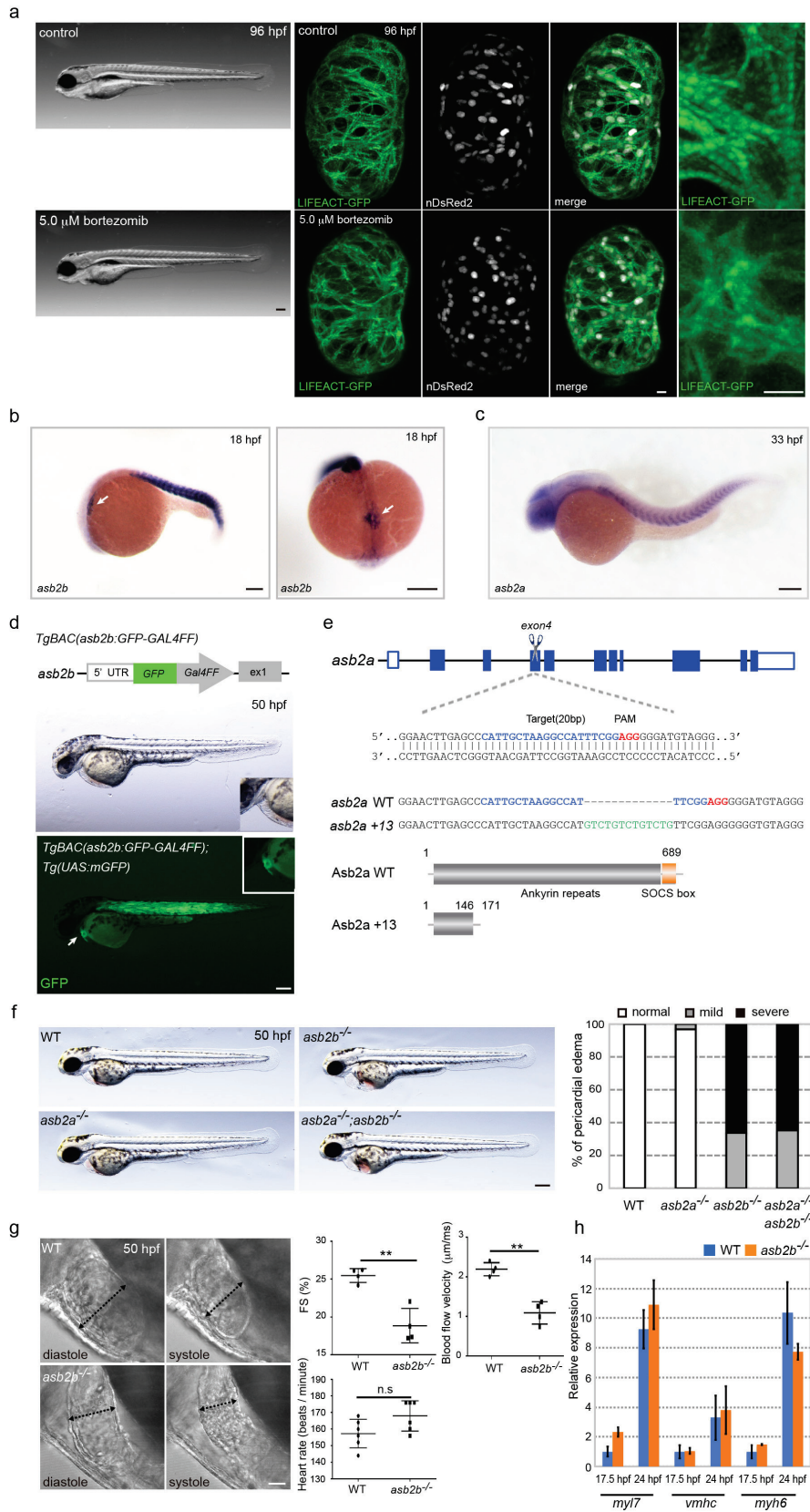
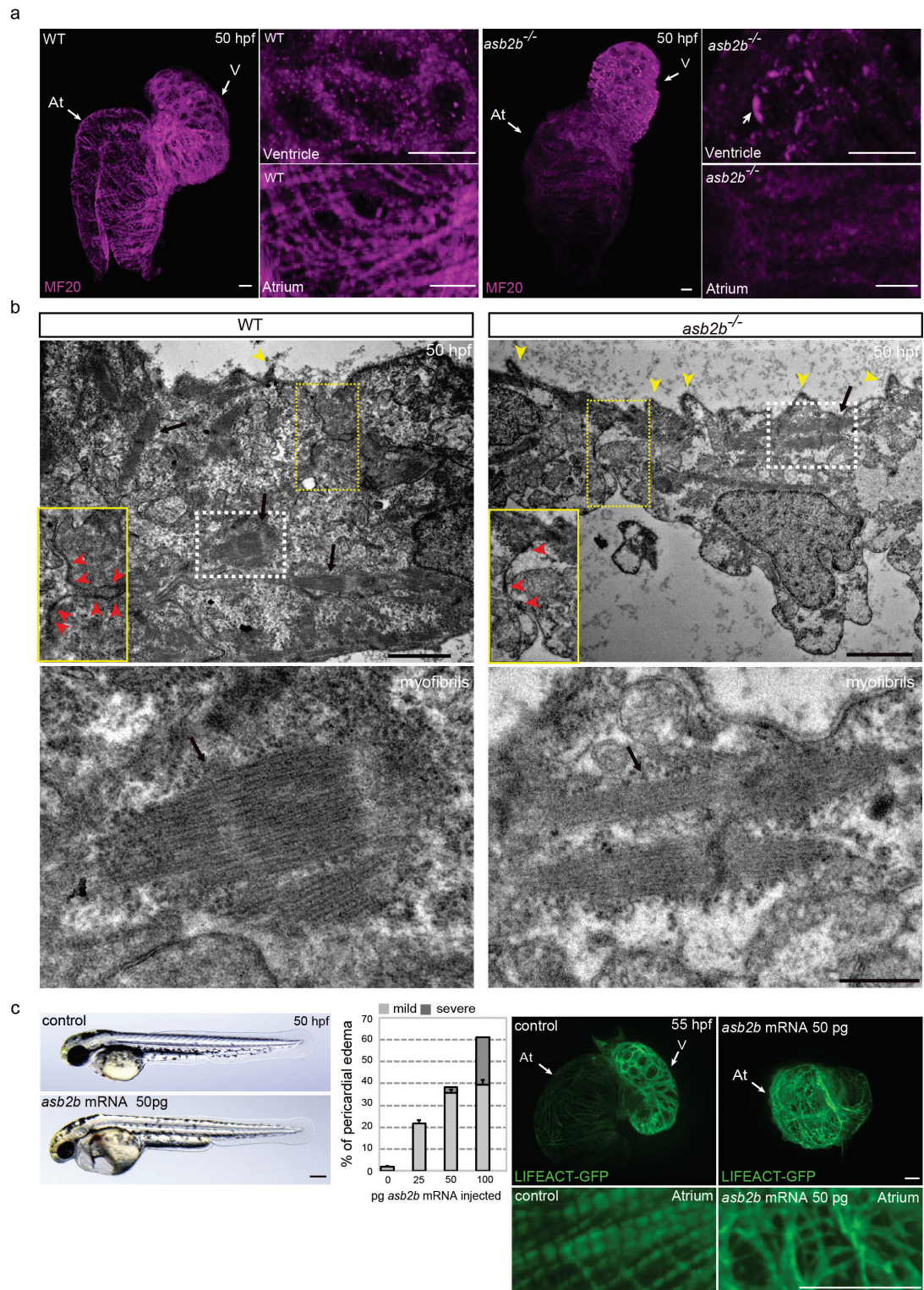


Supplementary Figures



Supplementary Figure 1 - *asb2b* mutant zebrafish exhibit cardiac defects. (a) Bright-field micrographs of 96 hpf larvae treated with DMSO (control) and bortezomib. Lateral

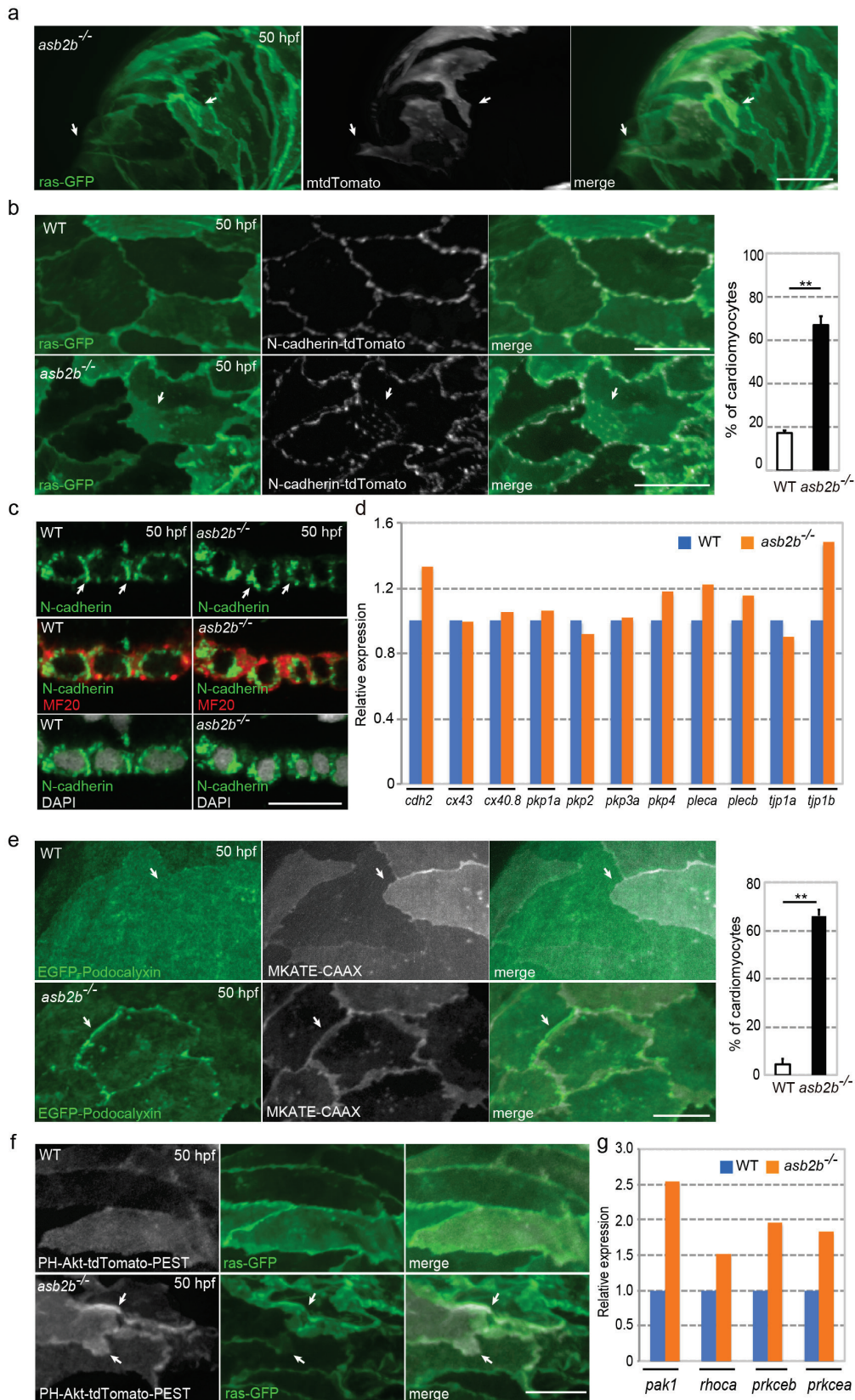
views, anterior to the left. Scale bar, 200 μm . 3D images of 96 hpf *Tg(myl7:LIFEACT-GFP)* hearts in animals treated with DMSO (control) and bortezomib. Animals were treated from 24 until 96 hpf. Scale bar, 20 μm . (b) *in situ* hybridization for *asb2b* expression in 18 hpf embryos. Lateral view, anterior to the left; dorsal view, anterior to the bottom. *asb2b* is expressed in the heart cone (arrows) as well as in the somites. (c) *in situ* hybridization for *asb2a* expression in 33 hpf embryos. Lateral view, anterior to the left. Scale bars, 200 μm . (d) Schematic representation of the reporter transgene: a *GFP-GAL4FF-polyA-Km^r* cassette was inserted at the ATG start codon in an *asb2b* BAC. Lateral views of a 50 hpf *Tg(asb2b:GFP-GAL4FF);Tg(UAS:mGFP)* embryo and close-up views of the heart; anterior to the left. An arrow points to the heart. GFP is expressed in the heart and somites. Scale bar, 200 μm . (e) Schematic representation of *asb2a* locus and the gRNA. Target sequence of gRNA and PAM are highlighted in blue and red, respectively. Inserted nucleotides are indicated in green. Predicted structure of WT and *Asb2a +13* mutant proteins. *asb2a +13* allele is predicted to encode a truncated polypeptide containing 25 incorrect amino acids (147-171 aa). (f) Bright-field micrographs of 50 hpf WT, *asb2a* mutant, *asb2b* mutant, and *asb2a;asb2b* double mutant embryos in lateral views. Scale bar, 200 μm . percentage of WT and mutant embryos exhibiting pericardial edema ($n=50$ fish). (g) Lateral close-up views of 50 hpf WT and *asb2b* mutant hearts in diastole and systole. Anterior to the left. Two-headed arrows indicate width of the ventricle. Quantification of ventricular fractional shortening (FS) and aortic blood flow velocity in WT and *asb2b* mutants. Heart rate measurement in 50 hpf WT and *asb2b* mutants ($n=4$ to 6 fish). Scale bar, 20 μm . (h) Relative mRNA expression (qPCR) of *myl7*, *vmhc* and *myh6* expression at 17.5 and 24 hpf in WT and *asb2b* mutants ($n=2$ technical replicates, RNA samples were obtained from 50 embryos). ** $P<0.01$ by one-way analysis of variance (ANOVA) followed by Tukey's HSD test. Error bars, SEM.



Supplementary Figure 2 - *asb2b* is involved in myofilament organization.

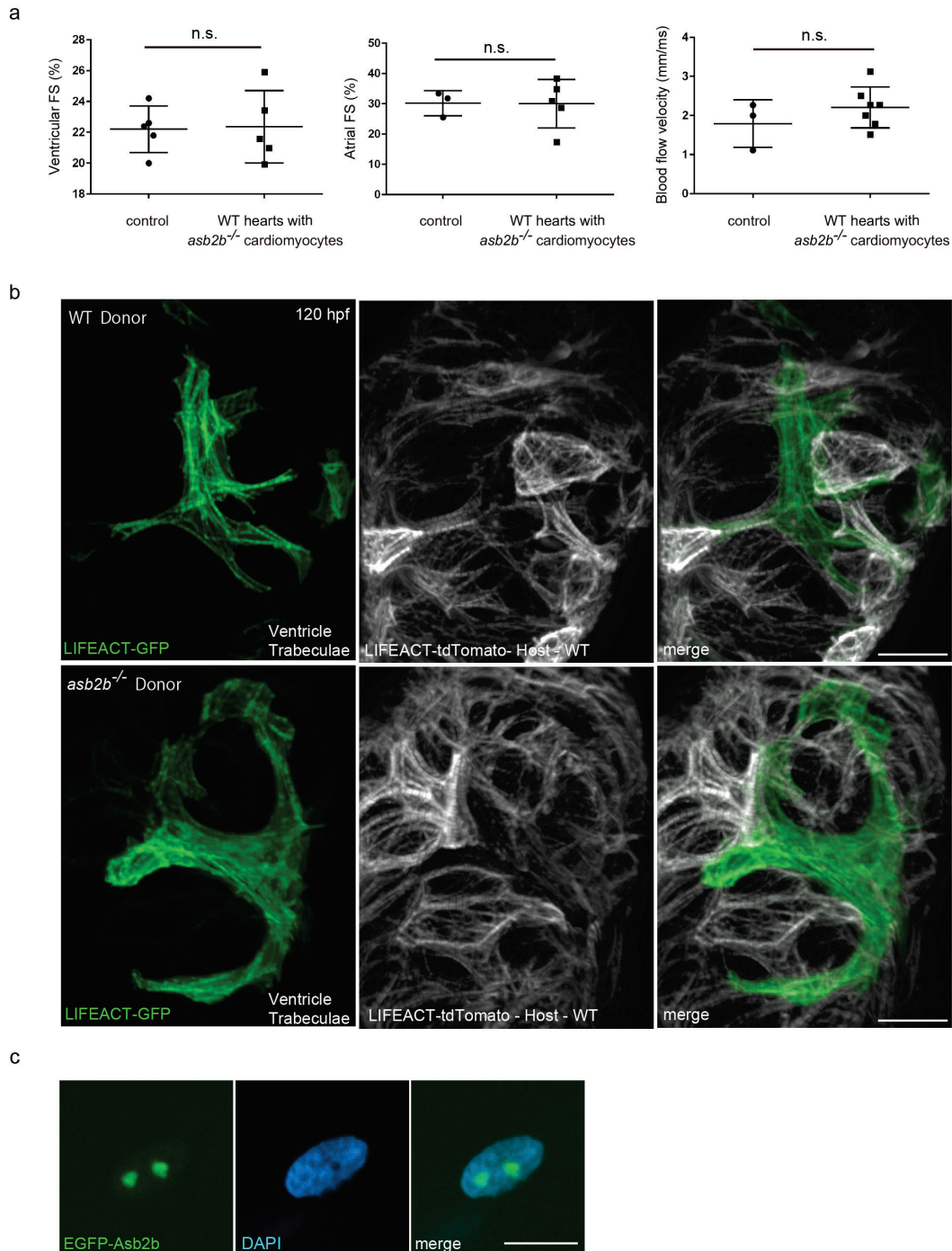
(a) 3D images of 50 hpf WT and *asb2b* mutant hearts stained with myosin heavy chain antibody (MF20). At, atrium; V, ventricle. Cardiomyocytes in the WT ventricle exhibit cortical myosin localization, while those in *asb2b* mutants show less cortical localization of myosin and exhibit punctae of myosin (arrow). WT atrial cardiomyocytes exhibit clear myofilaments, whereas fewer myofilaments are observed in *asb2b* mutants. (b) Electron

micrographs of 50 hpf WT and *asb2b* mutant ventricles. Close-up views of the cardiomyocyte cell-cell contact regions in yellow dotted boxes are shown (yellow squares with red arrowheads). Ventricular cardiomyocytes in *asb2b* mutants exhibit fewer regions of cell-cell contact (red arrowheads). *asb2b* mutant cardiomyocytes exhibit many more membrane protrusions (yellow arrowheads) compared to WT. Scale bars, 2 μm . Magnified views of myofilaments in white dotted boxes are shown (black arrows). Scale bar, 0.5 μm . (c) Lateral views of a 50 hpf control embryo and an embryo injected with 50 pg *asb2b* mRNA. Anterior to the left. Scale bar, 200 μm . Number of fish showing pericardial edema ($n=60$ fish). 1-cell stage embryos were injected with different amounts of *asb2b* mRNA and analyzed at 55 hpf. 3D images and close-up views of *Tg(myl7:LIFEACT-GFP)* hearts of 55 hpf embryos injected with 50 pg *asb2b* mRNA. At, atrium; V, ventricle. Error bars, SEM. Scale bars, 20 μm .

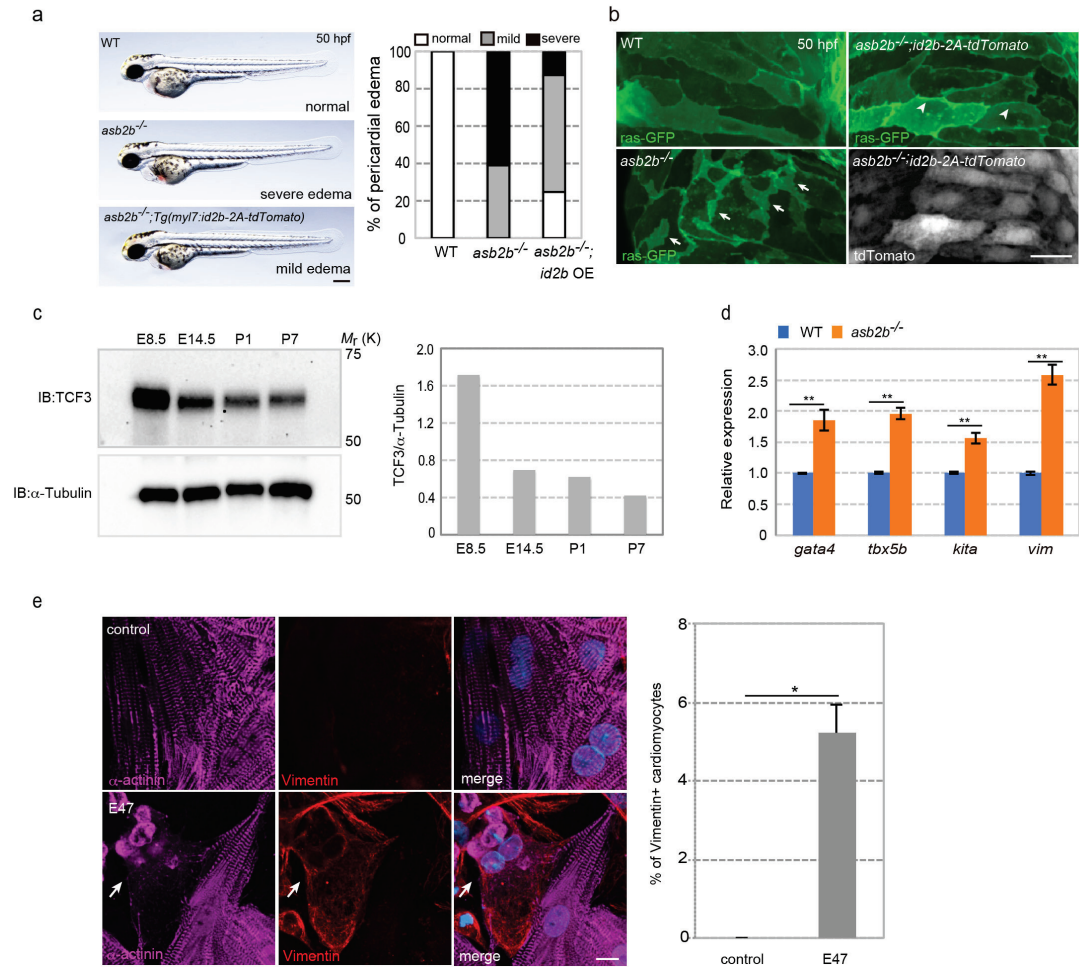


Supplementary Figure 3 – *asb2b* mutant cardiomyocytes exhibit irregular cell-cell junctions. (a) 3D images of 50 hpf *Tg(myl7:ras-GFP) asb2b* mutant atria mosaically expressing membrane-tdTomato (*mtdTomato*). Mosaic expression of *mtdTomato* was achieved by injecting a *myl7:mtdTomato* plasmid into *Tg(myl7:ras-GFP) asb2b* mutant

one-cell stage embryos. Membrane protrusions of mtdTomato positive cardiomyocytes are shown (arrows). (b) 3D images of 50 hpf *Tg(myl7:ras-GFP);Tg(myl7:N-cadherin-tdTomato)* WT and *asb2b* mutant atria. N-cadherin-tdTomato molecules in WT cardiomyocytes exhibit lateral localization in cell-cell junctions, while those in *asb2b* mutants also exhibit localization in overlapping regions (arrows). Number of cardiomyocytes exhibiting N-cadherin-tdTomato punctae in overlapping regions with neighbouring cardiomyocytes in 50 hpf WT and *asb2b* mutant hearts ($n=5$ hearts, with averages taken from 25 cardiomyocytes per heart). (c) Co-staining for N-cadherin (green), Myosin heavy chain (MF20, red) and DAPI (white) in 50 hpf WT and *asb2b* mutant hearts, transverse views. No obvious differences in N-cadherin expression are observed (arrows). (d) Relative mRNA expression (microarray) of junction component genes in isolated 50 hpf WT and *asb2b* mutant hearts. Gene names can be found in Supplementary Table 1. (e) 3D images of 50 hpf *Tg(myl7:EGFP-Podocalyxin);Tg(myl7:MKATE-CAAX)* WT and *asb2b* mutant atria. In WT hearts, EGFP-Podocalyxin shows uniform distribution across cell-cell borders (arrows), while those in *asb2b* mutants exhibit peripheral localization (arrows). Number of cardiomyocytes exhibiting EGFP-Podocalyxin localization in cardiomyocyte cell-cell borders in 50 hpf WT and *asb2b* mutant hearts ($n=5$ hearts, with averages taken from 20 cardiomyocytes per heart). (f) 3D images of 50 hpf *Tg(myl7:ras-GFP)* WT and *asb2b* mutant atria expressing PH-Akt-tdTomato-PEST. *asb2b* mutant cardiomyocytes exhibit enriched localization of PH-Akt-tdTomato-PEST in membrane protrusions (arrows). (g) Relative mRNA expression (microarray) of *pak1*, *rhoa*, *prkceb* and *prkcea* in isolated 50 hpf WT and *asb2b* mutant hearts. Gene names can be found in Supplementary Table 1. ** $P<0.01$ by one-way analysis of variance (ANOVA) followed by Tukey's HSD test. Error bars, SEM. Scale bars, 20 μm .

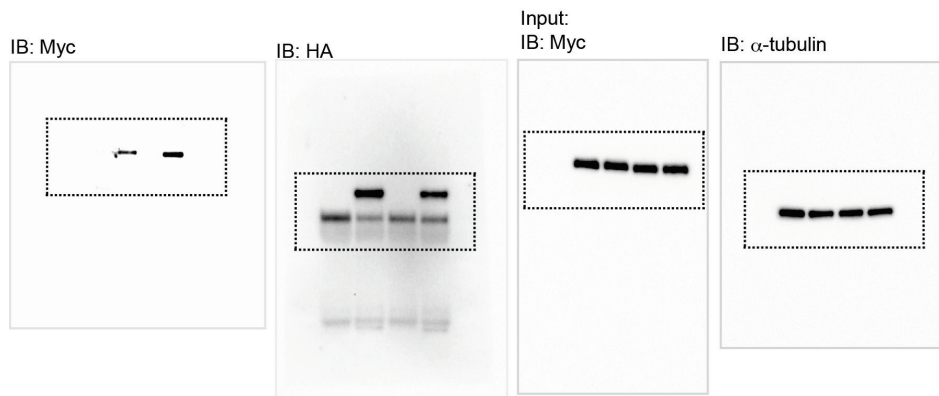


Supplementary Figure 4 – *asb2b* mutant cardiomyocytes in WT hearts exhibit disorganized myofilaments. (a) Quantification of fractional shortening (FS) in ventricles and atria in 50 hpf chimeric hearts, and aortic blood flow velocity. WT or *asb2b* mutant cells were transplanted into WT host embryos. No significant differences in FS or blood flow velocity were observed ($n=3$ to 7 fish). (b) 3D images of 120 hpf chimeric ventricles. *Tg(myf7:LIFEACT-GFP)* WT or *asb2b* mutant donor cells were transplanted into *Tg(myf7:LIFEACT-tdTomato)* WT host embryos. WT donor-derived cardiomyocytes exhibit mature sarcomeres in trabecular cardiomyocytes, while *asb2b* mutant-derived cardiomyocytes exhibit disorganized sarcomeres. (c) A 50 hpf *TgBAC(asb2b:GFP-asb2b)* heart stained with DAPI (blue). Scale bars, 20 μ m.

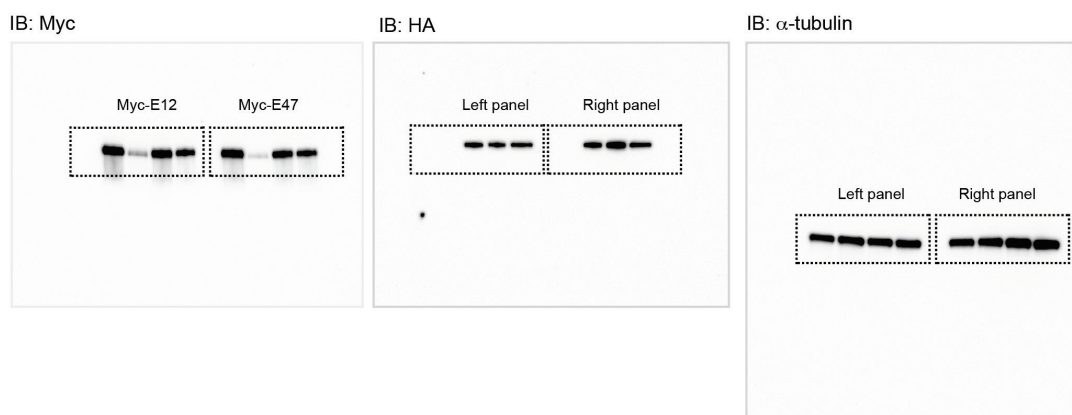


Supplementary Figure 5 - *asb2b* mutant and TCF3 overexpressing cardiomyocytes exhibit a number of phenotypes associated with cardiomyocyte dedifferentiation. (a) Brightfield micrographs of 50 hpf WT, *asb2b* mutant or *Tg(myl7:id2b-p2a-tdTomato)* *asb2b* mutant embryos in lateral views. Anterior to the left. Scale bar, 200 μ m. Number of fish exhibiting pericardial edema ($n=20$ fish). (b) 3D images of 50 hpf *Tg(myl7:ras-GFP)* WT and *asb2b* mutant atria as well as *Tg(myl7:ras-GFP);Tg(myl7:id2b-2A-tdTomato)* *asb2b* mutant atria. *asb2b* mutant cardiomyocytes exhibit irregular cell-cell borders and membrane protrusions (arrows), whereas *id2b*-overexpressing *asb2b* mutant cardiomyocytes exhibit less severe morphological phenotypes (arrowheads). (c) E8.5, E14.5, P1 and P7 mouse hearts were isolated and TCF3 protein levels were analyzed; relative expression levels shown on the right. (d) Relative mRNA expression (qPCR) of *gata4*, *tbx5b*, *kita* and *vim* in isolated 50 hpf WT (blue bar) and *asb2b* mutant (orange bar) hearts ($n=2$ technical replicates, RNA samples were obtained from 500 isolated hearts). (e) Rat NCMs transfected with control or *E47* adenovirus vectors, co-stained for α -actinin (magenta), Vimentin (red) and DAPI (blue). Arrow points to a Vimentin-positive cardiomyocyte. Number of Vimentin-positive Rat NCMs ($n=2$, with averages taken from 50 cardiomyocytes). * $P<0.05$, ** $P<0.01$ by one-way analysis of variance (ANOVA) followed by Tukey's HSD test. Error bars, SEM. Scale bars, 20 μ m.

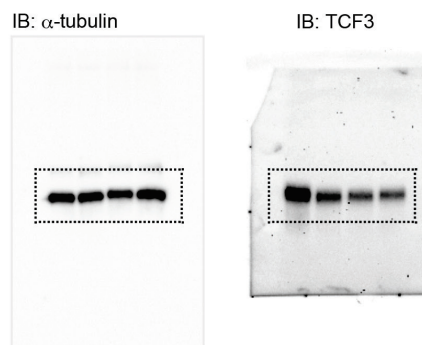
Uncropped images related to Fig. 4b



Uncropped images related to Fig. 4c



Uncropped images related to Supplementary Fig. 5c



Supplementary Figure 6 – Uncropped images related to western blotting data

Supplementary Tables

Cardiac differentiation marker			
ID	Gene Symbol	Gene Name	ratio (mutant/WT)
ENSDART00000102766	<i>ttni2a</i>	<i>troponin T type 2a</i>	1.20
ENSDART00000150079	<i>ttni1b</i>	<i>troponin I type 1b</i>	1.22
ENSDART00000138911	<i>actn2b</i>	<i>actinin, alpha 2b</i>	1.30
ENSDART00000017677	<i>ttna</i>	<i>titin, tandem duplicate 2</i>	1.87
ENSDART00000125953	<i>ttnb</i>	<i>titin, tandem duplicate 1</i>	1.36

Supplementary Fig. 3d - Junctional components			
ID	Gene Symbol	Gene Name	ratio (mutant/WT)
ENSDART00000024627	<i>cdh2</i>	<i>cadherin 2</i>	1.33
ENSDART00000047272	<i>cx43</i>	<i>connexin 43</i>	0.99
ENSDART00000149871	<i>cx40.8</i>	<i>connexin 40.8</i>	1.06
ENSDART00000123532	<i>pkp1a</i>	<i>plakophilin 1a</i>	1.06
ENSDART00000035899	<i>pkp2</i>	<i>plakophilin 2</i>	0.92
ENSDART00000139110	<i>pkp3a</i>	<i>plakophilin 3a</i>	1.02
ENSDART00000155685	<i>pkp4</i>	<i>plakophilin 4</i>	1.18
ENSDART00000019505	<i>pleca</i>	<i>plectin a</i>	1.22
ENSDART00000144558	<i>plecb</i>	<i>plectin b</i>	1.16
ENSDART00000108721	<i>tjp1a</i>	<i>tight junction protein 1a</i>	0.90

Supplementary Fig. 3g - actin regulators			
ID	Gene Symbol	Gene Name	ratio (mutant/WT)
ENSDART00000146147	<i>pak1</i>	<i>p21 protein (Cdc42/Rac)-activated kinase 1</i>	2.55
ENSDART00000090528	<i>rhoa</i>	<i>ras homolog family member Ca</i>	1.52
ENSDART00000045086	<i>prkceb</i>	<i>protein kinase C, epsilon b</i>	1.96
ENSDART00000105608	<i>prkcea</i>	<i>protein kinase C, epsilon a</i>	1.83

TCF3 targets			
ID	GeneSymbol	Gene Name	ratio (mutant/WT)
ENSDART00000138887	<i>cyp11a2</i>	<i>cytochrome P450, family 11, subfamily A, polypeptide 2</i>	3.46
ENSDART00000138702	<i>MEF2Cb</i>	<i>myocyte enhancer factor 2cb</i>	2.62
ENSDART00000125012	<i>caspl1</i>	<i>caspase 6, apoptosis-related cysteine peptidase, like 1</i>	2.60
ENSDART00000074181.2	<i>zp3.2</i>	<i>zona pellucida glycoprotein 3, tandem duplicate 2</i>	2.09
ENSDART00000108974	<i>kita</i>	<i>kit receptor a</i>	2.06
ENSDART00000149161	<i>abcc8</i>	<i>ATP-binding cassette, sub-family C (CFTR/MRP), member 8</i>	1.93
ENSDART00000055537	<i>socs1a</i>	<i>suppressor of cytokine signaling 1a</i>	1.84
ENSDART00000063775	<i>mal</i>	<i>mal, T-cell differentiation protein</i>	1.66
ENSDART00000140980	<i>sema3bl</i>	<i>sema domain, immunoglobulin domain (Ig), short basic domain, secreted, (semaphorin) 3bl</i>	1.62
ENSDART00000153127	<i>socs3b</i>	<i>suppressor of cytokine signaling 3b</i>	1.56
ENSDART00000137090	<i>id2a</i>	<i>inhibitor of DNA binding 2, dominant negative helix-loop-helix protein, a</i>	1.40

Supplementary Table 1 – Microarray data. 50 hpf WT and *asb2b* mutant hearts were isolated and RNA expression levels were analyzed by microarray.

Supplementary Fig. 1h

Sample Name	Assay Name	Ct Mean
17.5 hpf WT	<i>cmhc2</i>	26.76
17.5 hpf <i>asb2b</i> mutant	<i>cmhc2</i>	26.01
24 hpf WT	<i>cmhc2</i>	23.77
24 hpf <i>asb2b</i> mutant	<i>cmhc2</i>	23.64
17.5 hpf WT	<i>vmhc</i>	23.69
17.5 hpf <i>asb2b</i> mutant	<i>vmhc</i>	24.09
24 hpf WT	<i>vmhc</i>	22.19
24 hpf <i>asb2b</i> mutant	<i>vmhc</i>	22.10
17.5 hpf WT	<i>amhc</i>	29.45
17.5 hpf <i>asb2b</i> mutant	<i>amhc</i>	29.35
24 hpf WT	<i>amhc</i>	26.30
24 hpf <i>asb2b</i> mutant	<i>amhc</i>	26.84
17.5 hpf WT	<i>ef1α</i>	14.97
17.5 hpf <i>asb2b</i> mutant	<i>ef1α</i>	15.43
24 hpf WT	<i>ef1α</i>	15.19
24 hpf <i>asb2b</i> mutant	<i>ef1α</i>	15.30

Fig. 4a

Sample Name	Assay Name	Ct Mean
50 hpf WT hearts	<i>socs3b</i>	25.68
50 hpf <i>asb2b</i> mutant hearts	<i>socs3b</i>	24.29
50 hpf WT hearts	<i>id2a</i>	22.48
50 hpf <i>asb2b</i> mutant hearts	<i>id2a</i>	22.12
50 hpf WT hearts	<i>mef2cb</i>	28.83
50 hpf <i>asb2b</i> mutant hearts	<i>mef2cb</i>	27.55
50 hpf WT hearts	<i>ef1β</i>	23.79
50 hpf <i>asb2b</i> mutant hearts	<i>ef1β</i>	23.54

Supplementary Fig. 5d

Sample Name	Assay Name	Ct Mean
50 hpf WT hearts	<i>gata4</i>	21.08
50 hpf <i>asb2b</i> mutant hearts	<i>gata4</i>	20.51
50 hpf WT hearts	<i>tbx5b</i>	24.14
50 hpf <i>asb2b</i> mutant hearts	<i>tbx5b</i>	23.49
50 hpf WT hearts	<i>kita</i>	25.31
50 hpf <i>asb2b</i> mutant hearts	<i>kita</i>	24.98
50 hpf WT hearts	<i>vim</i>	24.83
50 hpf <i>asb2b</i> mutant hearts	<i>vim</i>	23.77
50 hpf WT hearts	<i>ef1β</i>	18.75
50 hpf <i>asb2b</i> mutant hearts	<i>ef1β</i>	19.07

Supplementary Table 2 – qPCR data. RNA expression levels were analyzed by RT-qPCR