

Fig. S1. Phenotypic characterization of S534A knock-in mice. (A) Close-up pictures of wild-type (WT) and S534A mice. (B) Two strategies for PCR-based genotyping of the mutant allele were used. Left: PCR amplification of a region flanking an Mfe I restriction site that was coupled to the mutation in the targeted allele. Right: Amplification of the remnant loxP site (after excision of the neomycin cassette) that was present on the mutant allele. (C and D) Spleen-body weight ratio and serum ALT levels were measured and H&E staining of the liver was performed on eight-week-old WT (n = 6) and S534A (n= 6) mice. (E and F) Spleen-body weight ratio and serum ALT were measured (E) and qPCR analysis of inflammatory gene expression in the liver (F) was performed on one-year-old male S534A (n = 5) and WT (n = 5) mice. Data are means \pm SD of 5 mice of each genotype.



Fig. S2. Determination of NF-κB activation in MEFs, spleens, and livers from WT and S534A mice. (A) WT and S534A MEFs were transduced with an adenoviral NF-κB reporter. The cells were left untreated (Ctrl) or were treated for 6 hours with TNF- α (30 ng/ml) or IL-1 β (5 ng/ml) before luciferase activity was measured. (B) WT and S534A MEFs were treated as described in (A) before being subjected to qPCR analysis of the abundances of the indicated mRNAs. Data in (A) and (B) are means ± SD of three independent experiments, each performed in triplicate. (C) WT (n = 9) and S534A (n = 9) mice were injected intravenously with LPS (1 mg/kg) and sacrificed 4 hours later. Splenocytes were analyzed by qPCR to determine the relative abundances of the indicated mRNAs. Data are means ± SD of 9 mice of each genotype. (D) WT (n = 8) and S534A (n = 10) mice were injected intravenously with TNF- α (5 µg/kg) and the mice were sacrificed 4 hours later. mRNA was extracted from the liver and analyzed by qPCR to determine the relative abundances of the indicated mRNAs. Data are means ± SD of 8 WT and 10 S534A mice. **P* < 0.001 vs. Ctrl.



Fig. S3. Analysis of gene expression in the livers of WT and S534A mice injected with low-dose LPS. WT (n=7) and S534A (n=7) mice were injected intravenously with LPS (1 μ g/kg) and sacrificed 4 hours later in one single experiment. Expression of *Ch25h*, *Il1a*, *Socs1*, and *Selp* mRNAs was determined by qPCR analysis and is expressed as fold induction in comparison to liver samples from untreated mice. Data are means ±SD of 7 mice per genotype .**P* < 0.05 vs. WT.



Fig. S4. Expression of NF- κ B-dependent genes in the spleens of WT and S534A mice at early and late time points after injection with low-dose LPS. (A and B) WT and S534A mice were injected intravenously with low-dose LPS (1 µg/kg) and spleens were collected 4 (A) and 8 (B) hours later. Expression of the indicated mRNAs was determined by qPCR analysis and is expressed as fold induction in comparison to liver samples from untreated mice. Data are means ± SD of 9 mice of each genotype (A) or of 6 WT and 10 S534A mice (B). *P < 0.05, **P < 0.01.



Fig. S5. Analysis of p65 protein abundance and NF- κ B target gene expression in macrophages from WT and S534A mice. (A) Bone marrow-derived macrophages (BMDM) from WT and S534A mice were left untreated or were treated with LPS (100 ng/ml) for the indicated times. Cells were analyzed by Western blotting with antibodies against the indicated proteins. (B) BMDM from WT (blue) and S534A (green) mice were treated with LPS for 6 hours. Expression of the indicated mRNAs was determined by qPCR analysis and is expressed as fold induction in comparison to untreated control macrophages. Data are means \pm SD of four experiments. * P<0.05; ** P<0.01; ns, non-significant



Fig. S6. IKK phosphorylates p65 at multiple sites. (A and B) p65 phosphorylation sites were analyzed by in vitro kinase assay in which recombinant IKK β protein was incubated with WT or mutant GST-p65 proteins (containing amino acid residues 354 to 551). The presence of equal amounts of GST-p65 substrate was confirmed by coomassie blue ("CBB") staining. The presence of IKK β was confirmed by detection of an autophosphorylation band of the appropriate size. (C) IKK phosphorylation sites in addition to the known Ser⁴⁶⁸ (S468) and Ser⁵³⁶ (S536) phosphorylation sites were investigated with different S536A constructs that had additional serine residues mutated. (D) Analysis of the in vitro phosphorylation of triple S468, S536A, and S547A mutants.