

Figure S1: UV-vis scan of 25 μ M s²c⁵Ura (left) or 25 μ M se²c⁵Ura (right) as a function of pH.

2

Notes to Supplemental Data: For each reaction described for the ¹H-NMR experiments in Figures 6-10, a separate experiment was conducted such that the reaction was allowed to proceed for 2 min, after which it was flash frozen and then lyophilized. The solid was then submitted for LCMS analysis. Each peak in the chromatogram corresponds to a compound identified by ESI-MS. Each peak is either labeled with the name of the compound along with the corresponding m/z value, or with a mass spectrogram shown as an inset figure with peaks labeled with the corresponding structure and m/z value. The legend to each figure identifies the identity of the compounds along with the corresponding m/z value. The red trace in the LC is a water blank and the **blue trace** in the LC represents the injection from the experiment. In each blue trace there is a peak at ~ 1.5 min that is from the buffer. A dehydration reaction (loss of 18 amu) occurs for all of the uracil derivatives as identified by the mass spectra.



Figure S2 (previous slide): 10 mM s²c⁵Ura was treated with 10 mM H₂O₂ in 100 mM potassium phosphate buffer pH 7.4, for 2 min and then flash frozen. The reaction was then lyophilized and submitted for LCMS analysis. The peak at 2.85 min in the HPLC trace was a mixture of c^{5} Ura (m/z = 157.1) and the decarboxylated c^5 Ura analog (m/z = 129.0 – structure shown) as identified in the mass spectrogram. The peak at 3.94 min in the HPLC trace contained a mixture of s^2c^5Ura (m/z = 173.0), the sulfenic acid form of s^2c^5 Ura (m/z = 189.0), and the disulfide form of s^2c^5Ura (m/z = 343.1) as identified in the mass spectrogram.

4



Figure S3 (previous slide): 10 mM se²c⁵Ura was treated with

10 mM H₂O₂ in 100 mM potassium phosphate buffer pH 7.4, for 2 min. The reaction was then flash frozen, lyophilized, and submitted for LCMS analysis. The peak at 2.94 min in the HPLC trace was identified in the mass spectrogram as c⁵Ura (m/ z = 157.1). The peak at 4.17 min in the HPLC trace contained a mixture of se^2c^5Ura (m/z = 221.1) and the diselende form of se^2c^5Ura (m/z = 439.1) as identified in the mass spectrogram. The unique isotopic pattern of selenium was clearly visible in the mass spectrogram. There were a number of other small peaks in the LC trace that could not be identified.



Figure S4 (previous slide): 10 mM s^2c^5 Ura was treated with 10 mM H_2O_2 in 100 mM potassium phosphate buffer, pH 7.4 for 2 min, after which 10 mM DTT (dissolved in the same buffer) was added and the reaction was allowed to proceed for a further 2 min. The reaction was then flash frozen, lyophilized, and then submitted for LCMS analysis. The peak at 2.90 min in the HPLC trace was identified in the mass spectrogram as $c^{5}Ura$ (m/z = 157.1). The peak at 4.02 min in the HPLC trace contains a mixture of s^2c^5 Ura (m/z = 173.0), the sulfenic acid form of s^2c^5Ura (m/z = 189.0), and the disulfide form of s^2c^5Ura (m/z = 343.1) as identified in the mass spectrogram. The peak at 10.56 min in the HPLC trace was identified as the oxidized form of DTT (m/z = 152.2 - mass spectrogram not shown). Last, the peak at 15.38 min in the HPLC trace is identified as the mixed disulfide between s^2c^5 Ura and DTT (m/z = 306.9).



Figure S5 (previous slide): 10 mM se²c⁵Ura was treated with 10 mM H₂O₂ in 100 mM potassium phosphate buffer pH 7.4, for 2 min, after which 10 mM DTT (dissolved in the same buffer) was added and the reaction was allowed to proceed for a further 2 min. The reaction was then flash frozen, lyophilized, and submitted for LCMS analysis. The peak at 2.75 min in the HPLC trace was identified in the mass spectrogram as $c^{5}Ura$ (m/z = 157.1). The peak at 3.97 min in the HPLC trace contains a mixture of se^2c^5Ura (m/z = 221.0) and the diselende form of $se^{2}c^{5}Ura$ (m/z = 439.0) as identified in the mass spectrogram. The peak at 9.77 min in the HPLC trace was identified as the oxidized form of DTT (m/z = 152.2). The peak at 15 min in the HPLC trace was identified as the mixed selenosulfide between se^2c^5Ura and DTT (m/z = 355.1 as shown in the mass spectrogram of the inset).



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Figure S6 (previous slide): 10 mM s²c⁵Ura was treated with 10 mM H₂O₂ in 100 mM potassium phosphate buffer pH 7.4, for 2 min, after which 10 mM ascorbate (dissolved in the same buffer) was added and the reaction was allowed to proceed for a further 2 min. The reaction was then flash frozen, lyophilized, and then submitted for LCMS analysis. The peak at 1.94 min in the HPLC trace was identified as ascorbate (m/z = 177.1). The peak at 2.75 min in the HPLC trace was identified in the mass spectrogram as c^{5} Ura (m/z = 157.1). The peak at 3.91 min in the HPLC trace contained a mixture of s^2c^5Ura (m/z = 173.0), the sulfenic acid form of s^2c^5 Ura (m/z = 189.0), and the disulfide form of s^2c^5Ura (m/z = 343.1) as identified in the mass spectrogram.



Figure S7 (previous slide): 10 mM se²c⁵Ura was treated with 10 mM H_2O_2 in 100 mM potassium phosphate buffer pH 7.4, for 2 min, after which 10 mM ascorbate (dissolved in the same buffer) was added and the reaction was allowed to proceed for a further 2 min. The reaction was then flash frozen, lyophilized, and submitted for LCMS analysis. The peak at 2.10 min in the HPLC trace was identified as ascorbate (m/z = 177.1). The peak at 2.84 min in the HPLC trace was identified in the mass spectrogram as c^{5} Ura (m/z = 157.1). The peak at 4.05 min in the HPLC trace contains a mixture of se^2c^5Ura (m/z = 221.1) and the diselenide form of se^2c^5Ura (m/z = 439.1) as identified in the mass spectrogram. The peak at 10.85 min in the HPLC trace could not be identified by mass analysis.



m/z

Figure S8 (previous slide): 10 mM s²c⁵Ura that was **preincubated** with 10 mM GSH, was treated with 10 mM H_2O_2 in 100 mM potassium phosphate buffer pH 7.4, for 2 min. The reaction was then flash frozen, lyophilized, and submitted for LCMS analysis. The peak at 2.86 min in the HPLC trace was identified in the mass spectrogram as $c^{5}Ura$ (m/z = 157.1). The peak at 3.98 min in the HPLC trace contained a mixture of s^2c^5Ura (m/z = 173.0), the sulfenic acid form of s^2c^5Ura (m/z = 189.0), and the disulfide form of s^2c^5 Ura (m/z = 343.1) as identified in the mass spectrogram. The peak at 4.61 min in the HPLC trace was identified as a mixture of GSH (m/z = 307.2)and GSSG (m/z = 613.2). GSH = reduced glutathione and GSSG = oxidized glutathione.



Figure S9 (previous slide): 10 mM se²c⁵Ura that was preincubated with 10 mM GSH, was treated with 10 mM H_2O_2 in 100 mM potassium phosphate buffer pH 7.4, for 2 min. The reaction was then flash frozen, lyophilized, and submitted for LCMS analysis. The peak at 4.10 min in the HPLC trace contained a mixture of se^2c^5Ura (m/z = 221.0), the selenenic acid form of se^2c^5Ura (m/z = 237.1), and the diselenide form of se^2c^5Ura (m/z = 439.0) as identified in the mass spectrogram. The peak at 4.64 min in the HPLC trace was identified as a mixture of GSH (m/z = 307.2) and GSSG (m/z = 613.2). GSH = reduced glutathione and GSSG = oxidized glutathione. The peak at 14.46 min in the HPLC trace was tentatively identified as a trimer of se^2c^5Ura (m/z = 646.1). We cannot be definitive about the identification, although the selenium isotope pattern is clearly visible in the mass spectrogram.



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Figure S10 (previous slide): 10 mM s²c⁵Ura that was preincubated with 10 mM ascorbate, was treated with 10 mM H_2O_2 in 100 mM potassium phosphate buffer pH 7.4, for 2 min. The reaction was then flash frozen, lyophilized, and submitted for LCMS analysis. The peak at 2.03 min in the HPLC trace is identified as ascorbate (m/z = 177.1). The peak at 2.82 min in the HPLC trace was identified in the mass spectrogram as c⁵Ura (m/z = 157.1). The peak at 4.02 min in the HPLC trace is identified as a mixture of s^2c^5 Ura (m/z = 173.0), the sulfenic acid form of s^2c^5 Ura at (m/z = 189.0), and the disulfide form of s^2c^5 Ura (m/z = 343.1) as identified in the mass spectrogram.



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22

Figure S11 (previous slide): 10 mM se²c⁵Ura that was preincubated with 10 mM ascorbate was treated with 10 mM H_2O_2 in 100 mM potassium phosphate buffer pH 7.4, for 2 min. The reaction was then flash frozen, lyophilized, and submitted for LCMS analysis. The peak at 2.01 min in the HPLC trace was identified as ascorbate (m/z = 177.1). The peak at 4.16 min in the HPLC trace was identified as a mixture of se²c⁵Ura (m/z = 221.0), the selenenic acid form of se²c⁵Ura (m/z = 237.1), and the diselenide form of se²c⁵Ura (m/z = 439.0) in the mass spectrogram.



Figure S12: Full ⁷⁷Se-NMR spectrum of the reaction shown in Figure 12 of the text after 18 hours of reaction. Only the resonance corresponding to se^2c^5Ura is visible at 188.5 ppm.