

SUPPORTING INFORMATION

Photocontrol of Anion Binding Affinity to a Bis-Urea Receptor Derived from Stiff-Stilbene

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Experimental Section

General methods and materials:

THF, toluene and CH₂Cl₂ were dried by using an MBraun solvent purification system. Dry DMSO and DMSO-*d*₆ were purchased from Acros Organics and were stored under N₂ over molecular sieves (4Å). The degassing of solvents was carried out by purging with N₂ for 30 min. All other chemicals were commercial products and were used as received. Flash chromatography (FC) was performed using silica gel (SiO₂) purchased from Merck (type 9385, 230-400 mesh) and thin-layer chromatography (TLC) was carried out on aluminum sheets coated with silica 60 F254 obtained from Merck; compounds were visualized with a UV lamp (254 nm) or by staining with phosphomolybdic acid (PMA). Melting points (m.p.) were determined using a Büchi-B545 capillary melting point apparatus. ¹H and ¹³C spectra were recorded on Varian Mercury-Plus 400 and Varian Mercury-Plus 600 spectrometers at 298K unless indicated otherwise. Chemical shifts (δ) are denoted in parts per million (ppm) relative to DMSO-*d*₆ (for ¹H detection, δ = 2.50 ppm; for ¹³C detection, δ = 39.52 ppm). Due to poor solubility, DMSO and H₂O peaks in the NMR spectra are larger than the compound peaks. The splitting pattern of peaks is designated as follows: s (singlet), d (doublet), t (triplet), (multiplet), br (broad), or dd (doublet of doublets). High-resolution mass spectrometry (ESI-MS) was performed on a LTQ Orbitrap XL spectrometer with ESI ionization. UV-vis spectra were recorded on Specord S600 and Hewlett-Packard HP 8543 diode arrays, equipped with a Quantum Northwest Peltier controller, in a 1 cm or 1 mm quartz cuvette. Irradiation of UV-vis samples was carried out at 20 °C using a Thorlab model M365F1 high-power LED (4.1 mW) and a Thorlab model M385F1 high-power LED (10.7 mW) positioned at a distance of 1 cm. NMR irradiation studies were performed at 20 °C with a Thorlab model M365FP1 high-power LED (15.5 mW) and a Thorlab model M385FP1 high-power LED (23.2 mW) coupled to a 600 μ m optical fiber, which guided the light into the NMR tube inside the spectrometer.¹

(E)/(Z)-6,6'-dibromo-2,2',3,3'-tetrahydro-1,1'-biindenylidene [(E)/(Z)-2]:

Titanium(IV) chloride (3.30 mL, 30.0 mmol) was slowly added to zinc powder (3.93 g, 60.0 mmol) in THF (30 mL) under a N₂ atmosphere and under vigorous stirring. The solution was stirred at reflux for 2 h and cooled back to rt, after which solid 6-bromo-1-indanone (3.17 g, 15.0 mmol) was added to the black suspension. The mixture was stirred at reflux for a further 24 h, treated with a saturated aqueous NH₄Cl solution, and extracted with CHCl₃ (3 \times 100

mL). The volume of the combined extracts was reduced to 30 mL and the precipitate was filtered off and air-dried to afford (*E*)-**2** (842 mg, 29%) as an off-white solid; m.p. 240.7 – 242.6 °C; ¹H NMR (400 MHz, DMSO-*d*₆, (*E*)-geometry assignment based on a NOESY spectrum): 7.68 (d, *J* = 1.8 Hz, 2H; 2 arom. H), 7.42 (dd, *J* = 8.0, 1.7 Hz, 2H; 2 arom. H), 7.33 (d, *J* = 8.0 Hz, 2H; 2 arom. H), 3.14-3.02 (m, br. 8H; 4 CH₂); too insoluble for a ¹³C NMR measurement. The filtrate was concentrated and FC (SiO₂, pentane) afforded (*Z*)-**2** (673 mg, 23%) as a white solid; m.p. 137.6 – 139.9 °C; ¹H NMR (400 MHz, DMSO-*d*₆): 8.01 (s, 2H; 2 arom. H), 7.40 (d, *J* = 8.0, 2H; 2 arom. H), 7.32 (d, *J* = 8.2 Hz, 2H; 2 arom. H), 2.95-2.89 (m, br. 4H; 2 CH₂), 2.92-2.75 (m, br. 4H; 2 CH₂); ¹³C NMR (100 MHz, DMSO-*d*₆): 147.6, 141.8, 135.1, 130.1, 127.5, 125.0, 118.7, 34.4, 29.6.

(*E*)-2,2',3,3'-tetrahydro-(1,1'-biindenylidene)-6,6'-diamine [(*E*)-3**]:**

Compound (*E*)-**2** (960 mg, 2.46 mmol), palladium(II) acetate (44 mg, 0.20 mmol), DPPF (136 mg, 0.25 mmol) and sodium tert-butoxide (473 mg, 4.92 mmol) were brought under a N₂ atmosphere in a Schlenk tube via three vacuum/N₂ cycles. Then degassed toluene (12 mL) was added, followed by benzophenone imine (1.03 mL, 6.15 mmol). The mixture was stirred at 90 °C for 20 h, cooled to rt and diluted with water (10 mL). The aqueous layer was extracted with CHCl₃ (3 × 25 mL) and the combined organic phases were dried over Na₂SO₄ and concentrated. FC (SiO₂, 0.1% NEt₃ in CH₂Cl₂) afforded the imine intermediate as a yellow solid, which was redissolved in THF (100 mL). A 2M aqueous HCl solution (50 mL) was added and after 1.5 h of stirring, the solution was made basic by addition of a saturated aqueous KHCO₃ solution (pH ~ 10). The mixture was extracted with EtOAc (3 × 50 mL) and the combined organic phases were dried over Na₂SO₄ and concentrated. The product was precipitated in Et₂O, filtered off and air-dried to afford (*E*)-**3** (503 mg, 78%) as a light-brown solid; m.p. 260 °C (decomp); ¹H NMR (400 MHz, DMSO-*d*₆): 6.97 (d, *J* = 8.0 Hz, 2H; 2 arom. H), 6.87 (d, *J* = 2.0 Hz, 2H; 2 arom. H), 6.45 (dd, *J* = 8.0, 1.9 Hz, 2H; 2 arom. H), 4.89 (s, 4H; 2 NH₂), 3.03-2.84 (m, br. 8H; 4 CH₂); ¹³C NMR (100 MHz, DMSO-*d*₆): 147.2, 143.4, 134.7, 134.1, 124.7, 113.6, 109.8, 31.9, 29.5; HRMS (ESI) *m/z*: 263.1544 ([M + H]⁺, calcd for C₁₈H₁₉N₂⁺: 263.1543).

(*Z*)-2,2',3,3'-tetrahydro-(1,1'-biindenylidene)-6,6'-diamine [(*Z*)-3**]:**

Compound (*Z*)-**2** (735 mg, 1.88 mmol), palladium(II) acetate (34 mg, 0.15 mmol), DPPF (104 mg, 0.19 mmol) and sodium tert-butoxide (362 mg, 3.77 mmol) were brought under a N₂ atmosphere in a Schlenk tube via three vacuum/N₂ cycles. Then degassed toluene (9 mL) was added, followed by benzophenone imine (0.79 mL, 4.71 mmol). The mixture was stirred at 90

°C for 16 h, cooled to rt and diluted with water (10 mL). The aqueous layer was extracted with CH₂Cl₂ (3 × 25 mL) and the combined organic phases were dried over Na₂SO₄ and concentrated. FC (SiO₂, 0.1% NEt₃ in CH₂Cl₂) afforded the imine intermediate as a yellow oil, which was redissolved in THF (20 mL). A 2M aqueous HCl solution (10 mL) was added and after 30 min of stirring, the mixture was diluted with water (10 mL) and extracted with Et₂O (4 × 10 mL). The water layer was then made basic by addition of a saturated aqueous KHCO₃ solution (pH ~ 10) and extracted with EtOAc (3 × 25 mL). The combined extracts were dried over Na₂SO₄ and concentrated to afford (*Z*)-**3** (332 mg, 67%) as a light-brown solid; m.p. 92.0 – 92.9 °C; ¹H NMR (400 MHz, DMSO-*d*₆): 7.27 (d, *J* = 2.1 Hz, 2H; 2 arom. H), 6.96 (d, *J* = 8.0 Hz, 2H; 2 arom. H), 6.45 (dd, *J* = 8.0, 2.0 Hz, 2H; 2 arom. H), 4.84 (s, 4H; 2 NH₂), 2.78-2.64 (m, br. 8H; 4 CH₂); ¹³C NMR (100 MHz, DMSO-*d*₆): 146.5, 140.5, 135.7, 134.6, 125.0, 114.2, 109.1, 34.8, 29.0; HRMS (ESI) *m/z*: 263.1543 ([M +H]⁺, calcd for C₁₈H₁₉N₂⁺: 263.1543).

(*E*)-1,1'-(2,2',3,3'-tetrahydro-(1,1'-biindenylidene)-6,6'-diyl)bis(3-phenylurea) [(*E*)-1]:

Phenyl isocyanate (66 μL, 0.60 mmol) was added to compound (*E*)-**3** (72 mg, 0.27 mmol) in CH₂Cl₂ (40 mL) under a N₂ atmosphere. The mixture was stirred at reflux for 3 d, after which the white precipitate was filtered off. The product was recrystallized from DMSO, washed vigorously with CH₂Cl₂ and air-dried to afford (*E*)-**1** (78 mg, 57%) as a beige solid; m.p. 260 °C (dec); ¹H NMR (400 MHz, DMSO-*d*₆): 8.71 (s, 2H; 2 NH), 8.63 (s, 2H; 2 NH), 7.89 (s, 2H; 2 arom. H), 7.46 (d, *J* = 8.0 Hz, 4H; 4 arom. H), 7.31-7.20 (m, 8H; 8 arom. H), 6.97 (t, *J* = 7.4 Hz, 2H; 2 arom. H), 3.15-3.01 (m, br. 8H; 4 CH₂); too insoluble for a ¹³C NMR measurement; HRMS (ESI) *m/z*: 501.2275 ([M +H]⁺, calcd for C₃₂H₂₉N₄O₂⁺: 501.2285).

(*Z*)-1,1'-(2,2',3,3'-tetrahydro-(1,1'-biindenylidene)-6,6'-diyl)bis(3-phenylurea) [(*Z*)-1]:

Phenyl isocyanate (86 μL, 0.79 mmol) was added to compound (*Z*)-**3** (103 mg, 0.39 mmol) in CH₂Cl₂ (5 mL) under a N₂ atmosphere. The solution was stirred for 16 h, after which the white precipitate was filtered off and air-dried to afford (*Z*)-**1** (137 mg, 70%) as a white solid; m.p. 340 °C (dec); ¹H NMR (400 MHz, DMSO-*d*₆): 8.74 (s, 2H; 2 NH), 8.50 (s, 2H; 2 NH), 8.11 (d, *J* = 2.1 Hz, 2H; 2 arom. H), 7.46-7.40 (m, 6H; 6 arom. H), 7.23 (d, *J* = 8.2 Hz, 2H; 2 arom. H), 7.10 (t, *J* = 7.8 Hz, 4H; 4 arom. H), 6.83 (t, *J* = 7.4 Hz, 2H; 2 arom. H), 2.93-2.75 (m, br. 8H; 4 CH₂); ¹³C NMR (100 MHz, DMSO-*d*₆): 152.6, 141.3, 140.1, 139.8, 137.5, 134.8, 128.6, 125.1, 121.5, 118.1, 117.7, 113.6, 34.4, 29.4; HRMS (ESI) *m/z*: 501.2275 ([M +H]⁺, calcd for C₃₂H₂₉N₄O₂⁺: 501.2285); elemental analysis calcd (%) for C₃₂H₂₈N₄O₂·²/₅C₂H₆SO: C 74.07, H 5.76, N 10.53; found: C 74.01, H 5.77, N 10.51.

^1H and ^{13}C NMR Spectra of new compounds

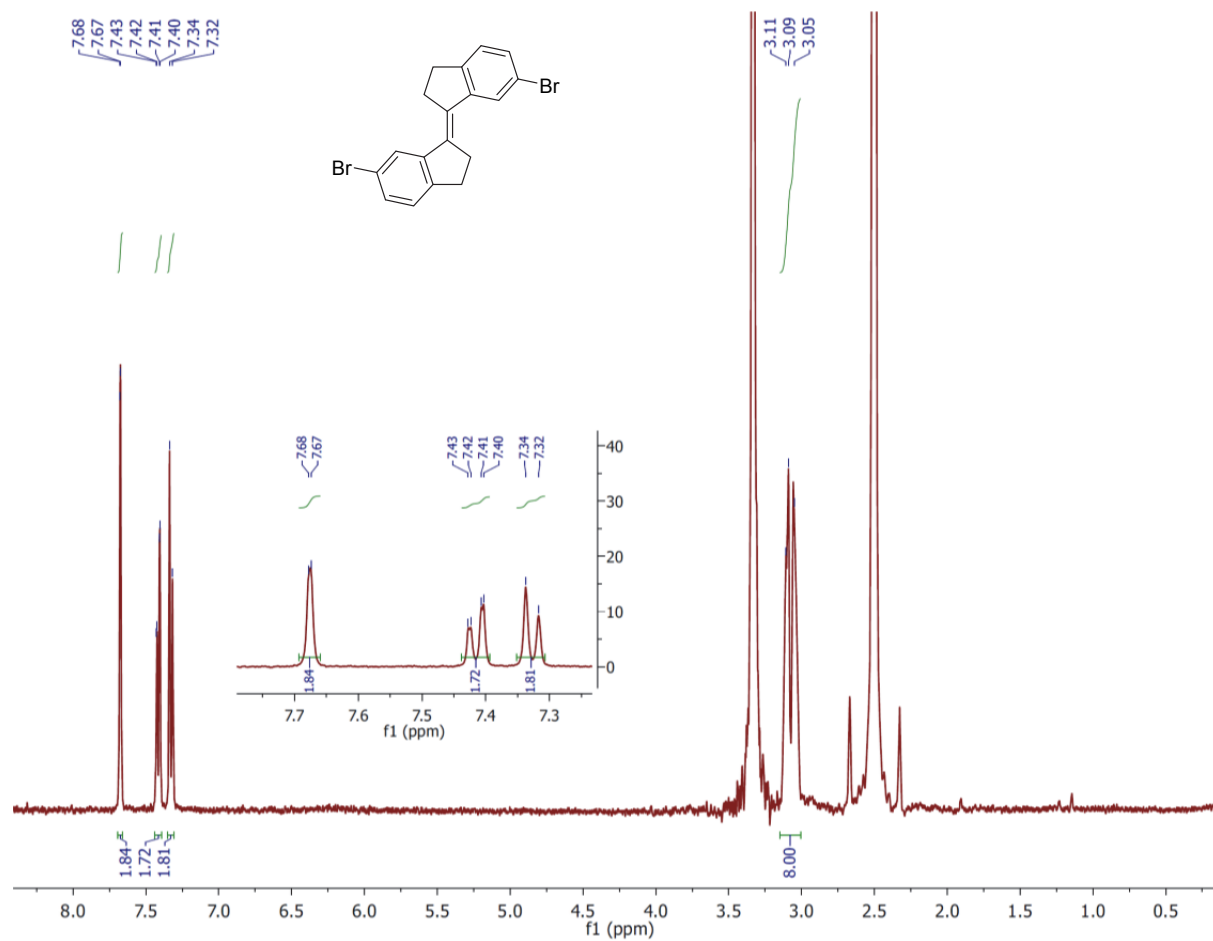


Figure S1. 400 MHz ^1H NMR spectrum of *(E)*-2 measured at 298 K in $\text{DMSO-}d_6$.

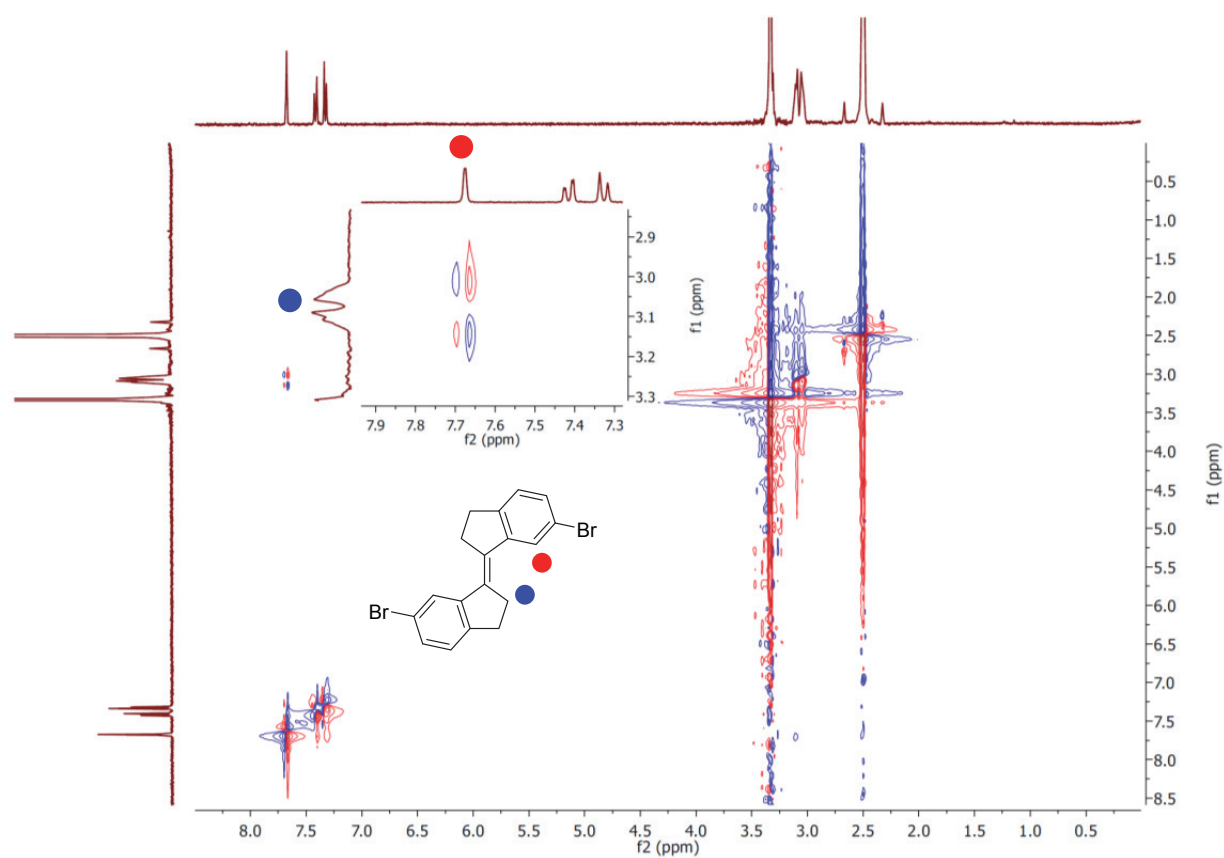


Figure S2. NOESY NMR spectrum of *(E)*-2 measured at 298 K in DMSO-*d*₆.

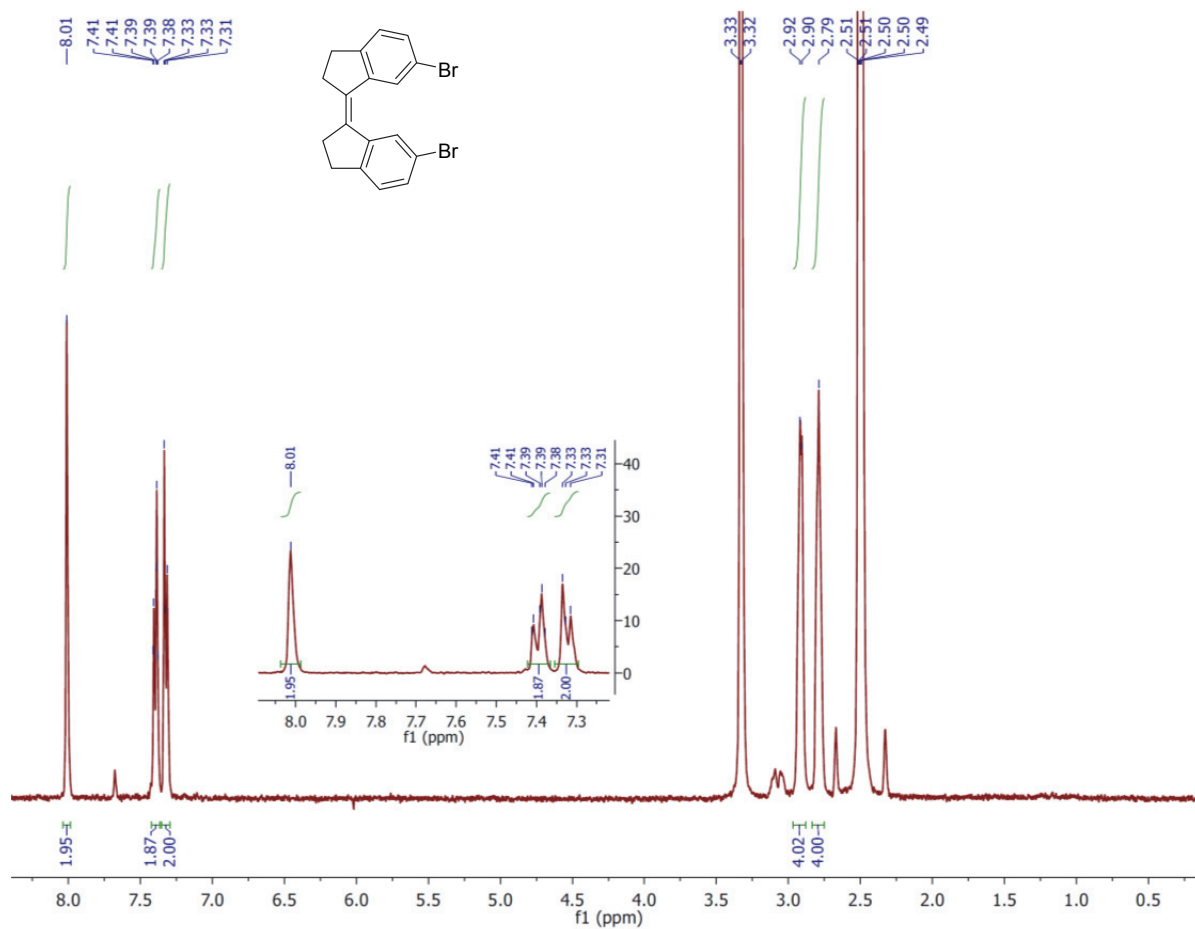


Figure S3. 400 MHz ^1H NMR spectrum of (Z)-2 measured at 298 K in $\text{DMSO-}d_6$.

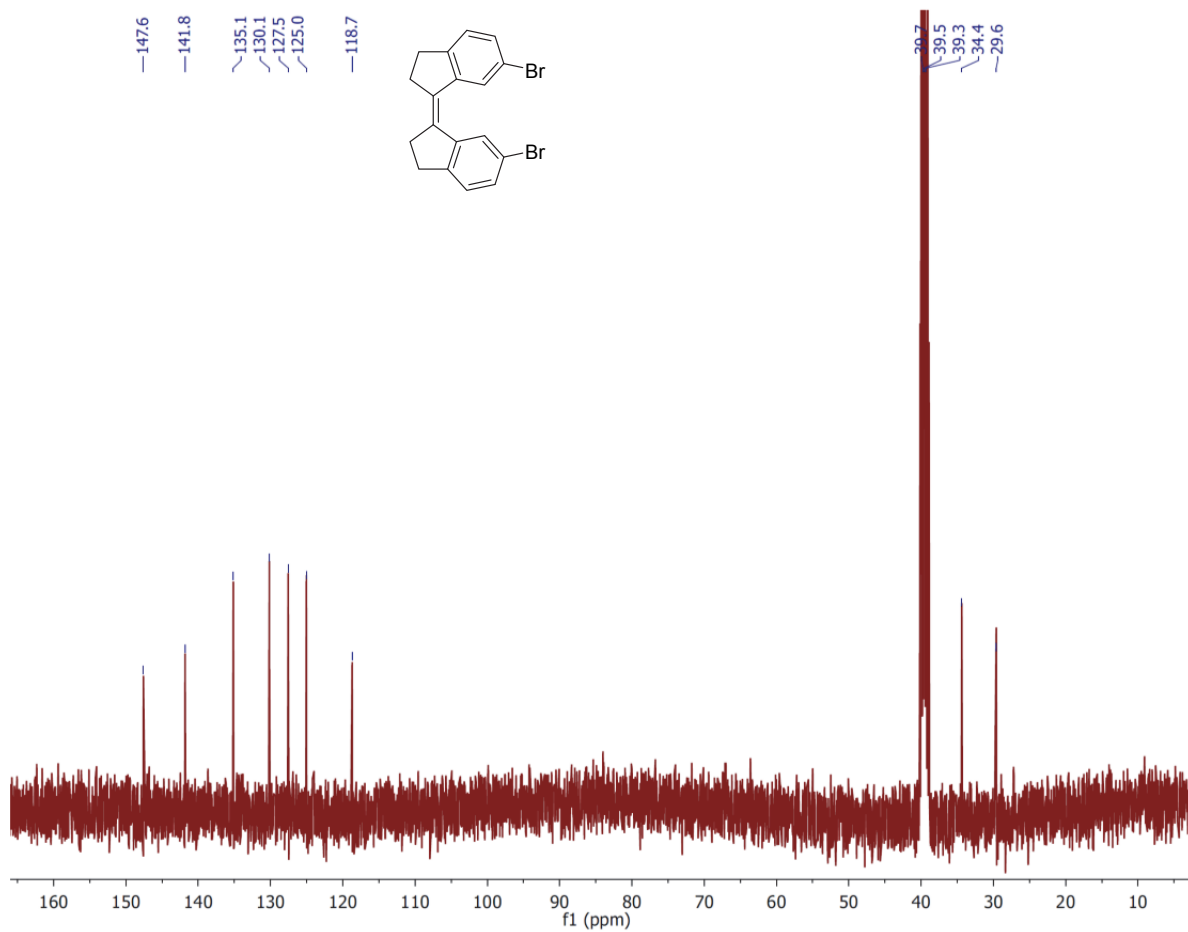


Figure S4. 100 MHz ^{13}C NMR spectrum of (Z)-2 measured at 298 K in $\text{DMSO-}d_6$.

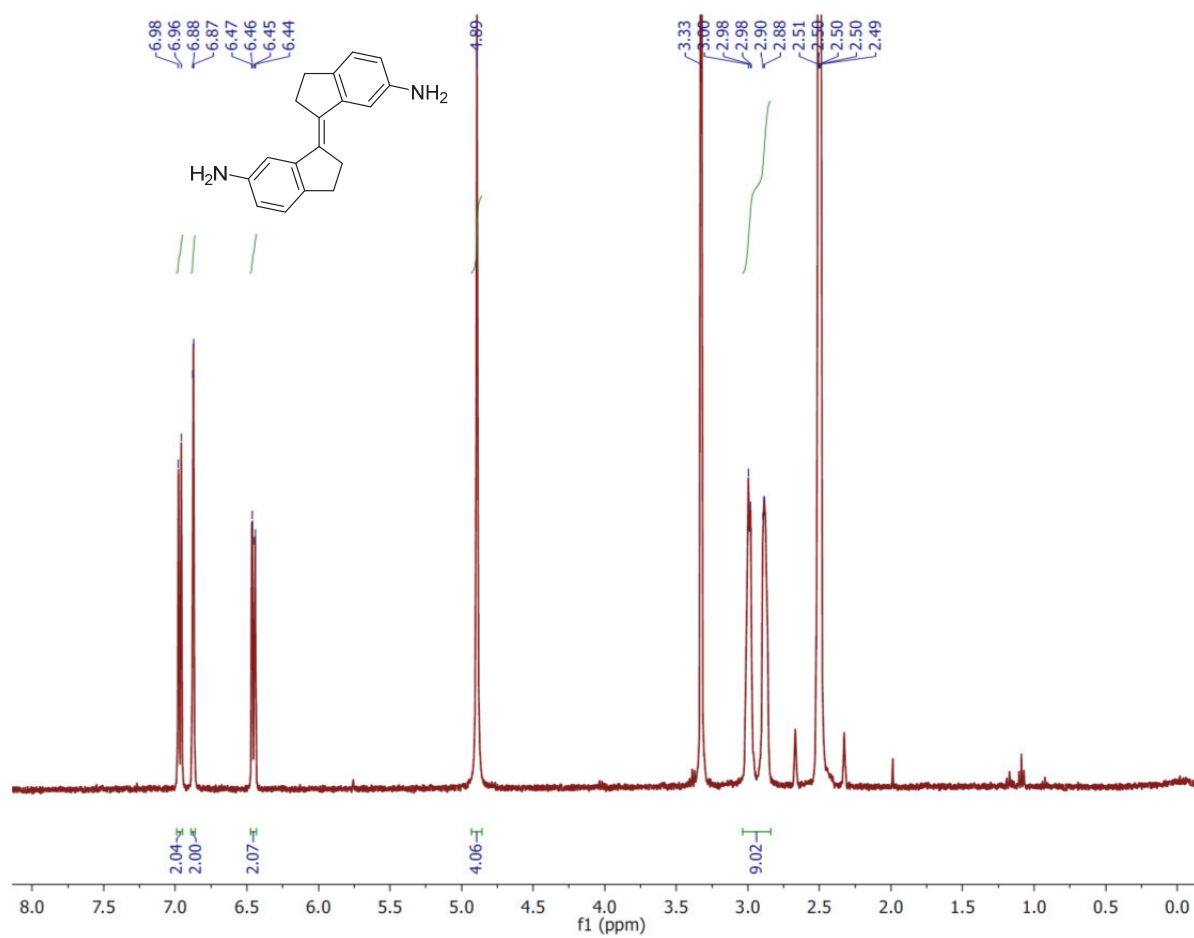


Figure S5. 400 MHz ¹H NMR spectrum of (*E*)-**3** measured at 298 K in DMSO-*d*₆.

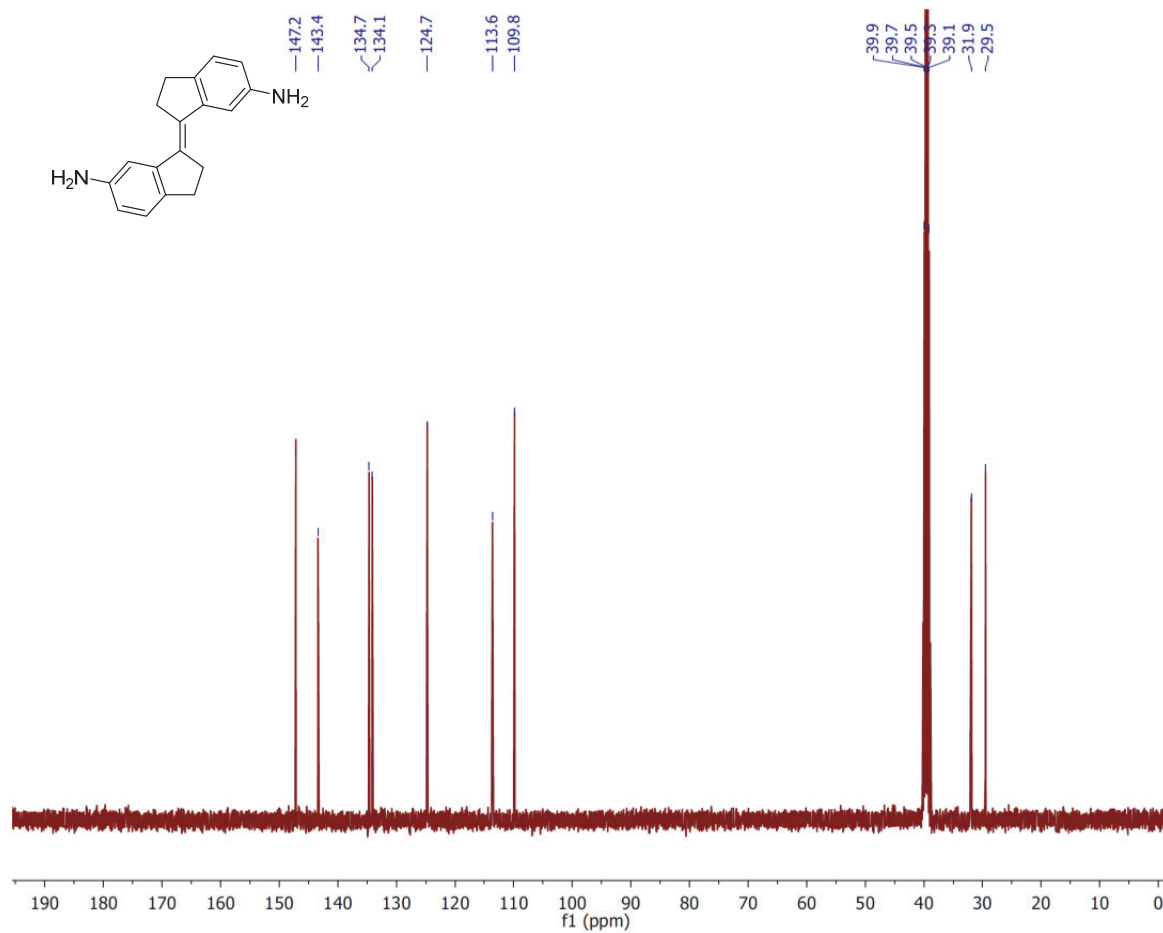


Figure S6. 100 MHz ¹³C NMR spectrum of (*E*)-**3** measured at 298 K in DMSO-*d*₆.

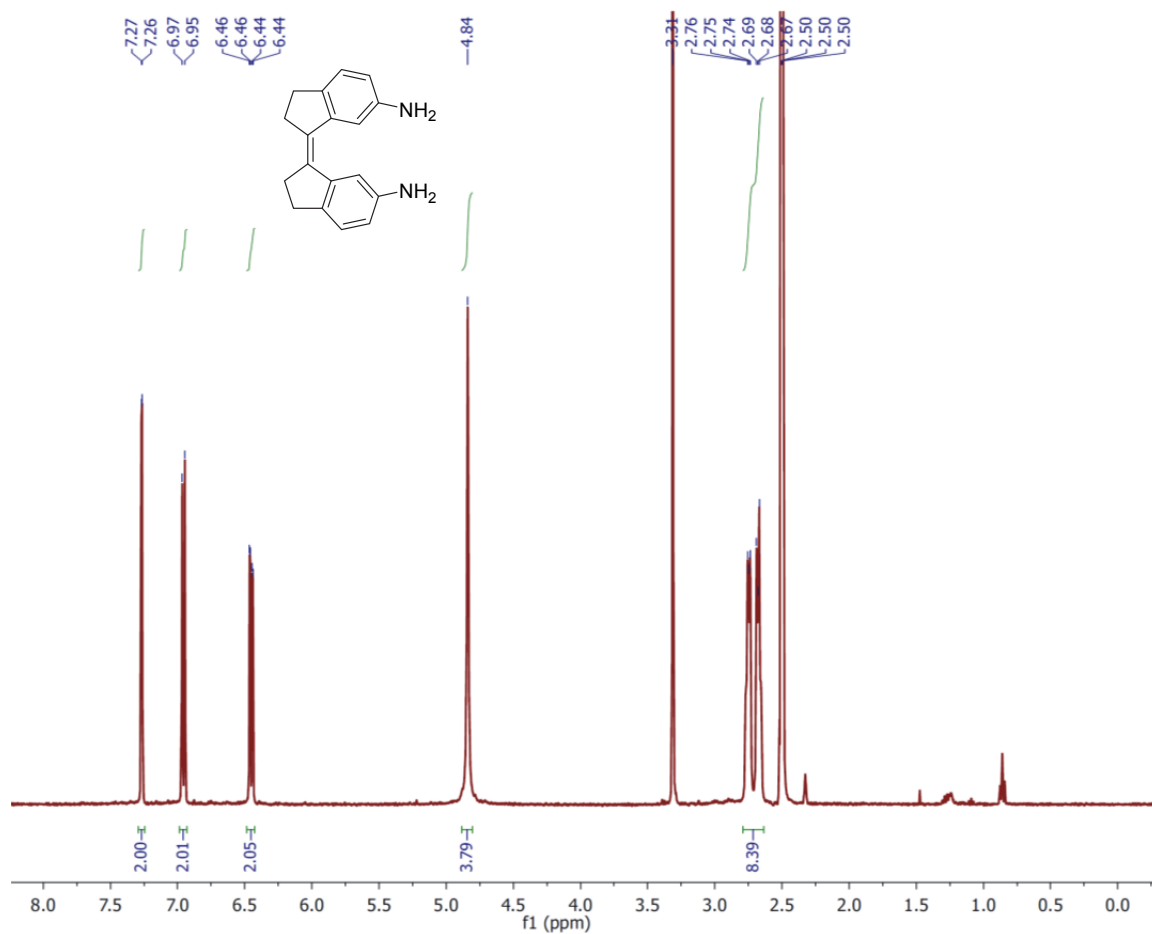


Figure S7. 400 MHz ^1H NMR spectrum of (Z)-3 measured at 298 K in $\text{DMSO-}d_6$.

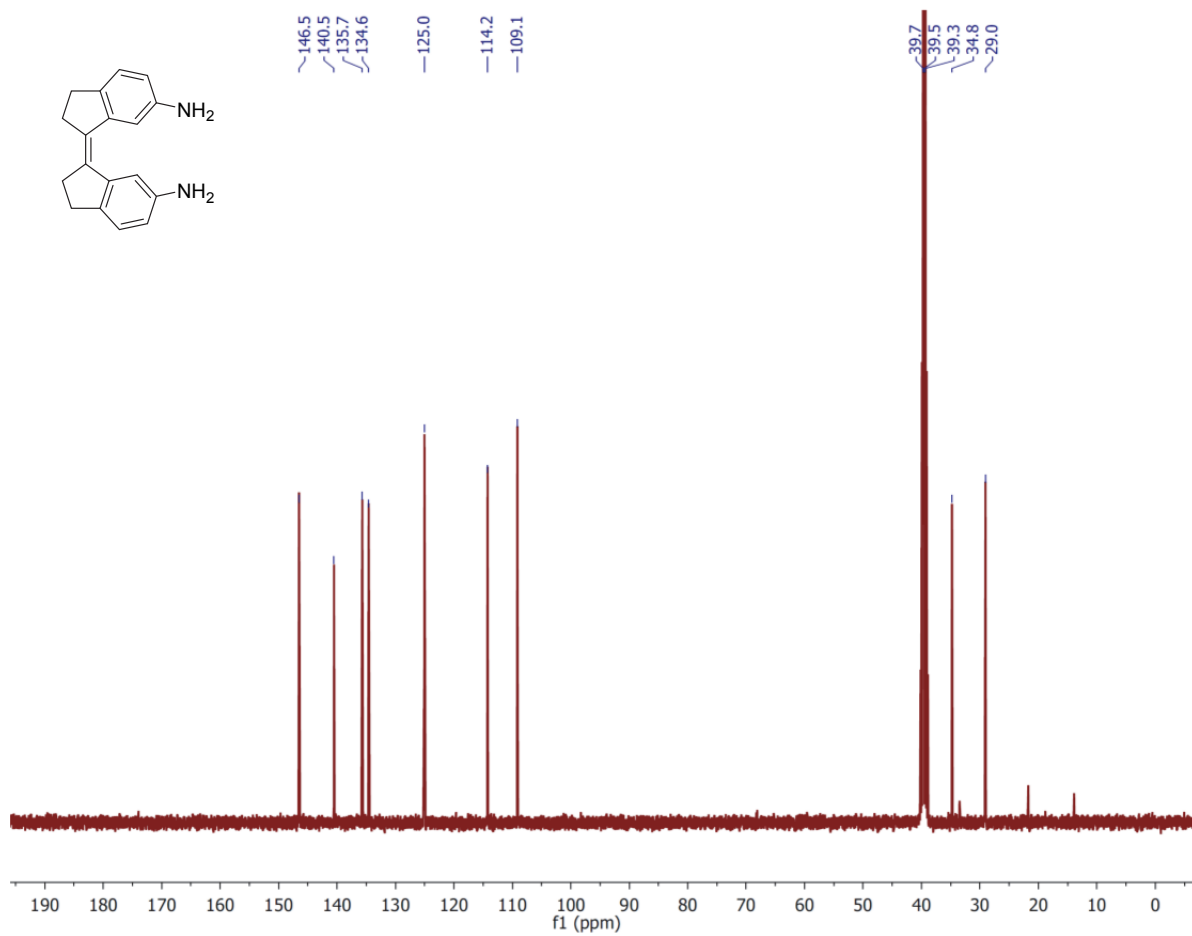


Figure S8. 100 MHz ¹³C NMR spectrum of (Z)-**3** measured at 298 K in DMSO-*d*₆.

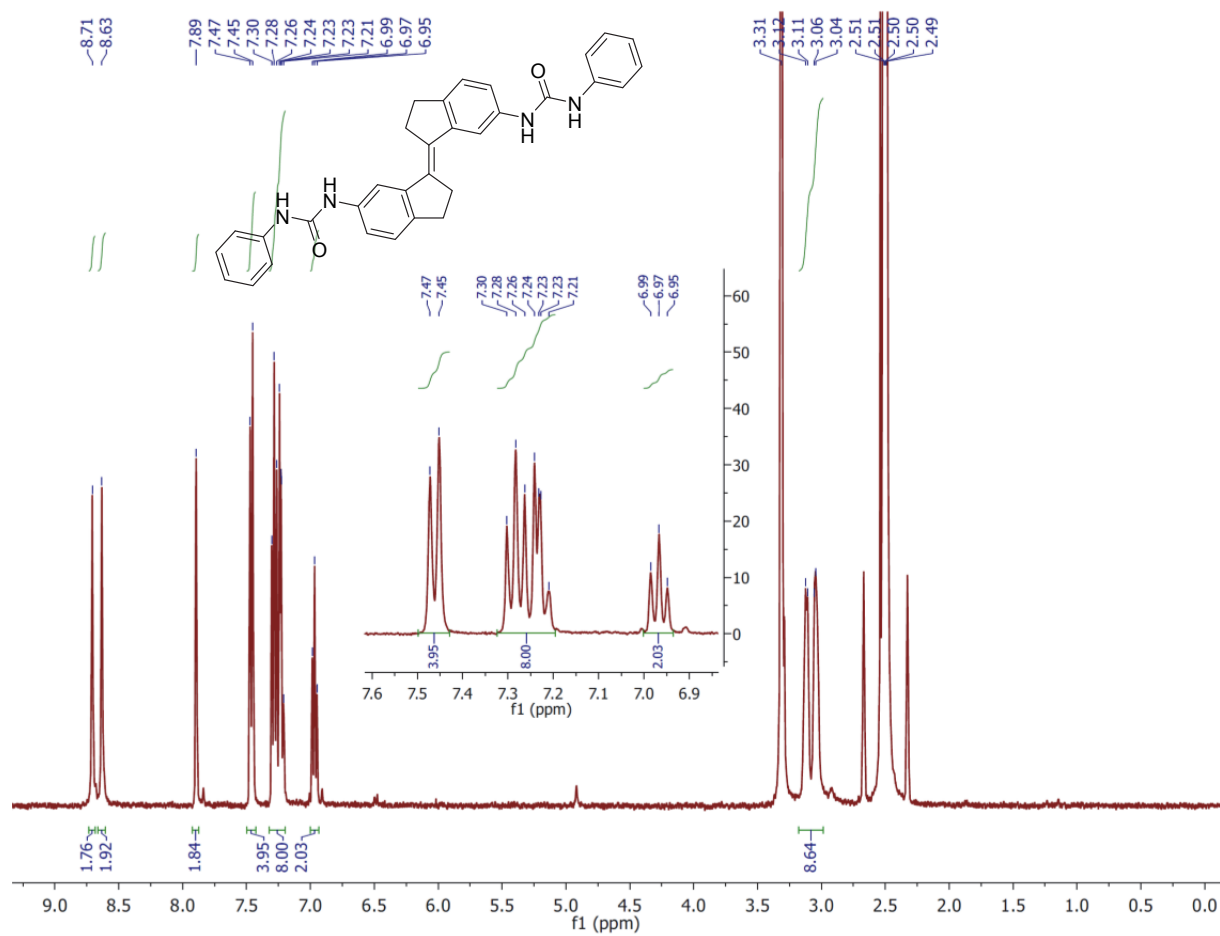


Figure S9. 400 MHz ^1H NMR spectrum of (*E*)-1 measured at 298 K in $\text{DMSO-}d_6$.

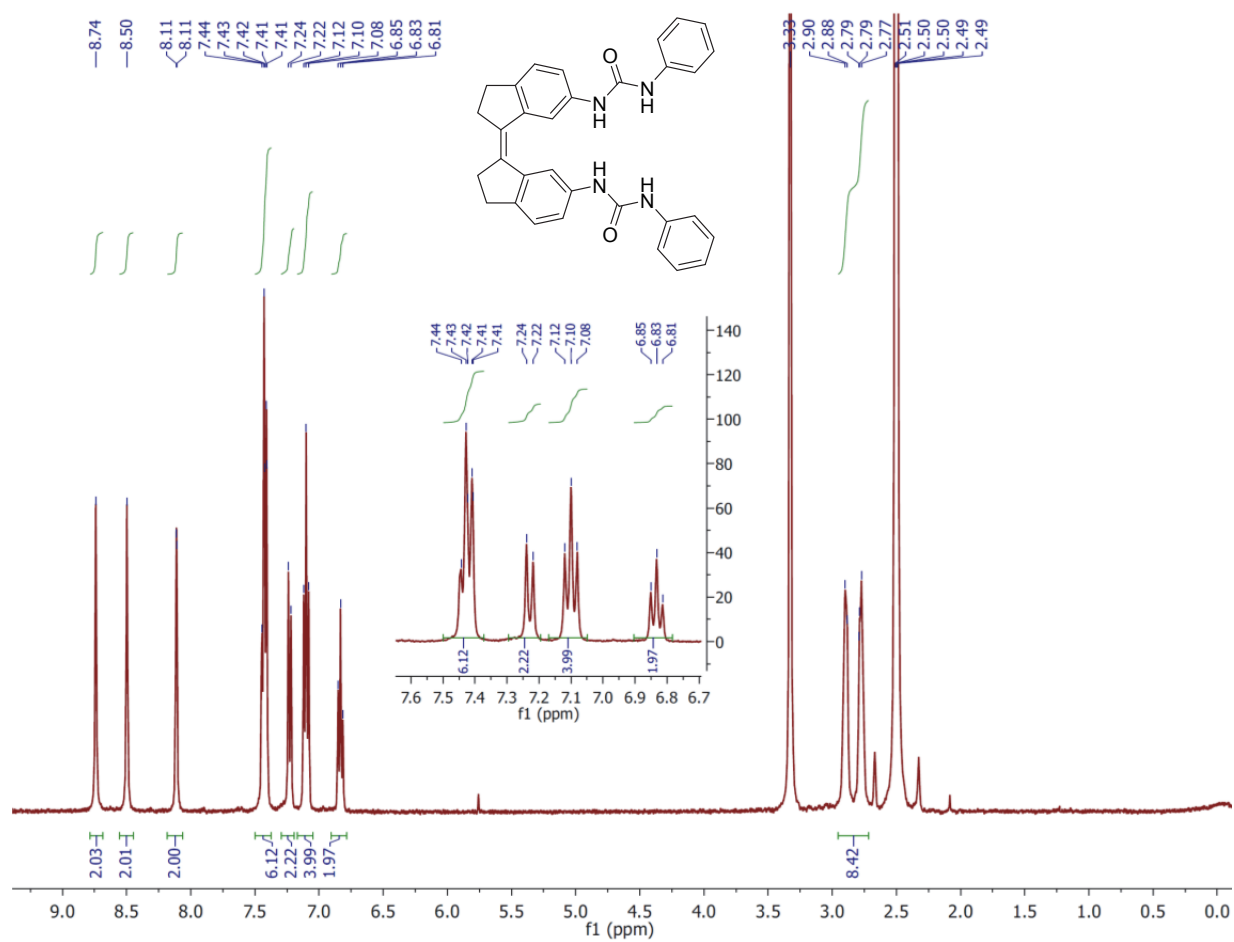


Figure S10. 400 MHz ¹H NMR spectrum of (Z)-1 measured at 298 K in DMSO-*d*₆.

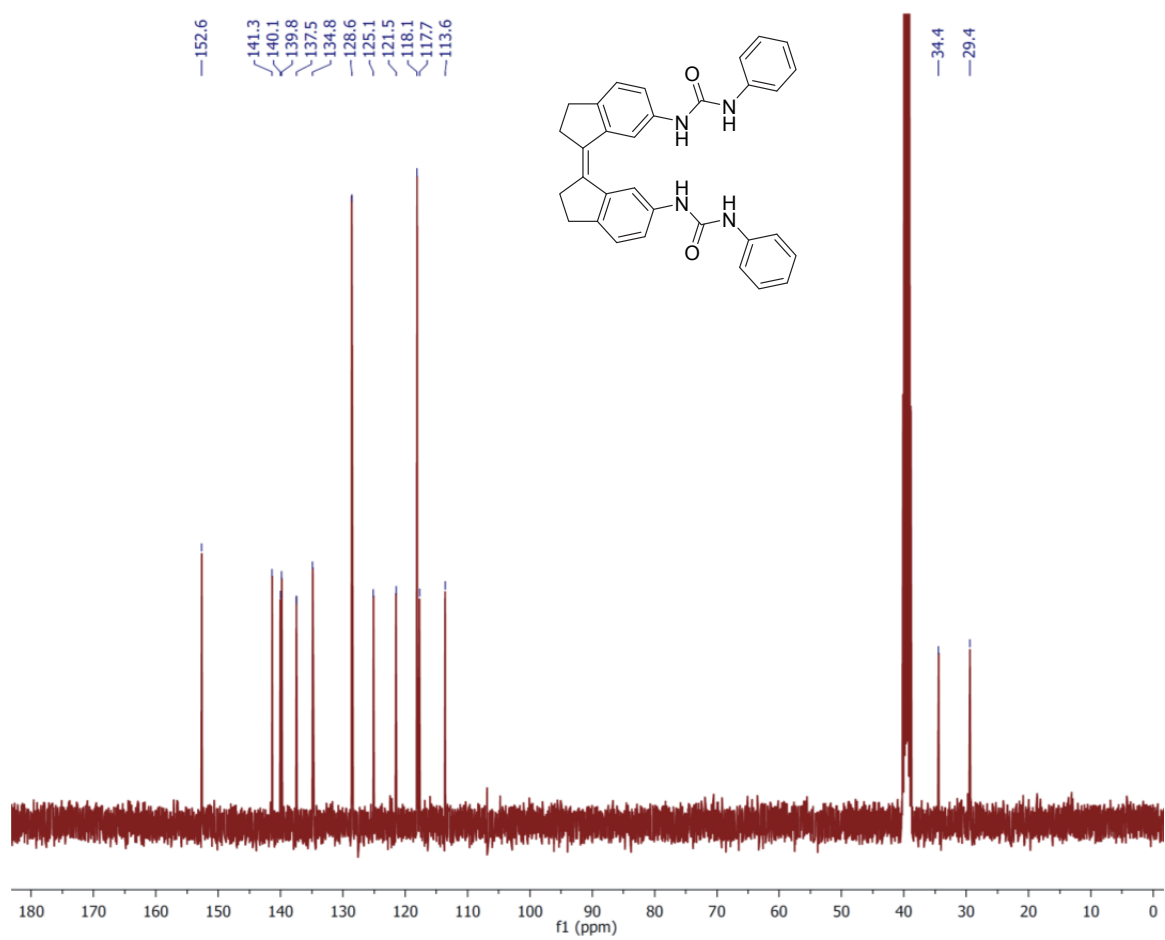


Figure S11. 100 MHz ¹³C NMR spectrum of (Z)-1 measured at 298 K in DMSO-*d*₆.

^1H NMR photoisomerization studies

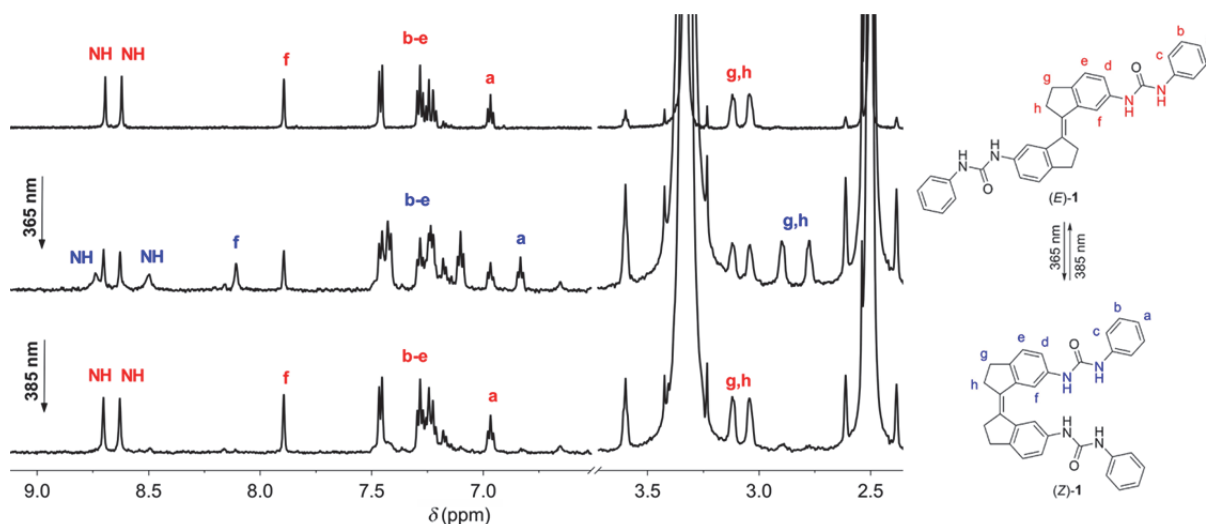


Figure S12. Aromatic and aliphatic region in the ^1H NMR spectrum (600 MHz, 293 K) of (*E*)-**1** in degassed $\text{DMSO-}d_6$ (2.5×10^{-4} M) (top) and spectral changes after irradiation with 365 nm light for 1 h (middle) followed by irradiation with 385 nm light for 1 h (bottom) at 20 °C. For determination of the PSS ratios, the integrals of the NH, H_a and H_f signals were averaged giving a PSS_{365} ratio (*E/Z*) of 49:51 and a PSS_{385} ratio (*E/Z*) of 93:7.

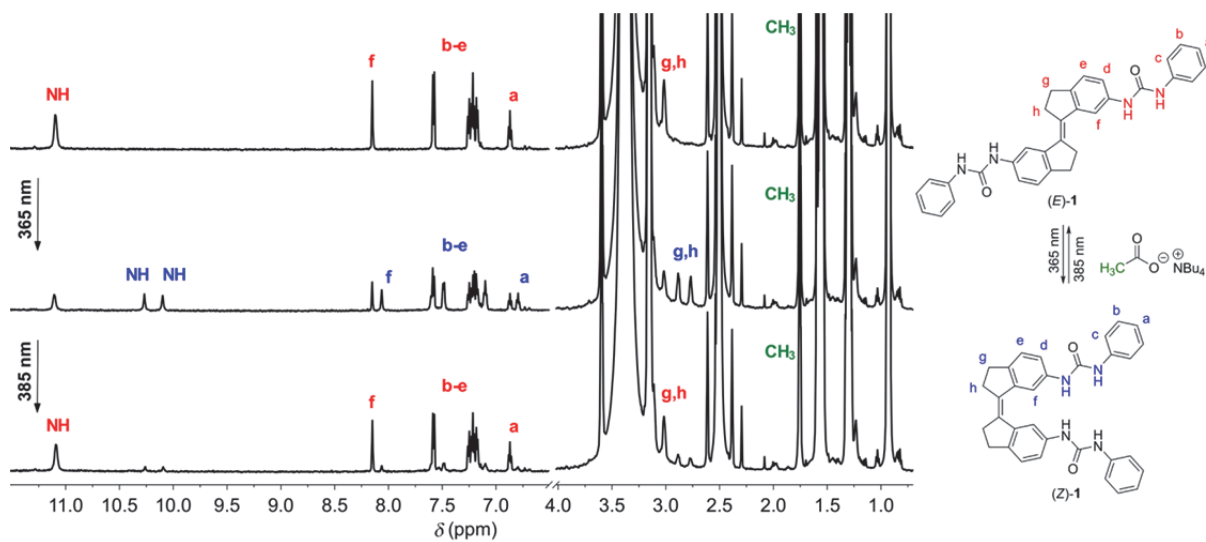


Figure S13. Aromatic and aliphatic region in the ^1H NMR spectrum (600 MHz, 293 K) of a mixture of (*E*)-**1** (2.5×10^{-4} M) and $[\text{Bu}_4\text{N}]^+[\text{CH}_3\text{CO}_2]^-$ (2.5×10^{-3} M) in degassed $\text{DMSO-}d_6/0.5\%$ H_2O (top) and spectral changes after irradiation with 365 nm light for 1 h (middle) followed by irradiation with 385 nm light for 1 h (bottom) at 20 °C. For determination of the PSS ratios, the integrals of the NH, H_a and H_f signals were averaged giving a PSS_{365} ratio (*E/Z*) of 48:52 and a PSS_{385} ratio (*E/Z*) of 88:12.

Quantum yield determination

The photon flux of the Thorlab model F365F1 high-power LED ($\lambda_{\max} = 365$ nm) was estimated by measuring the production of ferrous ions from potassium ferrioxalate.² The rate (r) of Fe^{2+} ion formation was: $8.81 \times 10^{-7} \text{ M s}^{-1}$. Hence, the moles of photons absorbed per time unit ($Nh\nu/t = \text{moles of Fe}^{2+}/\phi t$) in a 2 mL solution can be calculated using the reported quantum yield of ferrioxalate ($\phi = 1.21$),³ which gives: $1.46 \times 10^{-9} \text{ mol s}^{-1}$.

A solution of (*E*)-**1** in DMSO ($1 \times 10^{-4} \text{ M}$) in a 1 cm quartz cuvette was irradiated at 20 °C with the Thorlab model F365F1 high-power LED. The concentration was high enough to absorb all incident light ($\text{Abs}_{365} \geq 2$). The absorbance increase at $\lambda = 380$ nm was monitored over time by UV-vis spectroscopy and the molar absorptivities at this wavelength ($\epsilon_E = 2468.4 \text{ M}^{-1} \text{ cm}^{-1}$, $\epsilon_Z = 7365.5 \text{ M}^{-1} \text{ cm}^{-1}$) were used to calculate the concentration of (*Z*)-**1**. The slopes of the plots of the concentration increase versus time (Figure S12), at low conversion, represent the rate of formation (r). These were obtained by linear fitting to the equation $y = ax + b$ using Origin software. The photochemical quantum yield was obtained by comparison of the rate of formation of (*Z*)-**1** ($r = 3.64 \times 10^{-8} \text{ M s}^{-1}$) with the rate of Fe^{2+} formation from potassium ferrioxalate giving $\phi_{Z \rightarrow E} = 5.0 \pm 0.15\%$. The quantum yield for the ‘backward’ reaction is defined as: $\phi_{Z \rightarrow E} = \phi_{E \rightarrow Z} \epsilon_E n_E / \epsilon_Z n_Z$, where ϵ_E, ϵ_Z are the molar absorptivities at $\lambda = 365$ nm ($\epsilon_E = 27945 \text{ M}^{-1} \text{ cm}^{-1}$, $\epsilon_Z = 16834 \text{ M}^{-1} \text{ cm}^{-1}$) and n_E, n_Z are the amount of (*E*)-**1** and (*Z*)-**1** at the photostationary state ($\text{PSS}_{365} E/Z = 49:51$), giving $\phi_{Z \rightarrow E} = 8.0 \pm 0.23\%$.

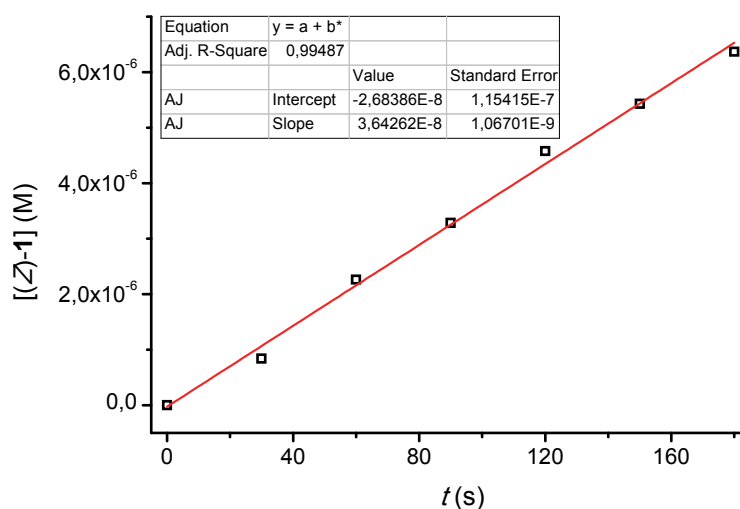


Figure S14. Concentration of (*Z*)-**1** as a function of time during $\lambda = 365$ nm irradiation of a $1 \times 10^{-4} \text{ M}$ solution (2 mL) obtained by measuring the absorbance increase at $\lambda = 380$ nm.

¹H NMR titration experiments

First, 5 mM solutions of receptors (*E*)-**1** and (*Z*)-**1** were prepared in DMSO-*d*₆/0.5%*H*₂O (v/v). The tetrabutylammonium anion was dissolved in the receptor solution and was present at a 50 mM concentration. The 50 mM anion solution was added stepwise to 0.5 mL of the 5 mM receptor solution and a ¹H NMR spectrum (400 MHz) was recorded after each addition.

Addition of [Bu₄N]⁺[Cl]⁻ to (*Z*)-**1**:

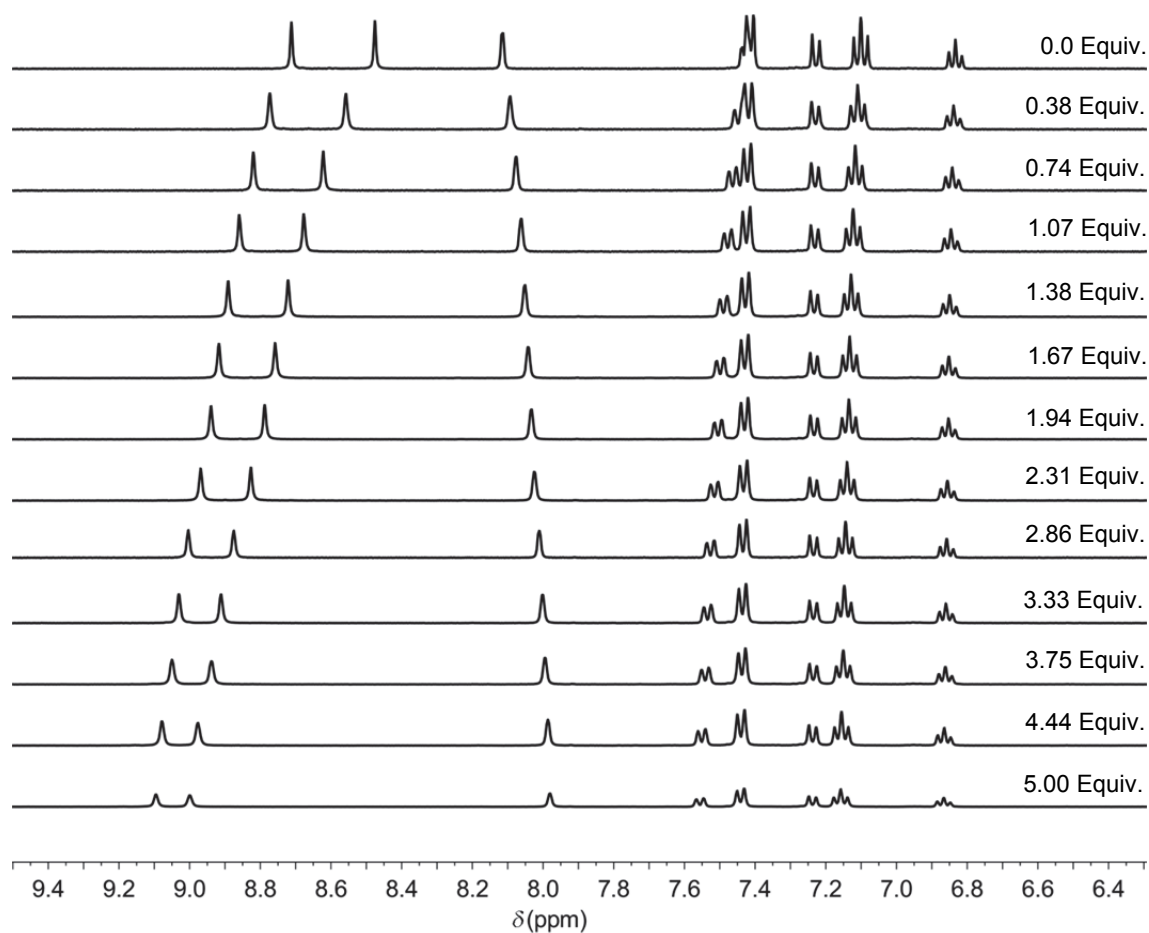


Figure S15. ¹H NMR spectral changes (400 MHz, 293 K) in the aromatic region of (*Z*)-**1** (5×10^{-3} M in DMSO-*d*₆/0.5%*H*₂O) upon the stepwise addition of [Bu₄N]⁺[Cl]⁻ (from top to bottom: 0.0, 0.38, 0.74, 1.07, 1.38, 1.67, 1.94, 2.31, 2.86, 3.33, 3.75, 4.44, 5.00 equivalents).

Addition of $[\text{Bu}_4\text{N}]^+[\text{Br}]^-$ to (Z)-1:

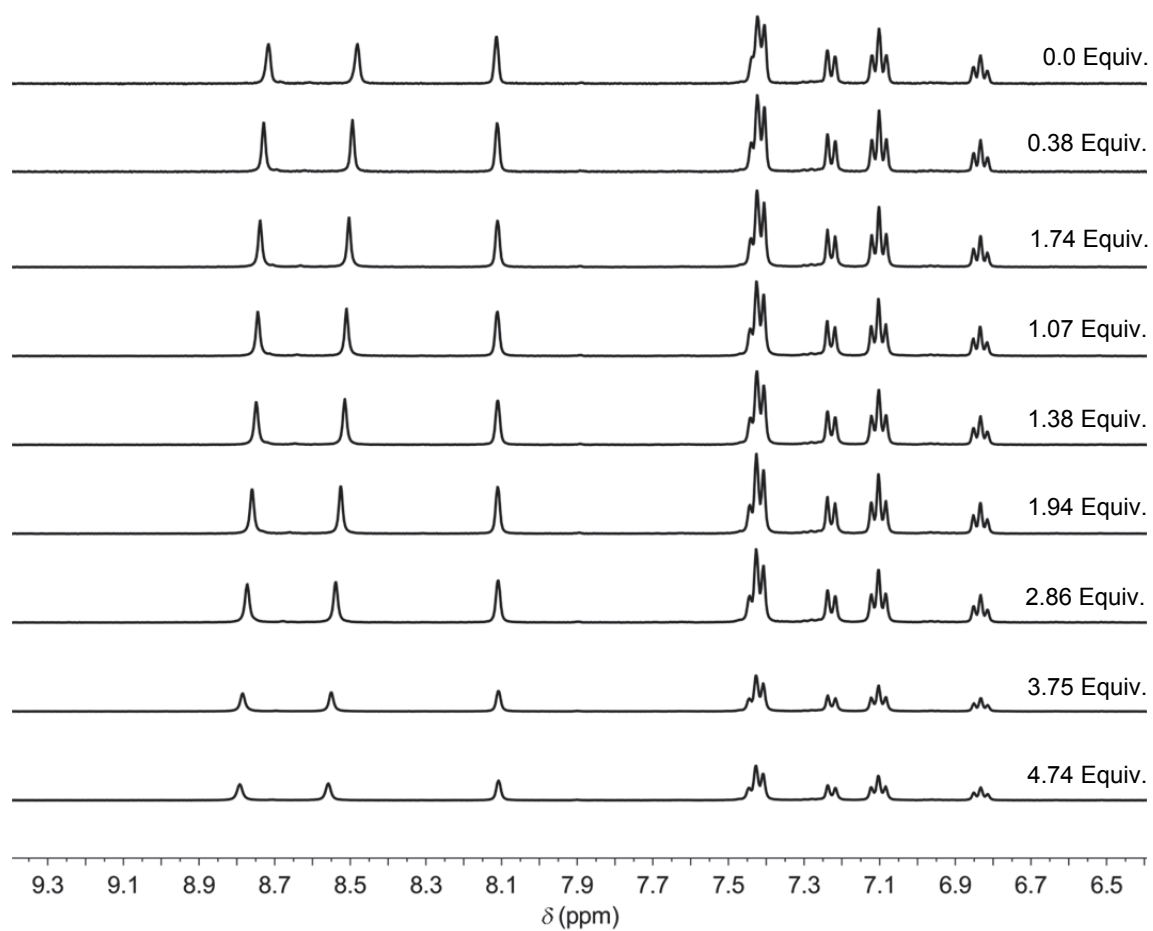


Figure S16. ^1H NMR spectral changes (400 MHz, 293 K) in the aromatic region of (Z)-1 (5×10^{-3} M in $\text{DMSO-}d_6/0.5\%\text{H}_2\text{O}$) upon the stepwise addition of $[\text{Bu}_4\text{N}]^+[\text{Br}]^-$ (from top to bottom: 0.0, 0.38, 0.74, 1.07, 1.38, 1.94, 2.86, 3.75, 4.74 equivalents).

Addition of $[\text{Bu}_4\text{N}]^+[\text{NO}_3]^-$ to (Z)-1:

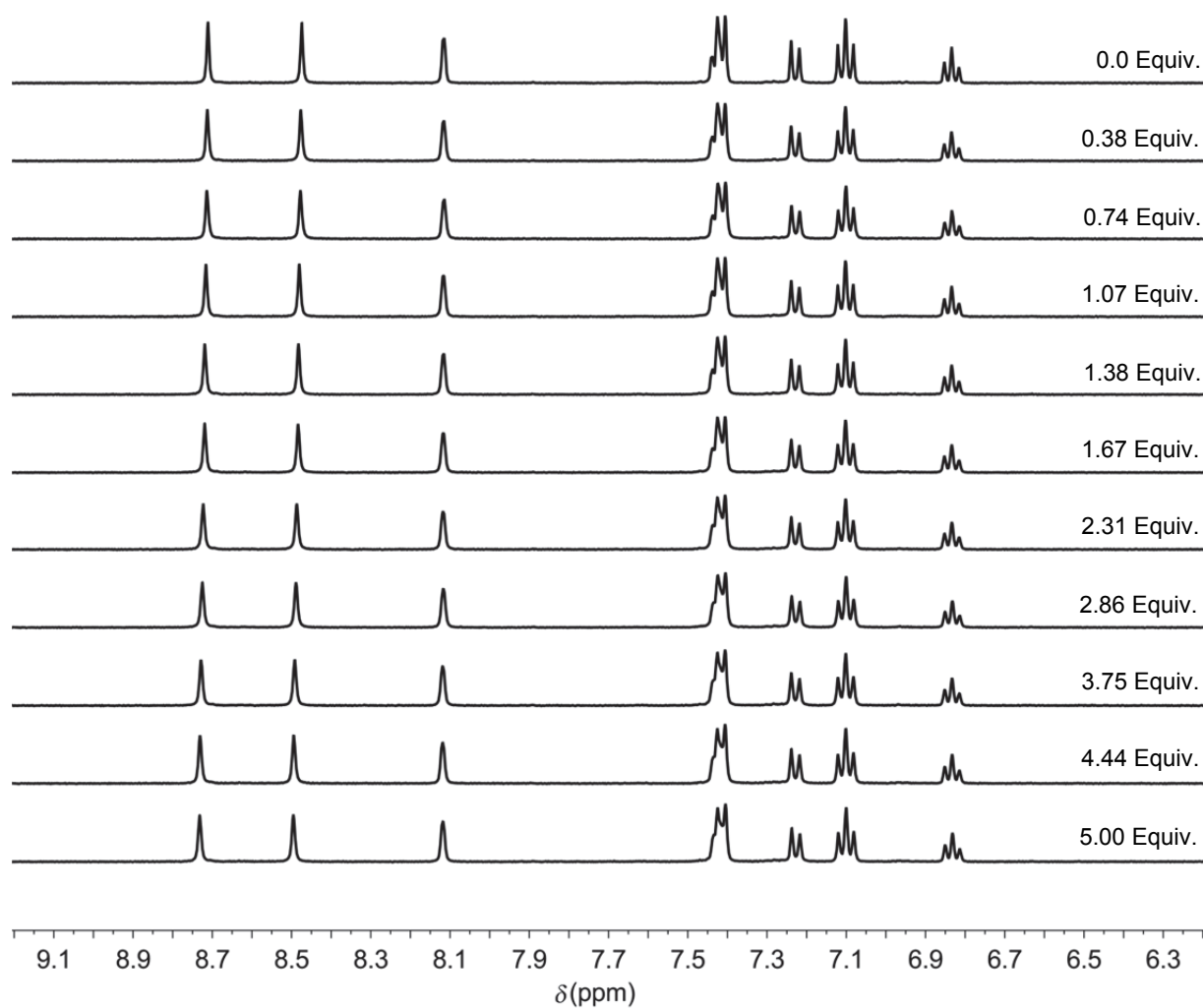


Figure S17. ^1H NMR spectral changes (400 MHz, 293 K) in the aromatic region of (Z)-1 (5×10^{-3} M in $\text{DMSO}-d_6/0.5\% \text{H}_2\text{O}$) upon the stepwise addition of $[\text{Bu}_4\text{N}]^+[\text{NO}_3]^-$ (from top to bottom: 0.0, 0.38, 0.74, 1.07, 1.38, 1.67, 2.31, 2.86, 3.75, 4.44, 5.00 equivalents).

Addition of $[\text{Bu}_4\text{N}]^+[\text{CH}_3\text{CO}_2]^-$ to (Z)-1:

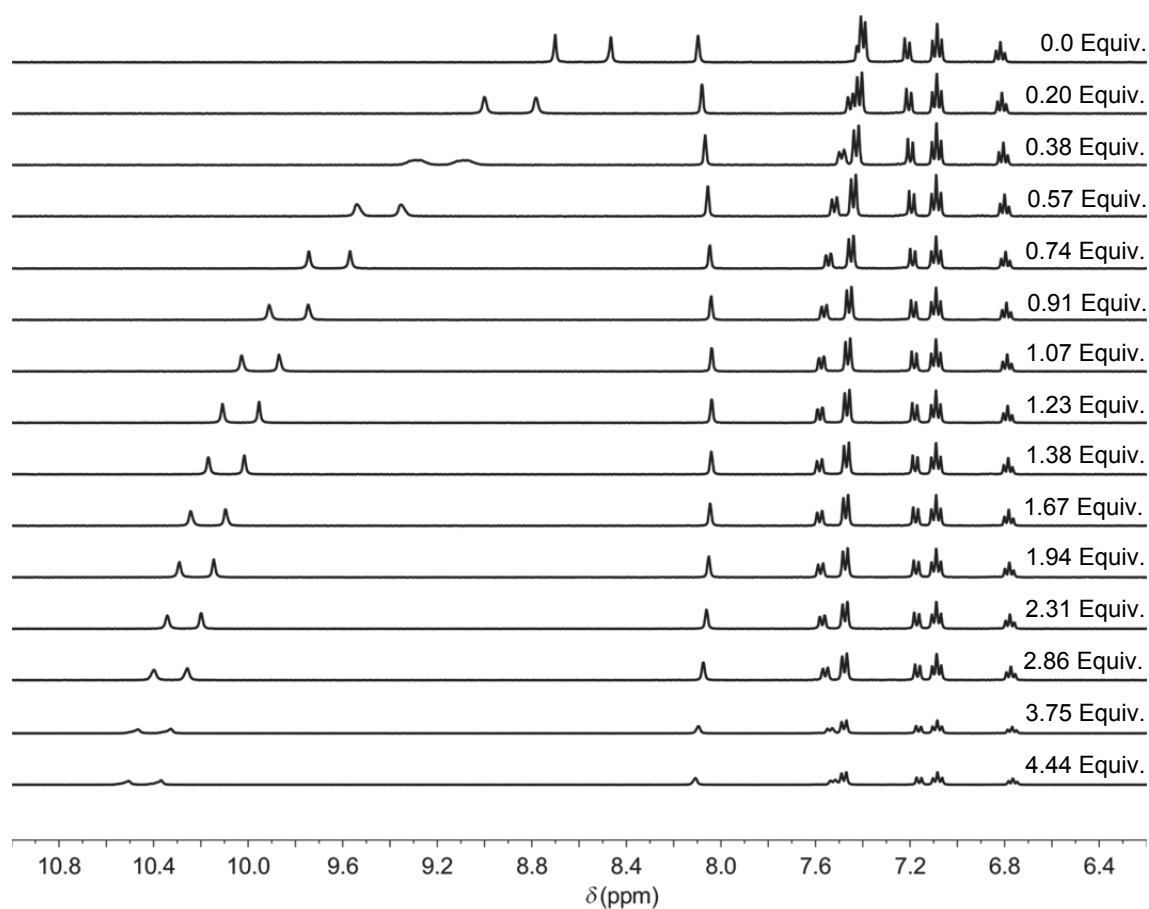


Figure S18. ^1H NMR spectral changes (400 MHz, 293 K) in the aromatic region of (Z)-1 (5×10^{-3} M in $\text{DMSO-}d_6/0.5\% \text{H}_2\text{O}$) upon the stepwise addition of $[\text{Bu}_4\text{N}]^+[\text{CH}_3\text{CO}_2]^-$ (from top to bottom: 0.0, 0.20, 0.38, 0.57, 0.74, 0.91, 1.07, 1.23, 1.38, 1.67, 1.94, 2.31, 2.86, 3.75, 4.44 equivalents).

Addition of $[\text{Bu}_4\text{N}]^+[\text{H}_2\text{PO}_4]^-$ to (Z)-1:

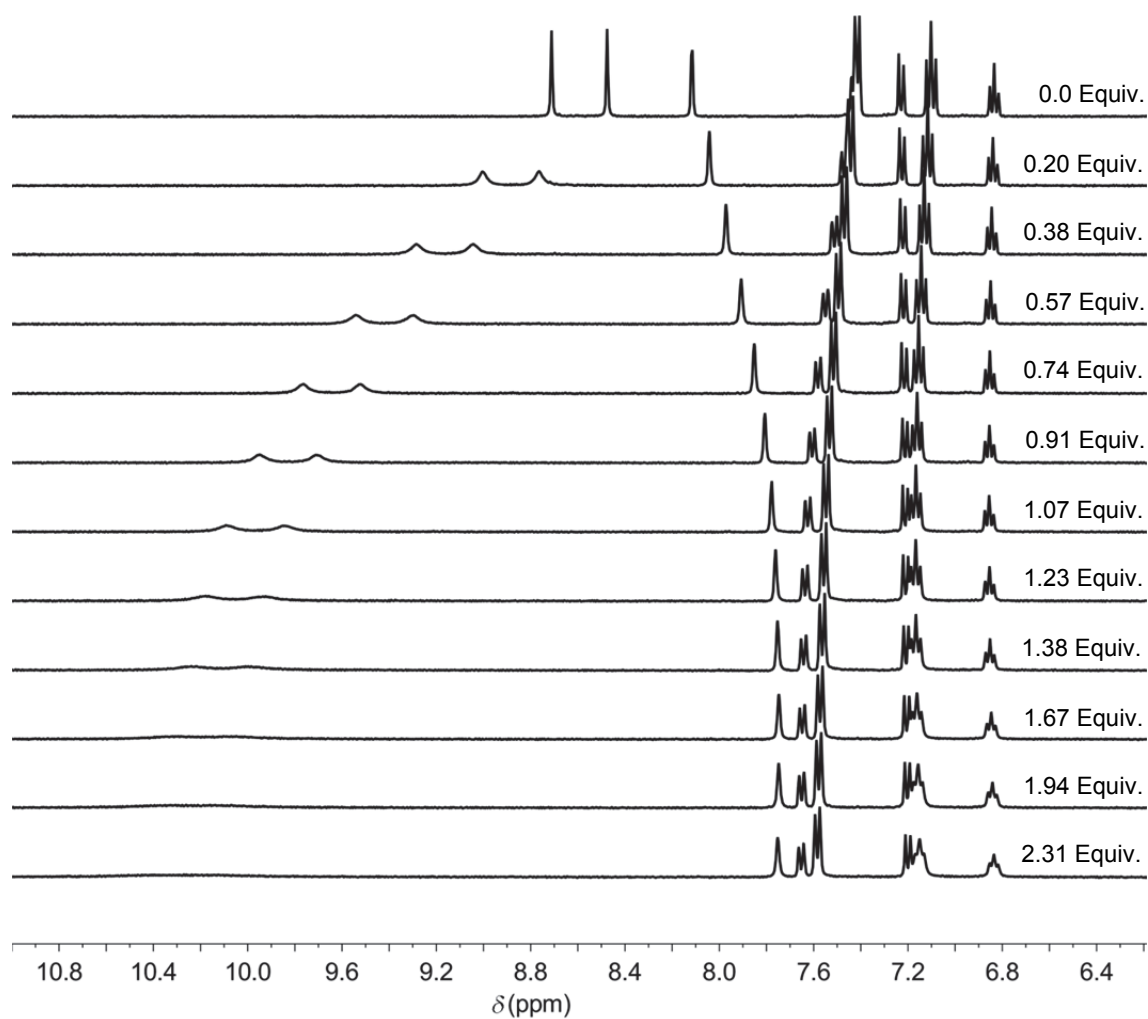


Figure S19. ^1H NMR spectral changes (400 MHz, 293 K) in the aromatic region of (Z)-1 (5×10^{-3} M in $\text{DMSO}-d_6/0.5\% \text{H}_2\text{O}$) upon the stepwise addition of $[\text{Bu}_4\text{N}]^+[\text{H}_2\text{PO}_4]^-$ (from top to bottom: 0.0, 0.20, 0.38, 0.57, 0.74, 0.91, 1.07, 1.23, 1.38, 1.67, 1.94, 2.31 equivalents).

Addition of $[\text{Bu}_4\text{N}]^+[\text{HSO}_4]^-$ to (Z)-1:

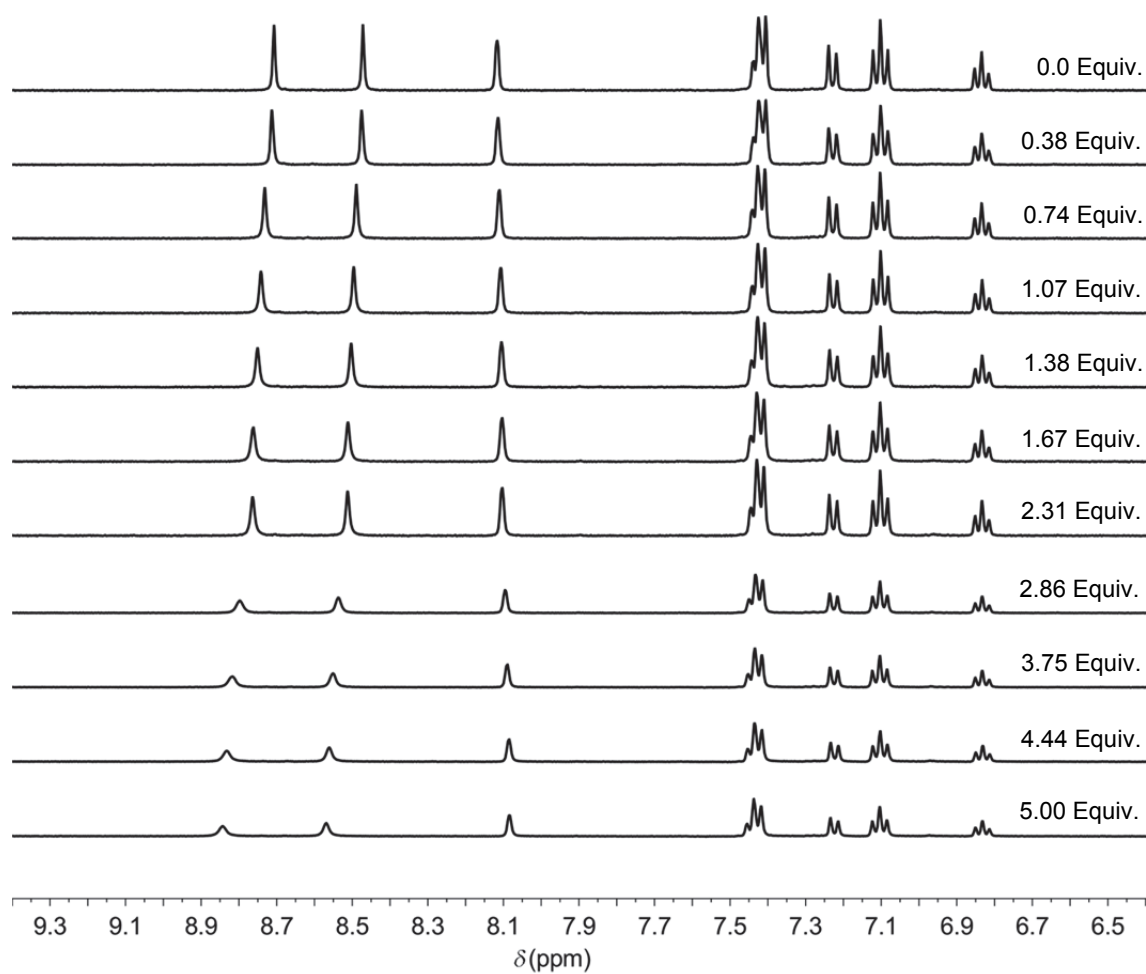


Figure S20. ^1H NMR spectral changes (400 MHz, 293 K) in the aromatic region of (Z)-1 (5×10^{-3} M in $\text{DMSO}-d_6/0.5\% \text{H}_2\text{O}$) upon the stepwise addition of $[\text{Bu}_4\text{N}]^+[\text{HSO}_4]^-$ (from top to bottom: 0.0, 0.38, 0.74, 1.07, 1.38, 1.67, 2.31, 2.68, 3.75, 4.44, 5.00 equivalents).

Addition of $[\text{Bu}_4\text{N}]^+[\text{Cl}]^-$ to (*E*)-1:

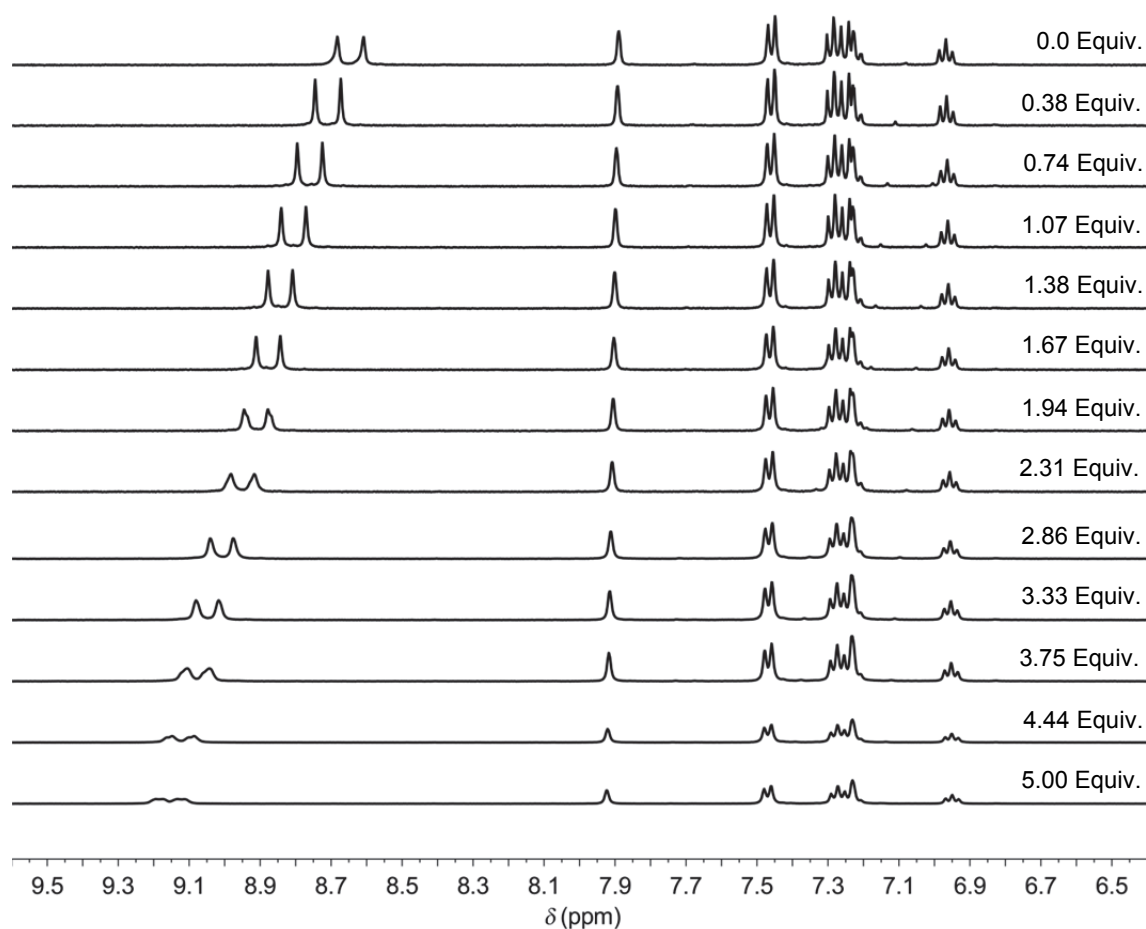


Figure S21. ^1H NMR spectral changes (400 MHz, 293 K) in the aromatic region of (*E*)-1 (5×10^{-3} M in $\text{DMSO}-d_6/0.5\% \text{H}_2\text{O}$) upon the stepwise addition of $[\text{Bu}_4\text{N}]^+[\text{Cl}]^-$ (from top to bottom: 0.0, 0.38, 0.74, 1.07, 1.38, 1.67, 1.94, 2.31, 2.86, 3.33, 3.75, 4.44, 5.00 equivalents).

Addition of $[\text{Bu}_4\text{N}]^+[\text{CH}_3\text{CO}_2]^-$ to (*E*)-1:

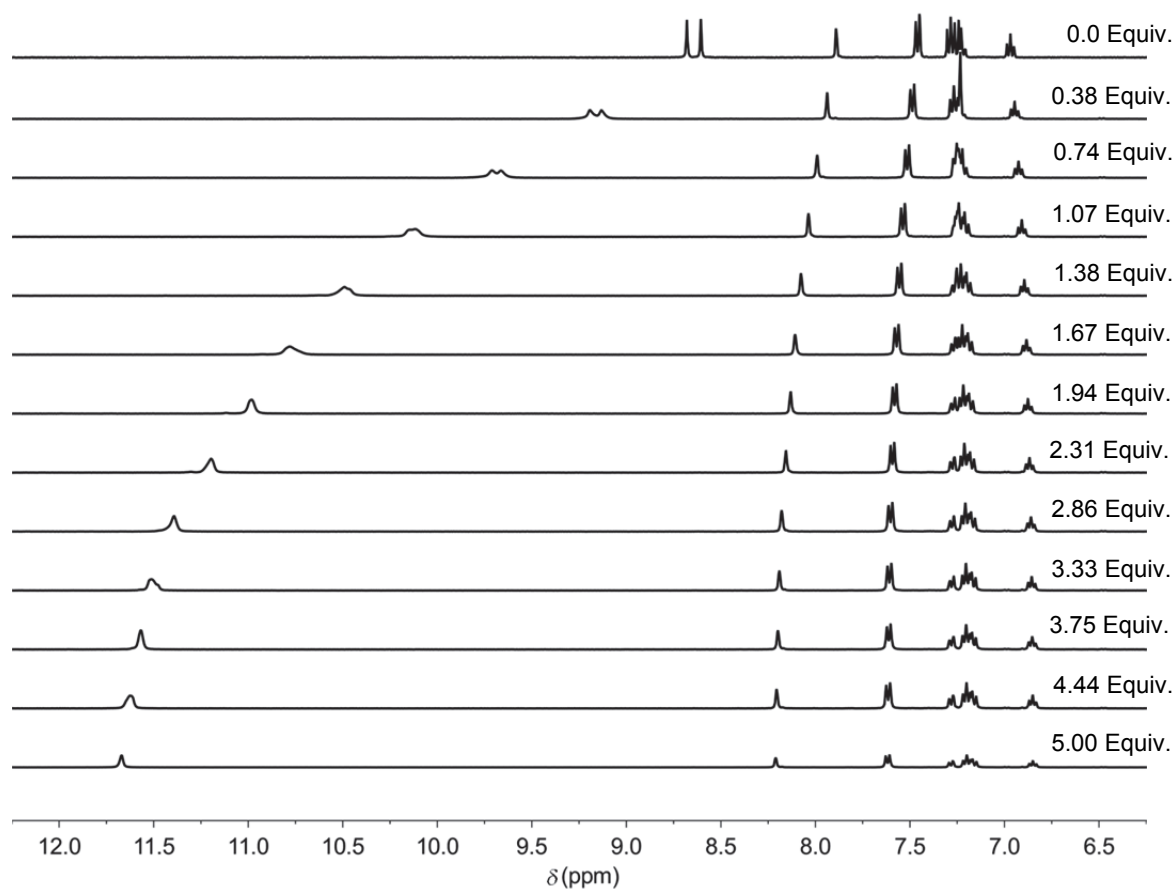


Figure S22. ^1H NMR spectral changes (400 MHz, 293 K) in the aromatic region of (*E*)-1 (5×10^{-3} M in $\text{DMSO}-d_6/0.5\% \text{H}_2\text{O}$) upon the stepwise addition of $[\text{Bu}_4\text{N}]^+[\text{CH}_3\text{CO}_2]^-$ (from top to bottom: 0.0, 0.38, 0.74, 1.07, 1.38, 1.67, 1.94, 2.31, 2.86, 3.33, 3.75, 4.44, 5.00 equivalents).

Addition of $[\text{Bu}_4\text{N}]^+[\text{H}_2\text{PO}_4]^-$ to (*E*)-1:

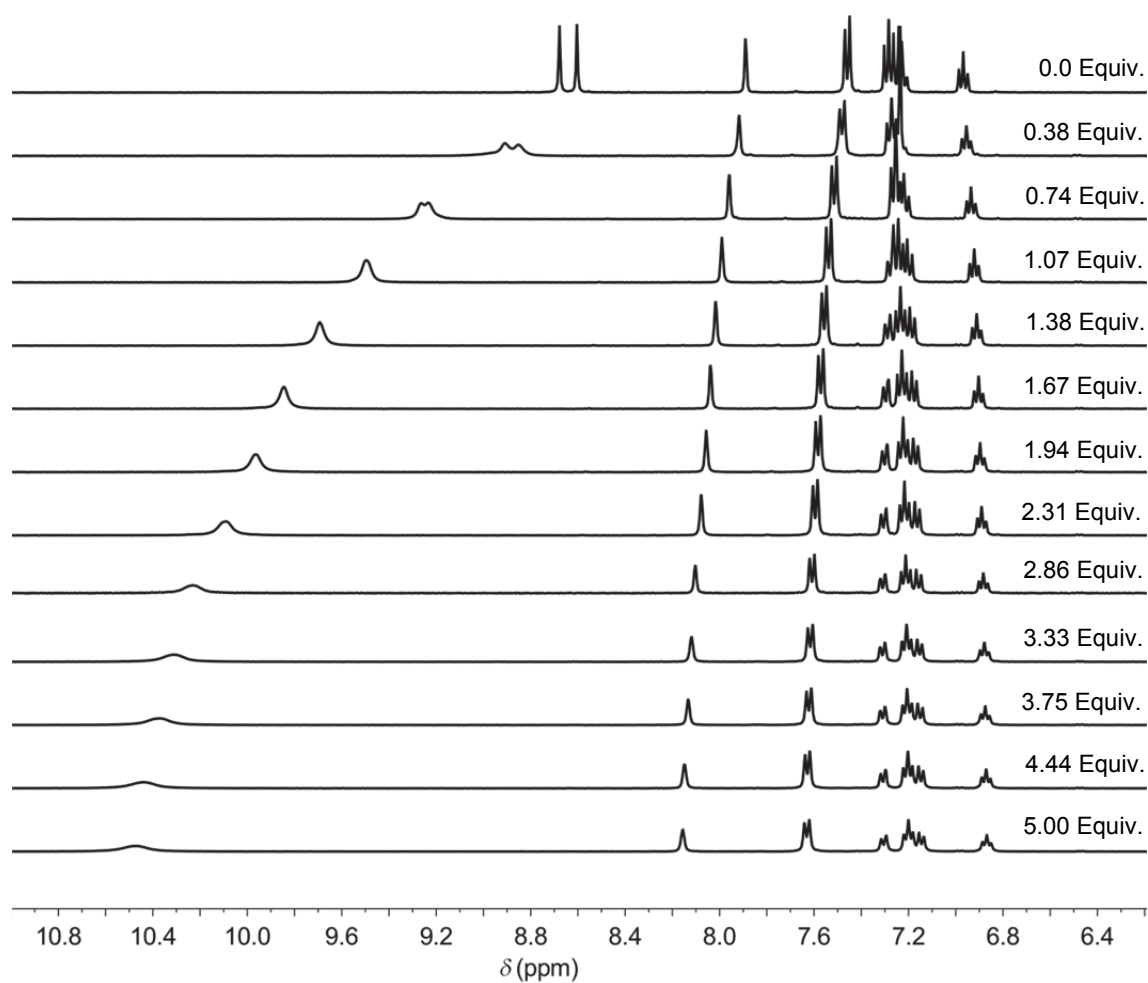
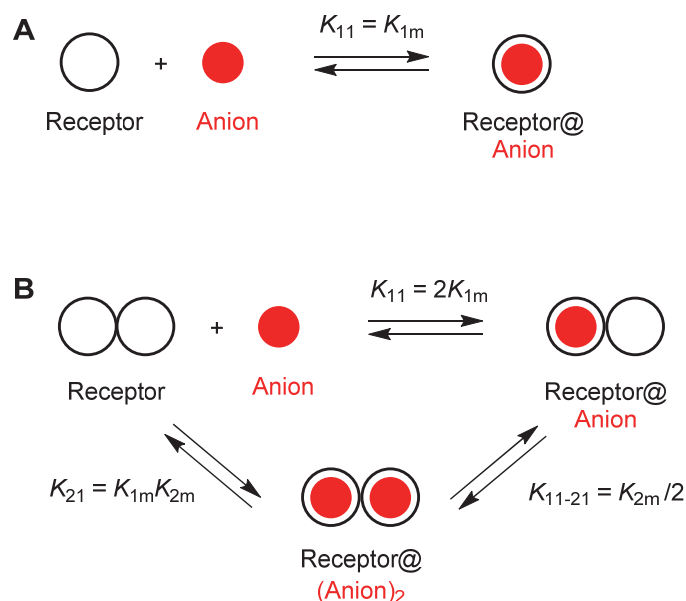


Figure S23. ^1H NMR spectral changes (400 MHz, 293 K) in the aromatic region of (*E*)-1 (5×10^{-3} M in $\text{DMSO}-d_6/0.5\% \text{H}_2\text{O}$) upon the stepwise addition of $[\text{Bu}_4\text{N}]^+[\text{H}_2\text{PO}_4]^-$ (from top to bottom: 0.0, 0.38, 0.74, 1.07, 1.38, 1.67, 1.94, 2.31, 2.86, 3.33, 3.75, 4.44, 5.00 equivalents).

Job plot and titration curve analysis



Scheme S1. Schematic representation of the species and equilibria involved in the formation of a 1:1 anion/receptor complex (A) and the stepwise formation of a 2:1 anion/receptor complex (B). The stability constant of the 1:1 complex (K_{11}), 2:1 complex (K_{21}) and the stepwise constant (K_{11-21}) are shown and related to the microscopic binding constants (K_{1m} , K_{2m}). The data were fitted using HypNMR software.⁴

It should be noted that no clear distinction can be made between the ¹H NMR signals of the 1:1 and 2:1 complexes throughout the titration. In the data analysis using a 2:1 model, both urea binding sites were therefore treated as equal, i.e. $K_{1m} = K_{2m}$ (cooperativity factor $\alpha = 1$). Furthermore, the data was initially fitted to a 1:1 model to afford K_{11} , after which the stability constant of the 2:1 complex [$K_{21} = (K_{11}/2)^2$] was included.

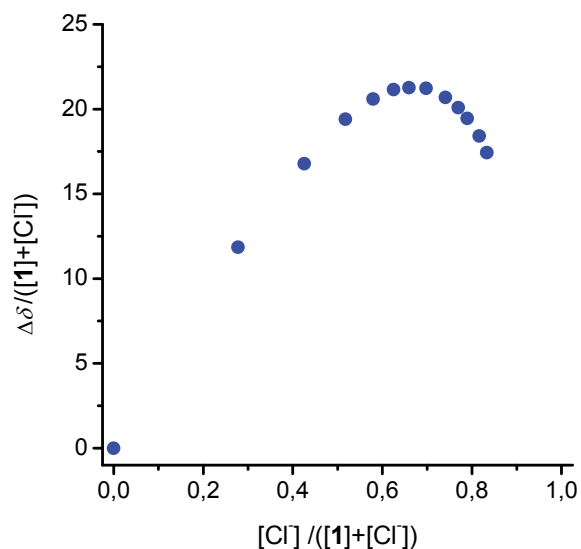


Figure S24. Job plot analysis generated from the addition of $[Bu_4N]^+[Cl]^-$ to (Z)-1 indicating a 2:1 binding stoichiometry.

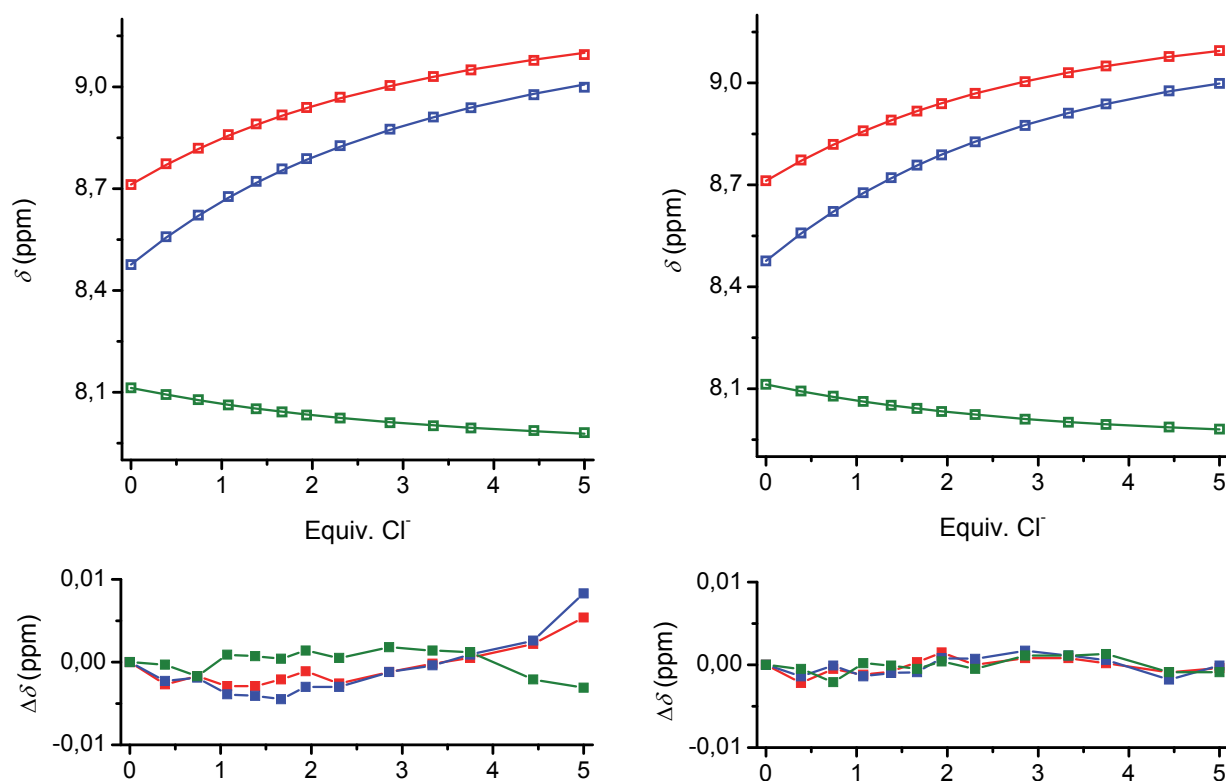


Figure S25. Titration curves for the addition of $[Bu_4N]^+[Cl]^-$ to (Z)-1 and data fitting obtained by simultaneous analysis of the urea-NH (red, blue) and arom-H (green) signals using either a 1:1 (left) or a 2:1 (right) binding model. Based on the residuals analysis, the latter binding model is more likely to be correct: $K_{1m} = 66 \text{ M}^{-1}$ (for 1:1); $K_{1m} = 33 \text{ M}^{-1}$ (for 2:1).

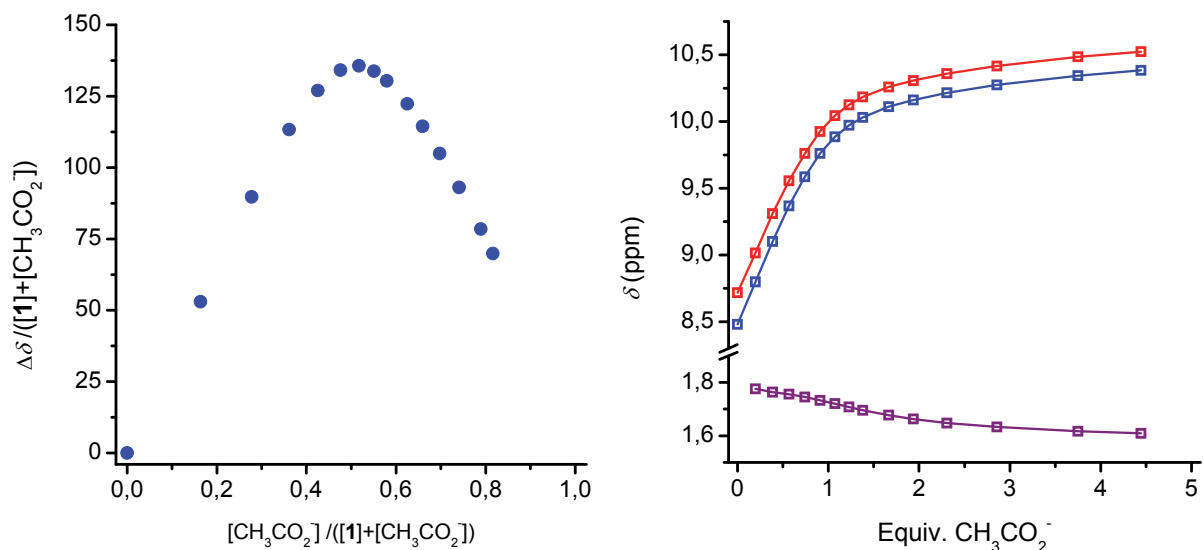


Figure S26. (left) Job plot analysis generated from the addition of $[\text{Bu}_4\text{N}]^+[\text{CH}_3\text{CO}_2]^-$ to (Z)-1 indicating 1:1 binding. (right) Titration curves and data fitting to a 1:1 model obtained by analysis of the urea-NH (red, blue) and CH_3CO_2^- (purple) signals: $K_{1m} = 1.40 \times 10^3 \text{ M}^{-1}$.

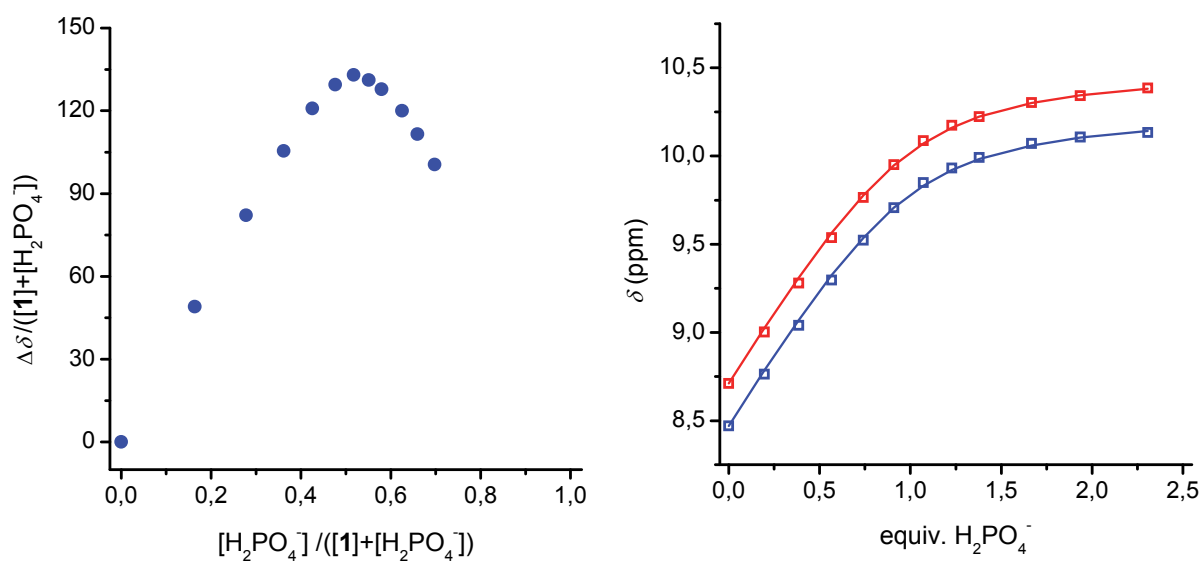


Figure S27. (left) Job plot analysis generated from the addition of $[\text{Bu}_4\text{N}]^+[\text{H}_2\text{PO}_4]^-$ to (Z)-1 indicating 1:1 binding. (right) Titration curves and data fitting to a 1:1 model obtained by analysis of the urea-NH (red, blue) signals: $K_{1m} = 2.02 \times 10^3 \text{ M}^{-1}$.

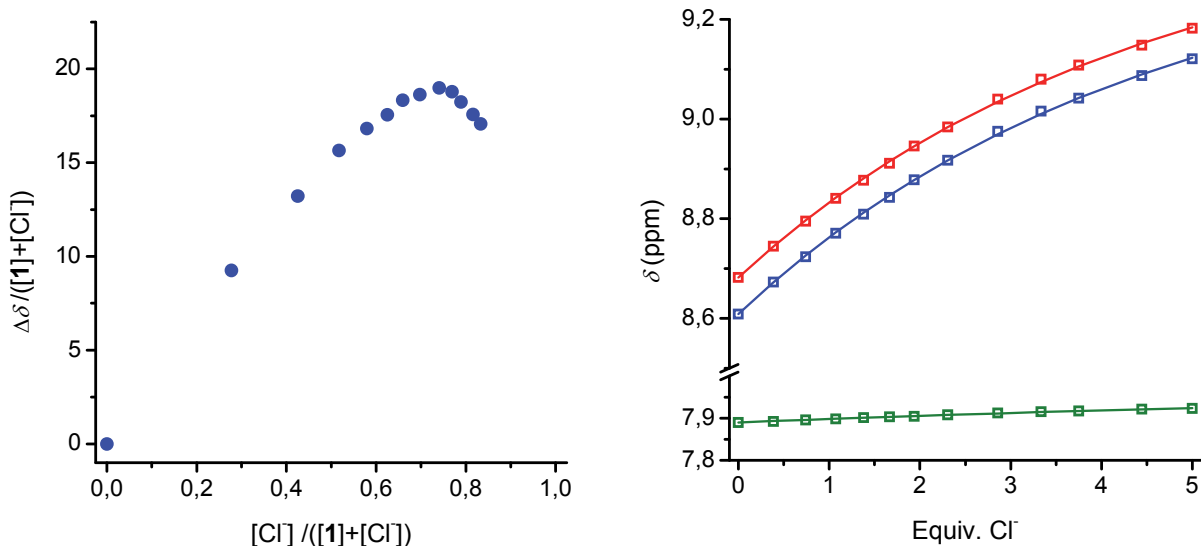


Figure S28. (left) Job plot analysis generated from the addition of $[Bu_4N]^+[Cl]^-$ to (*E*)-**1** indicating 2:1 binding. (right) Titration curves and data fitting to a 2:1 model obtained by simultaneous analysis of the urea-NH (red, blue) and arom-H (green) signals: $K_{1m} = 17 \text{ M}^{-1}$.

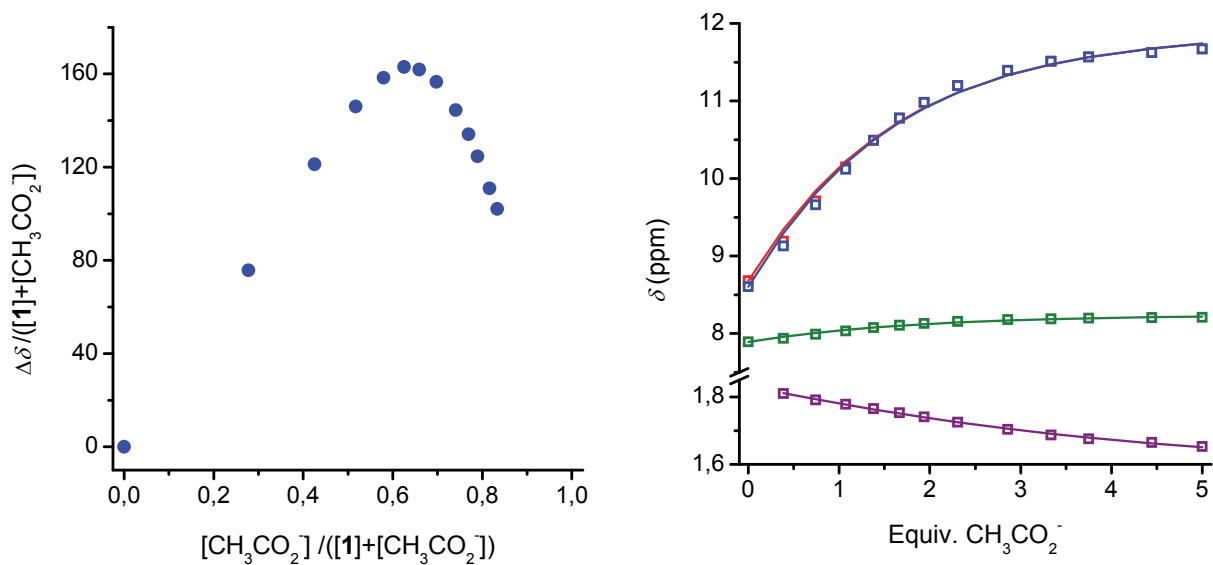


Figure S29. (left) Job plot analysis generated from the addition of $[Bu_4N]^+[CH_3CO_2]^-$ to (*E*)-**1** indicating 2:1 binding. (right) Titration curves and data fitting to a 2:1 model obtained by simultaneous analysis of the urea-NH (red, blue), arom-H (green) and $CH_3CO_2^-$ (purple) signals: $K_{1m} = 1.04 \times 10^2 \text{ M}^{-1}$.

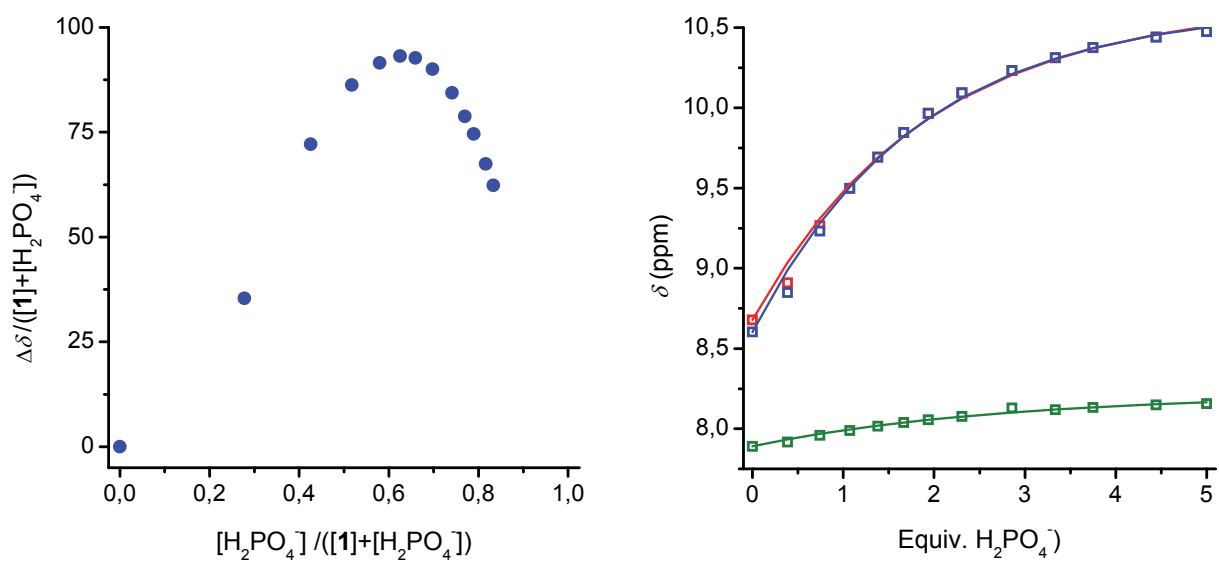
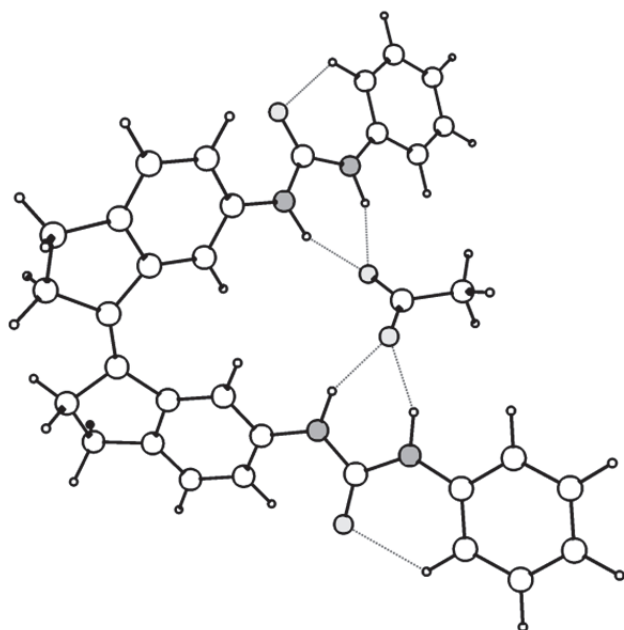


Figure S30. (left) Job plot analysis generated from the addition of $[Bu_4N]^+[H_2PO_4]^-$ to (*E*)-**1** indicating 2:1 binding. (right) Titration curves and data fitting to a 2:1 model obtained by simultaneous analysis of the urea-NH (red, blue) and arom-H (green) signals: $K_{1m} = 77 \text{ M}^{-1}$.

Geometry optimizations by DFT

The Gaussian 09 program⁵ was used for geometry optimizations and the calculation of energies. Initially, different input geometries were optimized at the semi-empirical PM6 level to find the global minima. Further geometry optimizations were performed at the DFT B3LYP/6-31G++(d,p) level using tight convergence criteria and using an IEFPCM DMSO solvation model. The DFT geometries were found to have zero imaginary frequencies.

Table S1. Cartesian coordinates of (Z)-**1**⊃CH₃CO₂⁻.



symmetry c1

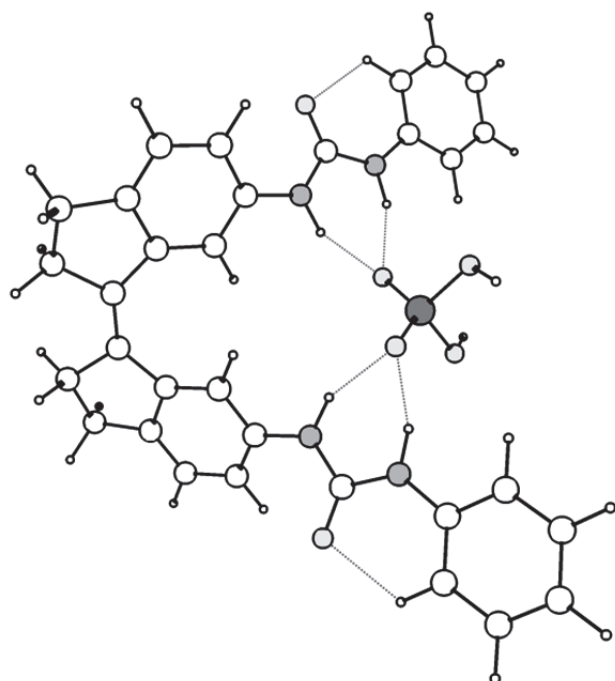
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C	-1.596919000	2.212073000	-0.365030000

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Sum of electronic and zero-point Energies= -1834.007803

Table S2. Cartesian coordinates of (Z)-**1**-H₂PO₄⁻.



symmetry c1

atom	X	Y	Z
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Sum of electronic and zero-point Energies= -2249.117127

References

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