Identification of serum protein biomarkers for utrophin based DMD therapy

Simon Guiraud¹*, Benjamin Edwards¹, Sarah E. Squire¹, Arran Babbs¹, Nandini Shah¹, Adam Berg¹, Huijia Chen¹, Kay E. Davies¹*

¹ Medical Research Council Functional Genomics Unit at the University of Oxford, Department of Physiology, Anatomy and Genetics, Oxford OX1 3PT, United Kingdom

*Co-correspondence

Address: Medical Research Council Functional Genomics Unit, Department of Physiology, Anatomy and Genetics, University of Oxford, Oxford OX1 3PT, United Kingdom Telephone: +44 (0)1865 285880

Emails: simon.guiraud@dpag.ox.ac.uk and kay.davies@dpag.ox.ac.uk

Supplemental data - Legends

Figure S1: Utrophin increased rescue *mdx* phenotype. 7 week old C57, *mdx* and Fiona (n=7) were sacrificed and serum and muscles collected (A) H&E-stained transverse muscle sections of TA muscle in *mdx* mice showing necrotic areas and regenerating fibres compared to wild-type tissue. In Fiona mice, these pathological features are fully rescued due to high level of utrophin protein expression at the sarcolemma. Scale bar: 100 μ m. (B) Western blots for utrophin protein showing a 2 fold increase in mdx related to C57 and a 4 fold increase in Fiona mice compared to *mdx*. (C) Quantification of centronucleation necrotic muscle area in EDL showing increased in *mdx* and full rescue in Fiona mice compared to the wild-type animals. (D) Necrosis quantification, increased in *mdx* compared to C57 and rescue to normal level in Fiona animals. (E). The difference in force produced between the first and fifth stretch is represented as a sensitive indicator of the resistance of the muscle to stretch-induced damage. EDL muscles were stretched at 15% of their fibre length whilst contracting tetanically. Values are mean \pm SEM of n=7 per group; (*) p<0.05, (**) p<0.01, (***) p<0.001.

Figure S2: Quality control of SOMAscan data. (A) Table of all hybridization normalization and median normalization. (B) Hybridization normalization scale per experimental groups. (C) Median normalization scale factors per dilutions and groups

Table S1: Serum *mdx* **protein biomarkers identified by SOMAscan.** High significant increased and decreased protein were defined using Mann-Witney U test (p<0.005; q<0.01).

Table S2: Serum *mdx* protein biomarkers and rescue in *mdx* transgenic Fiona mice. High significant increased and decreased protein were defined using Kruskal-Wallis one-way ANOVA (p<0.005; q<0.01). (+++) restored to wild-type levels (recovery score >70%), (++) restored towards

wild-type levels (50< recovery score <70, (+) low and inconsistent restoration towards wild-type (recover score <50, (-) not restored towards wild-type levels.

 Table S3: Details of the ELISA assays used in the study. All the ELISA kits were purchased from

 Antibodies-online.

Supplemental data – Methods

Histological analyses

Muscle samples were frozen in liquid nitrogen-chilled isopentane, and stored at -80°C. Transverse tibialis anterior (TA), were stained with Haematoxylin and Eosin solution (H&E). The Axioplan 2 Microscope System (Carl Zeiss, Germany) was used to obtain the images. The proportion of centrally nucleated fibres was determined by analysing the H&E images. Areas of necrosis were quantified based on the DMD_M.1.2.007 MDC1A_M.1.2.004 TREAT-NMD SOPS and performed with the Fiji ImageJ 1.49i software [1] on the whole TA sections.

Immunofluorescence

Frozen transverse TA muscle sections were blocked in 10% foetal bovine serum/PBS for 20 minutes, incubated with primary antibodies overnight at 4°C, washed in PBS and incubated with suitable Alexa Fluor secondary antibodies for 1 hour at room temperature. Sections were examined under an Axioplan 2 Microscope System (Carl Zeiss, Germany). The following antibodies and dilutions were used: goat monoclonal anti-utrophin (1:2000, UtroA), mouse monoclonal anti-Laminin- α 2 (1:500, sc-59854, Santa Cruz Biotechnology, Inc.).

Protein analyses

Western blots were performed according to standard procedures. Briefly, muscles were homogenized in RIPA buffer (R0278, Sigma-Aldrich) supplemented with protease inhibitors (1:100; P8340, Sigma-Aldrich). 30 µg of total protein were heat-denatured for 5 minutes at 100°C before loading onto NuPAGE 3– 8% TRIS Acetate Midi Gel (Novex, Life Technologies) and transferred to PVDF membranes (Millipore). Membranes were blocked for 1 hour with 10% skimmed milk in 0.1% PBS-Tween20 and then incubated with primary antibodies in 0.1% PBS-T for 1 hour at room temperature. Primary antibodies used were: mouse monoclonal anti-utrophin (1:50, MANCHO3(84A), gift from G.E. Morris) and goat polyclonal α-actinin Antibody (N-19) (1:30000, sc-7453, Santa Cruz). HRPlabelled secondary antibodies (Amersham) were incubated for 1 hour at room temperature. Immunoreactive bands were detected using ECL Western Blot Detection Reagents (SuperSignal West Pico Chemiluminescent Substrate, Thermo Scientific). The relative expression of the target proteins was estimated by densitometry using Actinin N-19 as references on a Xograph Compact X4 developer and quantified with the Fiji ImageJ 1.49i software.

Isolated muscle function analysis

Peak force, specific force, and force drop were measured from the extensor digitorum longus muscle of the treated and control mice. During dissection and experiments, muscles were bathed in oxygenated (95% O₂–5% CO₂) Krebs–Hensley solution composed of (mmol/l): NaCl, 118; NaHCO₃, 24.8, KCl, 4.75; KH₂PO₄, 1.18; MgSO₄, 1.18; CaCl₂, 2.54; glucose, 10 [2]. Contractile properties were measured as previously described [3]. In brief, isolated EDL were attached to a lever arm connected to a force transducer (model 300B); and stimulator (model 701B); the equipment was controlled using the signal interface (model 604A) and results were recorded by the DMC software (version 4.1.4; Aurora Scientific, Aurora, Ontario, Canada). The muscle was stimulated by single pulses of 0.2 milliseconds at 30 V while the optimum length (L_0) was determined. Optimum fibre length ($L_{\rm f}$) was calculated by multiplying $L_{\rm o}$ by the predetermined fibre length to muscle length ratio of 0.44 [4]. A force-frequency curve was generated and the maximum isometric force calculated. Absolute force (P_0) are normalized to specific force (sP_0 ; kN/m^2) using the equation (muscle mass/L_f x1.06 (the density of mammalian muscle). Percentage force drop were calculated by comparing maximum force between the first (ECC0) and fifth eccentric (ECC5) contractions, expressed as a percentage of ECC0. The muscle was stimulated into tetanus at the frequency required to generate the P_{0} , while in tetanic state the muscle was stretched at a rate of one fibre length per second for 0.15 seconds, equating to a total stretch of 15% of fibre length. All data were digitized and analysed using the DMA software (version 3.2; Aurora Scientific).

Supplemental data – Figures/Tables



Figure S1: Utrophin increased rescue *mdx* phenotype.

(A)

				Median	Median Normalisation Scale Factor			
Group	Sample	HybControlNormScale	RowCheck	Dilution 5%	Dilution 2%	Dilution 0.5%		
	C57 - 1	0.958	PASS	1.058	1.035	1.009		
	C57 - 2	0.993	PASS	1.037	1.06	1.033		
	C57 - 3	0.903	PASS	0.954	0.929	0.957		
	C57 - 4	1.011	PASS	1.009	1.018	1.018		
C57	C57 - 5	1.12	PASS	1.044	1.064	1.026		
	C57 - 6	1.001	PASS	0.938	0.931	0.977		
	C57 - 7	0.956	PASS	0.935	0.936	0.967		
	AVG	0.992		0.996	0.996	0.998		
	STDev	0.067	PASS	0.053	0.062	0.031		
	mdx - 1	1.022	PASS	1.271	1.223	1.227		
	mdx - 2	0.99	PASS	1.006	1.006	0.979		
	mdx - 3	1.032	PASS	0.9	0.956	1.014		
	mdx - 4	1	PASS	1.124	1.125	1.087		
mdx	mdx - 5	1.008	PASS	0.974	0.953	0.947		
	mdx - 6	0.959	PASS	0.913	0.938	0.932		
	mdx - 7	1.088	PASS	0.896	0.933	0.818		
	AVG	1.014		1.012	1.019	1.001		
	STDev	0.040	PASS	0.140	0.112	0.129		
	Fiona - 1	1.008	PASS	0.999	0.959	1.002		
	Fiona - 2	0.984	PASS	0.901	0.944	0.994		
	Fiona - 3	1.022	PASS	1.175	1.207	1.034		
	Fiona - 4	0.988	PASS	0.738	0.769	0.859		
Fiona	Fiona - 5	0.957	PASS	1.08	1.085	1.073		
	Fiona - 6	1.04	PASS	1.107	1.119	1.114		
	Fiona - 7	1.118	PASS	1.012	1.033	0.931		
	AVG	1.017	PASS	1.002	1.017	1.001		
	STDev	0.052		0.146	0.142	0.086		



S2: Quality control of SOMAscan data.

ank	Protein	Target	Uniprot	Dilution	Fold change (mdx vs C57)	pValu
1	Tumor necrosis factor receptor superfamily member 18	TNFRSF18/GITR	Q9Y5U5	5	25.4	0.000
2	Troponin I, fast skeletal muscle	TNNI2	P48788	5	10.2	0.000
3	Myosin-binding protein C, slow-type	MYBPC1	Q00872	5	7.7	0.000
-	Fibrinogen	FGA	P02671	0.5	6.5	0.000
•	Calcium/calmodulin-dependent protein kinase type II subunit beta	CAMK2B	Q13554	5	5.9	0.000
	Fibrinogen gamma chain	FGG	P02679	0.5	4.8	0.000
	Adenylate kinase isoenzyme 1	AK1	P00568	2	4.7	0.000
	Acidic leucine-rich nuclear phosphoprotein 32 family member B	ANP32B	Q92688	2	4.6	0.000
	Troponin T, cardiac muscle	TNNT2	P45379	5	4.6	0.001
)	Thrombospondin-4	THBS4	P35443	0.5	4.5	0.000
12	Calcium/calmodulin-dependent protein kinase type II subunit delta	CAMK2D	Q13557	5	4.5	0.000
1	Serine protease HTRA2, mitochondrial	HTRA2	043464	5	4.2	0.000
	Calcium/calmodulin-dependent protein kinase type II subunit alpha	CAMK2A	Q9UQM7	5	4.2	0.000
	Tyrosine-protein kinase Lyn	LYN	P07948	5	4.0	0.000
	Tyrosine-protein kinase Lyn, isoform B	LYNB	P07948	5	4.0	0.000
	Proliferating cell nuclear antigen	PCNA	P12004	5	4.0	0.001
	Eukaryotic translation initiation factor 4 gamma 2	EIF4G2	P78344	5	4.0	0.001
	60 kDa heat shock protein, mitochondrial	HSP60	P10809	5	3.7	0.000
	Tyrosine-protein kinase Fyn	FYN	P06241	5	3.5	0.000
	Tyrosine-protein kinase Yes	YES1	P07947	5	3.5	0.000
	Creating kinase M.tvog	CKM	P06732	2	2.5	0.001
	Mitoree activated protein kinase 9	MADEO	P00752	É.	3.3	0.001
	Nitogen-activated protein kinase a	MAFNO	F45565	5	5.4	0.000
	L-ractate denyorogenase 8 chain	LUND DDCD	20/192	0.5	3.3	0.000
	405 ribosomal protein 53	KPS3	P23396	5	3.2	0.000
	PIK3CA/PIK3R1	PIK3CA PIK3R1	P42336	5	3.1	0.000
	40S ribosomal protein S3a	RPS3A	P61247	5	3.1	0.000
	Cytochrome c	CYCS	P99999	5	3.0	0.001
	Proto-oncogene tyrosine-protein kinase Src	SRC	P12931	5	3.0	0.000
	Protein kinase C alpha type	PRKCA	P17252	5	3.0	0.000
	D-dimer	FGA FGB FGG	P02671	0.5	2.9	0.000
	Calpain I	CAPN1 CAPNS1	P07384	2	2.9	0.000
	RAC-beta serine/threonine-protein kinase	AKT2	P31751	5	2.8	0.001
	Mitogen-activated protein kinase 14	MAPK14	016539	2	2.8	0.000
	Pentidyl-prolyl cis-trans isomerase D	PPID	008752	5	2.8	0.000
	Troponin L cardiac muscle	TNNIB	P19429	5	2.8	0.003
	RAC-aloba /bata /samma serine /threonine-protein kinase	AKT1 AKT2 AKT3	P31749	2	2.7	0.000
	Called sail demain asstration asstration 80	000000	0761406	5	2.7	0.000
	Coned-con domain-containing protein 80	DTDNAA	005104	2	2.7	0.000
	Tyrosine-protein phosphatase non-receptor type 11	PIPNII	Q06124	2	2.6	0.001
	40S ribosomal protein S7	RPS7	P62081	5	2.5	0.000
	Heat shock protein HSP 90-alpha/beta	HSP90AA1/AB1	P07900	2	2.5	0.000
	Ribosomal protein S6 kinase alpha-3	RPS6KA3	P51812	5	2.5	0.000
	cAMP-dependent protein kinase catalytic subunit alpha	PRKACA	P17612	5	2.5	0.001
	Dual specificity mitogen-activated protein kinase kinase 1	MAP2K1	Q02750	5	2.4	0.000
	Dual specificity protein phosphatase 3	DUSP3	P51452	5	2.4	0.002
	Pro-opiomelanocortin	POMC	P01189	5	2.4	0.000
	Heat shock 70 kDa protein 1A	HSPA1A	PODMV8	5	2.4	0.000
	Casein kinase II 2-alpha': 2-beta heterotetramer	CSNK2A2 CSNK2B	P19784	5	2.4	0.004
	Glycogen synthase kinase-3 alpha/beta	GSK3A GSK3B	P49840	5	2.4	0.001
	Mitogen-activated protein kinase 12	MAPK12	P53778	5	2.4	0.000
	3-phosphoinositide-dependent protein kinase 1	PDPK1	015530	5	23	0.002
	Heat shock compate 71 kDa protain	LICDAO	P11142	2	2.2	0.000
	ATD dependent PNA holisers DDV10D	DOV100	00000000	É.	2.5	0.000
	AIP-dependent KNA neilcase DUX198	DDX19B	Q90MR2	5	2.3	0.000
	14-5-5 protein family	TWHAB	P31946	2	2.3	0.000
	Protein FAM3D	FAMBD	Q96BQ1	5	2.2	0.000
	NAD-dependent protein deacetylase sirtuin-2	SIRT2	Ø 8IX16	5	2.2	0.001
	Xaa-Pro aminopeptidase 1	XPNPEP1	Q9NQW7	5	2.2	0.000
	Receptor-type tyrosine-protein kinase FLT3	FLT3	P36888	5	2.2	0.001
	Protein kinase C beta type (splice variant beta-II)	PRKCB	P05771	2	2.2	0.001
	Tumor necrosis factor receptor superfamily member 27	EDA2R	Q9HAV5	5	2.2	0.000
	Osteocalcin	BGLAP	P02818	2	2.2	0.001
	Histone H2A.z	H2AFZ	POCOS5	5	2.2	0.000
	Tyrosine-protein kinase CSK	CSK	P41240	2	2.1	0.002
	Mothers against decapentablegic homolog 3	SMAD3	P84022	5	2.1	0.001
	Secreted and transmembrane protein 1	SECTM1	08WVN6	2	2 1	0.000
	Follistatin-related protein 1	ESTI 1	012941	2	2 1	0.000
	hata-adrenergic recentor kinase 1	ADRBK1	P25000	2	2.1	0.001
	Hampinyalia	LEE2	067/000	2	2.1	0.001
	nemojovelih Istorallulas adbasias malanda 2	ICANO	Q6ZVN8	2	2.0	0.001
	Intercentular adhesion molecule 2	ICAM2	P13598	2	2.0	0.000
	Alpha-soluble NSF attachment protein	NAPA	P54920	2	2.0	0.000
	Protein \$100-A12	\$100A12	P80511	2	2.0	0.001
	MAP kinase-activated protein kinase 3	МАРКАРКЗ	Q16644	2	2.0	0.003
	Repulsive guidance molecule A	RGMA	Q96886	2	2.0	0.000
	Mitochondrial import inner membrane translocase subunit TIM14	DNAJC19	Q96DA6	5	2.0	0.000
	Tyrosine-protein kinase Fer	FER	P16591	5	2.0	0.001
	Triosephosphate isomerase	TPI1	P60174	2	2.0	0.003
	Growth factor receptor-bound protein 2	GRB2	P62993	5	2.0	0.000
	Hsp90 co-chaperone Cdc37	CDC37	016543	5	2.0	0.001
	Methionine aminonentidase 2	METAP2	P50579	5	2.0	0.001
	Nideran 1	NID1	P14545	2	2.0	0.001
	Ridogen-1	CNITNI	F14545		2.0	0.000
	contactin-1	CNINI	012860	0.5	0.7	0.001
	Growth/differentiation factor 8	MSTN	014793	0.5	0.7	0.003
	Tumor necrosis factor receptor superfamily member 25	TNFRSF25	Q93038	5	0.7	0.00
	Desmoglein-2	DSG2	Q14126	0.5	0.4	0.000

Table S1: Serum mdx protein biomarkers identified by SOMAscan.

Rank	Protein	Target	Uniprot	Dilution	Fold change (mdx vs C57)	pValue	Recovery score (Fiona)
1	Tumor necrosis factor receptor superfamily member 18	TNFRSF18/GITR	Q9Y5U5	5	25.4	0.0001	+++
2	Troponin I, fast skeletal muscle	TNNI2	P48788	5	10.2	0.0002	**
4	Fibringen	FGA	Q00872 P02671	0.5	65	0.0003	
5	Calcium/calmodulin-dependent protein kinase type II subunit beta	CAMK2B	Q13554	5	5.9	0.0001	**
6	Fibrinogen gamma chain	FGG	P02679	0.5	4.8	0.0001	-
7	Adenylate kinase isoenzyme 1	AK1	P00568	2	4.7	0.0007	**
8	Acidic leucine-rich nuclear phosphoprotein 32 family member B	ANP32B	Q92688	2	4.6	0.0003	***
10	Troponini, cardiac muscle	THRSA	P35443	0.5	4.6	0.0011	
11	Calcium/calmodulin-dependent protein kinase type II subunit delta	CAMK2D	013557	5	4.5	0.0001	++
12	Serine protease HTRA2, mitochondrial	HTRA2	043464	5	4.2	0.0001	+++
13	Calcium/calmodulin-dependent protein kinase type II subunit alpha	CAMK2A	Q9UQM7	5	4.2	0.0003	***
14	Tyrosine-protein kinase Lyn	LYN	P07948	5	4.0	0.0003	**
15	Tyrosine-protein kinase Lyn, isoform B	LYN	P07948	5	4.0	0.0003	***
16	Proliferating cell nuclear antigen	PCNA EIE4G2	P12004	5	4.0	0.0012	***
18	60 kDa heat shock protein, mitochondrial	HSP60	P10809	5	3.7	0.0006	+++
19	Tyrosine-protein kinase Fyn	FYN	P06241	5	3.5	0.0004	++++
20	Myoglobin	MB	P02144	2	3.5	0.0011	+++
21	Tyrosine-protein kinase Yes	YES1	P07947	5	3.5	0.0003	**
22	Creatine kinase M-type	CKM	P06732	2	3.5	0.0011	+
25	I-lactate debydrogenase Bichain	IDHR	P07195	0.5	3.4	0.0007	÷
25	40S ribosomal protein S3	RPS3	P23396	5	3.2	0.0005	-
26	PIK3CA/PIK3R1	PIK3CA PIK3R1	P42336	5	3.1	0.0001	++
27	40S ribosomal protein S3a	RPS3A	P61247	5	3.1	0.0006	***
28	Cytochrome c	CYCS	P99999	5	3.0	0.0013	+++
29	Proto-oncogene tyrosine-protein kinase Src	SRC	P12931	5	3.0	0.0007	**
30	D-dimer	FGA FGB FGG	P02671	0.5	2.9	0.0001	
31	Calpain I	CAPNI CAPNSI	PU/384	2	2.9	0.0001	
33	Mitogen-activated protein kinase 14	MAPK14	016539	2	2.8	0.0005	+
34	Peptidyl-prolyl cis-trans isomerase D	PPID	Q08752	5	2.8	0.0008	1.4
35	Troponin I, cardiac muscle	TNNI3	P19429	5	2.8	0.0036	-
36	RAC-alpha/beta/gamma serine/threonine-protein kinase	AKT1 AKT2 AKT3	P31749	2	2.7	0.0007	-
37	Coiled-coil domain-containing protein 80	CCDC80	Q76M96	2	2.7	0.0002	**
38	Tyrosine-protein phosphatase non-receptor type 11	PTPN11	Q06124	5	2.6	0.0010	**
39	40S ribosomal protein S7	KPS/	P62081	5	2.5	0.0009	
40	Ribosomal protein 55 kinase alpha-3	RPSEKA3	P0/900 P51812	2	2.5	0.0004	Ţ
42	cAMP-dependent protein kinase catalytic subunit alpha	PRKACA	P17612	5	2.5	0.0012	-
43	Dual specificity mitogen-activated protein kinase kinase 1	MAP2K1	002750	5	2.4	0.0001	-
44	Dual specificity protein phosphatase 3	DUS3	P51452	5	2.4	0.0020	+++
45	Heat shock 70 kDa protein 1A	HSPA1A	PODMV8	5	2.4	0.0004	+
46	Casein kinase II 2-alpha':2-beta heterotetramer	CSNK2A2 CSNK2B	P19784	5	2.4	0.0043	····
4/	Glycogen synthase kinase-3 alpha/beta	GSK3A GSK3B	P49840	5	2.4	0.00011	
49	3-phosphoinositide-dependent protein kinase 1	PDPK1	015530	5	2.4	0.0004	
50	Heat shock cognate 71 kDa protein	HSPAS	P11142	2	2.3	0.0003	**
51	ATP-dependent RNA helicase DDX19B	DDX19B	Q9UMR2	5	2.3	0.0006	+
52	14-3-3 protein family	YWHAB	P31946	2	2.3	0.0003	**
53	Protein FAM3D	FAM3D	Q96BQ1	5	2.2	0.0001	+++
54	NAD-dependent protein deacetylase sirtuin-2	SIRT2	QSIXJ6	5	2.2	0.0014	***
55	xaa-Pro aminopeptidase 1 Recentor-type tyrosine-protein kinase ELT3	ELT3	Q9NQW7	5	2.2	0.0008	
57	Protein kinase C beta type (splice variant beta-II)	PRKCB	P05771	2	2.2	0.0016	2
58	Tumor necrosis factor receptor superfamily member 27	EDA2R	Q9HAV5	5	2.2	0.0001	+++
59	Osteocalcin	BGLAP	P02818	2	2.2	0.0012	***
60	Histone H2A.z	H2AFZ	POCOS5	5	2.2	0.0001	+++
61	Tyrosine-protein kinase CSK	CSK	P41240	2	2.1	0.0027	-
62	Mothers against decapentaplegic homolog 3	SMAD3	P84022	5	2.1	0.0016	***
64	Follistatio-related protein 1	ESTI 1	012841	2	2.1	0.0003	
65	beta-adrenergic receptor kinase 1	ADRBK1	P25098	5	2.1	0.0012	+++
66	Hemojuvelin	HFE2	Q6ZVN8	2	2.0	0.0010	+++
67	Intercellular adhesion molecule 2	ICAM2	P13598	2	2.0	0.0001	+++
68	Alpha-soluble NSF attachment protein	NAPA	P54920	2	2.0	0.0006	+
69	Protein \$100-A12	\$100A12	P80511	2	2.0	0.0013	***
70	MAP kinase-activated protein kinase 3	MAPKAPK3	Q16644	2	2.0	0.0031	
72	Mitochondrial import inner membrane translocase subunit TIM14	DNAIC19	096046	2	2.0	0.0005	
73	Triosephosphate isomerase	TPI1	P60174	2	2.0	0.0034	++
74	Growth factor receptor-bound protein 2	GRB2	P62993	5	2.0	0.0008	1
75	Casein kinase II 2-alpha:2-beta heterotetramer	CSNK2A1 CSNK2B	P68400 P67870	5	2.0	0.0004	+++
76	Hsp90 co-chaperone Cdc37	CDC37	Q16543	5	2.0	0.0012	-
77	Methionine aminopeptidase 2	METAP2	P50579	5	2.0	0.001	***
- 78	Nidogen-1	NID1	P14543		2.0	0.0003	···
/9	ContactIR-1 Growth/differentiation factor 11/0	GDE11 MSTN	Q12860	0.5	0.7	0.0013	
81	Bcl-2-like protein 2	BCL2L2	092843	5	0.7	0.0036	
82	beta-nerve growth factor	NGF	P01138	5	0.7	0.0006	12
83	E3 ubiquitin-protein ligase ZNRF3	ZNRF3	Q9ULT6	5	0.7	0.0009	-
84	Growth/differentiation factor 8	MSTN	014793	0.5	0.7	0.0033	+++
85	Tumor necrosis factor receptor superfamily member 25	TNFRSF25	Q93038	5	0.7	0.005	
86	Neurogenic locus notch homolog protein 2	NOTCH2	Q04721	5	0.6	0.0005	-
87	Glucose-6-phosphate isomerase	GPI	P06744	5	0.6	0.0018	100
89	Desmoglein-2	DSG2	Q14126	0.5	0.4	0.00012	
5.70.1	2 19 19 2 19 19 19 19 19 19 19 19 19 19 19 19 19	~cccm0600810	5475 S (547) S (547) S (57)	10000	2000		

Table S2: Serum *mdx* protein biomarkers and rescue in *mdx* transgenic Fiona mice.

Target	Catalog Number	Detection range	Sensitivity	Comments		
ANP32B	ABIN1745100		0.1ng/ml			
CAMK2A	ABIN426487	0.31-20 ng/mL	0.118 ng/mL	Sample below detection limit		
GITR	ABIN1672796	62.5-4000 pg/mL	<10pg/mL			
SIRT2	ABIN416762	1.56-100 ng/mL	0.65 ng/mL			
THBS4	ABIN426909	0.312-20 ng/mL	0.64 ng/mL			
TNNT2	ABIN426397	15.63-1000 pg/mL	6.2 pg/mL			

Table S3: Details of the ELISA assays used in the study.

References

- 1 Schindelin, J. *et al.* Fiji: an open-source platform for biological-image analysis. *Nature methods* **9**, 676-682, doi:10.1038/nmeth.2019 (2012).
- 2 Barclay, C. J., Woledge, R. C. & Curtin, N. A. Effects of UCP3 genotype, temperature and muscle type on energy turnover of resting mouse skeletal muscle. *Pflugers Archiv : European journal of physiology* **457**, 857-864, doi:10.1007/s00424-008-0552-z (2009).
- Lynch, G. S., Hinkle, R. T. & Faulkner, J. A. Power output of fast and slow skeletal muscles of mdx (dystrophic) and control mice after clenbuterol treatment. *Experimental physiology* 85, 295-299 (2000).
- 4 Brooks, S. V. & Faulkner, J. A. Contractile properties of skeletal muscles from young, adult and aged mice. *The Journal of physiology* **404**, 71-82 (1988).