

**Analytical and Bioanalytical Chemistry**

**Electronic Supplementary Material**

**Hydrophobic ionic liquids for quantitative bacterial cell lysis with subsequent DNA quantification**

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## Quantitative PCR setup

### *Salmonella* assay

The *fimA*-qPCR assay is targeting the major fimbrial subunit encoding gene *fimA* of *Salmonella* and amplifying a 84-bp fragment. The assay consists of the forward primer (*fimAF1*: 5'-CCTTTCTCCATCGTCCTGAA-3'), the reverse primer (*fimAR1*: TGGTGTTATCTGCCTGA-3') and the FAM-labelled TaqMan® probe (*fimAS1*: 5'-FAM-TGCGATCCGAAAGTGGCGG-BHQ1-3') (Eurofins MWG Operon, Ebersberg, Germany). All qPCRs were performed with a volume of 25 µl/PCR reaction containing a final concentration of 1× PCR buffer, 3.5 mM MgCl<sub>2</sub>, 300 nM of each primer and the probe, 200 µM of dNTPs, 1.5 U Platinum® Taq DNA polymerase (Invitrogen, Vienna, Austria) and 5 µl of DNA template. A qPCR temperature profile with an initial denaturation step at 94 °C for 2 min and an amplification protocol with 45 cycles (94 °C for 30 s and 60 °C for 1 min) was used.

### *Listeria monocytogenes* assay

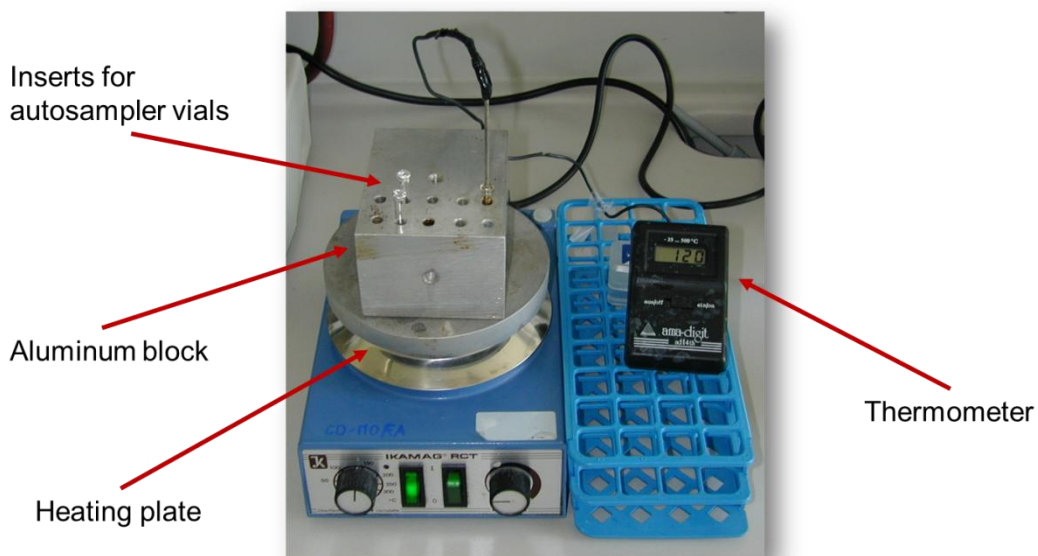
The *prfA*-qPCR assay is targeting a 274 bp fragment of the *prfA* gene of *Listeria monocytogenes*. The assay consists of the forward primer (*LIP1*: 5'-GATACAGAAACATCGGTTGGC-3'), the reverse primer (*LIP2*: 5'-GTGTAATCTTGATGCCATCAGG-3') and the FAM-labelled TaqMan® probe (*LIP probe 2*: (5'-FAM-CAGGATTAAGTTGACCGCA-MGB-3') (Eurofins MWG Operon, Ebersberg, Germany).

All qPCRs were performed with a volume of 25 µl/PCR reaction containing 1x PCR buffer, 3.5 mM MgCl<sub>2</sub>, 500 nM of each primer, 250 nM of each probe, 200 µM (of dNTPs, 1.5 U of Platinum® *Taq* DNA polymerase (Invitrogen, Vienna, Austria) and 5 µl DNA template. Amplification following initial denaturation at 94 °C for 2 min was performed in 45 cycles, at 94 °C for 15 s, and 64 °C for 1 min.

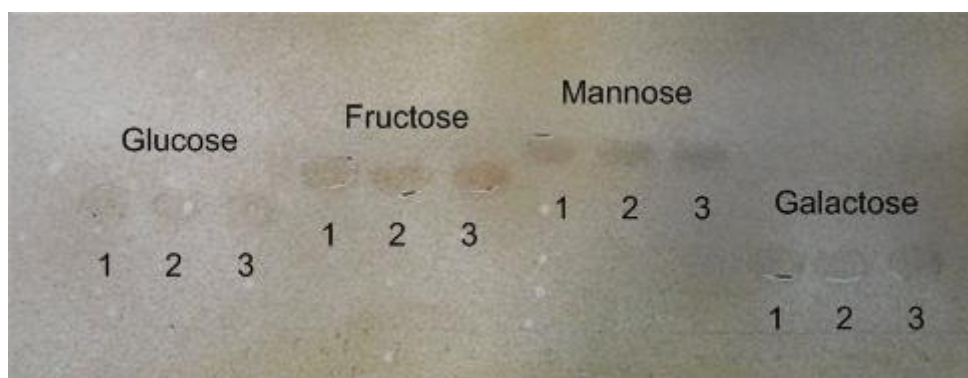
### *Escherichia coli* assay

The *sfmD*-qPCR assay is targeting a 106 bp fragment of *sfmD* gene, encoding a putative outer membrane export usher protein, of *E.coli*. The assay consists of the forward primer (*ERT2F*: 5'-ACTGGAATACTTCGGATTCAGATACGT -3'), the reverse primer (*ERT2R*: 5'-ATCCCTACAGATTCATTCCACGAAA-3') and the FAM-labelled TaqMan® probe (5'-FAM-CAGCAGCTGGGTTGGCATCAGTTATTCG-BHQ1-3') (Eurofins MWG Operon, Ebersberg, Germany).

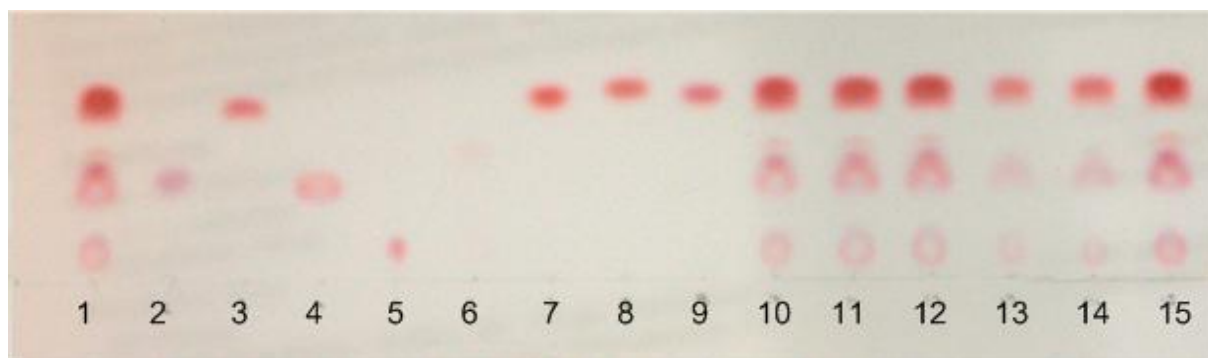
All qPCRs were performed with a volume of 25 µl/PCR reaction containing 1x PCR buffer, 3.5 mM MgCl<sub>2</sub>, 300 nM of each primer, 200 nM of each probe, 200 µM (of dNTPs, 1 U of Platinum® *Taq* DNA polymerase (Invitrogen, Vienna, Austria) and 5 µl DNA template. Amplification following initialdenaturation at 94 °C for 2 min was performed in 45 cycles, at 94 °C for 15 s, and 60 °C for 1 min.



**Fig S1** Aluminium block for DNA isolation experiments



**Fig. S2** Monosaccharides (glucose, fructose, mannose and galactose) applied directly (1), after extraction with  $[\text{BMPyr}^+][\text{Ntf}_2^-]$ , (2) or after extraction with  $[\text{BMPyr}^+][\text{Ntf}_2^-]$  and additional centrifugation (5 min,  $6,000 \times g$ ) (3)



**Fig. S3** Amino acids L-alanine (2), L-tyrosine (3), L-glutamine (4), L-histidine (5), L-cysteine (6), L-leucine (7), L-tryptophan (8), L-phenylalanine (9) applied directly, as a mixture containing all amino acids (1, 10 and 15), after extraction with  $[\text{BMPyr}^+][\text{Ntf}_2^-]$  (11), with additional centrifugation (5 min,  $6,000 \times g$ ) (12) or after extraction with  $[\text{BMPyr}^+][\text{Ntf}_2^-]$  with additional incubation for 2 min at  $140^\circ\text{C}$  (13) and additional centrifugation (5 min,  $6,000 \times g$ ) (14)