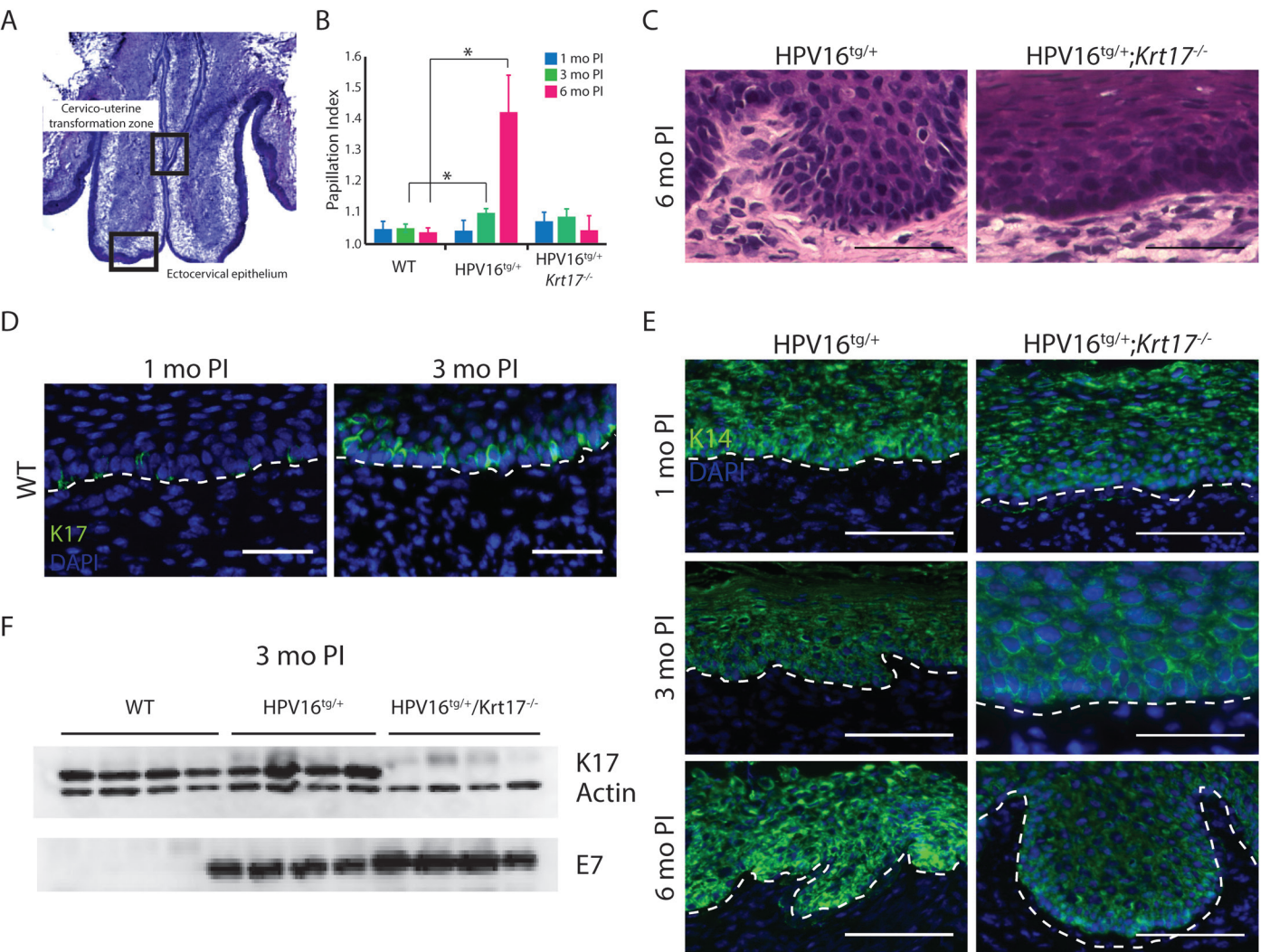
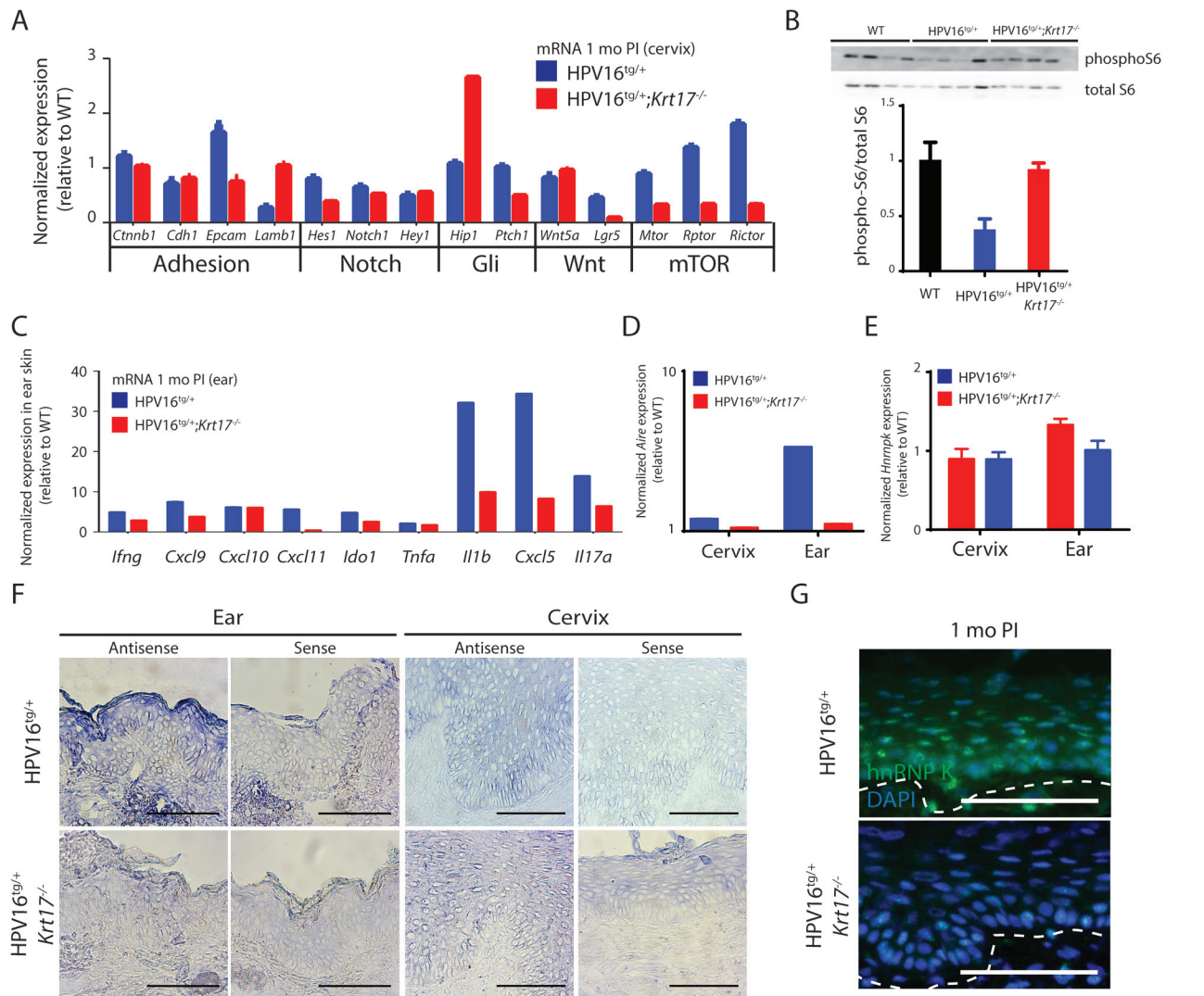


# Supplementary Fig. S1



**Supplementary Fig. S1: Characterization of cervical epithelium in HPV16<sup>tg/+</sup> mice.** **A)** Reconstructed image from low-magnification (5x) micrographs covering the entire mouse cervix. Boxed regions highlight the cervico-uterine transformation zone and ectocervical epithelium. **B)** Quantitation of epithelial papillation in transformation zone epithelia for WT, HPV16<sup>tg/+</sup>, and HPV16<sup>tg/+</sup>;Krt17<sup>-/-</sup> mice at 1-, 3-, and 6-mo post-implantation. Error bars are s.e.m. \*p<0.05 (Student's T-test). **C)** H&E staining of transformation zone epithelium for HPV16<sup>tg/+</sup> and HPV16<sup>tg/+</sup>;Krt17<sup>-/-</sup> mice at 6-mo post-implantation. Bar = 100  $\mu$ m. This panel accompanies Fig. 1A. **D)** Immunostaining for K17 in transformation zone epithelium of WT mice at 1-mo (left) and 3-mo (right) post-implantation. Nuclei are stained with DAPI (blue). Dotted line separates epithelium (above line) from underlying stroma (below line). Bar = 100  $\mu$ m. **E)** Immunostaining for K14 in transformation zone epithelium of HPV16<sup>tg/+</sup> (top) and HPV16<sup>tg/+</sup>;Krt17<sup>-/-</sup> (bottom) mice at 1-, 3-, and 6-mo post-implantation. Nuclei are stained with DAPI (blue). Dotted line separates epithelium (above line) from underlying stroma (below line). Bar = 100  $\mu$ m. **F)** Immunoblots for K17, Actin, and HPV E7 in cervical tissue of WT, HPV16<sup>tg/+</sup>, and HPV16<sup>tg/+</sup>;Krt17<sup>-/-</sup> mice at 3-mo post-implantation. Each lane represents a distinct biological replicate.

# Supplementary Fig. S2

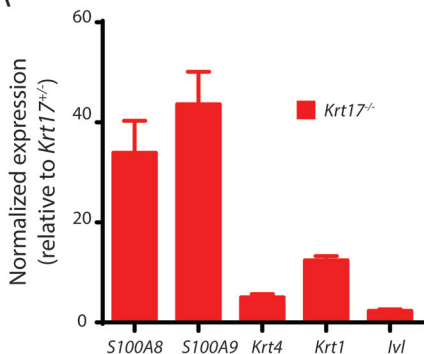


**Supplementary Fig. S2: Assessment of Aire and hnRNP K in cervical and ear tissues.**

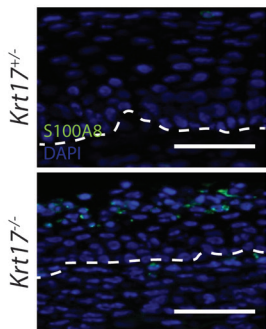
**A)** Normalized expression for gene transcripts associated with adhesion, Notch, Gli, Wnt, and mTOR signaling in cervical tissue of HPV16<sup>tg/+</sup> and HPV16<sup>tg/+</sup>;Krt17<sup>-/-</sup> mice at 1-mo post-implantation. Error bars are s.e.m. **B)** Immunoblots for phospho-S6 and total S6 in cervical tissue of WT, HPV16<sup>tg/+</sup>, and HPV16<sup>tg/+</sup>;Krt17<sup>-/-</sup> mice at 1-mo post-implantation. Each lane represents a distinct biological replicate. Densitometric quantification of phospho-S6/total S6 shown below. **C)** Normalized expression for inflammation associated gene transcripts in ear tissue of HPV16<sup>tg/+</sup> and HPV16<sup>tg/+</sup>;Krt17<sup>-/-</sup> mice at 1-mo post-implantation. Error bars are s.e.m. This panel accompanies Fig. 3B. **D)** Normalized expression for Aire transcript in cervical and ear tissues of HPV16<sup>tg/+</sup> and HPV16<sup>tg/+</sup>;Krt17<sup>-/-</sup> mice at 1-mo post-implantation. Error bars are s.e.m. **E)** Normalized expression for *Hnnpk* transcript in cervical and ear tissues of HPV16<sup>tg/+</sup> and HPV16<sup>tg/+</sup>;Krt17<sup>-/-</sup> mice at 1-mo post-implantation. Error bars are s.e.m. **F)** RNA *in situ* hybridization for *Aire* gene transcript in ear (left) and cervical (right) tissues of HPV16<sup>tg/+</sup> and HPV16<sup>tg/+</sup>;Krt17<sup>-/-</sup> mice at 1-mo post-implantation. Signals from both antisense and sense probes are shown. Bar = 100  $\mu$ m. **G)** Immunostaining for hnRNP K in transformation zone epithelium of HPV16<sup>tg/+</sup> and HPV16<sup>tg/+</sup>;Krt17<sup>-/-</sup> mice at 1-mo post-implantation. Nuclei are stained with DAPI (blue). Dotted line separates epithelium (above line) from underlying stroma (below line). Bar = 100  $\mu$ m.

# Supplementary Fig. S3

A



B



## Supplementary Fig. S3: Differentiation of cervical epithelia occurs in non-transgenic, unimplanted, aged mice in the absence of K17. A)

Normalized expression for differentiation-associated gene transcripts in cervical tissue of *Krt17*<sup>-/-</sup> mice relative to *Krt17*<sup>+/-</sup> littermates. Error bars are s.e.m.

B) Immunostaining for S100A8 in transformation zone epithelium of *Krt17*<sup>+/-</sup> (top) and *Krt17*<sup>-/-</sup> (bottom) mice. Nuclei are stained with DAPI (blue). Dotted line separates epithelium (above line) from underlying stroma (belowline). Bar = 100  $\mu$ m.

**Table S1. List of qRT-PCR primers.**

Target	Primer	Sequence (5'-->3')	Target	Primer	Sequence (5'-->3')
<i>18S</i>	Forward	CCTGTGCCTTCCTTGGA	<i>Inv</i>	Forward	ATGTCCCATCAACACACTG
<i>18S</i>	Reverse	CATTCGAACGTCTGCCCTATC	<i>Inv</i>	Reverse	TGGAGTTGGTTGCTTTGCTTG
<i>Actb</i>	Forward	GGCTGTATTCCCTCCATCG	<i>Krt1</i>	Forward	TGGGAGATTTTCAGGAGGAGG
<i>Actb</i>	Reverse	CCAGTTGGTAACAATGCCATGT	<i>Krt1</i>	Reverse	GCCACACTCTTGGAGATGCTC
<i>Aire</i>	Forward	AGGTCAGCTTCAGAGAAAACCA	<i>Krt4</i>	Forward	TCGGCAGCAGAAGTCTTTACA
<i>Aire</i>	Reverse	TCATTCCCAGCACTCAGTAGA	<i>Krt4</i>	Reverse	CAGCACCGTATCTCCAACG
<i>Cdh1</i>	Forward	CAGGTCTCCTCATGGCTTTGC	<i>Krt10</i>	Forward	ACGAGAAGCATGGCAACTCAA
<i>Cdh1</i>	Reverse	CTTCCGAAAAGAAGGCTGTCC	<i>Krt10</i>	Reverse	GCTCATCCAGTACCCTGCG
<i>Ctnnb1</i>	Forward	ATGGACGTGGGGCAACTTTTA	<i>Krt17</i>	Forward	ACCATCCGCCAGTTTACCTC
<i>Ctnnb1</i>	Reverse	CGCCATCCCTGTCAATAATCTG	<i>Krt17</i>	Reverse	CTACCCAGGCCACTAGCTGA
<i>Cxcl10</i>	Forward	CCAAGTGCTGCCGTCATTTTC	<i>Lamb1</i>	Forward	AGACTTTGGGGGTTTCATGTCA
<i>Cxcl10</i>	Reverse	GGCTCGCAGGGATGATTTCAA	<i>Lamb1</i>	Reverse	ATCGTCCCGTCTCCTTGTC
<i>Cxcl11</i>	Forward	GGCTTCCTTATGTTCAAACAGGG	<i>Mmp13</i>	Forward	TGTTTGACAGAGCACTACTTGAA
<i>Cxcl11</i>	Reverse	GCCGTTACTCGGGTAAATTACA	<i>Mmp13</i>	Reverse	CAGTACCTCTAAGCCAAAGAAA
<i>Cxcl5</i>	Forward	TCCAGCTCGCCATTCATGC	<i>Mmp9</i>	Forward	CTGGACAGCCAGACACTAAAG
<i>Cxcl5</i>	Reverse	TTGCGGCTATGACTGAGGAAG	<i>Mmp9</i>	Reverse	CTCGGGCAAGTCTTCAGAG
<i>Cxcl9</i>	Forward	TCCTTTTGGGCATCATCTTCC	<i>Mtor</i>	Forward	CAGTTCGCCAGTGGACTGAAG
<i>Cxcl9</i>	Reverse	TTGTAGTGGATCGTGCCTCG	<i>Mtor</i>	Reverse	GCTGGTCATAGAAGCGAGTAGAC
<i>Epcam</i>	Forward	GCGGCTCAGAGAGACTGTG	<i>Notch1</i>	Forward	GATGGCCTCAATGGGTACAAG
<i>Epcam</i>	Reverse	CCAAGCATTTAGACGCCAGTTT	<i>Notch1</i>	Reverse	TCGTTGTTGTTGATGTCACAGT
<i>Flg</i>	Forward	ATGTCCGCTCTCTGGAAAG	<i>Patch1</i>	Forward	CTTCCACCCACAGTCTCTCC
<i>Flg</i>	Reverse	TGGATTCTTCAAGACTGCCTGTA	<i>Patch1</i>	Reverse	GGGAAGGCTACTGGCCGG
<i>Hes1</i>	Forward	CCAGCCAGTGTCAACACGA	<i>Rictor</i>	Forward	GCTGCGTATCTCATCCAAGA
<i>Hes1</i>	Reverse	AATGCCGGGAGCTATCTTTCT	<i>Rictor</i>	Reverse	CTTTCTGACTAAGCGAAGGGC
<i>Hey1</i>	Forward	GCGCGGACGAGAATGGAAA	<i>Rptor</i>	Forward	TGTGAGAAAATCGAAGGCTCC
<i>Hey1</i>	Reverse	TCAGGTGATCCACAGTCATCTG	<i>Rptor</i>	Reverse	TTTGCACCGATGGTTTCCAGA
<i>Hip1</i>	Forward	CCACTTGATTGAACGCCTGTA	<i>S100a8</i>	Forward	AAATCACCATGCCCTCTACAAG
<i>Hip1</i>	Reverse	ACTCGCAGTCATCCATAGCCT	<i>S100a8</i>	Reverse	CCCACTTTTATCACCATCGCAA
<i>Hnrnpk</i>	Forward	ACTGATGAGATGGTTGAATTGCG	<i>S100a9</i>	Forward	ATACTCTAGGAAGGAAGGACACC
<i>Hnrnpk</i>	Reverse	CTGGCATTGTAGTCTGTACGG	<i>S100a9</i>	Reverse	TCCATGATGTCATTTATGAGGGC
<i>Ido1</i>	Forward	GCTTTGCTCTACCACATCCAC	<i>Sfn</i>	Forward	TTGGCTGAACAGGCCGAAC
<i>Ido1</i>	Reverse	CAGGCGCTGTAACCTGTGT	<i>Sfn</i>	Reverse	GTAAGCTACGGAAAGCAGGTTT
<i>lfng</i>	Forward	ATGAACGCTACACACTGCATC	<i>Tgfb</i>	Forward	CTCCCGTGGCTTCTAGTGC
<i>lfng</i>	Reverse	CCATCCTTTTGCCAGTTCCTC	<i>Tgfb</i>	Reverse	GCCTTAGTTTGGACAGGATCTG
<i>Il1b</i>	Forward	GAAATGCCACCTTTTGACAGT	<i>Tnfa</i>	Forward	CTGAACTTCGGGGTGATCGG
<i>Il1b</i>	Reverse	CTGGATGCTCTCATCAGGACA	<i>Tnfa</i>	Reverse	GGCTTGCTCACTCGAATTTGAGA
<i>Il17a</i>	Forward	TTTAACTCCCTTGGCGCAAAA			
<i>Il17a</i>	Reverse	CTTCCCTCCGATTGACAC			