

Supplemental Figures

FIGURE S1. Catalytic activities of MTSL-labeled ExoU variants. (A) Relative activity of each spin label derivative compared to unlabeled recombinant wild type ExoU. Black bars indicate spin-labeled variants with PED6 hydrolysis rates less than 0.25 wild type. Bars at the top of the Figure indicate locations of helices, lines represent the interhelical loops, and arrows the positions of variants with reduced activity. (B) Structural model of the C-terminal four-helix bundle indicating the position of derivatives with less than 25% activity as compared to wild type (blue spheres).

FIGURE S2. Far-UV circular dichroism spectra of ExoU and spin-labeled derivatives. Molar ellipticity (θ) in the region from 250-195 nm for wild type (WT) ExoU and several MTSL spin-labeled derivatives that exhibit reduced catalytic activity.

FIGURE S3A. EPR spectra of MTSL-labeled helical sites in the presence of liposomes. (A) Spectral overlays of spin labeled mutants in the first helix of the C-terminal four-helix bundle. Green spectra are in the presence of 100-fold excess liposome substrate and black spectra correspond to the apoenzyme state. (B-D): Spectral overlays of spin labeled sites in the second through fourth helices of the C-terminal bundle, respectively. Spectra were normalized to integrated intensity. Scaling factors for presentation are shown at the right of each spectrum. All scan widths are 100 G.

FIGURE S3B. EPR spectra MTSL-labeled loop sites in the presence of liposomes. (A) Spectral overlays of spin label mutants in the first loop of the helical bundle. Green spectra correspond to ExoU in the presence of 100-fold excess liposome substrate and black spectra correspond to the apoenzyme state. (B-D): Spectral overlays of spin label sites in the second and third interhelical loops of the C-Terminal bundle, respectively. Spectra were normalized to integrated intensity. Scaling factors for presentation are shown at the right of each spectrum. Scan widths 100 G.

FIGURE S4A. EPR spectral overlays of helical sites in the Apo and Holo states. (A) Spectral overlays of R1-labeled mutants in the first helix of the C-terminal four-helix bundle. Red spectra are in the presence of liposomes and diUb (Holo state) and black spectra correspond to the Apo state. (B-D): Spectral overlays of R1-labeled sites in the second through fourth helices of the C-terminal bundle, respectively. Spectra were normalized to integrated intensity. Scaling factors are shown at the right of each spectrum. Scan widths are 100 G.

FIGURE S4B. EPR spectral overlays of loop sites in the Apo and Holo states. (A) Spectral overlays of R1-labeled mutants in the first helix of the C-terminal four-helix bundle. Red spectra are in the presence of liposomes and diUb (Holo state) and black spectra correspond to the Apo state. (B-D): Spectral overlays of R1-labeled sites in the second through fourth helices of the C-terminal bundle, respectively. Spectra were normalized to integrated intensity. Scaling factors are shown at the right of each spectrum. Scan widths are 100 G.

FIGURE S5. Simulation of V602R1 EPR spectra in the apo (Left) and holo (Right) states. (Top) Overlay of the experimental (black) and fitted (red) spectra. (Bottom and middle) Individual components used to achieve the fit. A small (~2%) contribution from rapidly-tumbling was included to improve the overall fit in the apo state. The holo state could not be adequately fit with a single component. Component fractions, rotational correlation times, and order parameters are indicated in the *insets*. The order parameter was 0 unless otherwise noted.

FIGURE S6. Simulation of R611R1 EPR spectra in the apo (Left) and holo (Right) states. (Top) Overlay of the experimental (black) and fitted (red) spectra. (Bottom and middle) Individual components used to achieve the fit. Component fractions, rotational correlation times, and order parameters are indicated in the *insets*. The order parameter was 0 unless otherwise noted.

FIGURE S7. Experimentally determined Φ values and calculated distances from the lipid phosphates based on equation 4. (Open circles, apo), (green triangles, liposomes only), (orange squares, holo), (black squares, PCSL standards). Negative distances indicate sites exposed to the aqueous phase.

FIGURE S1.

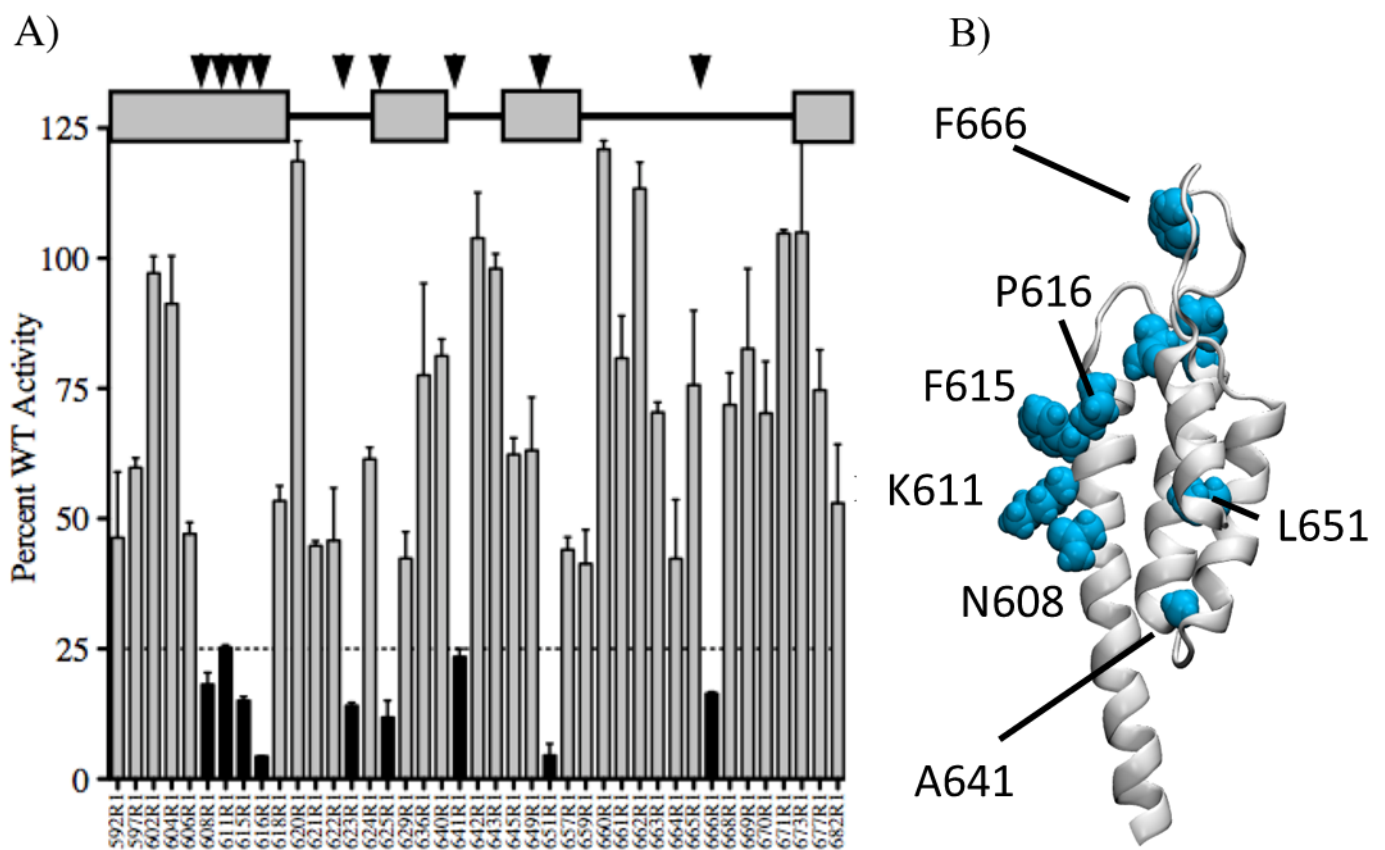


FIGURE S3A.

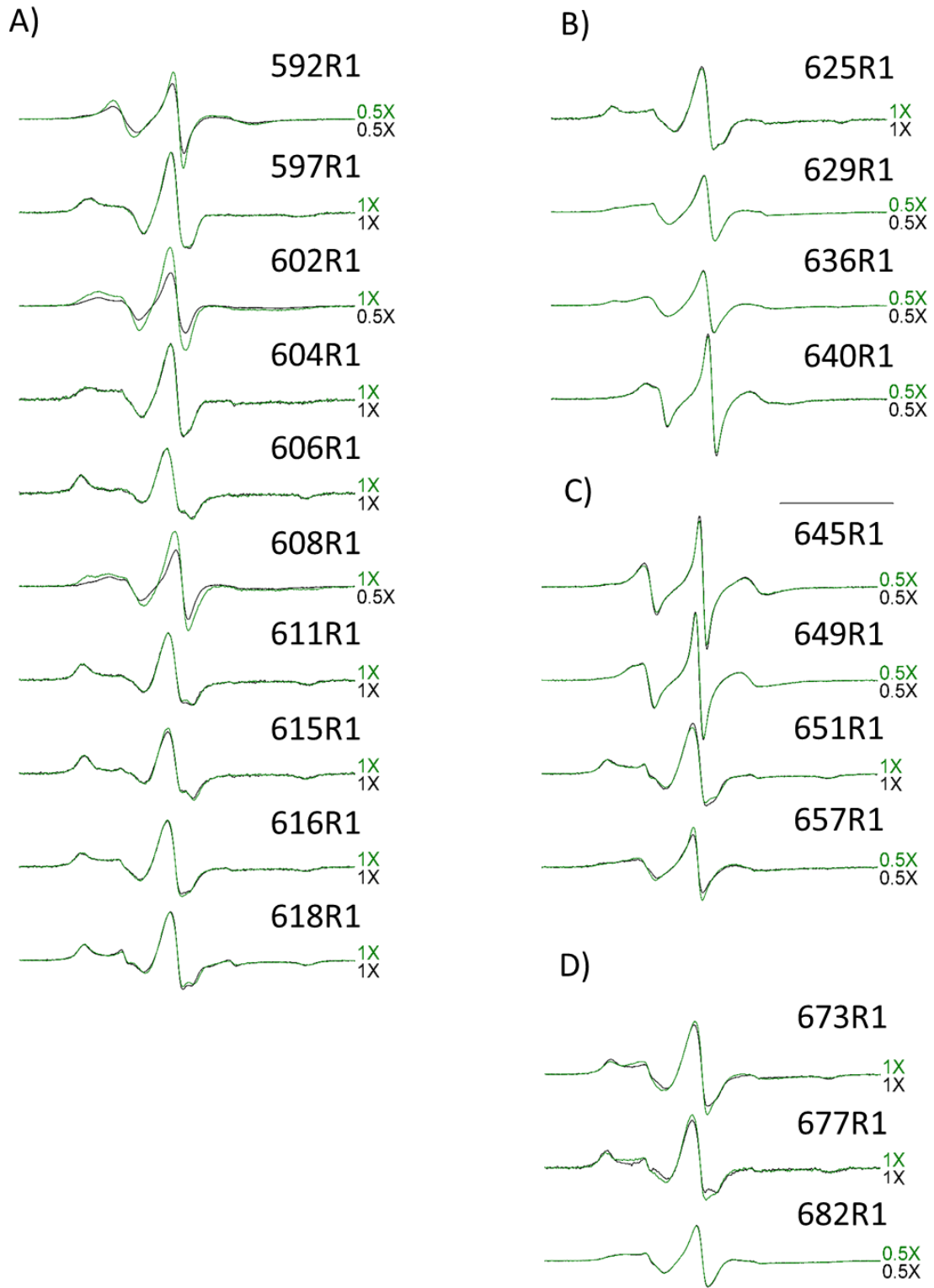


FIGURE S3B.

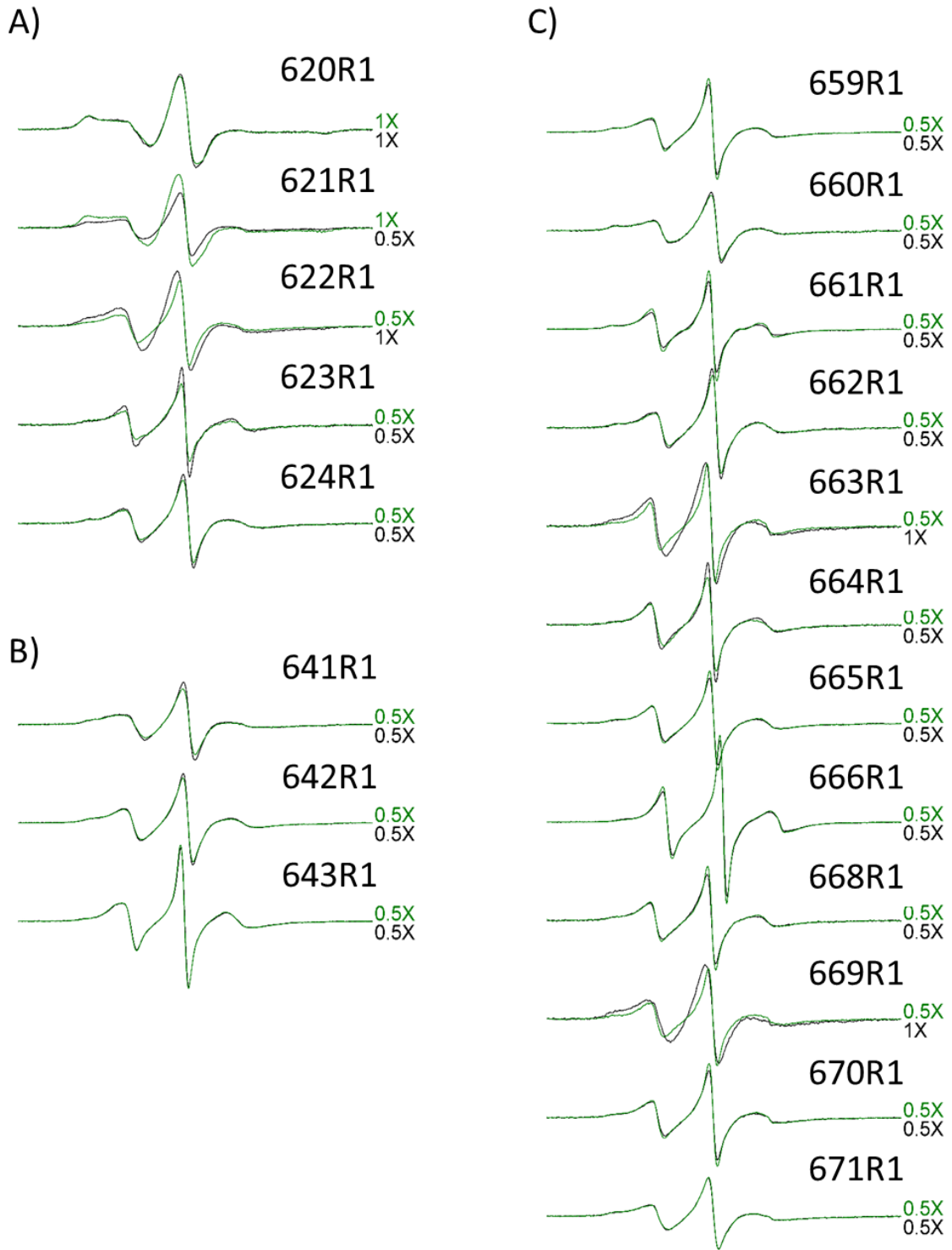


FIGURE S4A.

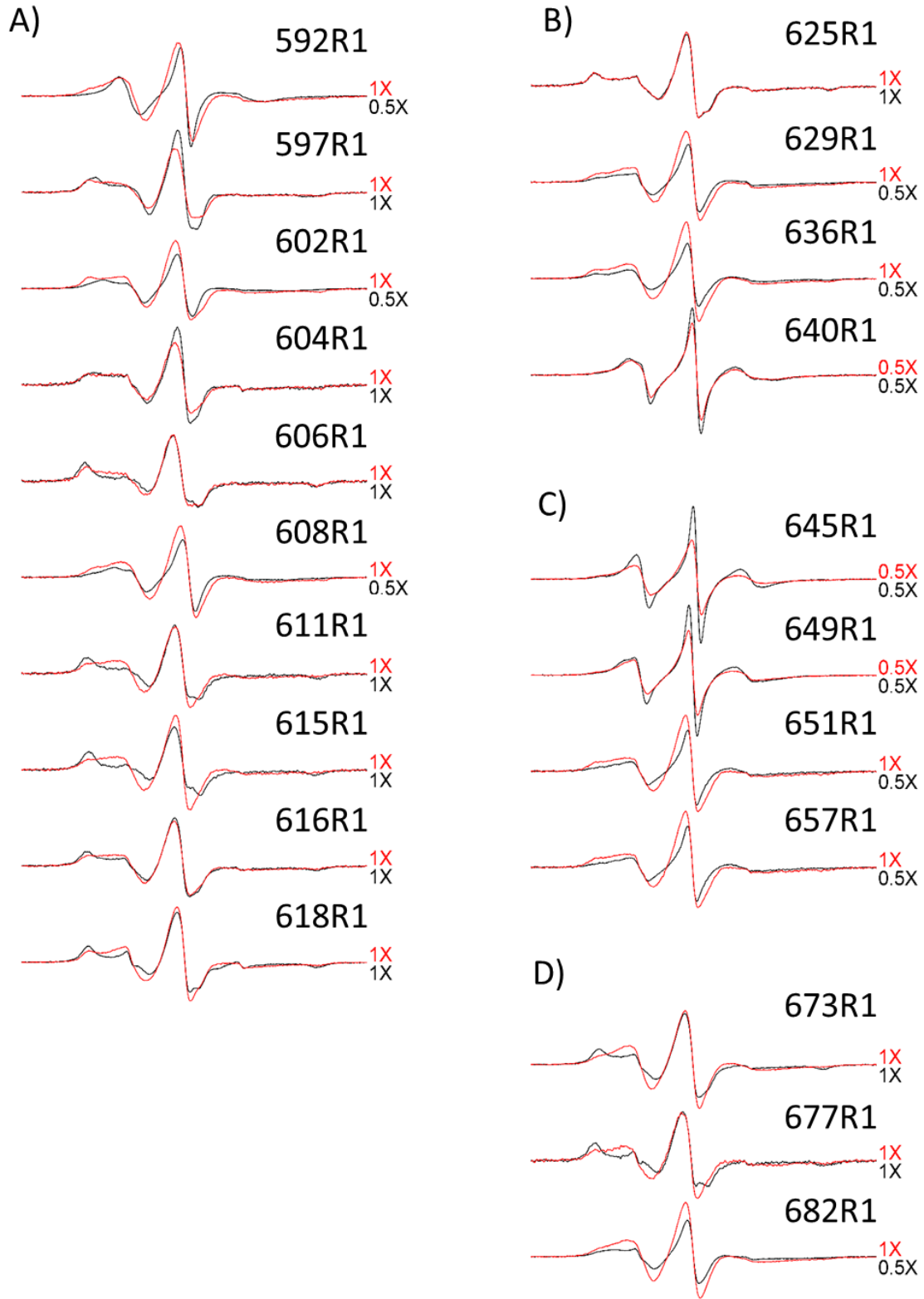


FIGURE S4B.

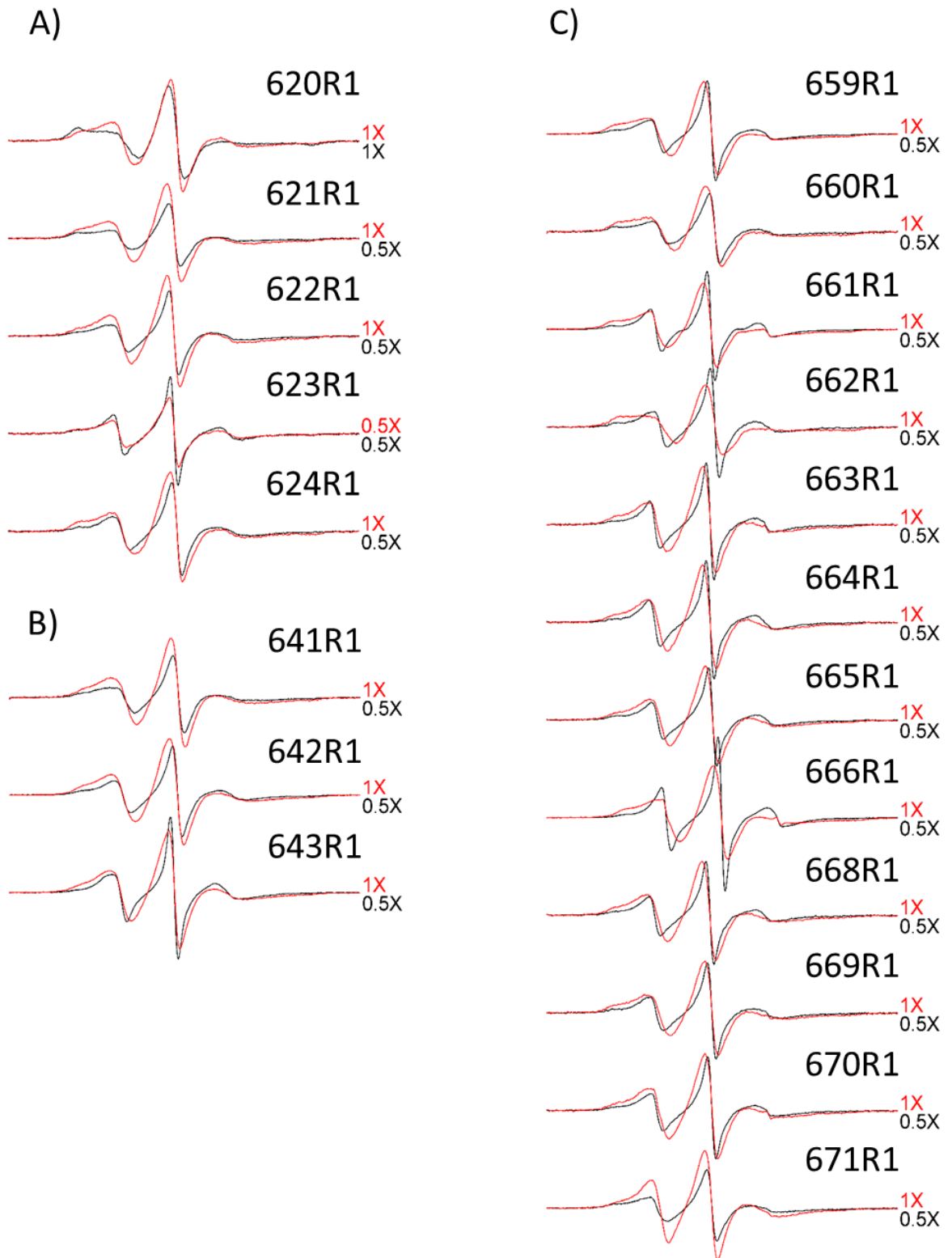


FIGURE S5.

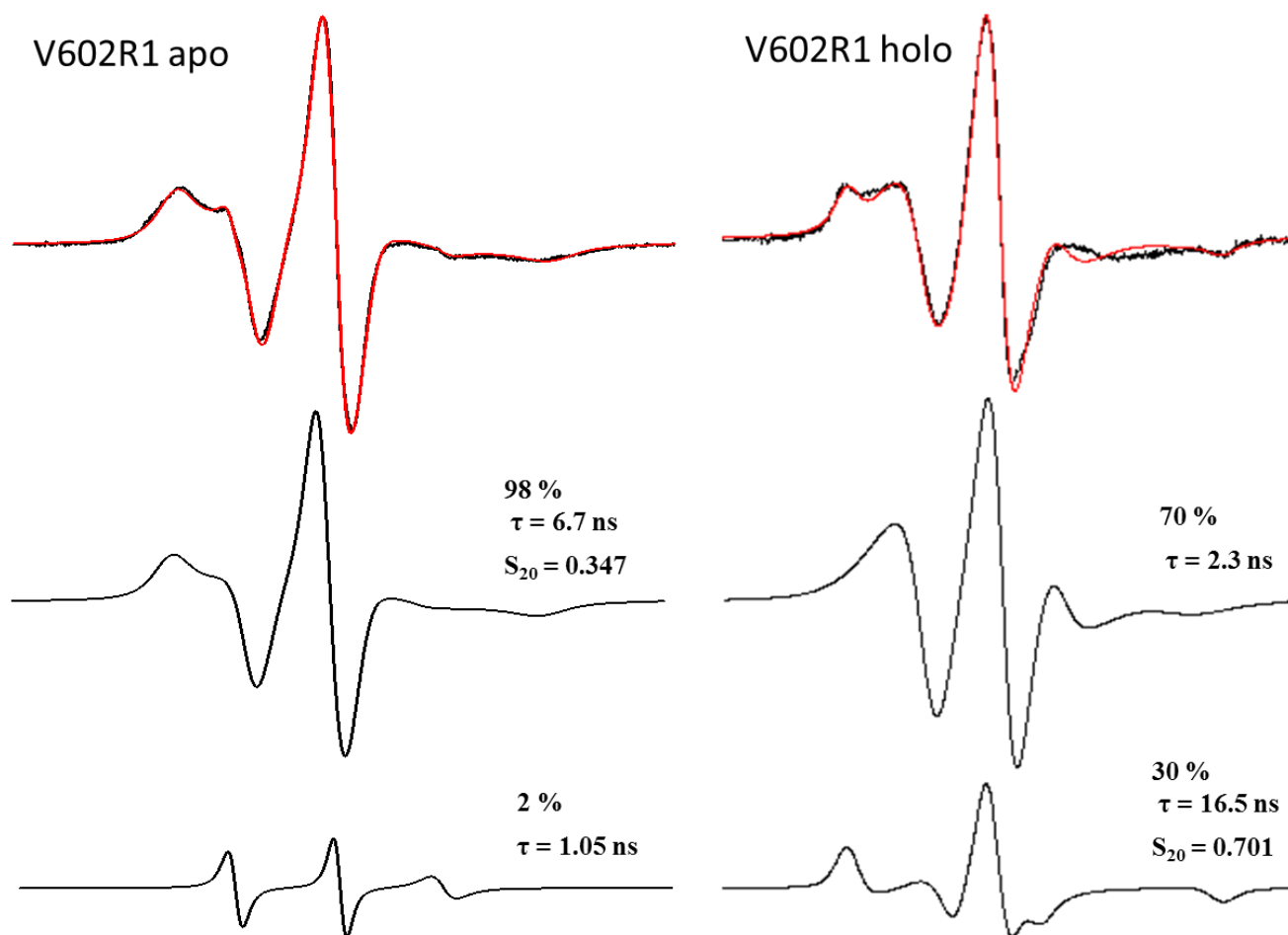


FIGURE S6.

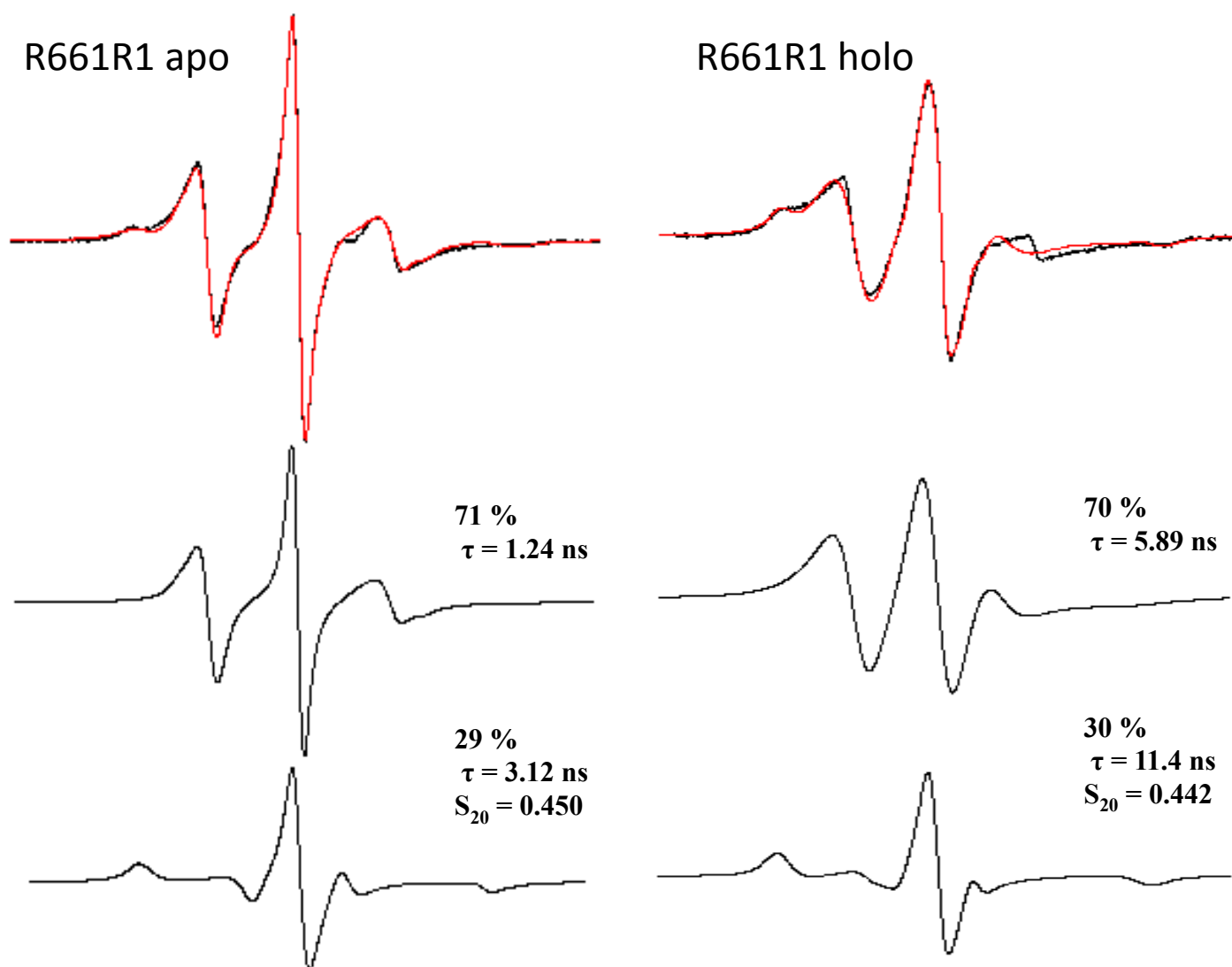


FIGURE S7.

