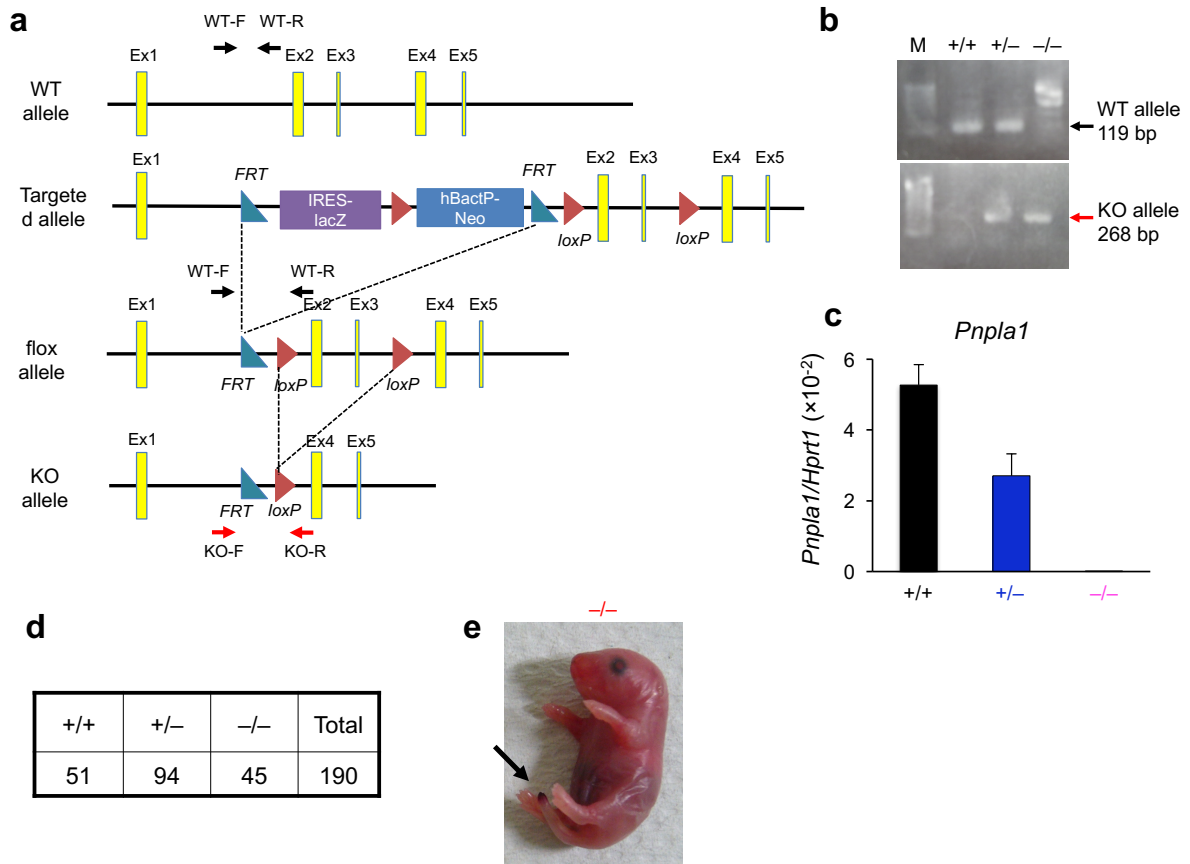


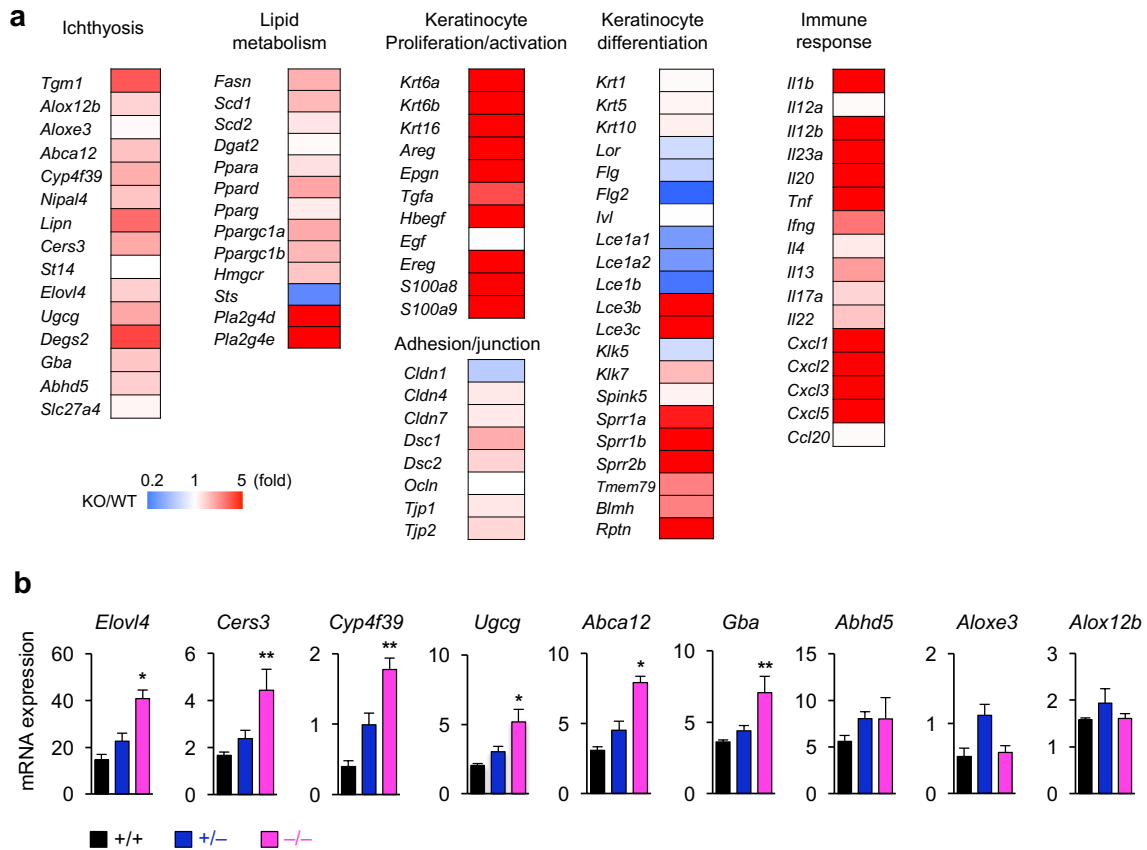
Supplementary Fig. 1. *Pnpla1* expression in the skin and differentiated cultured keratinocytes.

(a) qPCR analysis of *Pnpla1* expression in various tissues of 8-week-old male C57BL/6 mice, with *Hprt1* as an internal control (mean \pm s.e.m., $n = 6$ animals). (b) Immunofluorescent staining of newborn mouse skin sections with antibodies specific for PNPLA1 (green) and filaggrin, loricrin, keratin 1 or keratin 5 (red). Nuclei were counterstained with DAPI (blue). (c) Immunohistochemical staining of the back skin sections of adult C57BL/6 mice (8-week-old) with anti-PNPLA1 antibody (upper panel) or isotype-matched control antibody (lower panel). The sections were counterstained with hematoxylin. (d, e) qPCR analysis of expression levels of *Pnpla1*/*PNPLA1* and keratinocyte differentiation markers in mouse (d) and human (e) keratinocytes in culture with 1.2 mM CaCl_2 for the indicated periods, with *Hprt1* and *RPL13A*, respectively, as internal controls (mean \pm s.e.m., $n = 3$). Results are representative of two or three independent experiments. Scale bar, 20 μm .



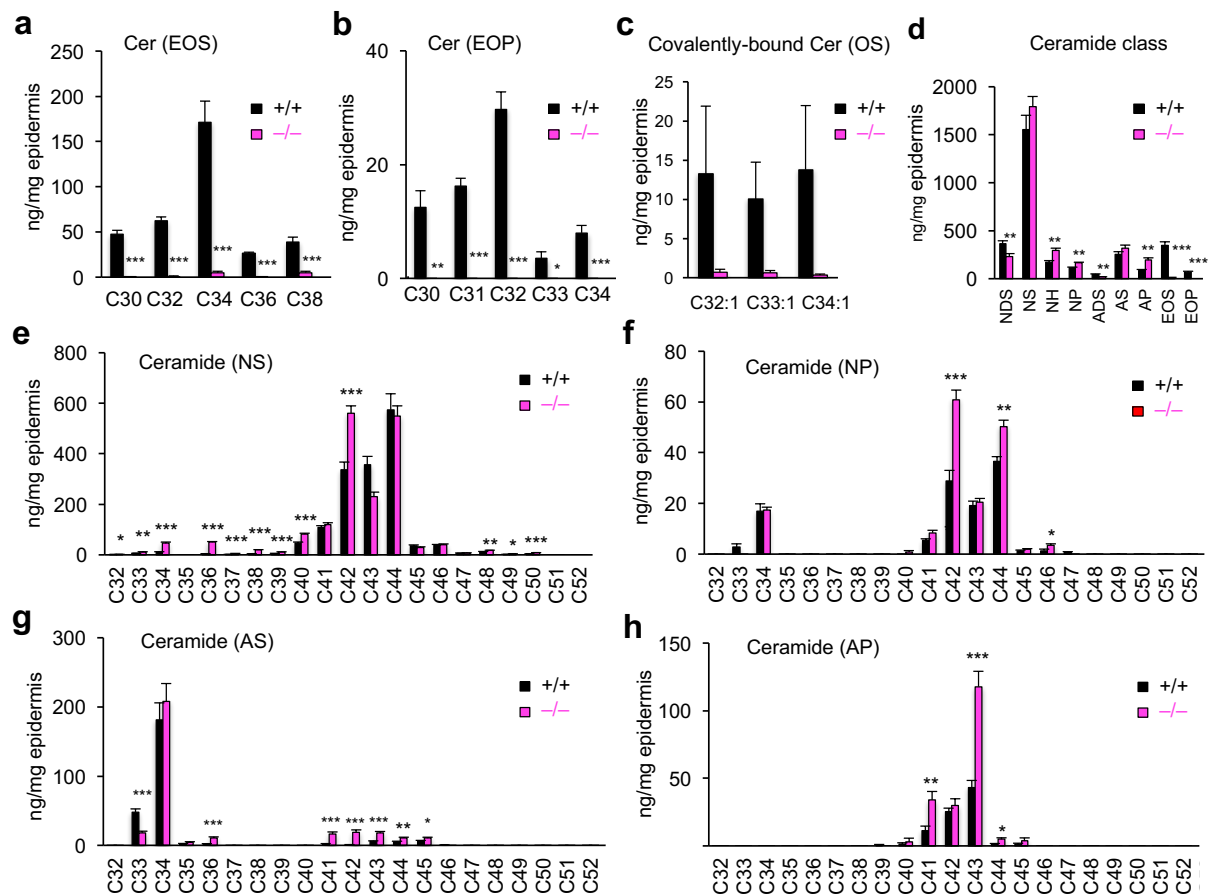
Supplementary Fig. 2. Generation of *Pnpla1*-deficient mice.

(a) Strategy of *Pnpla1* gene targeting. In the targeted allele, exons 2 and 3 were flanked by *loxP* sites, and the *IRES-LacZ* and *Neo* cassettes flanked by *FRT* sites were inserted between exons 1 and 2. *FRT* and *loxP* sites are depicted as blue and red triangles, respectively. The positions of primers for genotyping PCR (arrows labeled WT-F, WT-R, KO-F, and KO-R) are indicated. *Neo*, neomycin phosphotransferase gene. (b) PCR genotyping of tail DNA, amplifying 119-bp and 268-bp products of WT and KO alleles, respectively. (c) qPCR analysis of *Pnpla1* mRNA expression in the skin of *Pnpla1*^{+/+}, *Pnpla1*^{+/-} and *Pnpla1*^{-/-} mice at P0 (mean \pm s.e.m., n = 5 animals per group). (d) Frequencies of *Pnpla1* genotypes in pups derived from intercrosses between heterozygous mice. (e) Tail tip necrosis (arrow) in *Pnpla1*^{-/-} newborn mice. Representative (b, c, e) and compiled (d) results from multiple experiments are shown.



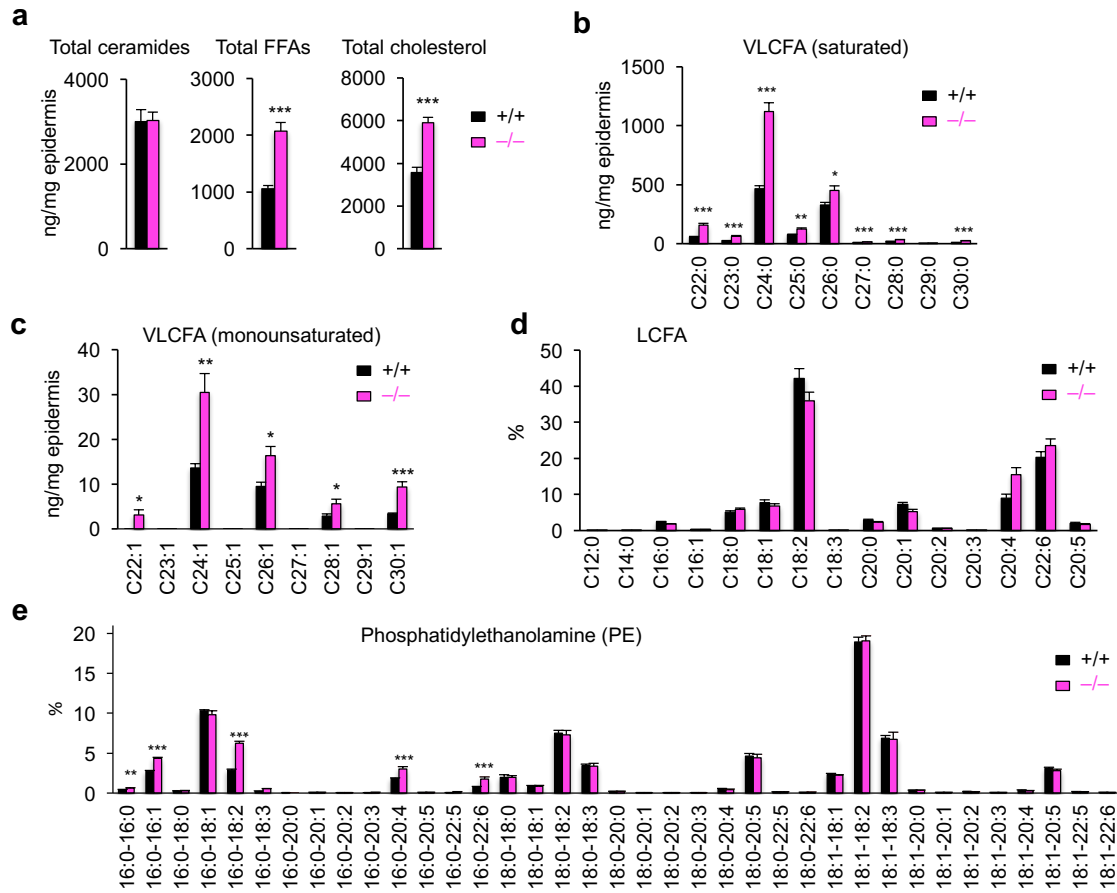
Supplementary Fig. 3. Changes in gene expression in *Pnpla1*^{-/-} epidermis.

(a) Altered expression profiles of genes associated with ichthyosis, lipid metabolism, keratinocyte proliferation/activation, adhesion/junction, keratinocyte differentiation, and immune response in *Pnpla1*^{-/-} (KO) epidermis relative to *Pnpla1*^{+/+} (WT) epidermis at P0, as assessed by DNA microarray analysis. Fold changes (KO/WT) are represented by color. Equal amounts of total RNA pooled from six mice for each genotype were used. (b) qPCR analysis of genes for epidermal ceramide metabolism in *Pnpla1*^{+/+}, *Pnpla1*^{+/-} and *Pnpla1*^{-/-} skin at P0 (mean ± s.e.m., n = 5 animals per genotype; **P* < 0.05 and ***P* < 0.01 versus *Pnpla1*^{+/+} mice). Representative results from two independent experiments are shown.



Supplementary Fig. 4. Effects of *Pnpl1* deletion on epidermal ceramides.

(a–h) LC-MS/MS quantification of EOS (a), EOP (b), protein-bound Cer OS (c), nine ceramide classes (d), NS (e), NP (f), AS (g), and AP (h) in the epidermis of *Pnpl1*^{+/+} and *Pnpl1*^{-/-} neonates (mean ± s.e.m., n = 5 animals; **P* < 0.05, ***P* < 0.01 and ****P* < 0.001 versus *Pnpl1*^{+/+} mice in an unpaired, two-tailed Student's *t*-test). Ceramide classes are abbreviated according to three different fatty acid components (N, non-hydroxy fatty acid; A, α -hydroxy fatty acid; EO, linoleic acid esterified ω -hydroxy fatty acid) linked via an amide bond to the sphingoid bases (S, sphingosine; DS, dehydrosphingosine; P, phytosphingosine; H, 6-hydroxysphingosine). Results are from one or two independent experiments.



Supplementary Fig. 5. Effects of *Pnpla1* deletion on epidermal FFAs, cholesterol, and phospholipids.

(a) Quantification of total contents of ceramides, FFAs, and cholesterol in $Pnpla^{+/+}$ and $Pnpla^{-/-}$ neonates by LC-MS. (b–e) Quantification of saturated and monounsaturated forms of VLCFA (b, c), LCFA (d), and phosphatidylethanolamine (e) in the epidermis of $Pnpla^{+/+}$ and $Pnpla^{-/-}$ neonates by LC-MS and LC-MS/MS. Data from one or two independent experiments are shown as the mean \pm s.e.m. (n = 5 animals; * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$ versus $Pnpla^{+/+}$ mice in an unpaired, two-tailed Student's *t*-test).

Supplementary Table 1. Primers for genotyping PCR.

Primer name	Annealing temperature (°C)	Product size (bp)	5'-3' sequence
<i>Pnpla1</i> WT-F	59	119	TGGCCTCTAAGCAAGTGCCCTTGT
<i>Pnpla1</i> WT-R			GCAGCTTCTTGTCCCTGCACCTGA
<i>Pnpla1</i> KO-F	59	330	TCTAAGGCGCATAACGATAC
<i>Pnpla1</i> KO-R			TCCTCATTTGTCTTAGGATGC
<i>Cre</i> -F	56.5	281	CCATCTGCCACCAGCCAG
<i>Cre</i> -R			TCGCCATCTTCCAGCAGG
<i>Abhd5</i> WT-F	57	417	AGGAGGAGGTGGACTCG
<i>Abhd5</i> COM-R			TTTCACAAAGTTTGCCAGG
<i>Abhd5</i> KO-F	57	626	TCCTCGTGCTTTACGGTATC
<i>Abhd5</i> COM-R			TTTCACAAAGTTTGCCAGG

Supplementary Table 2. Primers and UPL probes for qPCR (mouse).

Gene	Primer sequence	UPL probe
<i>Abca12</i>	CCTGCTAAACCAGACGATCC	#50
	ACTTGCACAAAGGGGTTC	
<i>Abhd5</i>	ATCTTTGGAGCCCGATCCT	#89
	CTTCTGGCTGATCTGCATACAC	
<i>Alox12b</i>	CTTTGGTCCTGATGGCAAC	#105
	GACAATCAGGCCCAGGAGT	
<i>Aloxe3</i>	GGCCTCACTGATCTTCAACG	#4
	GTCCAGGAGACCTCGAATCTT	
<i>Cers3</i>	CATGATATATCTGACATTTGGCTAGAG	#38
	GTGAAGATGAAGAACAGGGTGTTA	
<i>Cyp4f39</i>	AAAGAAGAAAGCCAAGAGAATTGA	#89
	TGCAACAGGTAGTCGGTGAG	
<i>Elovl4</i>	ACGACACCGTGGAGTTCTATC	#85
	GCGGCCAGTCTGCTACAC	
<i>Fabp5</i>	ACGGCTTTGAGGAGTACATGA	#46
	CTCGGTTTTGACCGTGATG	
<i>Flg</i>	GGAGGAAGAAACACTGAGCAA	#96
	CGATGTCTTGGTCATCTGGA	
<i>Gba</i>	GACCAACGCTTGCTGCTAC	#34
	ACAGCAATGCCATGAACGTA	
<i>Hbegf</i>	TCTTCTTGTCATCGTGGGACT	#55
	CACGCCAACTTCACTTTCT	
<i>Hprt1</i>	TGATAGATCCATTCCATGACTGTAGA	#22
	AAGACATTCTTTCCAGTTAAAGTTGAG	
<i>Krt1</i>	TGAGCTGAAGAACATGCAAGA	#31
	CATGTAAGCTGAATCCACATCC	
<i>Krt5</i>	CAGAGCTGAGGAACATGCAG	#22
	CATTCTCAGCCGTGGTACG	
<i>Krt6a</i>	GCCAAGGCAGACAGTCTAACA	#55
	CAGGCTACGGTTGTTGTCC	
<i>Krt6b</i>	GGAAATTGCCACCTACAGGA	#12
	GGTGGACTGCACCACAGAG	
<i>Lor</i>	GGTTGCAACGGAGACAACA	#11
	CATGAGAAAGTTAAGCCCATCG	
<i>Pnpl1</i>	TGCGGGATTGAGATGGAG	#94
	CGCATCATCTGTACCATCTTG	
<i>Ppard</i>	ATGGGGGACCAGAACACAC	#11
	GGAGGAATTCTGGGAGAGGT	
<i>Ugcg</i>	AGTTTCAATCCAGAATGATCAGG	#4
	CATTCTGAAATTGGCTCACAAAT	

Supplementary Table 3. Primers and UPL probes for qPCR (human).

<i>Gene</i>	Primer sequence	UPL probe
<i>FLG</i>	TGCTCCCGAGAAGATCCAT	#38
	GGACTCTGAGAGGCGATCTG	
<i>KRT1</i>	TCCGCTTGTTGATTTCATCC	#22
	ATCAATCTCGGTTGGATTTCG	
<i>KRT5</i>	CCACTTGGTGTCCAGAACCT	#1
	GCAGATCAAGACCCTCAACAAT	
<i>PNPLA1</i>	TGGTGGTTTCAGAGTTCACG	#63
	ACGAAGCAGCTGCAGTATAGG	
<i>RPL13A</i>	CTGGACCGTCTCAAGGTGTT	#18
	GCCCCAGATAGGCAAACCT	