

## Description of *U. maydis* strains used in this study.

Strain	Locus	Progenitor strain	Short description
FB1			<i>Wild type strain</i> [1]
FB2			<i>Wild type strain</i> [1]
AB33	<i>b</i>	FB2	<i>Pnar:bw2bE1</i> , expression of active b heterodimer under control of the <i>nar1</i> promoter, strain grows filamentously upon changing the nitrogen source. [2]
FB1aox1Δ	<i>aox1</i>	FB1	Carrying a deletion of <i>aox1</i> .
FB2aox1Δ	<i>aox1</i>	FB2	Carrying a deletion of <i>aox1</i> .
AB33aox1Δ	<i>aox1</i>	AB33	Carrying a deletion of <i>aox1</i> .
FB2aox1-Gfp	<i>aox1</i>	FB2aox1Δ	Expressing Aox1 C-terminally fused to eGfp.
AB33aox1-Gfp	<i>aox1</i>	AB33aox1Δ	Expressing Aox1 C-terminally fused to eGfp.
FB2P <sub>otef</sub> :aox1-Gfp	<i>ip<sup>S</sup></i>	FB2	<i>aox1-Gfp</i> is ectopically integrated in the defined <i>ip<sup>S</sup></i> locus and under control of the strong constitutive P <sub>otef</sub> promoter.
FB2P <sub>otef</sub> :5'UTR-aox1-Gfp	<i>ip<sup>S</sup></i>	FB2	<i>aox1-Gfp</i> is ectopically integrated in the defined <i>ip<sup>S</sup></i> locus and under control of the strong constitutive P <sub>otef</sub> promoter. In this construct the native 5'UTR is retained.
AB33P <sub>otef</sub> :aox1-Gfp	<i>ip<sup>S</sup></i>	AB33	<i>aox1-Gfp</i> is ectopically integrated in the defined <i>ip<sup>S</sup></i> locus and under control of the strong constitutive P <sub>otef</sub> promoter.
AB33P <sub>otef</sub> :5'UTR-aox1-Gfp	<i>ip<sup>S</sup></i>	AB33	<i>aox1-Gfp</i> is ectopically integrated in the defined <i>ip<sup>S</sup></i> locus and under control of the strong constitutive P <sub>otef</sub> promoter. In this construct the native 5'UTR is retained.

## Generation of *U. maydis* strains used in this study.

Strains	Relevant genotype	Uma	Reference	Transformed plasmid	Locus	Progenitor
AB33	<i>a2 P<sub>nar.</sub>:bW2 bE1</i>	133	[2]	pAB33	<i>b</i>	FB2
FB1 <i>aox1</i> Δ	<i>aox1</i> Δ	1328	this study	pAox1Δ_HygR (pUMa2163)	<i>aox1</i>	FB1
FB2 <i>aox1</i> Δ	<i>aox1</i> Δ	1329	this study	pAox1Δ_HygR (pUMa2163)	<i>aox1</i>	FB2
AB33 <i>aox1</i> Δ	<i>aox1</i> Δ	1330	this study	pAox1Δ_HygR (pUMa2169)	<i>aox1</i>	AB33
FB2 <i>aox1</i> -Gfp	<i>aox1-Gfp</i>	1333	this study	pAox1-Gfp_NatR (pUMa2169)	<i>aox1</i>	FB2 <i>aox1</i> Δ
AB33 <i>aox1</i> -Gfp	<i>aox1-Gfp</i>	1369	this study	pAox1-Gfp_NatR (pUMa2169)	<i>aox1</i>	AB33 <i>aox1</i> Δ
FB2P <sub>otef</sub> : <i>aox1</i> -Gfp	<i>aox1-Gfp</i>	1816	this study	pP <sub>otef</sub> : <i>aox1</i> -Gfp_CbxR (p2768)	<i>ip<sup>s</sup></i>	FB2
FB2P <sub>otef</sub> :5'UTR- <i>aox1</i> -Gfp	<i>aox1-Gfp</i>	1814	this study	pP <sub>otef</sub> :5'UTR- <i>aox1</i> -Gfp (p2767)	<i>ip<sup>s</sup></i>	FB2
AB33P <sub>otef</sub> : <i>aox1</i> -Gfp	<i>aox1-Gfp</i>	1817	this study	pP <sub>otef</sub> : <i>aox1</i> -Gfp_CbxR (p2768)	<i>ip<sup>s</sup></i>	AB33
AB33P <sub>otef</sub> :5'UTR- <i>aox1</i> -Gfp	<i>aox1-Gfp</i>	1815	this study	pP <sub>otef</sub> :5'UTR- <i>aox1</i> -Gfp (p2767)	<i>ip<sup>s</sup></i>	AB33

## Description of plasmids used for *U. maydis* strain generation.

Plasmid	pUMa	Resistance cassette	Short description
pAox1Δ_HygR	2163	SfiI-insert of MF1hs	Plasmid for generating deletion mutants of <i>aox1</i> . Resistance cassette is flanked by 1 kb upstream and 1 kb downstream region of <i>aox1</i> . Flanking regions were amplified by PCR using oRL1400/oRL1401 and oRL1402/oRL1403 and UM521 wild-type DNA as template. Plasmid was generated by Golden Gate cloning [3]
pAox1-Gfp_NatR	2169	SfiI-insert of pMF5-1n	Plasmid for generating eGfp fusions of <i>aox1</i> . A cassette containing Gfp, the T <sub>nos</sub> terminator and a nourseothricin resistance cassette is flanked by 2.4 kb upstream region including 1.3 kb of the <i>aox1</i> ORF and 1 kb downstream region of <i>aox1</i> . Flanking regions were amplified by PCR using oRL1400/oRL1425 and oRL1402/oRL1403 and UM521 wild-type DNA as template.
pP <sub>otef</sub> :5UTR- <i>aox1</i> -Gfp	2767	CbxR for integration at <i>ip<sup>S</sup></i> locus	Plasmid for ectopical integration and expression of <i>aox1-Gfp</i> . Contains 1.3 kb open reading frame (ORF) of <i>aox1</i> N-terminally fused with eGfp flanked by strong constitutively active promoter P <sub>otef</sub> and transcriptional terminator T <sub>nos</sub> upstream and downstream, respectively. This construct retains the 65 bp 5' UTR of <i>aox1</i> . The ORF and 5' UTR were amplified by PCR using oMF894/oDD808 and pAox1-Gfp_NatR as template.
pP <sub>otef</sub> : <i>aox1</i> -Gfp_CbxR	2768	CbxR for integration at <i>ip<sup>S</sup></i> locus	Plasmid for ectopical integration and expression of <i>aox1-Gfp</i> . Contains 1.3 kb ORF of <i>aox1</i> N-terminally fused with eGfp flanked by strong constitutively active promoter P <sub>otef</sub> and transcriptional terminator T <sub>nos</sub> upstream and downstream, respectively. The ORF were amplified by PCR using oMF894/oDD809 and pAox1-Gfp_NatR as template.

## DNA oligonucleotides used in this study.

Designation	Nucleotide sequence (5' --> 3')	Remarks
oRL1400	GGTCTCGCCTGCAATATTCCTGAGATAGTCGTTGAGG	<i>aox1</i> u2
oRL1401	GGTCTCCAGGCCGGTTACTGGCTTGGGCTG	<i>aox1</i> u3
oRL1402	GGTCTCCGGCCCTGCTTTCCAACCTGGATTCTG	<i>aox1</i> d1
oRL1403	GGTCTCGCTGCAATATTTTTCCCATGAGATGCTGC	<i>aox1</i> d2
oRL1425	AATGGCCGCGTTGGCCGCAGCGGTCTTTTCGGCCGC	<i>aox1</i> u3-fus
oMF894	TCGCAAGACCGCAACAG	
oDD808	GGCGAATTCAGACTTTTAGCAACCATAACAAAGC	Ectopic with 5'UTR
oDD809	CGGGAATTCATGTACGTTAGTACGCCCATC	Ectopic without 5'UTR
oRL1399	GTTCAACACGTCCGGAGG	<i>aox1</i> u1
oRL1404	CGCTGTTGCTCCATTCGG	<i>aox1</i> d3
oRL1405	AGGTACCCGGACCACAAC	<i>aox1</i> p1
oRL1406	GATTGAGCCAACCGTCGG	<i>aox1</i> p2

## Supplementary references

1. Banuett F, Herskowitz I. Different alleles of *Ustilago maydis* are necessary for maintenance of filamentous growth but not for meiosis. *Proceedings of the National Academy of Sciences of the United States of America*. 1989;86(15):5878-82.
2. Brachmann A, Weinzierl G, Kamper J, Kahmann R. Identification of genes in the bW/bE regulatory cascade in *Ustilago maydis*. *Molecular microbiology*. 2001;42(4):1047-63.
3. Terfruchte M, Joehnk B, Fajardo-Somera R, Braus GH, Riquelme M, Schipper K, et al. Establishing a versatile Golden Gate cloning system for genetic engineering in fungi. *Fungal genetics and biology : FG & B*. 2014;62:1-10.