

# Supporting Information

## 3D-e-Chem-VM: Structural Cheminformatics

## Research Infrastructure in a Freely Available

## Virtual Machine

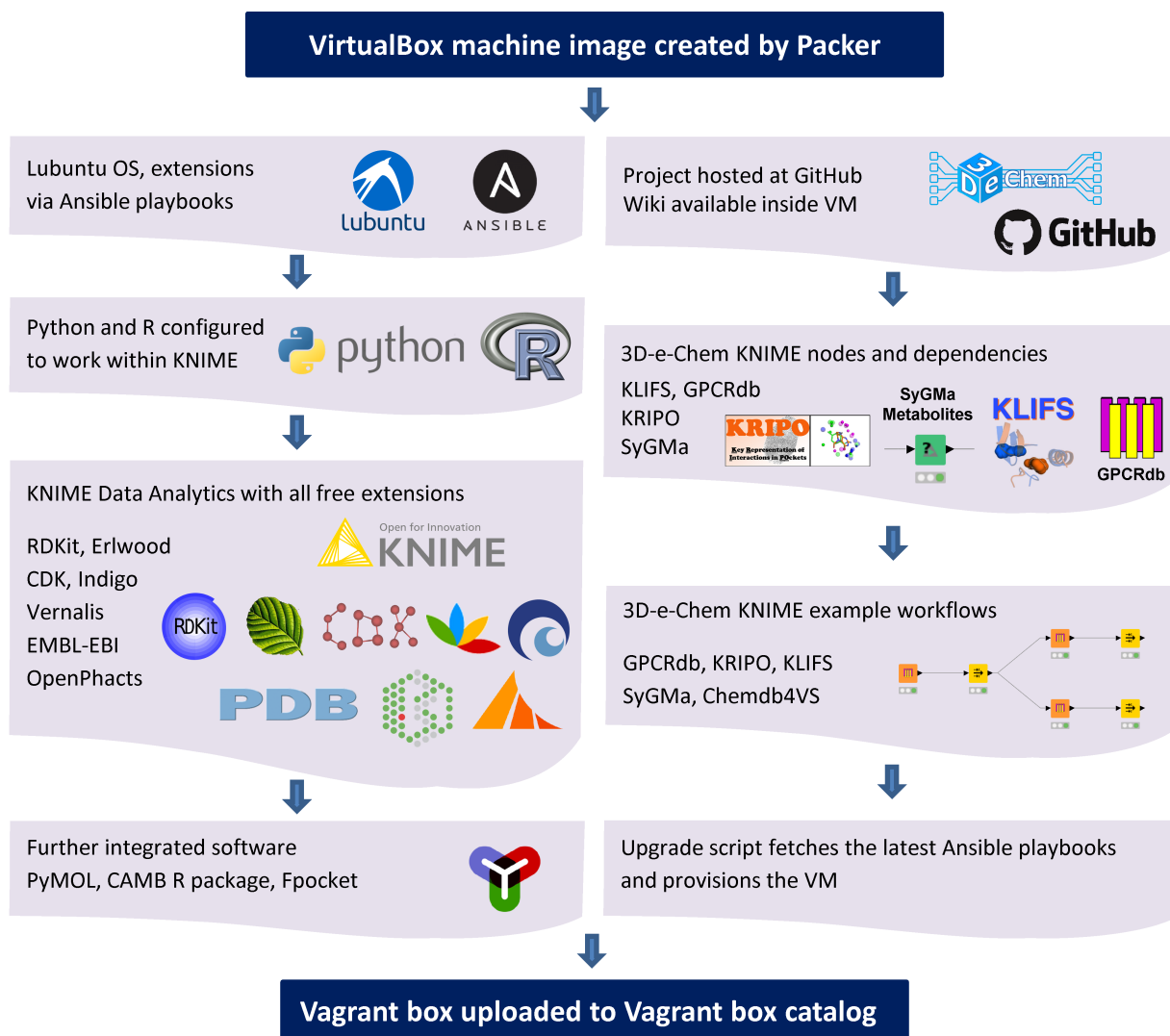
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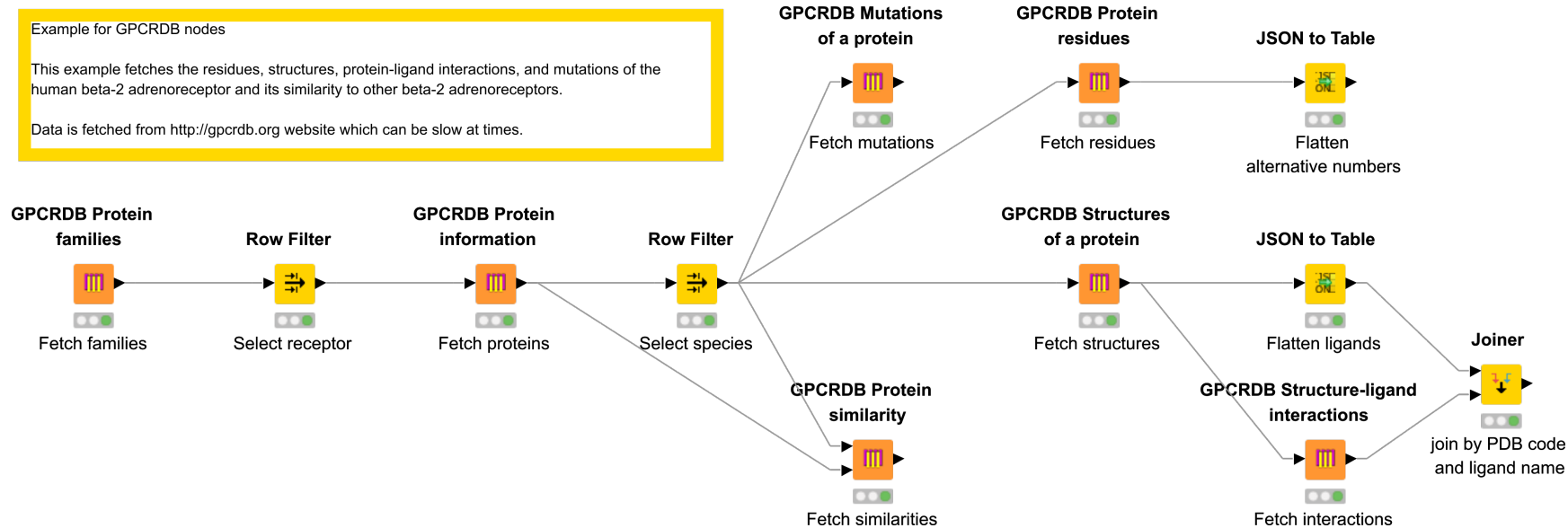
*# R.McG, S.V. and M.V. contributed equally*

Corresponding Author: \* R.McG. e-mail: ross.mcguire@bioaxisresearch.com; S.V. e-mail:

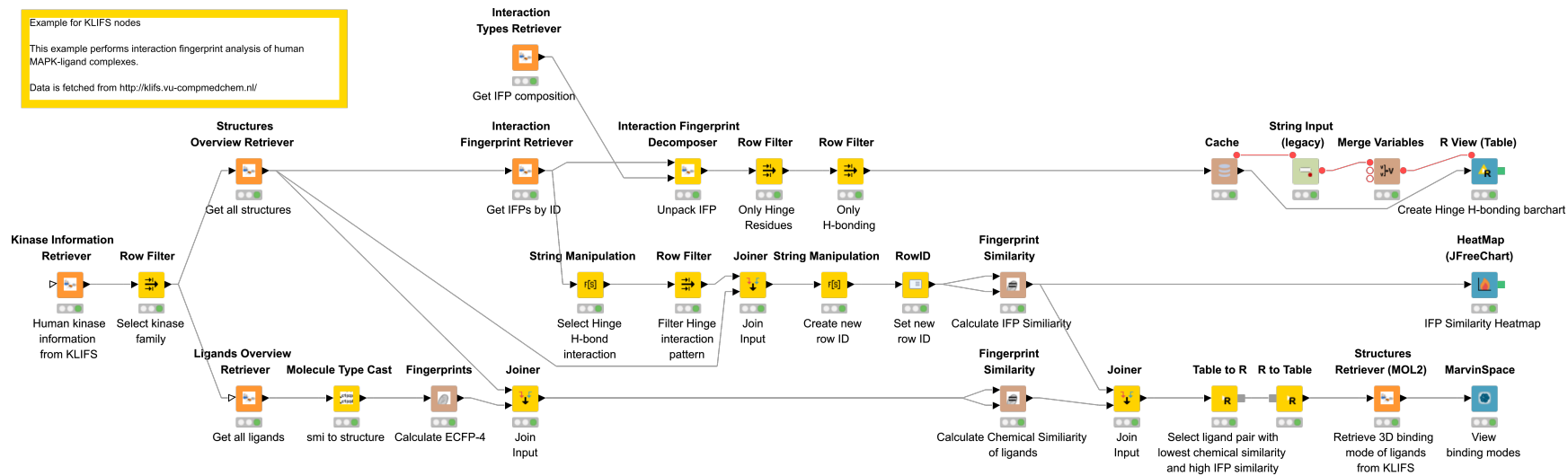
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**Figure S1** Contents of the 3D-e-Chem-VM virtual machine containing publicly available open source software and tools (left) complemented by novel KNIME nodes and workflows (right).



**Figure S2** Full version of GPCRdb nodes example workflow



**Figure S3** Full version of KLIFS nodes example workflow.

Example of using KRIPO data files using the KRIPObd python package.  
 Also shown how to use fragments db using database nodes.  
 For more information see <https://github.com/3D-e-Chem/kripodb>

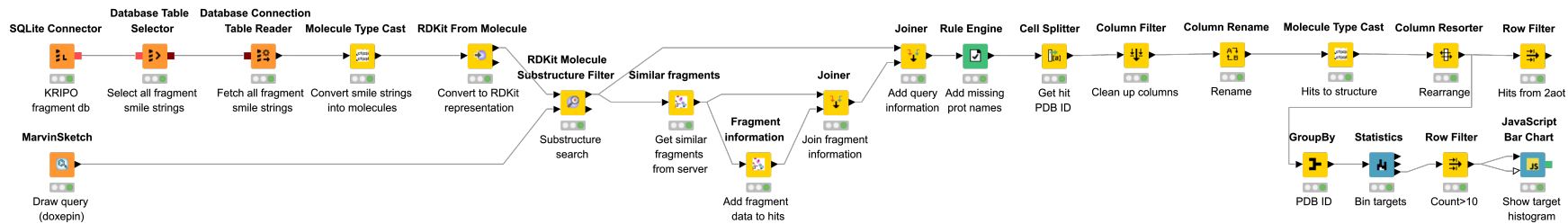
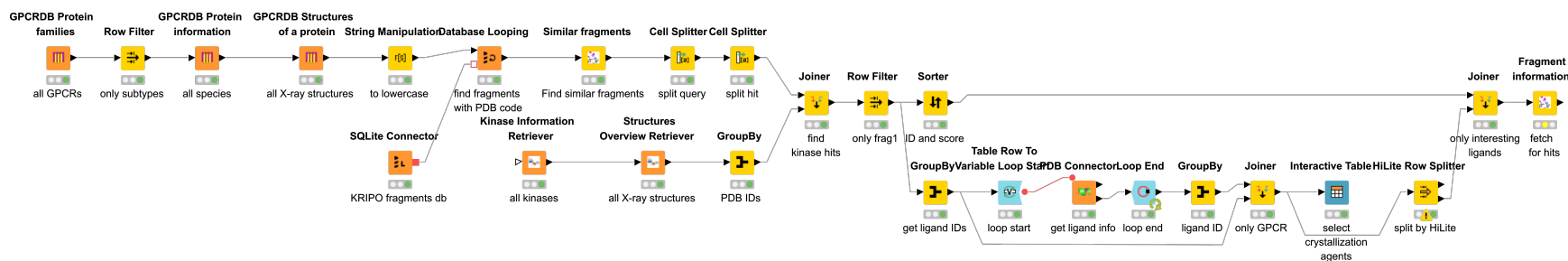
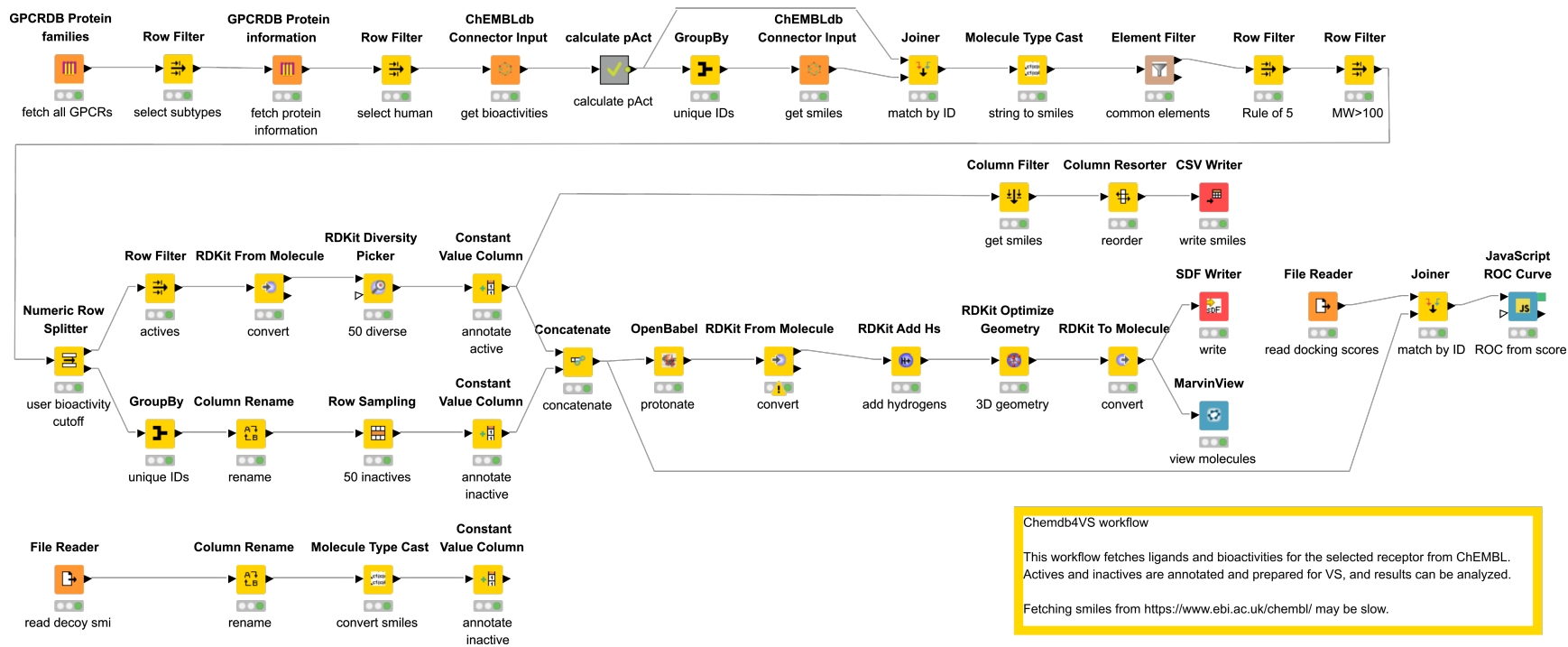


Figure S4 Full version of KRIPO nodes example workflow.



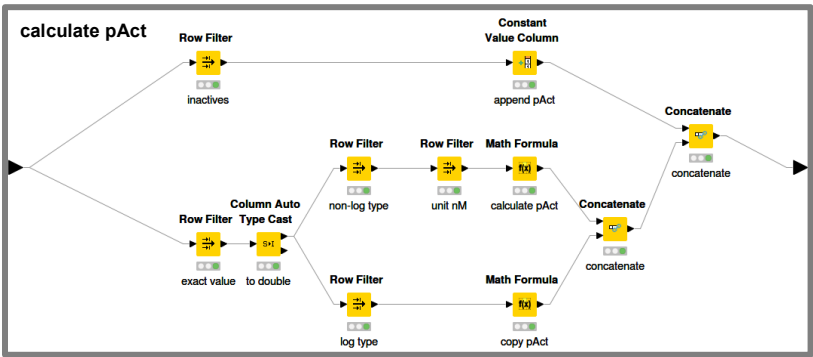
**Figure S5** Full version of an integrated 3D-e-Chem workflow for the identification of similar GPCR and kinase binding pockets. First the GPCRdb and KLIFS nodes are used to fetch all experimentally determined structures of ligand-protein complexes in the two drug target families. The KRIPO nodes are subsequently used to assess the structure-based pharmacophore similarity between all GPCR and kinase binding sites, yielding 1428 similar GPCR-kinase pairs (modified Tanimoto coefficient  $> 0.5$ , excluding crystallization agents). Although the scope of this work was not to examine all kinase-GPCR associations in depth, this analysis provides information on potential cross-reactivities between kinase and GPCR ligands that can be used for off-target identification, ligand repurposing, or the discovery of ligands with a desired GPCR-kinase polypharmacological profile.<sup>1</sup> The KRIPO pharmacophore analysis identified the similar ergotamine bound serotonin 5-HT<sub>2B</sub> receptor (PDB: 4ib4) and Sorafenib bound p38 $\alpha$  MAP kinase (PDB: 3heg) binding site pair (modified Tc = 0.55), which is consistent with the recent experimental identification of sorafenib as a high affinity 5-HT<sub>2B</sub> ligand ( $K_i = 56$  nM).<sup>2</sup> The pharmacophore-based similarity search furthermore identified doxepin bound histamine H<sub>1</sub> receptor binding site (PDB: 3rze) and the maternal embryonic leucine zipper kinase (MELK, PDB: 4umt) as targets that share acidic and aromatic/hydrophobic binding site features<sup>3, 4</sup> and can potentially be targeted by dual ligands.

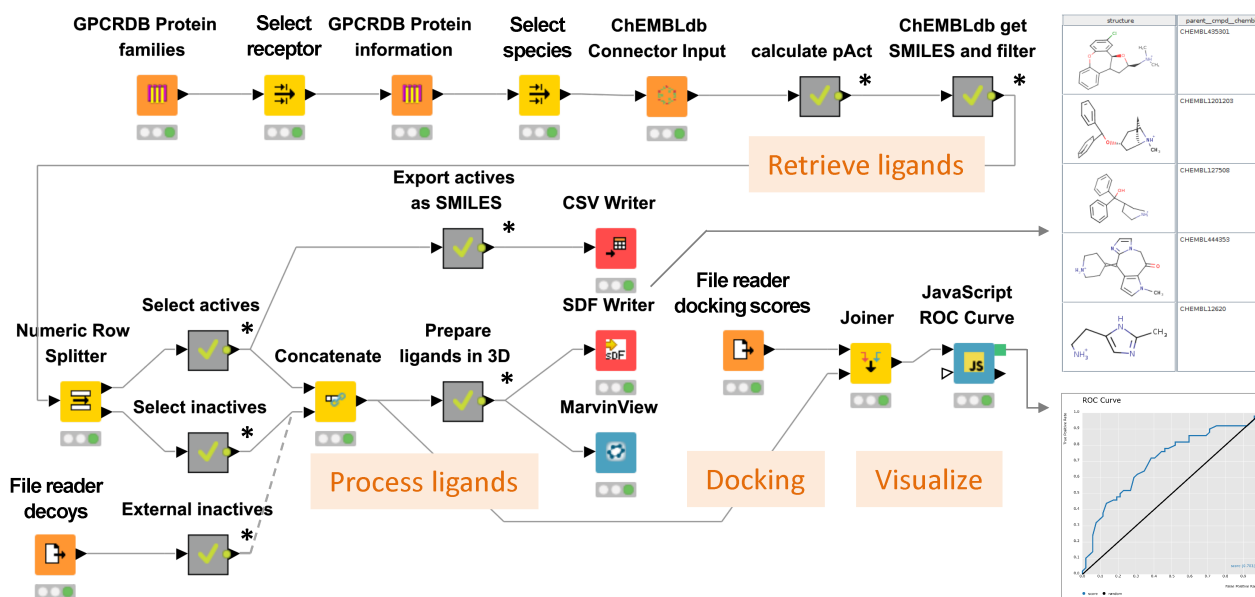


**Chemdb4VS workflow**

This workflow fetches ligands and bioactivities for the selected receptor from ChEMBL. Actives and inactive are annotated and prepared for VS, and results can be analyzed.

Fetching smiles from <https://www.ebi.ac.uk/chembl/> may be slow.





**Figure S6** Full and small versions of Chemdb4VS workflow and the “calculate pAct” metanode expanded. This workflow is designed to collect experimentally supported ligand data sets for the retrospective validation of virtual screening methods. In contrast to traditional decoy collection methods where decoys are only assumed to be inactive (as e.g. in the DUD-E<sup>5</sup>) the workflow collects both actives and known inactive from the ChEMBL database in order to use validated datasets for the evaluation of virtual screens. In the example workflow, the novel GPCRdb nodes were utilized in combination with a ChEMBLdb Connector Input node for the download of all bioactivity data available for a specified receptor. Bioactivity data are filtered and processed separately for non-logarithmic ( $EC_{50}$ ,  $IC_{50}$ ,  $K_i$ ,  $K_D$ ,  $K_B$ ) and logarithmic ( $pK_i$ ,  $\log K_i$  etc.) activity types and for DRUGMATRIX assays, and the pActivity value is calculated for the data points passing the filters. The user can set the active/inactive bioactivity cutoff (in nM) in a Numeric Row Splitter node. SMILES strings for the filtered ChEMBL IDs are downloaded and are filtered for druglike properties and their number is reduced to a user defined limit. Representative actives are selected by the RDKit Diversity Picker node and inactive are sampled randomly. Alternatively, external ligands can be imported and decoy sets can be created by for example using the DUD-E server.<sup>5</sup> Ligands are protonated using the OpenBabel node and their three-dimensional geometries are optimized by the RDKit Optimize Geometry node. The results of an external docking simulation of the generated sets are imported, and based on the active/inactive annotation a ROC curve is plotted using the JavaScript ROC Curve node.



## References

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