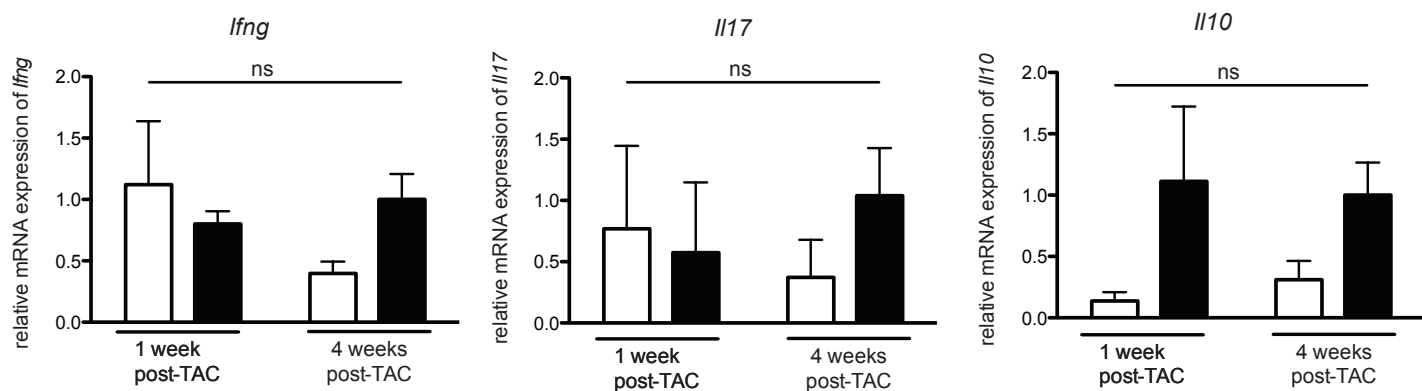
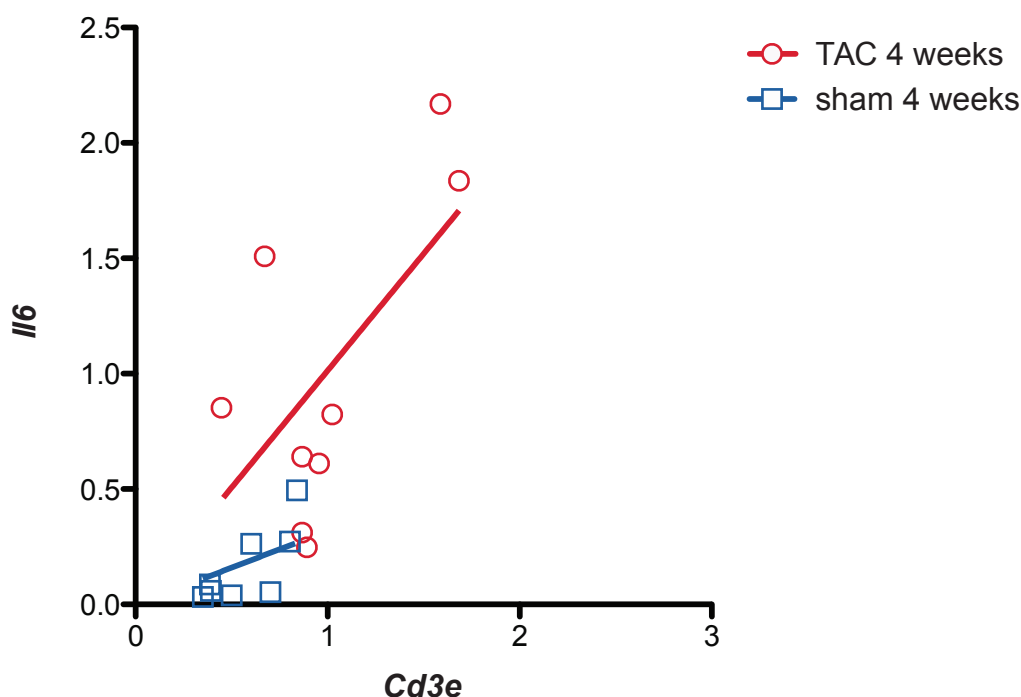
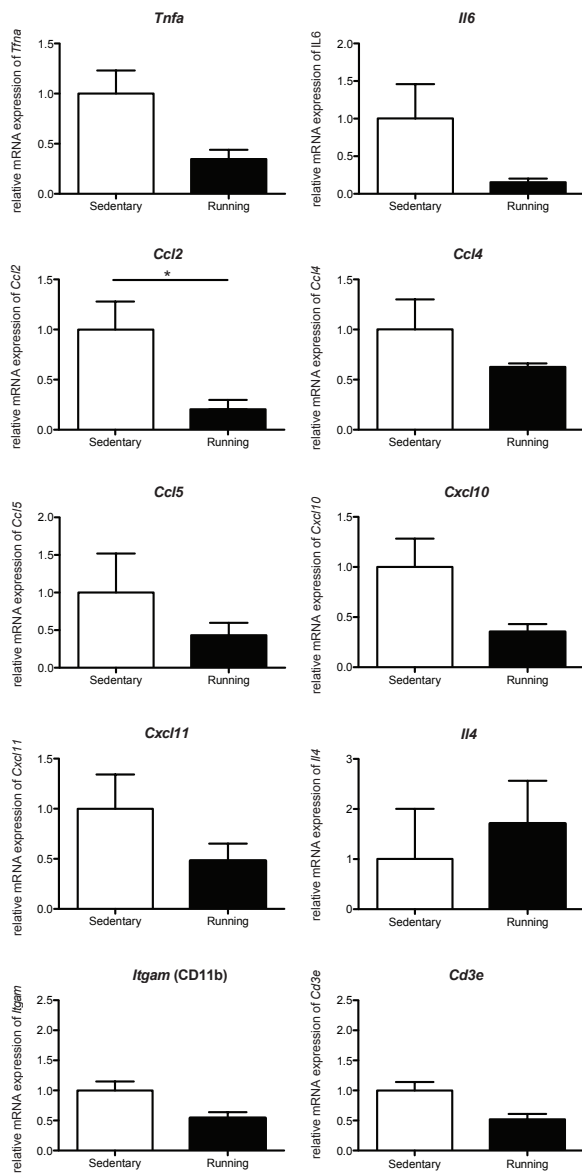


a**b****IL-6 / CD3e linear regression****Supplementary Figure 1**

(a) Characterization of the inflammatory signature in hypertrophic left ventricle of mice. Gene expression analysis (TaqMan real-time qPCR) of mediators of inflammation within the left ventricle of C57BL6/J mice. Relative mRNA expression in sham-operated control mice (white bars) and TAC-operated mice (black bars) at 1 and 4 weeks after surgery, internally normalized to 18s rRNA expression. *Ifng*, *Il17* and *Il10*. Values are mean \pm SEM (n=7-9). Two-way ANOVA, Bonferroni post-test; ns, not significant. **(b) Linear regression of IL-6 and CD3e mRNA expression in left ventricles of TAC and sham-operated mice.** red circles and line: Linear regression between *Cd3e* and *Il6* expression in left ventricle of C57BL6/J TAC-operated mice 4 weeks after operation (n=9). Linear regression test; p value = 0.0002. blue squares and line: Linear regression between *Cd3e* and *Il6* expression in left ventricle of C57BL6/J sham-operated mice 4 weeks after operation (n=7). Linear regression test; p value = 0.0037.

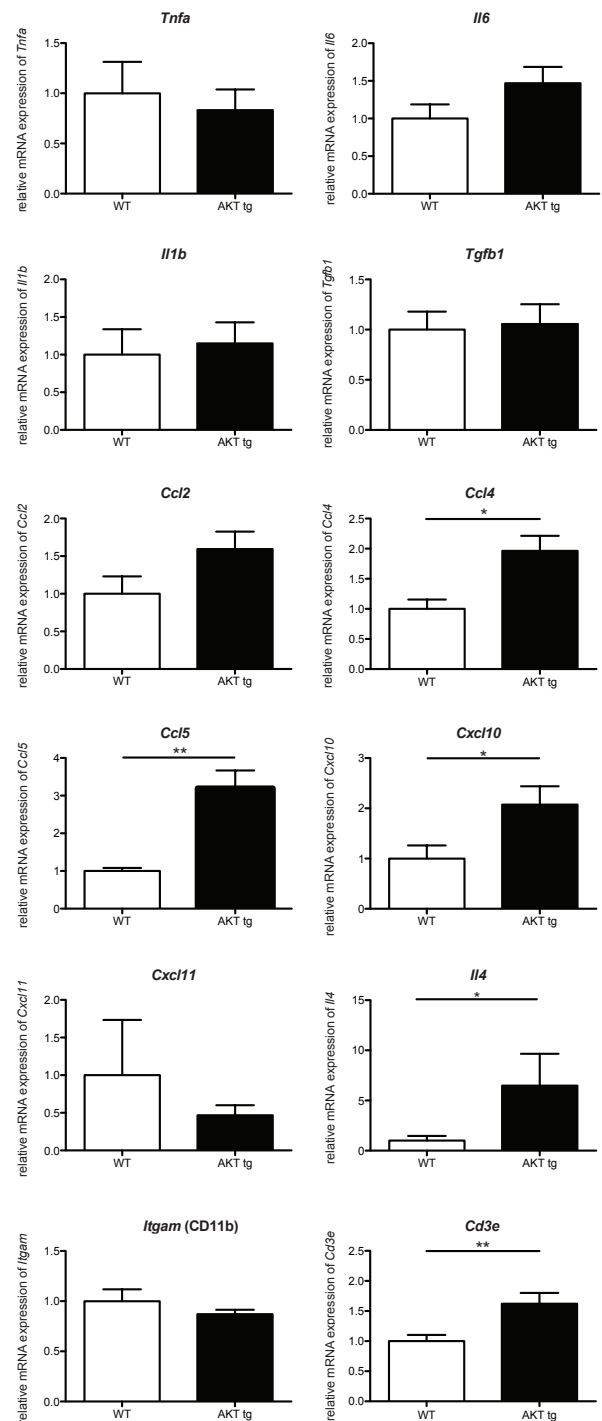
Exercise-trained mice

a



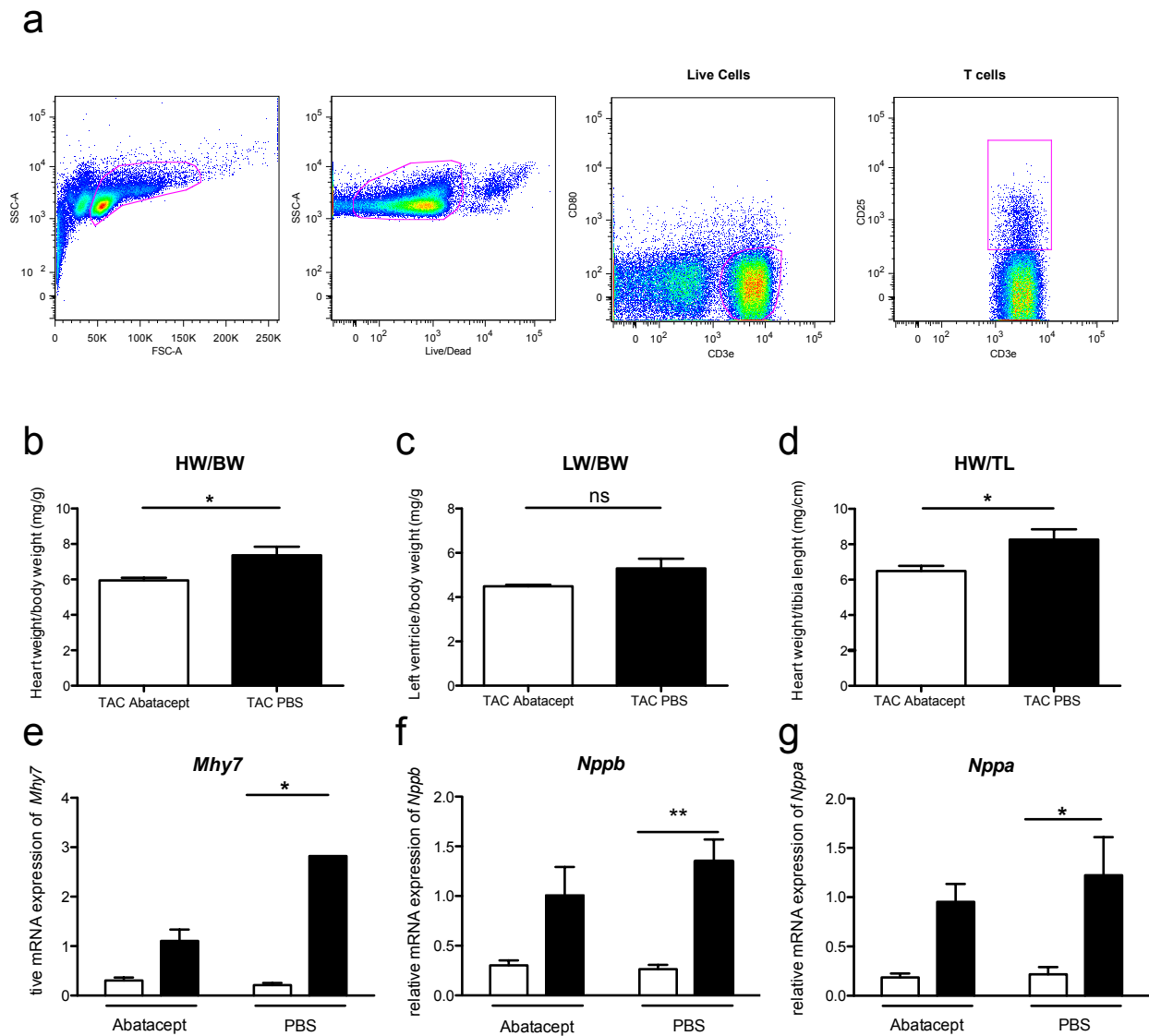
AKT-transgenic mice

b



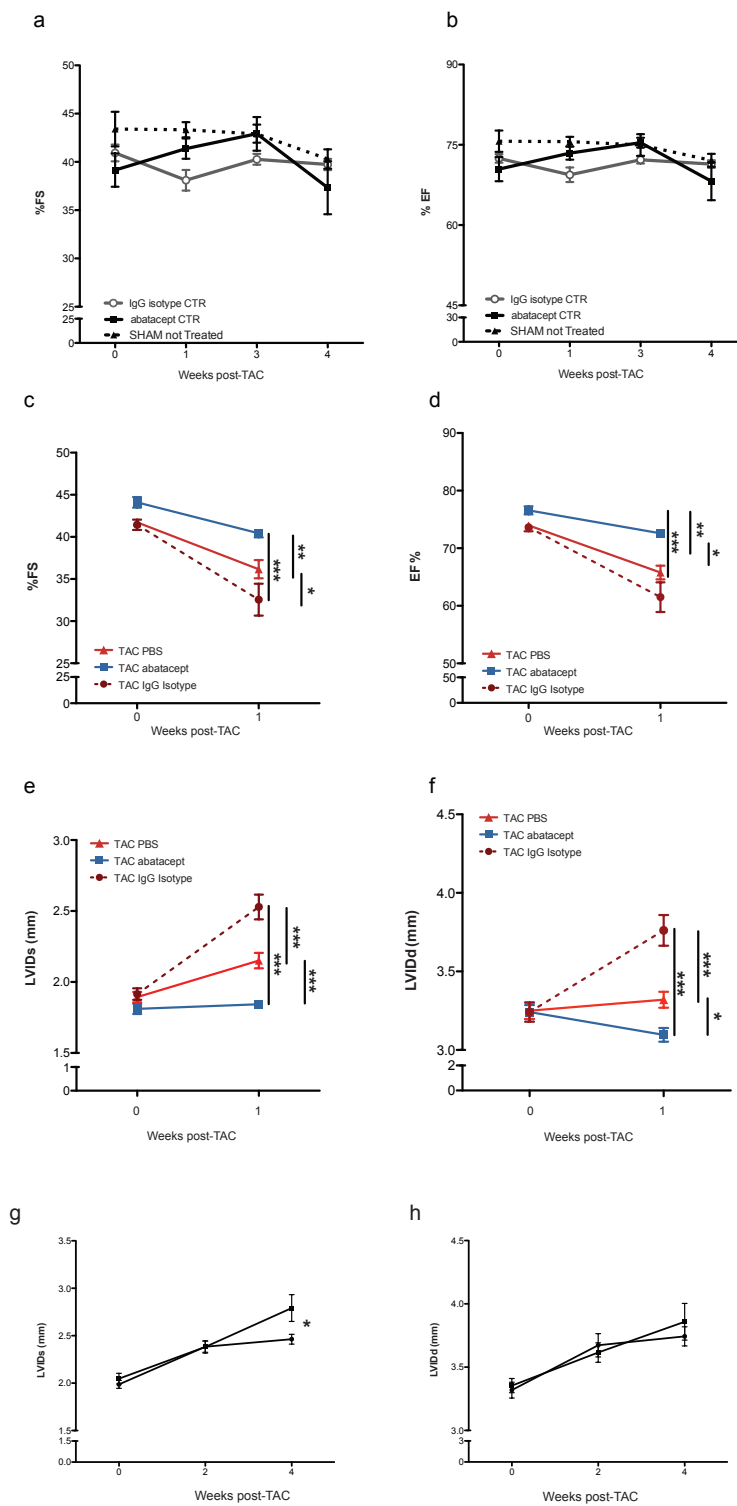
Supplementary Figure 2

Immune mediators in models of physiological hypertrophy (a) Gene expression analysis of mediators of inflammation in the left ventricle of exercise-trained mice (running) compared to sedentary mice (sedentary). Bars show relative mean mRNA expression, internally normalized to 18s rRNA expression. The expression of *Tnfa*, *Il6*, *Ccl4*, *Ccl5*, *Cxcl10*, *Cxcl11*, *Il4*, *Itgam* (CD11b), or *Cd3e* did not change significantly, whilst *Ccl2* significantly decreased in the trained mice group. Values are mean \pm SEM (n=3). Unpaired t-test: *, p-value <0.05. **(b)** Gene expression analysis of inflammatory mediators by TaqMan real-time qPCR in the left ventricle of 8-week-old Akt transgenic mice (Akt-Tg, white) (n=6) compared to wild-type mice (WT, black) (n=7). Bars show relative mean mRNA expression internally normalized to 18s rRNA expression. The relative expression of *Tnfa*, *Il6*, *Il1b*, *Tgfb1*, *Ccl2*, *Cxcl11*, and the innate cell marker *Itgam* (CD11b) did not increase, whilst *Ccl4*, *Ccl5*, *Cxcl10*, *Il4*, and *Cd3e* increased significantly. Values are indicated as mean \pm SEM. Mann-Whitney test: *, p-value <0.05; **, p-value <0.01.



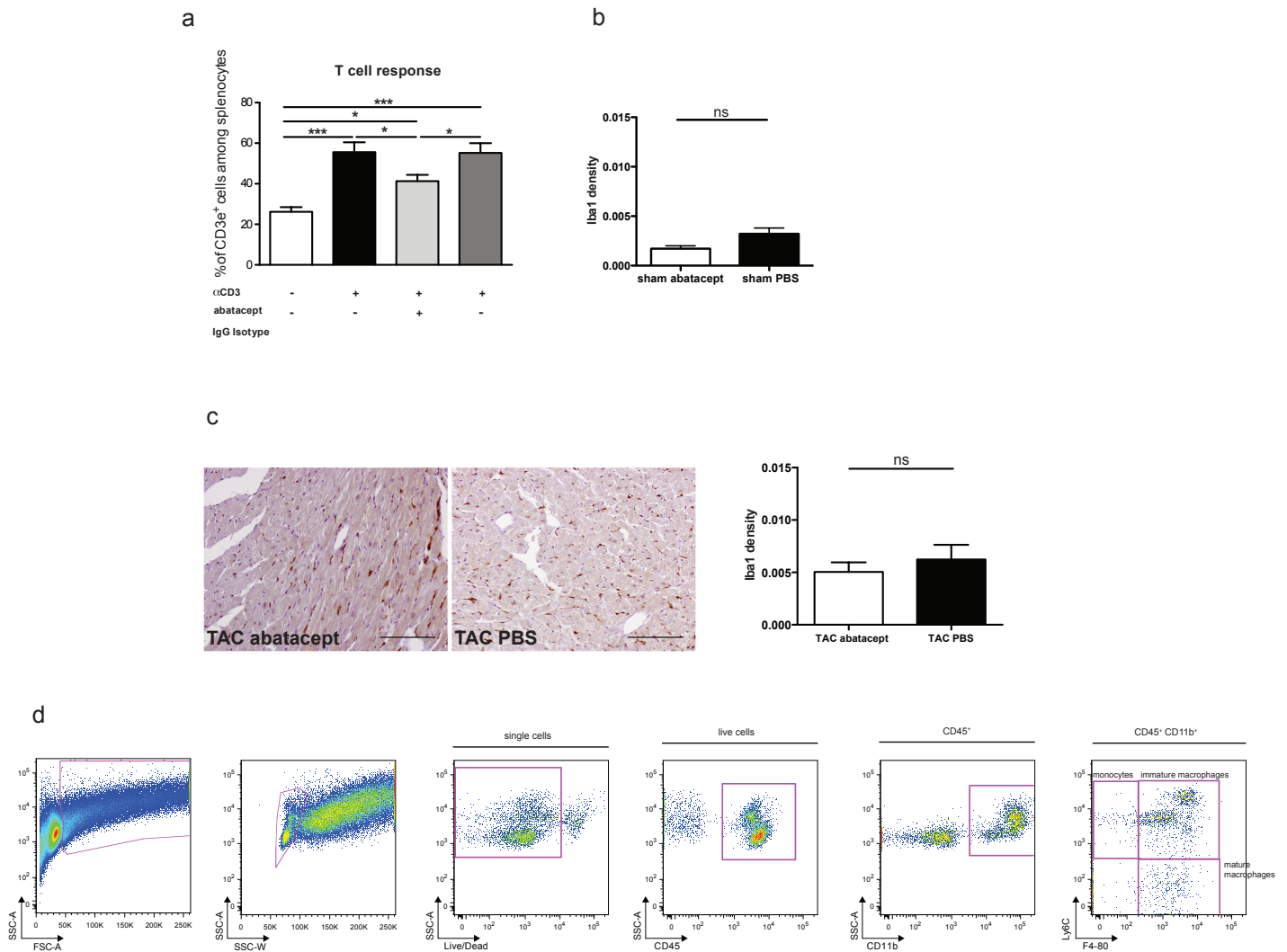
Supplementary Figure 3

Gating strategy for FACS analysis of T cell activation. (a) Cells were gated on forward and side scatter, and dead cells were excluded. The CD80⁺ population corresponded to autofluorescent cells, and hence was excluded. CD3e-expressing cells were selected and CD25 expression was assessed. **Progression of cardiac hypertrophy is limited by abatacept.** (b) Heart weight body weight (mg/g), (c) left ventricle body weight (mg/g), (d) heart weight tibia length (mg/cm) ratios in TAC-operated mice treated with abatacept or PBS, 4 weeks post operation. Values are indicated as mean \pm SEM (n=5-7). Unpaired t-test: *, p-value <0.05. Relative gene expression by real-time qPCR internally normalized to 18S for genes expressing (e) β -myosin (*Mhy7*), (f) brain natriuretic peptide (*Nppb*) and (g) atrial natriuretic factor (*Nppa*) in the same mice. Values are mean \pm SEM (n=4-7). Two-way ANOVA with Bonferroni post-test; *, p-value <0.05; **, p-value <0.01.



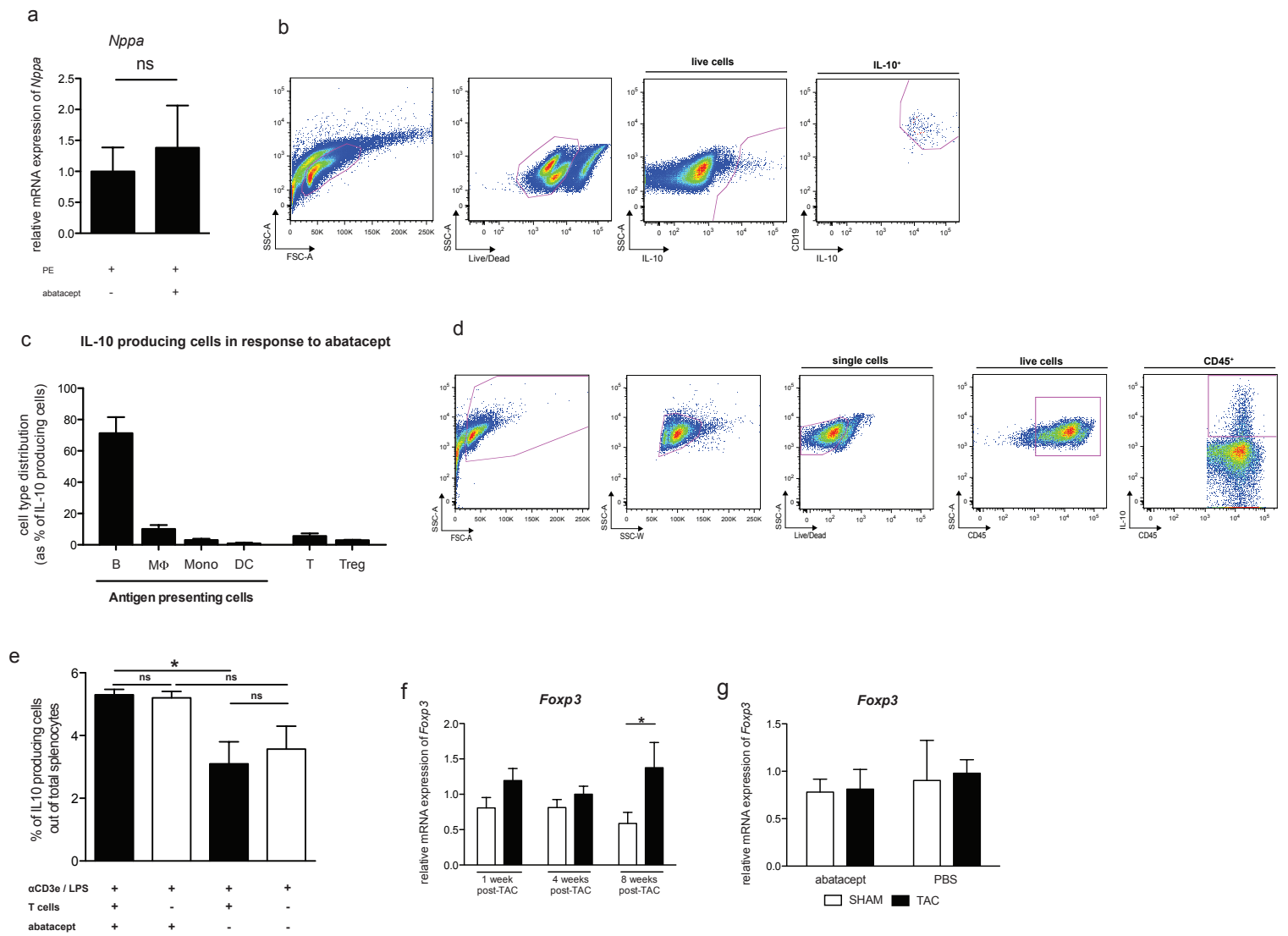
Supplementary Figure 4

Heart functionality is unchanged in non-operated mice after abatacept or human IgG isotype treatment. (a) Fractional shortening (%FS) and **(b)** ejection fraction (%EF) in non-operated mice treated with abatacept or human IgG isotype control, or sham-operated, non-treated mice (included for comparison). Values are mean \pm SEM (n=3–4). Two-way ANOVA with Bonferroni post-test; no significant differences observed. **(c)** Fractional shortening (%FS), **(d)** ejection fraction (%EF), **(e)** left ventricle internal dimension in systole (LVIDs) and **(f)** left ventricle internal dimension in diastole (LVIDd) in TAC-operated mice treated with abatacept, PBS or human IgG isotype control for 1 week starting 2 days after operation. Values are mean \pm SEM (n=6–9). Two-way ANOVA with Bonferroni post-test; *, p value <0.05; **, p value <0.01; ***, p value <0.001. **(g)** left ventricle internal dimension in systole (LVIDs) and **(h)** left ventricle internal dimension in diastole (LVIDd) in TAC-operated mice treated with abatacept or PBS for 2 weeks starting 2 weeks after operation. Values are mean \pm SEM (n=7–9). Two-way ANOVA with Bonferroni post-test; *, p value <0.05.



Supplementary Figure 5

Abatacept attenuates T cell responses *in vitro* (a) Total splenocytes of 8 week-old C57BL/6J mice activated with anti-CD3 and cultured with 20 μ g/ml abatacept or IgG isotype control for 72 hours were analyzed by FACS for CD3e expression. Bars show mean \pm SEM of 4 independent experiments (n=4). One way-ANOVA repeated measures test with Tukey's post-test; *, p-value <0.05; ***, p-value <0.001. (b) Statistical analysis of AIF-1(Iba1) immunohistochemical staining in left ventricles of sham-operated mice injected with abatacept (white bar) or PBS (black bar) at 4 weeks post-operation. AIF-1 density plotted as mean \pm SEM. Unpaired t-test. (n=2). (c) Representative images and statistical analysis of AIF-1 immunohistochemical staining (brown coloration; original magnification 20X; scale bar = 100 μ m) in left ventricles of TAC mice at 4 weeks post-operation, treated with abatacept or PBS. AIF-1 density plotted as mean \pm SEM; TAC abatacept (white bars); TAC PBS (black bars). Unpaired t-test. (n=2). (d) Gating strategy for flow cytometry analysis of cardiac single cell suspensions from TAC operated mice 1 week after operation, corresponding to data in Fig. 4d,e. Cells were gated based on forward and side scatter, doublets were excluded and live single cells were gated. CD45 and CD11b-expressing cells were selected and identified on the basis of Ly6C and F4-80 expression: Ly6C⁺ F4-80⁻ monocytes, Ly6C⁺ F4-80⁺ immature macrophages and Ly6C⁻ F4-80⁺ mature macrophages.



Supplementary Figure 6

(a) Evaluation of direct effects of abatacept on cardiomyocytes. Real-time qPCR of *Nppa*, in mouse neonatal cardiomyocytes cultured with 100 μ M of phenylephrine (PE) and 20 μ g/ml of abatacept for 48 hours. Bars show mean \pm SEM of relative mRNA expression, internally normalized to 18s rRNA (n=6). Unpaired t-test; ns, not significant. **Abatacept induces *in vitro* production of IL-10** **(b)** Gating strategy for FACS analysis of IL-10 producing cells in presence of abatacept. Cells were gated based on forward and side scatter, dead cells were excluded and IL-10 producing cells were gated. Specific population frequencies were then analyzed (CD19⁺ B cells, CD11c⁺ dendritic cells, CD11b⁺ monocytes and myeloid-derived cells, F4/80⁺ macrophages, CD3e⁺ T cells, and CD3e⁺ Foxp3⁺ Treg cells). **(c)** Total splenocytes of 8 week-old C57BL/6J mice activated with anti-CD3 and cultured with 20 μ g/ml abatacept were analyzed to identify the cellular source of IL-10 production, gated as in (b). Mean \pm SEM of 3 independent experiments (n=3). **(d)** Gating strategy for FACS analysis of IL-10 producing leukocytes in response to abatacept. Cells were gated based on forward and side scatter, doublets and dead cells were excluded. CD45⁺ cells were identified and CD45⁺ IL-10-producing cells were assessed. **(e)** Total splenocytes or T cell-depleted splenocytes of 8 week-old C57BL/6J mice were activated with 2 μ g/ml anti-CD3 and 5 μ g/ml of LPS. After 48 hours in culture with or without 20 μ g/ml abatacept were analyzed by flow cytometry for IL-10 production. Bars show mean \pm SEM of 3 independent experiments (n=3) of the percentage of IL10 producing cells out of total splenocytes. Two-way ANOVA with Bonferroni post-test; *, p-value <0.05. **Presence of regulatory T cells in TAC- and sham-operated mice. (f)** Real-time qPCR for *Foxp3* in the left ventricle of C57BL6/J mice, 1, 4, and 8 weeks after TAC or sham operation. Bars show mRNA expression internally normalized to 18s rRNA expression. Mean \pm SEM (n=7–9). Two-way ANOVA with Bonferroni post-test; *, p-value <0.05. **(g)** Real-time qPCR for *Foxp3* for the same samples as in Fig. 4f. Bars show mRNA expression internally normalized to 18s rRNA expression. Mean \pm SEM (n=7–9). Two-way ANOVA with Bonferroni post-test; ns, not significant.

SHAM

TAC

	Basal (n=3)	1 week post- sham (n=3)	3 weeks post- sham (n=3)	4 weeks post- sham (n=3)	Basal (n=7)	1 week post-TAC (n=7)	3 weeks post-TAC (n=7)	4 weeks post-TAC (n=7)
Age (wks)	8.7±0.0	9.6±0.1	11.7±0.1	12.6±0.0	8.6±0.1	9.6±0.1	11.6±0.1	12.6±0.0
Weight(g)	22.0±1.0	22.4±0.8	23.6±0.9	23.9±1.1	22.8±1.6	22.3±1.3	24.0±1.5	24.6±1.5
HR M - mode (bpm)	545.7±71.1	507.7±93.9	549.7±66.0	598.0±29.0	577.0±66.4	530.1±48.7	517.9±87.1	606.0±36.6
% FS	43.4±3.1	43.3±1.4	42.9±3.0	40.3±1.7	40.5±2.4	37.7±1.2 [‡]	33.2±1.5 ^{‡§}	30.6±2.2 ^{‡§}
% EF	75.7±3.5	75.6±1.5	75.0±3.5	72.1±2.1	72.3±3.6	69.0±1.6 [‡]	62.7±2.1 ^{‡§}	58.8±3.5 ^{‡§}
LVIDd (mm)	3.4±0.1	3.4±0.1	3.5±0.1	3.4±0.0	3.4±0.1	3.4±0.2	3.7±0.1 ^{*◇}	3.7±0.3 [‡]
LVIDs (mm)	1.9±0.1	1.9±0.1	2.0±0.2	2.0±0.1	2.0±0.2	2.1±0.1 [*]	2.5±0.1 [‡]	2.6±0.3 ^{*‡}
IVSd (mm)	0.7 ± 0.0	0.8 ± 0.1	0.7±0.1	0.8±0.1	0.7±0.0	0.9±0.1 ^{*‡}	0.9±0.1 ^{*‡}	1.0±0.1 ^{#‡}
IVSs (mm)	1.1±0.1	1.2±0.0	1.2±0.1	1.2±0.1	1.2±0.1	1.3±0.1 [‡]	1.3±0.1 ^{*◇}	1.4±0.1 ^{*◇}
LVPWd (mm)	0.7±0.1	0.8±0.0	0.8±0.1	0.8±0.1	0.8±0.1	0.9±0.1	0.9±0.1 ^{#‡}	0.9±0.1 [‡]
LVPWs (mm)	1.2±0.1	1.3±0.0	1.2±0.1	1.2±0.1	1.2±0.1	1.3±0.1	1.3±0.1 ^{*◇}	1.3±0.1 [‡]

- ‡ p<0.05 TAC versus sham at the same time point
* p<0.01 TAC versus sham at the same time point
◇ p<0.001 TAC versus sham at the same time point
‡ p<0.05 TAC basal versus TAC at each time point
§ p<0.01 TAC basal versus TAC at each time point
p<0.001 TAC basal versus TAC at each time point

Supplementary Table 1: Hemodynamic parameters for TAC- or sham-operated mice at 1, 3 and 4 weeks post-operation

	Sham								TAC							
	Abatacept				PBS				Abatacept				PBS			
	Basal (n=8)	1 week post-sham (n=7)	3 weeks post-sham (n=8)	4 weeks post-sham (n=8)	Basal (n=8)	1 week post-sham (n=5)	3 weeks post-sham (n=7)	4 weeks post-sham (n=7)	Basal (n=9)	1 week post-TAC (n=8)	3 weeks post-TAC (n=7)	4 weeks post-TAC (n=7)	Basal (n=10)	1 week post-TAC (n=10)	3 weeks post-TAC (n=10)	4 weeks post-TAC (n=10)
Age (wks)	9.0±0.3	9.9±0.0	12.0±0.0	12.9±0.0	9±0.3	10.1±0.0	12.0±0.1	13.0±0.0	8.9±0.0	9.9±0.0	11.9±0.0	12.9±0.0	9.0±0.3	10.1±0.0	12.1±0.0	12.9±0.0
Weight (g)	22.6±1.4	22.1±1.3	23.5±1.5	23.7±1.7	22.6±1.4	21.7±1.4	23.11±2.1	24.0±1.9	22.5±1.6	22.1±1.4	23.4±1.3	24±1.8	22.6±1.4	21.6±1.8	23.6±1.6	24.2±1.4
HR M-mode (bpm)	564.4±75.8	514.0±56.4	602.8±68.6	614.4±44.5	564.4±75.8	579.0±18.9	577.3±69.9	575.2±85.6	556.9±86.1	546.6±55.8	597.9±94.6	561.4±66.0	564.4±75.8	544.7±46.2	575.2±85.6	558.3±60.9
%FS	41.4±3.8	43.0±4.1	42.6±1.5	40.7±2.0	41.4±3.8	43.3±2.5	40.6±2.5	41.4±1.7	42.0±1.5	41.3±3.2*	37.3±1.6* [⊗]	36.5±3.7*	41.4±3.8	34.9±1.3 [◇]	30.7±3.0 [◇]	27.5±3.8 [◇]
%EF	73.2±4.8	75.2±4.8	74.9±1.6	72.8±2.4	73.2±4.8	75.8±2.6	72.7±3	73.8±1.9	74.1±1.8	73.4±3.9 [§]	68.3±2.2 [§] #	67.2±4.8*	73.2±4.8	65.2±1.8 [◇]	58.9±4.6 [◇]	53.9±6.2 [◇]
LVIDd (mm)	3.4±0.1	3.3±0.2	3.3±0.2	3.2±0.2	3.4±0.1	3.1±0.2	3.2±0.1	3.1±0.3	3.4±0.1	3.3±0.2	3.5±0.2 [†] ⊙	3.5±0.1*	3.4±0.1	3.5±0.2 ⁺	3.8±0.3 [◇]	3.8±0.2 [◇]
LVIDs (mm)	2±0.2	1.9±0.2	1.9±0.1	1.9±0.1	2.0±0.2	1.8±0.1	1.9±0.1	1.8±0.2	1.9±0.1	1.9±0.2*	2.2±0.2* [⊗]	2.2±0.2* [⊙]	2.0±0.2	2.3±0.1 [◇]	2.6±0.3 [◇]	2.8±0.3 [◇]
IVSd (mm)	0.8±0.0	0.7±0.1	0.8±0.0	0.8±0.0	0.8±0.0	0.8±0.1	0.8±0.0	0.8±0.0	0.8±0.1	0.9±0.1 [#]	0.9±0.1 [⊙]	1.0±0.0 [#]	0.8±0.0	0.9±0.1	0.9±0.1 ⁺	1.0±0.1 [◇]
IVSs (mm)	1.2±0.1	1.2±0.0	1.3±0.0	1.2±0.0	1.2±0.1	1.3±0.1	1.2±0.1	1.2±0.1	1.2±0.1	1.3±0.0 [#]	1.3±0.1	1.4±0.0 [#]	1.2±0.1	1.3±0.1	1.4±0.1 [▲]	1.3±0.1
LVPWd (mm)	0.8±0.1	0.8±0.1	0.8±0.1	0.8±0.0	0.8±0.1	0.8±0.0	0.8±0.1	0.9±0.1	0.7±0.1	0.9±0.1 [#]	1±0.1 [#]	0.9±0.0 [#]	0.8±0.1	0.9±0.1 [◇]	1.0±0.1 [◇]	1.0±0.1 [◇]
LVPWs (mm)	1.2±0.0	1.3±0.0	1.2±0.1	1.2±0.1	1.2±0.0	1.2±0.0	1.2±0.0	1.2±0.0	1.2±0.1	1.4±0.1 [#]	1.4±0.1 [#]	1.4±0.0 [#]	1.2±0.0	1.4±0.1 [◇]	1.4±0.0 [◇]	1.4±0.1 [◇]

[‡]p<0.05 TAC abatacept versus TAC PBS
[§]p<0.01 TAC abatacept versus TAC PBS
^{*}p<0.001 TAC abatacept versus TAC PBS
[⊙]p<0.05 TAC abatacept versus sham abatacept
[⊗]p<0.01 TAC abatacept versus sham abatacept
[#]p<0.001 TAC abatacept versus sham abatacept
[▲]p<0.05 TAC PBS versus sham PBS
⁺p<0.01 TAC PBS versus sham PBS
[◇]p<0.001 TAC PBS versus sham PBS

Supplementary Table 2: Hemodynamic parameters for TAC- or sham-operated mice treated with abatacept or PBS, at 1, 3 and 4 weeks post-operation

Gene	Forward	Reverse
β-Myosin (<i>Mhy7</i>)	5'-CGCATCAAGGAGCTCACC-3'	5'-CTGCAGCCGCAGTAGGTT-3'
Brain Natriuretic Peptide (<i>Nppb</i>)	5'-GTCAGTCGTTGGGCTGTAAC-3'	5'-AGACCCAGGCAGAGTCAGAA-3'
Atrial Natriuretic Peptide (<i>Nppa</i>)	5'-CACAGATCTGATGGATTCAAGA-3'	5'-CCTCATCTTCTACCGGCATC-3'
18S Ribosomal RNA (18S)	5'-AAATCAGTTATGGTTCCTTTGGTC-3'	5'-GCTCTAGAATTACCACAGTTATCCAA-3'

Supplementary Table 3 : List of primers used.