

SUPPLEMENTARY INFORMATION

Bacterial persistence induced by salicylate via reactive oxygen species
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SUPPLEMENTARY METHODS

Bacterial strains and growth conditions

ΔmarRAB Δrob ΔsoxSR strain

We generated a PCR fragment of the kanamycin resistant gene flanked by FRT regions from pKD13 with extensions homologous to the regions adjacent to the operon or gene ¹.

For deletion of the *marRAB* operon, we used the forward primer: **CCAGCGATCTGTTCAATGAAATTATCCATTGGGTCGCTTAATCCATATG**
GTGTAGGCTGGAGCTGCTTC
and the reverse primer:
AAGCCCCGAGATGTCGGGGCCAGAACAACACTACATAGCGTGTTGATTAT
AATTCCGGGGATCCGTCGACC.

For deletion of the *rob* gene, we used the forward primer:
CGACGGATCGGAATCAGCAGTTCACAGCGTAGATTAATTGGGCGATCTC
CGTGTAGGCTGGAGCTGCTTC
and the reverse primer:
ATGGATCAGGCCGGCATTATTCGCGACCTTTTAATCTGGCTGGAAGGTC
AATTCCGGGGATCCGTCGACC.

For deletion of the *soxSR* genes, we used the forward primer:
AATTACAGGCGGTGGCGATAATCGCTGGGAGTGCGATCAAACCTGCCGAC
GGTGTAGGCTGGAGCTGCTTC
and the reverse primer:
TGTTTATCTTCCAGCAAGCGTGCGCCGGTACCTTCTTCTCCTAAGCGGTC
ATTCCGGGGATCCGTCGACC.

Bold letters indicate the homologous recombination extensions. We then followed the protocol from ¹ using the phage lambda recombinase harbored on the low copy plasmid pSIM6. After recombination, the resistance marker gene was removed using the pCP20 helper plasmid encoding the FLP recombinase.

MarA⁺ strain

The transversion mutations were derived from ², where they are listed as “TV -14 to -18” and “TV +11 to 15.” To construct this strain we synthesized a DNA fragment containing a modified version of the *marRAB* promoter:

GCATCGCATTGAACAAAACCTTGAACCGATTTAGCAAAACGTGGCATCGGTCA
ATTCATTCATTTGACTTATACTTGCCTGTT**ACCTATTATCCCCTGCAACTAATT**
ACGGTAAAGGGCAACTAATGTGAAAAGTACCAGCGATCTGTTCAATGAAATT
ATT.

Underlined letters indicate the MarR binding sites and bold letters indicate the transversion mutations. We introduced this modified promoter via homologous recombination following ¹, then cured the resistance marker. To accomplish this, we ligated the modified *marRAB* promoter given above to the kanamycin resistance gene

flanked by FRT regions from pKD13. The ligated fragment containing the kanamycin resistance gene followed by the modified *marRAB* promoter was then amplified using the forward primer:

GGGGTAAACAAGGATAAAGTGTCACTCTTTAGCTAGCCTTGCATCGCAT
TGTGTAGGCTGGAGCTGCTTC

and the reverse primer:

AATAATTT**CATTGAACAGATCGCTGGT**.

Bold letters indicate the homologous recombination extensions. We followed the protocol for recombination and curing of the kanamycin marker as described above.

SUPPLEMENTARY FIGURES

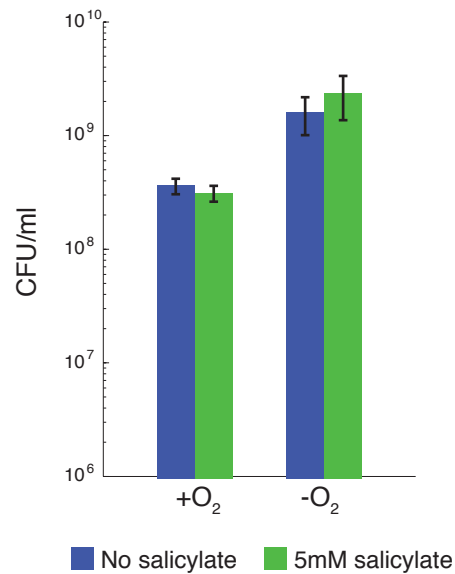


Figure S1: Salicylate exposure does not affect initial cell counts before addition of antibiotics.

Colony forming units per milliliter (CFU/ml) for cultures with and without 5 mM salicylate under aerobic conditions and anaerobic conditions before addition of antibiotics (t = 3 hours in aerobic conditions and t = 4 hours in anaerobic conditions). Error bars show standard errors from n = 12 biological replicates under aerobic conditions and from n = 6 biological replicates under anaerobic conditions.

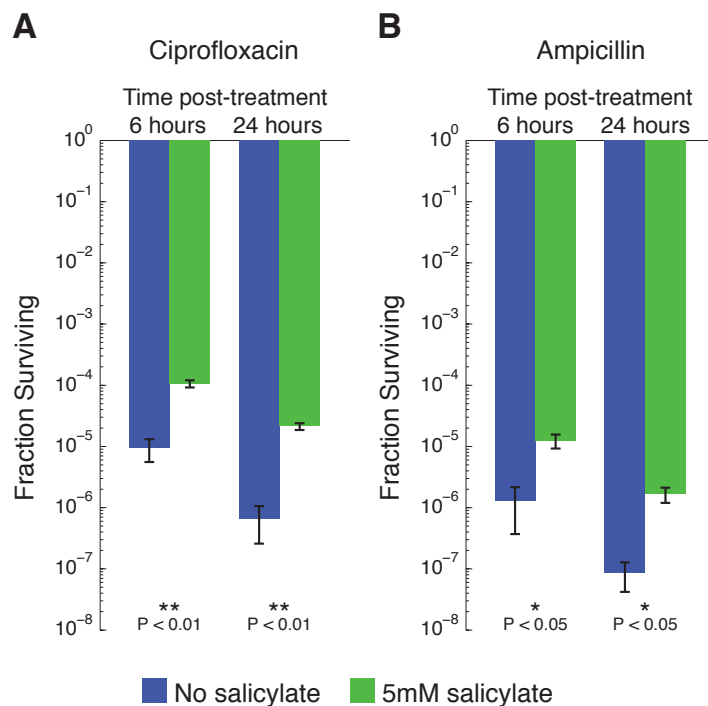


Figure S2: Salicylate increases survival rate after 6 and 24 hours of ciprofloxacin or ampicillin treatment.

Fraction of *E. coli* MG1655 surviving after 6 and 24 hours of (A) 5 $\mu\text{g/ml}$ ciprofloxacin or (B) 100 $\mu\text{g/ml}$ ampicillin treatment with and without 5 mM salicylate. Error bars show standard error of $n = 3$ biological replicates. We used the Student's t-test to test for statistical significance.

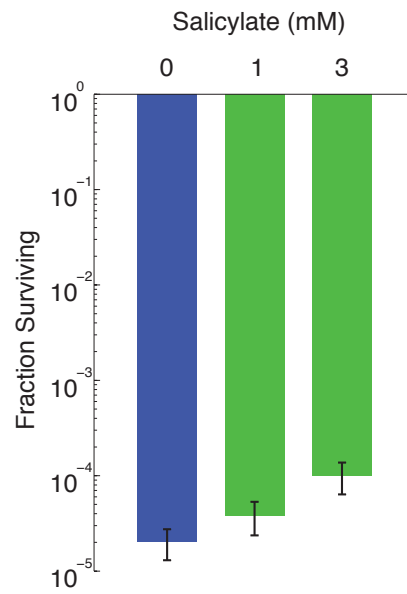


Figure S3: 1 and 3 mM salicylate increase persister levels.

Survival after 6 hours of 5 $\mu\text{g/ml}$ ciprofloxacin treatment with and without 1 or 3mM salicylate. Error bars show standard errors from n = 3 biological replicates.

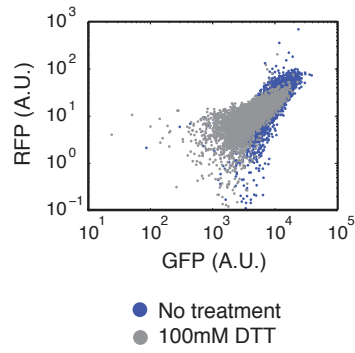


Figure S4: Effect of DTT on membrane potential.

Red vs. green fluorescence scatter plots showing individual cells treated with the membrane potential indicator DiOC2(3) with and without 100 mM DTT.

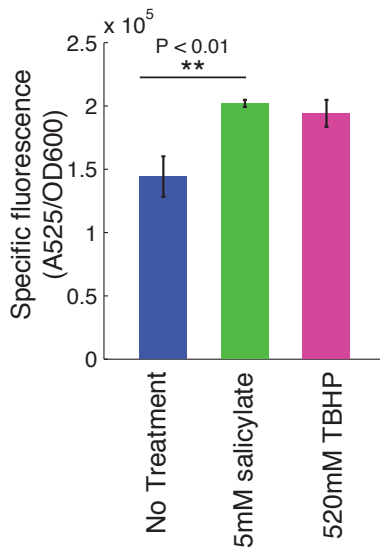


Figure S5: Salicylate induces ROS in $\Delta marRAB \Delta rob \Delta soxSR$ strain.

Green fluorescence divided by OD_{600nm} for $\Delta marRAB \Delta rob \Delta soxSR$ cultures treated with the general ROS indicator carboxy-H₂DCFDA. TBHP is a positive control for ROS. Error bars show standard deviation from $n = 3$ biological replicates. We used the Student's t-test to test for statistical significance.

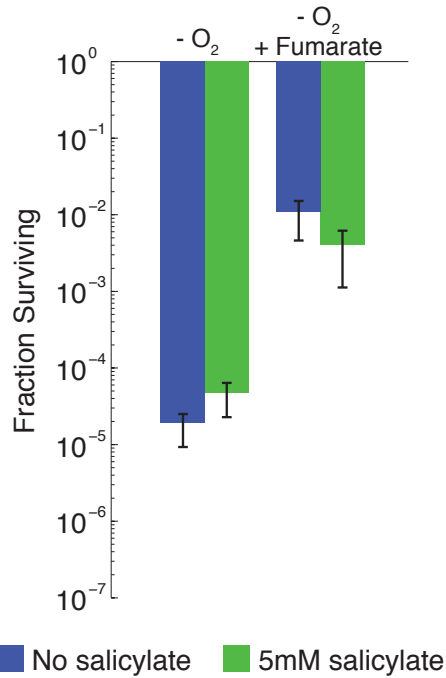


Figure S6: Salicylate-induced persistence in ampicillin requires oxygen.

Survival with and without 5 mM salicylate after 6 hours of 100 $\mu\text{g/ml}$ ampicillin treatment under anaerobic conditions with and without 40 mM fumarate. Error bars show standard errors from $n = 3$ biological replicates.

SUPPLEMENTARY REFERENCES

- 1 Datsenko, K. A. & Wanner, B. L. One-step inactivation of chromosomal genes in *Escherichia coli* K-12 using PCR products. *Proceedings of the National Academy of Sciences of the United States of America* **97**, 6640-6645, doi:10.1073/pnas.120163297 (2000).
- 2 Martin, R. G. & Rosner, J. L. Transcriptional and translational regulation of the *marRAB* multiple antibiotic resistance operon in *Escherichia coli*. *Molecular microbiology* **53**, 183-191, doi:10.1111/j.1365-2958.2004.04080.x (2004).