

Antimalarial Properties of Simplified Kalihinol Analogues

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Supporting Information

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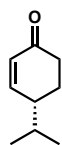
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II. General Experimental Methods

All reactions were performed under an inert atmosphere of argon using oven-dried or flame-dried glassware and Teflon® coated stir bars. Solvents were dried by passage through columns of activated alumina, and *tert*-butyl alcohol was distilled from calcium hydride prior to use. Trimethylsilyl cyanide and methyl vinyl ketone were purified by distillation prior to use. Commercial reagents were used as received unless noted otherwise, and all other reagents were prepared using known literature procedures. Reactions were monitored by thin-layer chromatography (TLC) performed on 250 μm silica gel 60 plates with 254 nm fluorescent indicator from EMD Chemicals using UV light as a visualizing agent and $\text{KMnO}_4/\text{H}_2\text{SO}_4$, *p*-anisaldehyde or ceric ammonium molybdate and heat as developing agents. Flash chromatography was performed on EMD Chemicals (40-63 μm) silica gel. NMR spectra were recorded on a Bruker 500 MHz or a Bruker 600 MHz spectrometer. Chemical shifts are reported in parts per million using residual non-deuterated solvent as an internal standard (CDCl_3 : 7.26 ppm for ^1H NMR and 77.16 ppm for ^{13}C NMR). Data are reported as follows: chemical shift, multiplicity (ap = apparent, s = singlet, d = doublet, t = triplet, q = quartet, quin = quintet, m = multiplet, br = broad), coupling constant(s) in Hz, integration. NMR spectra were obtained at 298 K unless otherwise noted. FT-IR spectra were recorded on a Varian 640-IR spectrometer and are reported in terms of frequency of absorption (cm^{-1}). Optical rotations were measured with a Jasco P-1010 polarimeter operating on the sodium D-line (589 nm) using a 50 mm path-length cell and are reported as: $[\alpha]_{\text{D}}^{\text{T}}$ (concentration in g/100 mL, solvent). Analytical chiral HPLC was performed with an Agilent 1100 Series HPLC using a Chiralpak AD-H column (4.6 mm x 25 cm) obtained from Daicel Chemical Industries Ltd. with visualization at 254 nm. High resolution mass spectra (HRMS) were recorded on a Waters LCT Premier spectrometer using ESI-TOF

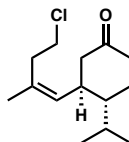
(electrospray ionization-time of flight) or CI-TOF (chemical ionization-time of flight). Melting points (mp) are uncorrected and were measured on a Mel-Temp II melting point apparatus.

III. Experimental Procedures and Characterization Data



9: (±)-cryptone

(±)-Cryptone (9). The title compound was prepared according to the literature procedure using a two-step Robinson annulation sequence.¹ The spectral data for this compound are consistent with those reported in the literature.² ¹H NMR (CDCl₃, 500 MHz) δ 6.89 (dt, *J* = 10.5, 2.0 Hz, 1H), 6.00 (dd, *J* = 10.5, 2.5 Hz, 1H), 2.50 (dt, *J* = 16.5, 4.0 Hz, 1H), 2.38 – 2.32 (m, 1H), 2.32 – 2.25 (m, 1H), 2.04 – 1.96 (m, 1H), 1.86 – 1.71 (m, 1H), 0.96 (t, *J* = 7.0 Hz, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 200.2, 154.5, 129.8, 42.6, 37.5, 31.6, 25.4, 19.8, 19.6.



(±)-10

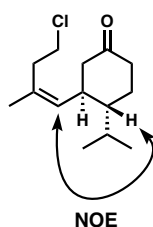
Ketone 10. To a solution of (*Z*)-4-chloro-1-iodo-2-methylbut-1-ene³ (0.346 g, 1.50 mmol) in Et₂O (6 mL, previously sparged with argon gas for 20 min) at –78 °C was added *tert*-butyllithium (2.1 mL, 1.45 M in pentane, 3.01 mmol) dropwise *via* syringe. After 20 min, freshly prepared MgBr₂•OEt₂ (2.2 mL, ~1 M suspension in Et₂O, 2.15 mmol) was added dropwise *via* syringe. After 20 min, CuBr•DMS (0.155 g, 0.752 mmol) was added in one portion. TMEDA

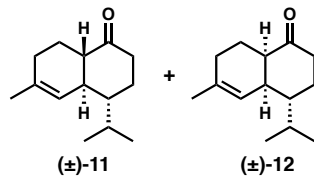
¹ Chen, K.; Ishihara, Y.; Galán, M. M.; Baran, P. S. *Tetrahedron* **2010**, *66*, 4738–4744.

² Hawley, R. C.; Schreiber, S. L. *Synth. Commun.* **1990**, *20*, 1159–1165.

³ (*Z*)-4-chloro-1-iodo-2-methylbut-1-ene was prepared according to the previously reported procedure: Daub, M. E.; Prudhomme, J.; Le Roch, K.; Vanderwal, C. D. *J. Am. Chem. Soc.* **2015**, *137*, 4912–4915.

(0.11 mL, 0.752 mmol) was added dropwise *via* syringe immediately afterwards. After 30 min at $-78\text{ }^{\circ}\text{C}$, TMSCl (0.11 mL, 0.859 mmol) was added dropwise *via* syringe followed by a solution of (\pm)-cryptone (**9**) (98.9 mg, 0.716 mmol) in Et_2O (0.5 mL). The transfer was completed with an additional portion of Et_2O (0.5 mL). After allowing the reaction mixture to stir for 2 h at $-78\text{ }^{\circ}\text{C}$, the reaction was quenched with 9:1 saturated NH_4Cl solution/ NH_4OH (15 mL). After warming to room temperature, the reaction mixture was extracted with Et_2O (3 x 10 mL). The combined organic extracts were washed with 1 M HCl (15 mL), saturated NaHCO_3 solution (15 mL), and brine (15 mL). The organic phase was dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 5% EtOAc /hexanes) to afford the title compound (138.2 mg, 80%) as a white solid (mp $55\text{--}57\text{ }^{\circ}\text{C}$). ^1H NMR (CDCl_3 , 600 MHz) δ 5.10 (d, $J = 9.6\text{ Hz}$, 1H), 3.54 (t, $J = 7.2\text{ Hz}$, 2H), 2.61 – 2.50 (m, 2H), 2.50 – 2.43 (m, 2H), 2.42 – 2.31 (m, 2H), 2.14 (ap t, $J = 13.2\text{ Hz}$, 1H), 2.02 – 1.88 (m, 2H), 1.73 (s, 3H), 1.52 – 1.40 (m, 2H), 0.96 (d, $J = 7.0\text{ Hz}$, 3H), 0.73 (d, $J = 7.0\text{ Hz}$, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 211.4, 131.8, 131.6, 47.9, 47.1, 42.4, 41.2, 40.6, 35.2, 27.9, 24.5, 22.9, 21.9, 16.1; IR (thin film) ν 2957, 2870, 1714, 1455, 1368, 1259, 1202, 1066 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{14}\text{H}_{23}\text{ClONa}$ ($\text{M} + \text{Na}$) $^+$ 265.1335, found 265.1329. ^1H -NOESY-2D (500 MHz, CDCl_3) spectra were obtained for ketone **10** and selected NOE interactions are shown.

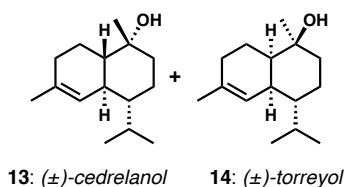




Decalones 11 and 12. To a solution of alkyl chloride **10** (0.292 g, 1.20 mmol) in *tert*-butyl alcohol (6.0 mL) at 30 °C was added potassium *tert*-butoxide (1.10 mL, 1.6 M in THF, 1.81 mmol) dropwise *via* syringe. After stirring for 6 h at 30 °C, the reaction was quenched with saturated NH₄Cl solution (6 mL). The aqueous layer was extracted with pentane (3 × 15 mL), and the combined organic extracts were washed with water (3 × 10 mL) and with brine (10 mL). The organic layer was dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 5% Et₂O/pentane) to afford a 1.1:1 mixture of *trans*- and *cis*-decalones (176.4 mg, 71%) as a colorless oil, which was characterized as a mixture. The spectral data for these compounds are consistent with those reported in the literature.^{4,5} ¹H NMR (CDCl₃, 500 MHz) δ 5.55 (s, 1H), 5.32 (s, 1H), 2.52 (br s, 1H), 2.49 – 2.11 (m, 7H), 2.11 – 1.95 (m, 8H), 1.95 – 1.85 (m, 1H), 1.85 – 1.78 (m, 1H), 1.68 (s, 3H), 1.64 (s, 3H), 1.62 – 1.38 (m, 6H), 1.00 (d, *J* = 5.0 Hz, 3H), 0.99 (d, *J* = 5.0 Hz, 3H), 0.90 (d, *J* = 7.0 Hz, 3H), 0.77 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 214.8, 213.1, 136.0, 135.1, 124.4, 121.6, 51.2, 46.9, 46.1, 45.2, 44.4, 41.2, 39.0, 38.4, 29.8, 28.6, 26.9, 26.5, 25.5, 24.0, 23.9, 23.8, 23.1, 22.0, 21.70, 21.65, 17.9, 15.1.

⁴ Nishikawa, K.; Nakahara, H.; Shirokura, Y.; Nogata, Y.; Yoshimura, E.; Umezawa, T.; Okino, T.; Matsuda, F. *J. Org. Chem.* **2011**, *76*, 6558–6573.

⁵ Taber, D. F.; Gunn, B. P. *J. Am. Chem. Soc.* **1979**, *101*, 3992–3993.



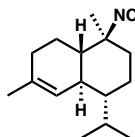
(±)-Cedrelanol (13) and (±)-torreyol (14). Methylmagnesium bromide (1.9 mL, 2.25 M in Et₂O, 4.27 mmol) was added dropwise to a solution of a mixture of decalones **11** and **12** (0.176 g, 0.855 mmol) in THF (8.6 mL) cooled to 0 °C. The reaction mixture was allowed to warm to room temperature for 30 min, and the reaction was quenched with saturated aqueous NH₄Cl solution (10 mL) at 0 °C. The aqueous layer was extracted with Et₂O (3 x 15 mL). The combined organic extracts were washed with brine (20 mL), dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 10% EtOAc/hexanes), yielding (±)-cedrelanol (**13**) (0.106 g, 55%) as a colorless oil and (±)-torreyol (**14**) (0.084 g, 44%) as a white solid (mp 109–110 °C, lit.⁵ mp 108.5–109 °C). The spectral data for these compounds are consistent with those reported in the literature.^{5,6,7}

(±)-Cedrelanol (**13**): ¹H NMR (CDCl₃, 600 MHz) δ 5.55 (br s, 1H), 2.22 – 2.15 (m, 1H), 2.04 – 1.89 (m, 4H), 1.74 (dt, *J* = 13.1, 2.9 Hz, 1H), 1.67 (s, 3H), 1.50 – 1.45 (m, 1H), 1.45 – 1.29 (m, 3H), 1.22 (s, 3H), 1.09 (t, *J* = 10.3 Hz, 1H), 1.01 (tt, *J* = 11.4, 3.2 Hz, 1H), 0.92 (d, *J* = 6.9 Hz, 3H), 0.79 (d, *J* = 6.9 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 134.5, 122.8, 70.8, 48.1, 46.8, 40.4, 37.9, 31.0, 28.6, 26.3, 23.9, 22.7, 21.5, 19.9, 15.3.

(±)-Torreyol (**14**): ¹H NMR (CDCl₃, 600 MHz) δ 5.52 (d, *J* = 4.3 Hz, 1H), 2.06 – 1.92 (m, 4H), 1.92 – 1.86 (m, 1H), 1.66 (s, 3H), 1.63 – 1.47 (m, 6H), 1.35 – 1.24 (m, 2H), 1.30 (s, 3H), 1.15 – 1.05 (m, 1H) 0.89 (d, *J* = 6.9 Hz, 3H), 0.77 (d, *J* = 6.9 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 134.5, 124.7, 72.7, 45.6, 44.2, 36.9, 35.4, 31.3, 28.1, 26.5, 23.8, 21.8, 21.7, 18.6, 15.4.

⁶ Pronin, S. V; Reiher, C. A.; Shenvi, R. A. *Nature* **2013**, *501*, 195–199.

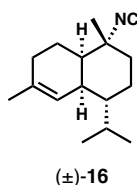
⁷ Borg-Karlson, A.-K.; Norin, T.; Talvitie, A. *Tetrahedron* **1981**, *37*, 425–430.



15: (±)-10-isocyano-4-cadinene

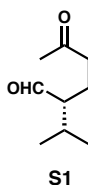
(±)-10-Isocyano-4-cadinene (15). The title compound was prepared according to the literature procedure.⁶ A mixture of (±)-cedrelanol (**13**) (28.3 mg, 0.127 mmol) and pyridine (50 μ L, 0.508 mmol) in CH_2Cl_2 (1.3 mL) at 0 $^\circ\text{C}$ was treated with trifluoroacetic anhydride (40 μ L, 0.255 mmol). After 15 minutes, the reaction was quenched with 1 M HCl (2 mL). The aqueous layer was extracted with CH_2Cl_2 (3 x 3 mL). The combined organic extracts were washed with water (5 mL), washed with saturated NaHCO_3 solution (5 mL), dried over MgSO_4 , filtered and concentrated *in vacuo*. A portion of the crude trifluoroacetate (15.9 mg, 0.0499 mmol) was dissolved in TMSCN (50 μ L), and a solution of scandium(III) trifluoromethanesulfonate (~0.7 mg, 0.0015 mmol) in TMSCN (50 μ L) was added. After 4 h at room temperature, the reaction was quenched with TMEDA (20 μ L) and the volatiles were removed under reduced pressure. The crude residue was dissolved in hexanes (5 mL), washed with saturated aqueous NaHCO_3 solution (5 mL), dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 30% CH_2Cl_2 /hexanes) to afford the title compound (3.9 mg, 34%) as a thin film along with a mixture of diastereomers (1.1 mg, 9%). The spectral data for this compound are consistent with those reported in the literature.⁶ ^1H NMR (CDCl_3 , 600 MHz) δ 5.46, (s, 1 H), 2.22 – 2.13 (m, 1H), 2.12 – 1.94 (m, 4H), 1.82 (td, J = 13.3, 4.0 Hz, 1H), 1.73 (t, J = 10.8 Hz, 1H), 1.68 (s, 3H), 1.59 (dq, J = 13.3, 3.3 Hz, 1H), 1.50 (t, J = 11.0 Hz, 1H), 1.34 (dd, J = 12.3, 5.8 Hz, 1H), 1.30 (s, 3H), 1.12 (qd, J = 13.0, 3.5 Hz, 1H), 1.06 (tt, J = 12.0, 3.0 Hz, 1H), 0.91 (d, J = 7.0 Hz, 3H), 0.76 (d, J = 7.0 Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ

152.1 (br t, $J = 4.3$ Hz), 135.5, 121.4, 60.9 (br t, $J = 5.1$ Hz), 48.1, 46.3, 40.7, 38.0, 30.8, 26.0, 23.9, 23.8, 21.5, 20.3, 20.1, 15.2.

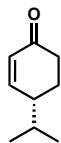


Isocyanide 16. The following procedure was adopted from Pronin *et al.*⁶ A mixture of (±)-torreyol (**14**) (30.8 mg, 0.139 mmol) and pyridine (50 μ L, 0.556 mmol) in CH_2Cl_2 (1.4 mL) at 0 °C was treated with trifluoroacetic anhydride (40 μ L, 0.278 mmol). After 15 minutes, the reaction was quenched with 1 M HCl (2 mL). The aqueous layer was extracted with CH_2Cl_2 (3 x 3 mL). The combined organic extracts were washed with water (5 mL), washed with saturated NaHCO_3 solution (5 mL), dried over MgSO_4 , filtered and concentrated *in vacuo*. The crude trifluoroacetate was dissolved in TMSCN (0.14 mL), and a solution of scandium(III) trifluoromethanesulfonate (3.4 mg, 0.0695 mmol) in TMSCN (0.14 mL) was added. After 3 h at room temperature, the reaction was quenched with TMEDA (20 μ L) and the volatiles were removed under reduced pressure. The crude residue was dissolved in hexanes (5 mL), washed with saturated aqueous NaHCO_3 solution (5 mL), dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 30% CH_2Cl_2 /hexanes) to afford the title compound (10.4 mg, 33%) as a thin film. ^1H NMR (CDCl_3 , 500 MHz) δ 5.56 (d, $J = 1.0$ Hz, 1H), 2.39 – 2.32 (m, 1H), 2.07 – 1.93 (m, 3H), 1.83 (d, $J = 13.0$ Hz, 1H), 1.74 (d, $J = 11.0$ Hz, 1H), 1.65 (s, 3H), 1.61 – 1.54 (m, 1H), 1.54 – 1.37 (m, 4H), 1.42 (s, 3H), 1.31 – 1.21 (m, 1H), 0.90 (d, $J = 7.0$ Hz, 3H), 0.87 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 153.3 (t, $J = 4.5$ Hz), 133.6, 124.4, 62.2 (t, $J = 4.4$ Hz), 44.6, 43.6, 35.0, 33.6, 31.0, 27.9, 26.7, 23.6, 21.7, 20.0, 19.4, 15.4; IR (thin film) ν 2956, 2893, 2872, 2832, 2126,

1452, 1384, 1154, 884 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{16}\text{H}_{25}\text{NNa}$ ($\text{M} + \text{Na}$)⁺ 254.1885, found 254.1880.



(R)-2-Isopropyl-5-oxohexanal (S1). The title compound was prepared according to the literature procedure.¹ The spectral data for this compound are consistent with those reported in the literature.^{8,9} $[\alpha]_{\text{D}}^{24} = -33.7$ ($c = 1.0$, CHCl_3), lit.⁹ $[\alpha]_{\text{D}} = +40$ ($c = 1.72$, CDCl_3) for the (*S*)-form; ^1H NMR (500 MHz, CDCl_3) δ 9.60 (d, $J = 3.0$ Hz, 1H), 2.50 (ddd, $J = 17.5, 8.8, 5.8$ Hz, 1H), 2.36 (ddd, $J = 17.8, 8.0, 6.8$ Hz, 1H), 2.12 (s, 3H), 2.09 – 1.98 (m, 2H), 1.88 – 1.71 (m, 2H), 0.99 (d, $J = 6.5$ Hz, 3H), 0.96 (d, $J = 6.5$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 208.2, 205.5, 57.7, 41.4, 30.2, 28.5, 20.4, 19.6, 19.5.

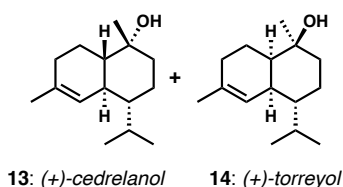


(-)-Cryptone (9). To a solution of (*R*)-2-isopropyl-5-oxohexanal (**S1**) (0.731 mg, 4.68 mmol) in 2-propanol (15.6 mL) was added a solution of lithium isopropoxide (0.35 mL, 0.66 M in THF, 0.234 mmol). After 1 h, the reaction was quenched with saturated NH_4Cl solution (16 mL). The aqueous phase was extracted with ether (3 x 40 mL). The combined organic extracts were washed with water (2 x 20 mL), brine (1 x 20 mL), dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 10% Et_2O /pentane) to afford (-)-cryptone (0.435 g, 66%) as a colorless oil. The spectral data for this compound are

⁸ Hagiwara, H.; Komatsubara, N.; Ono, H.; Okabe, T.; Hoshi, T.; Suzuki, T.; Ando, M. *J. Chem. Soc. Perkin Trans. 1* **2001**, 316–322.

⁹ Hudlicky, T.; Fleming, A.; Radesca, L. *J. Am. Chem. Soc.* **1989**, *111*, 6691–6707.

consistent with those reported in the literature.² $[\alpha]_D^{24} = -80.8$ ($c = 1.04$, CHCl_3), lit.¹⁰ $[\alpha]_D^{21} = -90$ ($c = 1.38$, CHCl_3); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 6.89 (dt, $J = 10.0, 2.0$ Hz, 1H), 6.01 (dd, $J = 10.0, 2.5$ Hz, 1H), 2.51 (dt, $J = 17.0, 4.0$ Hz, 1H), 2.38 – 2.32 (m, 1H), 2.32 – 2.25 (m, 1H), 2.04 – 1.96 (m, 1H), 1.86 – 1.71 (m, 1H), 0.97 (t, $J = 7.0$ Hz, 6H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 200.2, 154.5, 129.8, 42.6, 37.5, 31.6, 25.4, 19.8, 19.6. The enantiomeric excess of (–)-cryptone was determined to be 88% using chiral HPLC (Chiralpak AD-H column, 2% *i*PrOH in hexanes, flow rate of 0.5 mL/min).



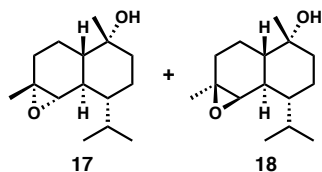
(+)-Cedrelanol (13) and (+)-torreyol (14). The title compounds were prepared from (–)-cryptone according to the three-step sequence described above. The spectral data for these compounds are consistent with those reported in the literature.^{5–7}

(+)-Cedrelanol (13): $[\alpha]_D^{24} = +2.2$ ($c = 1.0$, CHCl_3), lit.⁷ $[\alpha]_D = +3.4$ ($c = 1.2$, CHCl_3); $^1\text{H NMR}$ (CDCl_3 , 600 MHz) δ 5.55 (br s, 1H), 2.22 – 2.15 (m, 1H), 2.04 – 1.89 (m, 4H), 1.74 (dt, $J = 13.1, 2.9$ Hz, 1H), 1.67 (s, 3H), 1.50 – 1.45 (m, 1H), 1.45 – 1.29 (m, 3H), 1.22 (s, 3H), 1.09 (t, $J = 10.3$ Hz, 1H), 1.01 (tt, $J = 11.4, 3.2$ Hz, 1H), 0.92 (d, $J = 6.9$ Hz, 3H), 0.79 (d, $J = 6.9$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 134.5, 122.8, 70.8, 48.1, 46.8, 40.4, 37.9, 31.0, 28.6, 26.3, 23.9, 22.7, 21.5, 19.9, 15.3.

(+)-Torreyol (14): mp 109–110 °C, lit.⁵ mp 108.5–109 °C; $[\alpha]_D^{24} = +95.4$ ($c = 1.0$, CHCl_3), lit.⁷ $[\alpha]_D = -100.4$ ($c = 1.2$, CHCl_3) for (–)-torreyol; $^1\text{H NMR}$ (CDCl_3 , 600 MHz) δ 5.52 (d, $J = 4.3$ Hz, 1 H), 2.06 – 1.92 (m, 4H), 1.92 – 1.86 (m, 1H), 1.66 (s, 3H), 1.63 – 1.47 (m, 6H), 1.35 – 1.24

¹⁰ Gillespie, D. T. C.; Macbeth, A. K.; Mills, J. A. *J. Chem. Soc.* **1948**, 996–999.

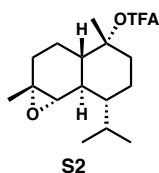
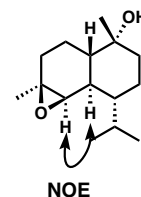
(m, 2H), 1.30 (s, 3H), 1.15 – 1.05 (m, 1H) 0.89 (d, $J = 6.9$ Hz, 3H), 0.77 (d, $J = 6.9$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 134.5, 124.7, 72.7, 45.6, 44.2, 36.9, 35.4, 31.3, 28.1, 26.5, 23.8, 21.8, 21.7, 18.6, 15.4.



Epoxides 17 and 18. To a solution of (+)-cedrelanol (**13**) (0.188 g, 0.845 mmol) in acetone (42 mL) was added saturated aqueous NaHCO_3 (28 mL). The resulting mixture was cooled to $0\text{ }^\circ\text{C}$ and a solution of Oxone $^\circledR$ (0.571 g, 0.930 mmol) in H_2O (2 mL) was added dropwise over 5 minutes. The reaction mixture was stirred vigorously for 30 min at $0\text{ }^\circ\text{C}$, diluted with H_2O (50 mL) and extracted with EtOAc (3 x 50 mL). The combined organic extracts were diluted with hexanes until cloudy, dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 10 \rightarrow 30% EtOAc/hexanes) to afford β -epoxide **18** (77.1 mg, 38%) as a white solid (mp $90\text{--}93\text{ }^\circ\text{C}$) and α -epoxide **17** (120.4 mg, 60%) as a colorless oil. The hydroxylic peaks in the ^1H NMR spectra for epoxides **17** and **18** could not be unambiguously identified.

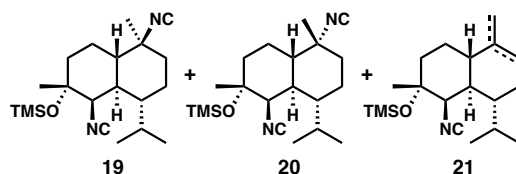
Epoxide **17**: $[\alpha]_D^{24} = +22.6$ ($c = 1.0$, CHCl_3); ^1H NMR (CDCl_3 , 500 MHz) δ 2.94 (s, 1H), 2.32 – 2.20 (m, 1H), 2.09 (dd, $J = 14.3, 3.8$ Hz, 1H), 1.73 – 1.46 (m, 5H), 1.46 – 1.32 (m, 2H), 1.30 (s, 3H), 1.27 – 1.11 (m, 2H), 1.17 (s, 3H), 0.96 (d, $J = 7$ Hz, 3H), 0.88 (d, $J = 7$ Hz, 3H), 0.90 – 0.85 (m, 1H); ^{13}C NMR (125 MHz, CDCl_3) δ 70.5, 61.5, 58.2, 47.5, 44.7, 39.9, 38.7, 30.8, 28.0, 26.4, 23.8, 21.6, 20.1, 19.7, 15.7; IR (thin film) ν 3459 (br), 2957, 2937, 2871, 2849, 1463, 1453, 1368, 1135, 1003, 876 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{15}\text{H}_{26}\text{O}_2\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 261.1830, found 261.1838. ^1H -NOESY-2D (500 MHz, CDCl_3) spectra were obtained for epoxide **17**.

Epoxide **18**: $[\alpha]_D^{24} = +10.7$ ($c = 1.0$, CHCl_3); $^1\text{H NMR}$ (CDCl_3 , 500 MHz) δ 3.08 (s, 1H), 2.29 – 2.19 (m, 1H), 1.91 – 1.80 (m, 2H), 1.80 – 1.72 (m, 1H), 1.72 – 1.66 (m, 1H), 1.64 – 1.57 (m, 1H), 1.54 – 1.49 (m, 1H), 1.44 – 1.33 (m, 3H), 1.31 (s, 3H), 1.19 – 1.08 (m, 2H), 1.16 (s, 3H), 0.95 (d, $J = 6.9$ Hz, 3H), 0.85 (d, $J = 6.9$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 70.9, 61.9, 58.7, 44.3, 42.4, 40.3, 38.4, 29.2, 29.1, 26.6, 25.0, 21.8, 21.4, 20.0, 15.6; IR (thin film) ν 3463 (br), 2956, 2934, 2872, 1464, 1375, 1209, 1142, 1022, 907, 845 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{15}\text{H}_{26}\text{O}_2\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 261.1830, found 261.1838. $^1\text{H-NOESY-2D}$ (500 MHz, CDCl_3) spectra were obtained for epoxide **18** and selected NOE interactions are shown.



Trifluoroacetate S2. A mixture of epoxide **17** (0.120 g, 0.505 mmol) and pyridine (0.16 mL, 2.02 mmol) in CH_2Cl_2 (5.1 mL) at 0 °C was treated with trifluoroacetic anhydride (0.14 mL, 1.01 mmol). After 30 minutes, the reaction was quenched with 1 M HCl (5 mL). The aqueous layer was extracted with CH_2Cl_2 (3 x 5 mL). The combined organic extracts were washed with water (10 mL), washed with saturated NaHCO_3 solution (10 mL), dried over MgSO_4 , filtered and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 50% CH_2Cl_2 /hexanes) to afford the title compound (0.114 g, 67%) as a colorless oil. $[\alpha]_D^{24} = -7.0$ ($c = 1.0$, CHCl_3); $^1\text{H NMR}$ (CDCl_3 , 500 MHz) δ 2.91 (s, 1H), 2.75 (dt, $J = 15.0, 3.0$ Hz, 1H), 2.29 – 2.22 (m, 1H), 2.12 (dd, $J = 13.8, 4.8$ Hz, 1H), 1.76 (t, $J = 11.7$ Hz, 1H), 1.67 – 1.54 (m, 3H), 1.57 (s, 3H), 1.40 – 1.24 (m, 3H), 1.31 (s, 3H), 1.14 (qd, $J = 13.8, 3.3$ Hz, 1H), 0.96 (d, $J = 6.6$ Hz, 3H), 0.92 (t, $J = 11.7$ Hz, 1H), 0.85 (d, $J = 6.6$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 156.2 (q,

$J = 41.0$ Hz), 114.6 (q, $J = 285.4$ Hz), 88.3, 61.4, 57.9, 48.6, 43.9, 38.4, 34.3, 30.7, 26.4, 23.8, 23.7, 21.5, 19.8, 19.6, 15.2; IR (thin film) ν 2960, 2873, 1778, 1453, 1373, 1218, 1155 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{17}\text{H}_{25}\text{F}_3\text{O}_3\text{Na}$ ($M + \text{Na}$)⁺ 357.1653, found 357.1647.



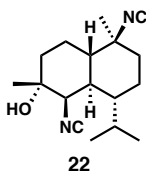
Isocyanides 19, 20, and 21. The following procedure was adopted from Pronin *et al.*⁶ A solution of trifluoroacetate **S2** (62.3 mg, 0.186 mmol) in TMSCN (0.19 mL) was cooled to 0 °C. A solution of scandium(III) trifluoromethanesulfonate (9.2 mg, 0.0186 mmol) in TMSCN (0.37 mL) was added, and the reaction mixture was allowed to warm to room temperature after 1 h. After 24 h at room temperature, the reaction was quenched with TMEDA (30 μL) and the volatiles were removed under reduced pressure. The crude residue was dissolved in hexanes (10 mL), washed with saturated aqueous NaHCO_3 solution (5 mL), dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 2% EtOAc/hexanes) to afford a 6.3:1 mixture of elimination products **21** (18.9 mg, 32%) as a thin film and a 1.9:1 mixture of isocyanides **19** and **20** (8.0 mg, 12%) as a thin film. Isocyanides **19** and **20** were separated by column chromatography (SiO_2 , 80% CH_2Cl_2 /hexanes). The 6.3:1 mixture of alkene isomers (**21**) was characterized as a mixture; only the resonances in the ^1H and ^{13}C NMR spectra for the major Δ^9 -isomer are listed.

Diisocyanide **19**: $[\alpha]_{\text{D}}^{22} = +14.9$ ($c = 0.66$, CHCl_3); ^1H NMR (CDCl_3 , 500 MHz) δ 3.55 (s, 1H), 2.06 (dt, $J = 13.0, 3.3$ Hz, 1H), 1.94 – 1.60 (m, 8H), 1.54 – 1.40 (m, 2H), 1.45 (s, 3H), 1.30 (s, 3H), 1.11 (qd, $J = 13.3, 3.5$ Hz, 1H), 0.95 (d, $J = 7.0$ Hz, 3H), 0.72 (d, $J = 7.0$ Hz, 3H), 0.13 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 158.5 (br t, $J = 4.9$ Hz), 153.1 (br t, $J = 4.4$ Hz), 73.4, 61.6

(br t, $J = 5.2$ Hz), 60.3 (br t, $J = 5.3$ Hz), 42.6, 42.1, 40.4, 35.9, 32.6, 27.5, 25.5, 21.4, 21.2, 20.6, 19.2, 14.9, 2.3; IR (thin film) ν 2956, 2874, 2131, 1456, 1384, 1252, 1184, 1038, 842 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{20}\text{H}_{34}\text{N}_2\text{OSiNa}$ ($\text{M} + \text{Na}$) $^+$ 369.2338, found 369.2343.

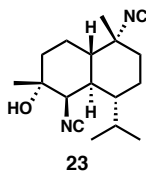
Diisocyanide **20**: $[\alpha]_{\text{D}}^{24} = -7.7$ ($c = 0.34$, CHCl_3); ^1H NMR (CDCl_3 , 500 MHz) δ 3.59 (s, 1H), 2.10 – 2.01 (m, 1H), 1.97 (ap d, $J = 13.5$ Hz, 1H), 1.92 (quin of d, $J = 6.8, 3.0$ Hz, 1H) 1.75 – 1.65 (m, 5H), 1.54 – 1.34 (m, 2H), 1.44 (s, 3H), 1.41 (br s, 3H), 1.34 – 1.24 (m, 2H), 0.96 (d, $J = 6.5$ Hz, 3H), 0.79 (d, $J = 6.5$ Hz, 3H), 0.16 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 157.8 (br t), 155.3 (br t), 72.9, 61.6 (br t, $J = 5.3$ Hz), 61.5 (br t, $J = 4.9$ Hz), 42.3, 42.0, 39.1, 36.0, 32.3, 27.8, 27.6, 25.7, 21.3, 21.1, 19.1, 15.0, 2.4; IR (thin film) ν 2957, 2874, 2127, 1454, 1379, 1252, 1184, 1043, 869, 841 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{20}\text{H}_{34}\text{N}_2\text{OSiNa}$ ($\text{M} + \text{Na}$) $^+$ 369.2338, found 369.2332.

Isocyanide **21**: $[\alpha]_{\text{D}}^{23} = +25.3$ ($c = 0.90$, CHCl_3); ^1H NMR (CDCl_3 , 500 MHz) δ 5.42 (s, 1H), 3.56 (s, 1H), 2.10 – 1.50 (m, 9H) 1.64 (s, 3H), 1.45 (s, 3H), 1.28 – 1.16 (m, 1H), 0.94 (d, $J = 6.9$ Hz, 3H), 0.74 (d, $J = 6.9$ Hz, 3H), 0.12 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 157.3 (br t, $J = 4.8$ Hz), 135.1, 121.8, 73.7, 61.7 (br t, $J = 5.2$ Hz), 38.9, 38.4, 37.6, 34.4, 27.7, 25.6, 25.3, 23.8, 21.2, 21.0, 14.6, 2.5; IR (thin film) ν 2959, 2894, 2872, 2133, 1455, 1379, 1251, 1052, 842 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{19}\text{H}_{33}\text{NOSiNa}$ ($\text{M} + \text{Na}$) $^+$ 342.2229, found 342.2215.



Diisocyanide 22. To a solution of TMS ether **19** (6.6 mg, 0.0190 mmol) in THF (0.2 mL) was added TBAF (40 μL , 1.0 M in THF, 0.0381 mmol). After 1 h, the reaction was quenched with water (1 mL). The aqueous layer was extracted with ether (3 \times 2 mL). The combined organic

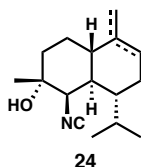
extracts were dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 20% EtOAc/hexanes) to afford diisocyanide **22** (3.5 mg, 67%) as a thin film. The spectral data for this compound are consistent with those reported in the literature.¹¹ [α]_D²³ = +23.3 (c = 0.35, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 3.63 (s, 1H), 2.07 (dt, *J* = 13.0, 3.3 Hz, 1H), 1.95 – 1.60 (m, 8H), 1.58 – 1.43 (m, 2H), 1.45 (s, 3H), 1.33 (br t, *J* = 2.0 Hz, 3H), 1.26 (s, 1H), 1.13 (qd, *J* = 12.8, 3.3 Hz, 1H), 0.96 (d, *J* = 7.0 Hz, 3H), 0.75 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 158.7 (br t, *J* = 4.5 Hz), 153.1 (br t, *J* = 4.6 Hz), 70.5, 60.9 (br t, *J* = 5.4 Hz), 60.3 (br t, *J* = 5.1 Hz), 42.8, 42.4, 40.4, 36.2, 32.8, 28.9, 25.5, 21.4, 21.2, 20.9, 19.2, 15.2; IR (thin film) ν 3414 (br), 2955, 2873, 2133, 1452, 1386, 1267, 1180, 1126, 1002, 760 cm⁻¹; HRMS (ESI) *m/z* calcd for C₁₇H₂₆N₂ONa (M + Na)⁺ 297.1943, found 297.1949.



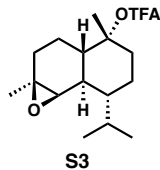
Diisocyanide 23. To a solution of TMS ether **20** (3.4 mg, 0.0098 mmol) in THF (0.2 mL) was added TBAF (20 μ L, 1.0 M in THF, 0.0196 mmol). After 1 h, the reaction was quenched with water (1 mL). The aqueous layer was extracted with ether (3 x 2 mL). The combined organic extracts were dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 15% EtOAc/hexanes) to afford diisocyanide **23** (1.9 mg, 70%) as a thin film. [α]_D²³ = -5.9 (c = 0.19, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 3.68 (s, 1H), 2.04 – 1.88 (m, 3H), 1.87 – 1.60 (m, 5H), 1.57 – 1.28 (m, 5H), 1.44 (s, 3H), 1.43 (br s, 3H), 0.97 (d, *J* = 7.0 Hz, 3H), 0.81 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 158.1 (br t, *J* =

¹¹ White, R. D.; Wood, J. L. *Org. Lett.* **2001**, *3*, 1825–1827.

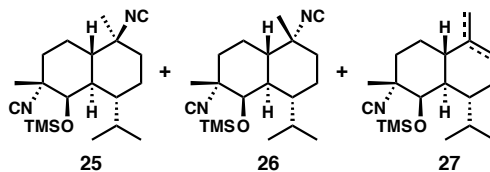
5.5 Hz), 155.4 (br t, $J = 4.4$ Hz), 70.1, 61.3 (br t, $J = 4.4$ Hz), 60.9 (br t, $J = 5.2$ Hz), 42.5, 42.1, 39.2, 36.4, 32.4, 28.6, 27.9, 25.7, 21.4, 21.0, 19.1, 15.2; IR (thin film) ν 3428 (br), 2957, 2874, 2133, 1455, 1377, 1276, 1181, 1001, 833 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{17}\text{H}_{26}\text{N}_2\text{ONa}$ ($\text{M} + \text{Na}$)⁺ 297.1943, found 297.1934.



Isocyanide 24. To a solution of TMS ether **21** (9.0 mg, 0.0282 mmol) in THF (0.28 mL) was added TBAF (60 μL , 1.0 M in THF, 0.0563 mmol). After 1 h, the reaction was quenched with water (1 mL). The aqueous layer was extracted with ether (3 \times 2 mL). The combined organic extracts were dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 2.5:47.5:50 EtOAc/ CH_2Cl_2 /hexanes) to afford a 7:1 mixture of isocyanides **24** (5.0 mg, 71%) as a thin film. The alkene isomers were characterized as a mixture; only the resonances in the ^1H and ^{13}C NMR spectra for the major Δ^9 -isomer are listed. $[\alpha]_{\text{D}}^{24} = +64.4$ ($c = 0.50$, CHCl_3); ^1H NMR (CDCl_3 , 500 MHz) δ 5.43 (s, 1H), 3.63 (s, 1H), 2.15 – 1.78 (m, 8H), 1.71 – 1.51 (m, 2H), 1.65 (s, 3H), 1.45 (s, 3H), 1.24 (qd, $J = 13.2, 4.0$ Hz, 1H), 0.95 (d, $J = 6.9$ Hz, 3H), 0.77 (d, $J = 6.9$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 157.6 (br t, $J = 4.4$ Hz), 134.6, 122.0, 70.9, 60.8 (br t, $J = 5.4$ Hz), 39.2, 38.6, 37.6, 34.0, 28.5, 25.6, 25.1, 23.8, 21.2, 20.9, 14.9; IR (thin film) ν 3422 (br), 2960, 2934, 2892, 2871, 2153, 2134, 1450, 1378, 1207, 1012 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{16}\text{H}_{25}\text{NONa}$ ($\text{M} + \text{Na}$)⁺ 270.1834, found 270.1833.



Trifluoroacetate S3. A mixture of epoxide **18** (77.1 mg, 0.323 mmol) and pyridine (0.10 mL, 0.647 mmol) in CH₂Cl₂ (3.2 mL) at 0 °C was treated with trifluoroacetic anhydride (90 μL, 0.647 mmol). After 30 minutes, the reaction was quenched with 1 M HCl (3 mL). The aqueous layer was extracted with CH₂Cl₂ (3 x 5 mL). The combined organic extracts were washed with water (10 mL), washed with saturated NaHCO₃ solution (10 mL), dried over MgSO₄, filtered and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 40% CH₂Cl₂/hexanes) to afford the title compound (70.1 mg, 65%) as a white solid (mp 86–88 °C). $[\alpha]_D^{24} = -12.5$ (c = 1.0, CHCl₃); ¹H NMR (CDCl₃, 600 MHz) δ 3.11 (s, 1H), 2.75 (dt, *J* = 15.0, 3.0 Hz, 1H), 2.25 (quin of d, *J* = 6.9, 3.3 Hz, 1H), 1.92 – 1.80 (m, 3H), 1.69 – 1.64 (m, 1H), 1.61 – 1.55 (m, 1H), 1.56 (s, 3H), 1.45 (tt, *J* = 11.7, 3.3 Hz, 1H), 1.38 (td, *J* = 14.4, 4.2 Hz, 1H), 1.33 (s, 3H), 1.30 – 1.20 (m, 2H), 1.13 (qd, *J* = 13.2, 3.6 Hz, 1H), 0.95 (d, *J* = 7.2 Hz, 3H), 0.83 (d, *J* = 7.2 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 156.0 (q, *J* = 40.8 Hz), 114.5 (q, *J* = 285.6 Hz), 89.5, 61.5, 58.6, 43.8, 43.4, 37.9, 34.4, 28.8, 26.5, 24.9, 24.1, 21.5, 21.3, 19.6, 15.1; IR (thin film) ν 2957, 2868, 1777, 1452, 1372, 1207, 1162 cm⁻¹; HRMS (CI) *m/z* calcd for C₁₅H₂₅O (M – C₂O₂F₃)⁺ 221.1905, found 221.1907.



Isocyanides 25, 26, and 27. The following procedure was adopted from Pronin *et al.*⁶ A solution of trifluoroacetate **S3** (18.5 mg, 0.0553 mmol) in TMSCN (50 μL) was cooled to 0 °C. A

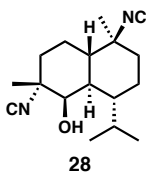
solution of scandium(III) trifluoromethanesulfonate (2.7 mg, 0.00553 mmol) in TMSCN (0.11 mL) was added, and the reaction mixture was allowed to warm to room temperature after 1 h. After 24 h at room temperature, the reaction was quenched with TMEDA (20 μ L) and the volatiles were removed under reduced pressure. The crude residue was dissolved in hexanes (5 mL), washed with saturated aqueous NaHCO₃ solution (3 mL), dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 3% EtOAc/hexanes) to afford a 5.7:1 mixture of elimination products **27** (6.8 mg, 38%) as a thin film, diisocyanide **25** (4.3 mg, 22%) as thin film, and diisocyanide **26** (3.8 mg, 20%) as a thin film. The 5.7:1 mixture of alkene isomers (**27**) was characterized as a mixture; only the resonances in the ¹H and ¹³C NMR spectra for the major Δ^9 -isomer are listed.

Diisocyanide **25**: $[\alpha]_D^{23} = -3.3$. (c = 0.73, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 3.83 (s, 1H), 2.04 (dt, *J* = 13.0, 3.3 Hz, 1H), 1.95 – 1.79 (m, 3H), 1.79 – 1.64 (m, 4H), 1.63 – 1.48 (m, 2H), 1.40 (br s, 3H), 1.33 (br s, 3H), 1.30 (dt, *J* = 11.5, 3.8 Hz, 1H), 1.11 (qd, *J* = 13.5, 3.3 Hz, 1H), 0.94 (d, *J* = 6.9 Hz, 3H), 0.81 (d, *J* = 6.9 Hz, 3H), 0.16 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 157.0 (br t, *J* = 4.6 Hz), 152.5 (br t, *J* = 4.6 Hz), 73.2, 61.5 (br t, *J* = 5.1 Hz), 60.5 (br t, *J* = 5.1 Hz), 41.4, 40.44, 40.37, 38.7, 31.4, 27.5, 25.7, 22.2, 21.3, 20.7, 18.9, 15.0, 1.1; IR (thin film) ν 2957, 2900, 2875, 2127, 1456, 1385, 1252, 1131, 1114, 883, 841 cm⁻¹; HRMS (ESI) *m/z* calcd for C₂₀H₃₄N₂OSiNa (M + Na)⁺ 369.2338, found 369.2338.

Diisocyanide **26**: $[\alpha]_D^{24} = -13.4$ (c = 0.51, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 3.89 (s, 1H), 1.99 – 1.88 (m, 3H), 1.80 – 1.66 (m, 4H), 1.63 – 1.46 (m, 3H), 1.40 (br s, 6H), 1.37 – 1.20 (m, 2H), 0.95 (d, *J* = 7.0 Hz, 3H), 0.88 (d, *J* = 7.0 Hz, 3H), 0.15 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 157.6 (br t, *J* = 4.9 Hz), 155.6 (br t, *J* = 4.7), 73.3, 61.2 (br t, *J* = 4.6 Hz), 60.9 (br t, *J* = 5.3 Hz), 40.9, 40.7, 39.1, 38.9, 31.3, 28.1, 27.7, 25.9, 22.1, 21.2, 18.8, 15.1, 1.1; IR (thin film) ν

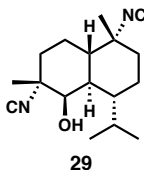
2957, 2930, 2898, 2875, 2132, 1451, 1370, 1252, 1134, 1058, 840 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{20}\text{H}_{34}\text{N}_2\text{OSiNa}$ ($\text{M} + \text{Na}$)⁺ 369.2338, found 369.2331.

Isocyanide **27**: $[\alpha]_{\text{D}}^{23} = -20.5$ ($c = 1.27$, CHCl_3); ^1H NMR (CDCl_3 , 500 MHz) δ 5.40 (s, 1H), 3.89 (s, 1H), 2.17 (t, $J = 11.3$ Hz, 1H), 1.98 – 1.03 (complex, 9H), 1.64 (s, 3H), 1.40 (s, 3H), 0.92 (d, $J = 6.9$ Hz, 3H), 0.85 (d, $J = 6.9$ Hz, 3H), 0.15 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 156.2 (br t, $J = 4.9$ Hz), 135.4, 121.6, 73.3, 61.7 (br t, $J = 5.1$ Hz), 41.5, 37.4, 36.1, 33.1, 27.6, 26.03, 25.99, 23.6, 21.1, 21.0, 14.7, 0.96; IR (thin film) ν 2959, 2907, 2127, 1452, 1368, 1252, 1127, 887, 841 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{19}\text{H}_{33}\text{NOSiNa}$ ($\text{M} + \text{Na}$)⁺ 342.2229, found 342.2240.

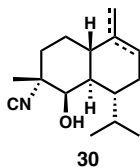


Diisocyanide 28. To a solution of TMS ether **25** (7.3 mg, 0.0211 mmol) in THF (0.21 mL) was added TBAF (40 μL , 1.0 M in THF, 0.0421 mmol). After 1 h, the reaction was quenched with water (1 mL). The aqueous layer was extracted with ether (3 x 2 mL). The combined organic extracts were dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 15% EtOAc/hexanes) to afford diisocyanide **28** (3.3 mg, 57%) as a thin film. $[\alpha]_{\text{D}}^{23} = +37.5$ ($c = 0.33$, CHCl_3); ^1H NMR (CDCl_3 , 500 MHz) δ 3.79 (d, $J = 4.7$ Hz, 1H), 2.06 (dt, $J = 12.8, 3.3$ Hz, 1H), 1.99 – 1.89 (m, 2H), 1.84 – 1.51 (m, 8H), 1.48 (br t, $J = 1.8$ Hz, 3H), 1.44 (ap tt, $J = 12.0, 3.3$ Hz, 1H), 1.35 (br s, 3H), 1.15 (qd, $J = 13.5, 3.5$ Hz, 1H), 0.94 (d, $J = 6.9$ Hz, 3H), 0.70 (d, $J = 6.9$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 156.8 (br t), 152.7 (br t), 71.0, 61.0 (br t, $J = 5.3$ Hz), 60.5 (br t, $J = 5.3$ Hz), 41.8, 41.5, 40.6, 38.1, 31.1, 26.6, 25.8, 22.0, 21.5, 20.6, 19.4, 15.3; IR (thin film) ν 3394 (br), 2955, 2873, 2128,

1455, 1386, 1127, 1032, 759 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{17}\text{H}_{26}\text{N}_2\text{ONa}$ ($\text{M} + \text{Na}$)⁺ 297.1943, found 297.1954.

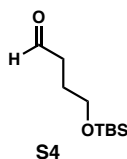


Diisocyanide 29. To a solution of TMS ether **26** (5.1 mg, 0.0147 mmol) in THF (0.2 mL) was added TBAF (30 μL , 1.0 M in THF, 0.0294 mmol). After 1 h, the reaction was quenched with water (1 mL). The aqueous layer was extracted with ether (3 x 2 mL). The combined organic extracts were dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 20% EtOAc/hexanes) to afford diisocyanide **29** (2.9 mg, 73%) as a thin film. $[\alpha]_{\text{D}}^{24} = -22.6$ ($c = 0.29$, CHCl_3); ^1H NMR (CDCl_3 , 500 MHz) δ 3.86 (s, 1H), 2.01 – 1.90 (m, 3H), 1.83 – 1.60 (m, 6H), 1.59 – 1.50 (m, 1H), 1.48 (br t, $J = 1.8$ Hz, 3H), 1.43 – 1.33 (m, 3H), 1.41 (br t, $J = 1.8$ Hz, 3H), 0.95 (d, $J = 7.0$ Hz, 3H), 0.88 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 157.5 (br t), 155.8 (br t), 71.1, 61.2 (br t, $J = 4.4$ Hz), 60.4 (br t, $J = 5.4$ Hz), 41.7, 41.2, 39.3, 38.4, 31.1, 28.0, 26.8, 26.0, 21.8, 21.5, 19.3, 15.4; IR (thin film) ν 3391 (br), 2956, 2939, 2873, 2151, 2128, 1454, 1386, 1179, 1130, 1032 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{17}\text{H}_{26}\text{N}_2\text{ONa}$ ($\text{M} + \text{Na}$)⁺ 297.1943, found 297.1931.



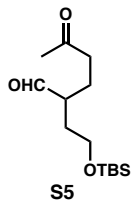
Isocyanide 30. To a solution of TMS ether **27** (12.7 mg, 0.0397 mmol) in THF (0.40 mL) was added TBAF (80 μL , 1.0 M in THF, 0.0795 mmol). After 1 h, the reaction was quenched with water (1 mL). The aqueous layer was extracted with ether (3 x 3 mL). The combined organic

extracts were dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 1:49:50 EtOAc/CH₂Cl₂/hexanes) to afford a 5.6:1 mixture of isocyanides **30** (5.8 mg, 59%) as a thin film. The alkene isomers were characterized as a mixture; only the resonances in the ¹H and ¹³C NMR spectra for the major Δ⁹-isomer are listed. [α]_D²⁴ = -4.1 (c = 0.58, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 5.42 (s, 1H), 3.86 (d, *J* = 4.5 Hz, 1H), 2.11 – 1.51 (m, 10H), 1.65 (s, 3H), 1.49 (t, *J* = 1.8 Hz, 3H), 1.36 (qd, *J* = 13.0, 4.0 Hz, 1H), 0.94 (d, *J* = 7.0 Hz, 3H), 0.84 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 156.0 (br t, *J* = 4.4 Hz), 134.9, 122.0, 71.6, 61.1 (br t, *J* = 5.2 Hz), 41.0, 38.5, 36.5, 32.9, 26.7, 26.1, 25.7, 24.1, 21.3, 21.0, 15.3; IR (thin film) ν 3412 (br), 2957, 2932, 2870, 2151, 2128, 1451, 1387, 1367, 1038 cm⁻¹; HRMS (ESI) *m/z* calcd for C₁₆H₂₅NONa (M + Na)⁺ 270.1834, found 270.1841.

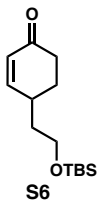


Aldehyde S4. The title compound was prepared according to the literature procedure.¹² The spectral data for this compound are consistent with those reported in the literature.¹² ¹H NMR (500 MHz, CDCl₃) δ 9.78 (t, *J* = 1.8 Hz, 1H), 3.64 (t, *J* = 6.0 Hz, 2H), 2.49 (td, *J* = 7.0, 1.5 Hz, 2H), 1.85 (quin, *J* = 6.6 Hz, 2H), 0.88 (s, 9H), 0.03 (s, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 202.8, 62.2, 40.9, 26.0, 25.6, 18.4, -5.3.

¹² Kwan, E. E.; Scheerer, J. R.; Evans, D. A. *J. Org. Chem.* **2013**, *78*, 175–203.

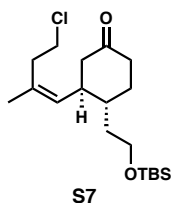


Keto-aldehyde S5. The following procedure was adapted from Hagiwara *et al.*⁸ Diethylamino(trimethyl)silane (90 μ L, 0.473 mmol) was added to a solution of aldehyde **S4** (0.478 g, 2.36 mmol) in MeCN (7.9 mL). The reaction mixture was cooled to 0 $^{\circ}$ C and methyl vinyl ketone (0.29 mL, 3.54 mmol) was added dropwise *via* syringe. The reaction mixture was heated at reflux for 24 h, cooled to room temperature, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 10% EtOAc/hexanes) to afford the title compound (0.561 g, 87%) as a colorless oil. The spectral data for this compound are consistent with those reported in the literature.¹⁸ ¹H NMR (CDCl₃, 500 MHz) δ 9.57 (d, J = 2.5 Hz, 1H), 3.68 – 3.59 (m, 2H), 2.56 – 2.32 (m, 3H), 2.13 (s, 3H), 1.96 – 1.84 (m, 2H), 1.77 – 1.65 (m, 2H), 0.86 (s, 9H), 0.02 (s, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 208.1, 204.4, 60.6, 48.7, 40.9, 32.7, 30.2, 26.0, 22.3, 18.4, –5.4.



Enone S6. To a solution of keto-aldehyde **S5** (2.43 g, 8.92 mmol) in Et₂O (83 mL) and THF (28 mL) was added aqueous KOH solution (89 mL, 0.3 N) and *n*Bu₄NOH (2.9 mL, 40% aq) in one portion. The reaction mixture was heated at reflux for 8 h and cooled to room temperature, at which point additional KOH (1.50 g, 26.7 mmol) was added in one portion. The reaction mixture was heated at reflux for 15 h, and cooled to room temperature. The biphasic reaction mixture was separated, and the aqueous layer was extracted with Et₂O (2 x 100 mL). The combined organic

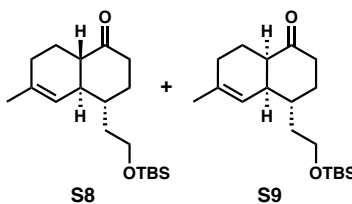
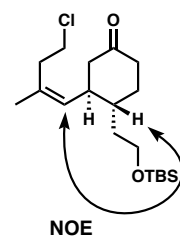
extracts were washed with brine (100 mL), dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 10% EtOAc/hexanes) to afford the title compound (1.48 g, 65%) as a colorless oil. The spectral data for this compound are consistent with those reported in the literature.¹³ ¹H NMR (CDCl₃, 500 MHz) δ 6.90 (ddd, *J* = 10.0, 2.8, 1.3 Hz, 1H), 5.97 (ddd, *J* = 10.0, 2.5, 0.75 Hz, 1H), 3.79 – 3.67 (m, 2H), 2.68 – 2.57 (m, 1H), 2.49 (dt, *J* = 16.8, 4.8 Hz, 1H), 2.36 (ddd, *J* = 17.0, 12.3, 5.0 Hz, 1H), 2.17 – 2.08 (m, 1H), 1.79 – 1.66 (m, 2H), 1.66 – 1.56 (m, 1H), 0.89 (s, 9H), 0.06 (s, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 200.0, 155.4, 129.0, 60.5, 37.4, 37.0, 33.1, 28.7, 26.0, 18.4, –5.21, –5.24.



Ketone S7. To a solution of (*Z*)-4-chloro-1-iodo-2-methylbut-1-ene³ (0.313 g, 1.36 mmol) in Et₂O (2.3 mL, previously sparged with argon gas for 20 min) at –78 °C was added *tert*-butyllithium (1.70 mL, 1.60 M in pentane, 2.72 mmol) dropwise *via* syringe. After 20 min, lithium 2-thienylcyanocuprate solution (5.43 mL, 0.25 M in THF, 1.36 mmol) was added dropwise *via* syringe. The reaction mixture was allowed to stir for 1 h at –78 °C. A solution of enone **S6** (0.230 g, 0.905 mmol) in Et₂O (1.0 mL) was added dropwise *via* syringe. The transfer was completed with additional portions of Et₂O (2 x 0.5 mL). After allowing the reaction mixture to warm to –40 °C and stir for 1 h, the reaction was quenched with 9:1 saturated NH₄Cl solution/NH₄OH (10 mL). After warming to room temperature, the solution turned a deep blue color. The aqueous layer was extracted with Et₂O (3 x 15 mL). The combined organic extracts were washed with 9:1 saturated NH₄Cl solution/NH₄OH (2 x 10 mL), washed with brine (10

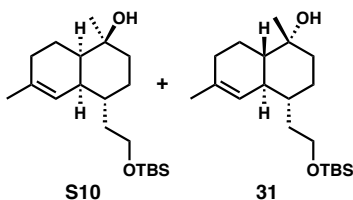
¹³ Houjeiry, T. I.; Poe, S. L.; McQuade, D. T. *Org. Lett.* **2012**, *14*, 4394–4397.

mL), dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 5% EtOAc/hexanes) to afford the title compound (0.298 g, 92%) as a colorless oil. ¹H NMR (CDCl₃, 500 MHz) δ 5.09 (d, *J* = 9.5 Hz, 1H), 3.71 – 3.60 (m, 2H), 3.56 – 3.48 (m, 2H), 2.55 – 2.24 (m, 6H), 2.24 – 2.10 (m, 2H), 1.88 – 1.79 (m, 1H), 1.73 (s, 3H), 1.69 – 1.59 (m, 1H), 1.39 (qd, *J* = 13.0, 4.8 Hz, 1H), 1.22 – 1.13 (m, 1H), 0.89 (s, 9H), 0.04 (s, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 211.0, 132.0, 131.7, 61.2, 47.6, 43.4, 42.4, 41.0, 38.1, 36.3, 35.3, 31.1, 26.1, 23.0, 18.4, –5.1, –5.2; IR (thin film) ν 2954, 2930, 2897, 2857, 1717, 1255, 1096, 835, 775 cm⁻¹; HRMS (CI) *m/z* calcd for C₁₉H₃₅ClO₂SiNH₄ (M + NH₄)⁺ 376.2439, found 376.2437. ¹H-NOESY-2D (500 MHz, CDCl₃) spectra were obtained for ketone **S7** and selected NOE interactions are shown.



Decalones S8 and S9. To a solution of alkyl chloride **S7** (0.298 g, 0.829 mmol) in *tert*-butyl alcohol (8.3 mL) at 30 °C was added potassium *tert*-butoxide (0.62 mL, 1.6 M in THF, 0.995 mmol) dropwise *via* syringe. After stirring for 6 h at 30 °C, the reaction was quenched with saturated NH₄Cl solution (16 mL). The aqueous layer was extracted with pentane (3 x 20 mL), and the combined organic extracts were washed with water (4 x 20 mL) and brine (20 mL). The organic layer was dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 5% EtOAc/hexanes) to afford a 1:1 mixture of *trans*- and *cis*-decalones (176.4 mg, 71%) as a colorless oil, which was characterized as a mixture. ¹H NMR (CDCl₃, 500 MHz) δ 5.54 (s, 1H), 5.30 (s, 1H), 3.78 – 3.62 (m, 4H), 2.51 – 2.34 (m, 4H), 2.33 – 2.25 (m, 2H), 2.22 – 1.77 (m, 14H), 1.70 – 1.29 (m, 6H), 1.67 (s, 3H), 1.63 (s, 3H), 0.897

(s, 9H), 0.895 (s, 9H), 0.059 (s, 6H), 0.058 (s, 6H); ^{13}C NMR (125 MHz, CDCl_3) δ 214.0, 212.8, 135.7, 124.5, 122.3, 61.6, 61.1, 51.2, 47.1, 46.1, 42.2, 41.1, 38.1, 38.0, 35.9, 35.3, 34.9, 32.7, 39.8, 28.2, 28.1, 26.1, 23.81, 23.80, 23.0, 21.9, 18.4, -5.12, -5.14, -5.17. IR (thin film) ν 2953, 2928, 2857, 1713, 1472, 1254, 1100, 836, 776 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{19}\text{H}_{34}\text{O}_2\text{SiNa}$ ($\text{M} + \text{Na}$) $^+$ 345.2226, found 345.2226.

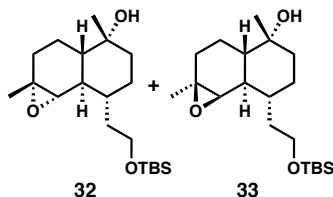


***cis*-Decalin S10 and *trans*-decalin 31.** Methylmagnesium bromide (1.54 mL, 3.0 M in Et_2O , 4.62 mmol) was added dropwise to a solution of a mixture of decalones **S8** and **S9** (0.213 g, 0.923 mmol) in THF (9.2 mL) cooled to 0 $^\circ\text{C}$. The reaction mixture was allowed to warm to room temperature for 30 min, and the reaction was quenched with saturated aqueous NH_4Cl solution (12 mL) at 0 $^\circ\text{C}$. The aqueous layer was extracted with Et_2O (3 x 20 mL). The combined organic extracts were washed with brine (20 mL), dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 5 \rightarrow 10% EtOAc /hexanes), yielding *trans*-decalin **31** (0.103 g, 46%) and *cis*-decalin **S10** (0.101 g, 45%) as colorless oils.

cis-Decalin **S10**: ^1H NMR (CDCl_3 , 500 MHz) δ 5.57 (d, $J = 4.0$ Hz, 1H), 3.71 – 3.64 (m, 1H), 3.63 – 3.56 (m, 1H), 2.03 – 1.94 (m, 2H), 1.91 – 1.81 (m, 3H), 1.72 – 1.36 (m, 6H), 1.65 (s, 3H), 1.35 – 1.22 (m, 2H), 1.30 (s, 3H), 1.10 (qd, $J = 13.0, 4.0$ Hz, 1H), 0.89 (s, 9H), 0.04 (s, 6H); ^{13}C NMR (125 MHz, CDCl_3) δ 134.7, 124.9, 72.6, 61.7, 45.5, 40.1, 36.4, 36.2, 35.3, 31.2, 29.4, 28.0, 26.1, 23.7, 18.7, -5.08, -5.12; IR (thin film) ν 3380 (br), 2956, 2929, 2893, 2857, 1462, 1254,

1095, 835, 774 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{20}\text{H}_{38}\text{O}_2\text{SiNa}$ ($\text{M} + \text{Na}$)⁺ 361.2539, found 361.2547.

trans-Decalin **31**: ^1H NMR (CDCl_3 , 500 MHz) δ 5.53 (s, 1H), 3.74 – 3.67 (m, 1H), 3.67 – 3.59 (m, 1H), 2.06 – 1.86 (m, 4H), 1.85 – 1.76 (m, 1), 1.72 – 1.60 (m, 2H), 1.66 (s, 3H), 1.47 – 1.28 (m, 4H), 1.22 (s, 3H), 1.17 – 1.00 (m, 3H), 0.90 (s, 9H), 0.06 (s, 6H); ^{13}C NMR (125 MHz, CDCl_3) δ 134.3, 123.5, 70.7, 61.5, 47.8, 40.5, 40.1, 38.5, 36.1, 31.0, 28.6, 27.9, 26.1, 23.8, 22.6, 18.5, –5.07, –5.11; IR (thin film) ν 3455 (br), 2954, 2928, 2857, 1462, 1375, 1255, 1096, 835, 774 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{20}\text{H}_{38}\text{O}_2\text{SiNa}$ ($\text{M} + \text{Na}$)⁺ 361.2539, found 361.2544.

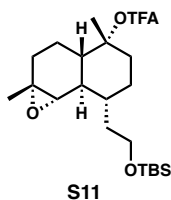


Epoxides 32 and 33. To a solution of *trans*-decalin **31** (0.109 g, 0.321 mmol) in acetone (16.1 mL) was added saturated aqueous NaHCO_3 (10.7 mL). The resulting mixture was cooled to 0 °C and a solution of Oxone® (0.217 g, 0.353 mmol) in H_2O (2 mL) was added dropwise over 5 minutes. The reaction mixture was stirred vigorously for 30 min at 0 °C, diluted with H_2O (20 mL) and extracted with EtOAc (4 x 20 mL). The combined organic extracts were diluted with hexanes until cloudy, dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 10 → 20% EtOAc/hexanes) to afford β -epoxide **33** (25.7 mg, 23%) and α -epoxide **32** (72.4 mg, 64%) as colorless oils.

Epoxide **33**: ^1H NMR (CDCl_3 , 500 MHz) δ 3.78 – 3.71 (m, 1H), 3.71 – 3.63 (m, 1H), 3.17 (s, 1H), 2.02 – 1.93 (m, 1H), 1.90 – 1.78 (m, 2H), 1.69 – 1.56 (m, 4H), 1.55 – 1.38 (m, 4H), 1.30 (s, 3H), 1.20 – 1.06 (m, 2H), 1.16 (s, 3H), 0.99 (s, 1H), 0.89 (s, 9H), 0.05 (s, 6H); ^{13}C NMR (125 MHz, CDCl_3) δ 70.8, 62.1, 61.3, 58.5, 41.9, 41.1, 40.1, 36.2, 35.9, 29.1, 29.0, 27.7, 26.1, 24.9,

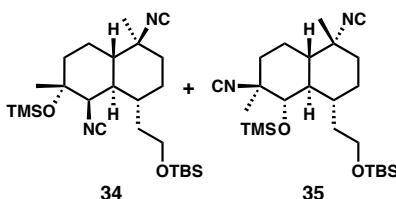
21.7, 18.5, -5.12, -5.16; IR (thin film) ν 3480 (br), 2929, 2857, 1462, 1377, 1255, 1096, 836, 775 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{20}\text{H}_{38}\text{O}_3\text{SiNa}$ ($\text{M} + \text{Na}$)⁺ 377.2488, found 377.2488.

Epoxide **32**: ¹H NMR (CDCl_3 , 500 MHz) δ 3.78 – 3.70 (m, 1H), 3.70 – 3.61 (m, 1H), 2.90 (s, 1H), 2.13 – 2.01 (m, 2H), 1.78 – 1.70 (m, 1H), 1.68 – 1.54 (m, 3H), 1.52 – 1.44 (m, 1H), 1.41 – 1.32 (m, 4H), 1.29 (s, 3H), 1.18 (s, 3H), 1.17 – 1.06 (m, 1H), 1.04 (s, 1H), 0.89 (s, 9H), 0.85 (t, J = 11.8 Hz, 1H), 0.05 (s, 6H); ¹³C NMR (125 MHz, CDCl_3) δ 70.4, 62.4, 61.1, 58.2, 47.3, 41.4, 39.7, 36.4, 36.3, 30.8, 28.6, 28.2, 26.1, 23.7, 19.6, 18.4, -5.11, -5.18; IR (thin film) ν 3481 (br), 2954, 2928, 2856, 1450, 1378, 1255, 1095, 836, 775 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{20}\text{H}_{38}\text{O}_3\text{SiNa}$ ($\text{M} + \text{Na}$)⁺ 377.2488, found 377.2483.



Trifluoroacetate S11. A mixture of epoxide **32** (79.3 mg, 0.224 mmol) and pyridine (60 μL , 0.672 mmol) in CH_2Cl_2 (2.2 mL) at 0 °C was treated with trifluoroacetic anhydride (50 μL , 0.335 mmol). After 30 minutes, the reaction was quenched with 1 M HCl (3 mL). The aqueous layer was extracted with CH_2Cl_2 (3 x 3 mL). The combined organic extracts were washed with water (5 mL), washed with saturated NaHCO_3 solution (5 mL), dried over MgSO_4 , filtered and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 5% EtOAc/hexanes) to afford the title compound (82.3 mg, 82%) as a colorless oil. ¹H NMR (CDCl_3 , 500 MHz) δ 3.76 – 3.69 (m, 1H), 3.67 – 3.59 (m, 1H), 2.88 (s, 1H), 2.70 (dt, J = 14.8, 3.3 Hz, 1H), 2.15 – 2.02 (m, 2H), 1.86 – 1.79 (m, 1H), 1.68 – 1.52 (m, 3H), 1.57 (s, 3H), 1.49 – 1.21 (m, 4H), 1.30 (s, 3H), 1.05 (qd, J = 14.0, 3.5 Hz, 1H), 0.91 (t, J = 11.0 Hz, 1H), 0.88 (s, 9H), 0.04 (s, 6H); ¹³C NMR (125 MHz, CDCl_3) δ 156.2 (q, J = 41 Hz), 114.5 (q, J = 285.4 Hz),

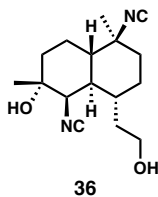
88.2, 62.0, 60.7, 58.0, 48.6, 41.2, 36.2, 35.7, 34.3, 30.8, 27.7, 26.1, 23.73, 23.65, 19.6, 18.4, –5.17, –5.24; IR (thin film) ν 2956, 2930, 2859, 1779, 1452, 1374, 1255, 1157, 1092, 837, 777 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{22}\text{H}_{37}\text{F}_3\text{O}_4\text{SiNa}$ ($\text{M} + \text{Na}$)⁺ 473.2311, found 473.2301.



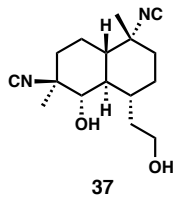
Diisocyanides 34 and 35. The following procedure was adopted from Pronin *et al.*⁶ A solution of trifluoroacetate **S11** (50.7 mg, 0.113 mmol) in TMSCN (0.11 mL) was cooled to 0 °C. A solution of scandium(III) trifluoromethanesulfonate (5.5 mg, 0.0113 mmol) in TMSCN (0.23 mL) was added, and the reaction mixture was allowed to warm to room temperature after 1 h. After 24 h at room temperature, the reaction was quenched with TMEDA (30 μL) and the volatiles were removed under reduced pressure. The crude residue was dissolved in hexanes (5 mL), washed with saturated aqueous NaHCO_3 solution (3 mL), dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 3 \rightarrow 6% EtOAc/hexanes) to afford diisocyanide **34** (5.0 mg, 10%) as thin film and diisocyanide **35** (6.0 mg, 11%) as a 4:1 mixture of C10 epimers.

Diisocyanide **34**: ^1H NMR (CDCl_3 , 500 MHz) δ 3.71 – 3.57 (m, 3H), 2.02 (dt, $J = 13.5, 4.0$ Hz, 1H), 1.90 – 1.09 (m, 12H), 1.45 (s, 3H), 1.31 (br s, 3H), 0.90 (s, 9H), 0.13 (s, 9H), 0.06 (s, 6H); ^{13}C NMR (125 MHz, CDCl_3) δ 159.0 (br t), 153.3 (br t), 73.5, 61.9 (br t, $J = 5.1$ Hz), 60.7, 60.2 (br t, $J = 4.9$ Hz), 42.6, 40.4, 38.7, 34.8, 34.6, 32.6, 27.6, 26.8, 26.1, 21.1, 20.5, 18.5, 2.4, –5.16, –5.19; IR (thin film) ν 2952, 2935, 2857, 2130, 1462, 1384, 1252, 1103, 1039, 840 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{25}\text{H}_{46}\text{N}_2\text{O}_2\text{Si}_2\text{Na}$ ($\text{M} + \text{Na}$)⁺ 485.2996, found 485.2990.

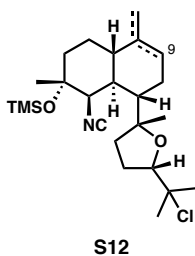
Diisocyanide **35**: ^1H NMR (CDCl_3 , 500 MHz) δ 3.71 (d, $J = 9.0$ Hz, 1H), 3.69 – 3.56 (m, 2H), 2.39 – 2.29 (m, 1H), 2.13 (dt, $J = 10.0, 3.5$ Hz, 1H), 1.98 – 1.73 (m, 4H), 1.51 – 1.06 (m, 7H), 1.41 (s, 3H), 1.40 (s, 3H), 0.88 (s, 9H), 0.28 (s, 9H), 0.03 (s, 6H); ^{13}C NMR (125 MHz, CDCl_3) δ 156.1 (br t), 155.5 (br t), 81.0, 63.0 (br t, $J = 5.1$ Hz), 60.8, 60.6 (br t, $J = 4.3$ Hz), 47.8, 44.4, 38.4, 38.0, 37.9, 37.3, 28.5, 26.7, 26.1, 21.8, 20.6, 18.4, 1.5, -5.2 , -5.3 ; IR (thin film) ν 2952, 2930, 2897, 2858, 2127, 1463, 1384, 1253, 1104, 840, 775 cm^{-1} ; HRMS (ESI) m / z calcd for $\text{C}_{25}\text{H}_{46}\text{N}_2\text{O}_2\text{Si}_2\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 485.2996, found 485.3009.



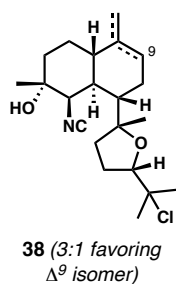
Diisocyanide 36. To a solution of silyl ether **34** (7.8 mg, 0.0169 mmol) in THF (0.2 mL) was added TBAF (90 μL , 1.0 M in THF, 0.0845 mmol). After 24 h, the reaction was quenched with saturated NaHCO_3 solution (1 mL). The reaction mixture was extracted with ether (3 \times 2 mL). The combined organic extracts were washed with water (5 mL), washed with brine (5 mL), dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 2 \rightarrow 3% $\text{MeOH}/\text{CH}_2\text{Cl}_2$) to afford diisocyanide **36** (2.7 mg, 57%) as a thin film. ^1H NMR (CDCl_3 , 500 MHz) δ 3.78 – 3.64 (s, 3H), 2.05 (dt, $J = 13.0, 3.3$ Hz, 1H), 1.96 – 1.59 (m, 9H), 1.59 – 1.31 (m, 3H), 1.45 (s, 3H), 1.35 (s, 3H), 1.29 – 1.17 (m, 2H); ^{13}C NMR (125 MHz, CDCl_3) δ 159.0 (br t), 153.2 (br t), 70.5, 61.2 (br t, $J = 5.4$ Hz), 60.21, 60.17 (br t, $J = 5.3$ Hz), 42.8, 40.2, 38.8, 34.8, 34.3, 32.7, 28.8, 26.6, 21.1, 20.8; IR (thin film) ν 3390 (br), 2937, 2871, 2134, 1451, 1384, 1122, 1036, 1003, 733 cm^{-1} ; HRMS (ESI) m / z calcd for $\text{C}_{16}\text{H}_{24}\text{N}_2\text{O}_2\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 299.1736, found 299.1733.



Diisocyanide 37. To a solution of silyl ether **35** (8.5 mg, 0.0184 mmol) in THF (0.2 mL) was added TBAF (0.10 mL, 1.0 M in THF, 0.0918 mmol). After 24 h, the reaction was quenched with saturated NaHCO₃ solution (1 mL). The reaction mixture was extracted with ether (3 x 2 mL). The combined organic extracts were washed with water (5 mL), washed with brine (5 mL), dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 3% MeOH/CH₂Cl₂) to afford diisocyanide **37** (3.9 mg, 76%) as a 4:1 mixture of C10 epimers. ¹H NMR (CDCl₃, 500 MHz) δ 4.67 (br s, 1H), 3.90 (td, *J* = 10.0, 3.0 Hz, 1H), 3.76 (ddd, *J* = 10.0, 6.5, 4.0 Hz, 1H), 3.55 (d, *J* = 10.0 Hz, 1H), 2.56 (br s, 1H), 2.29 – 2.19 (m, 1H), 2.12 (dt, *J* = 13.5, 3.8 Hz, 1H), 2.06 – 1.17 (m, 10H), 1.42 (br s, 3H), 1.40 (br s, 3H), 1.06 (t, *J* = 12.8 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 155.3 (br t), 153.1 (br t), 79.7, 62.8 (br t, *J* = 5.6 Hz), 60.8 (br t, *J* = 4.6 Hz), 60.1, 48.7, 42.1, 39.4, 39.1, 36.8, 36.3, 28.8, 28.2, 21.6, 19.3; IR (thin film) ν 3357 (br), 2938, 2870, 2130, 1448, 1384, 1116, 1041, 731 cm⁻¹; HRMS (ESI) *m/z* calcd for C₁₆H₂₄N₂O₂Na (M + Na)⁺ 299.1736, found 299.1727.



Isocyanide S12. The title compound was isolated as a side product in our previous synthesis of kalihinol B.¹⁴ The crude product was fully characterized after fluoride-mediated removal of the silyl ether.

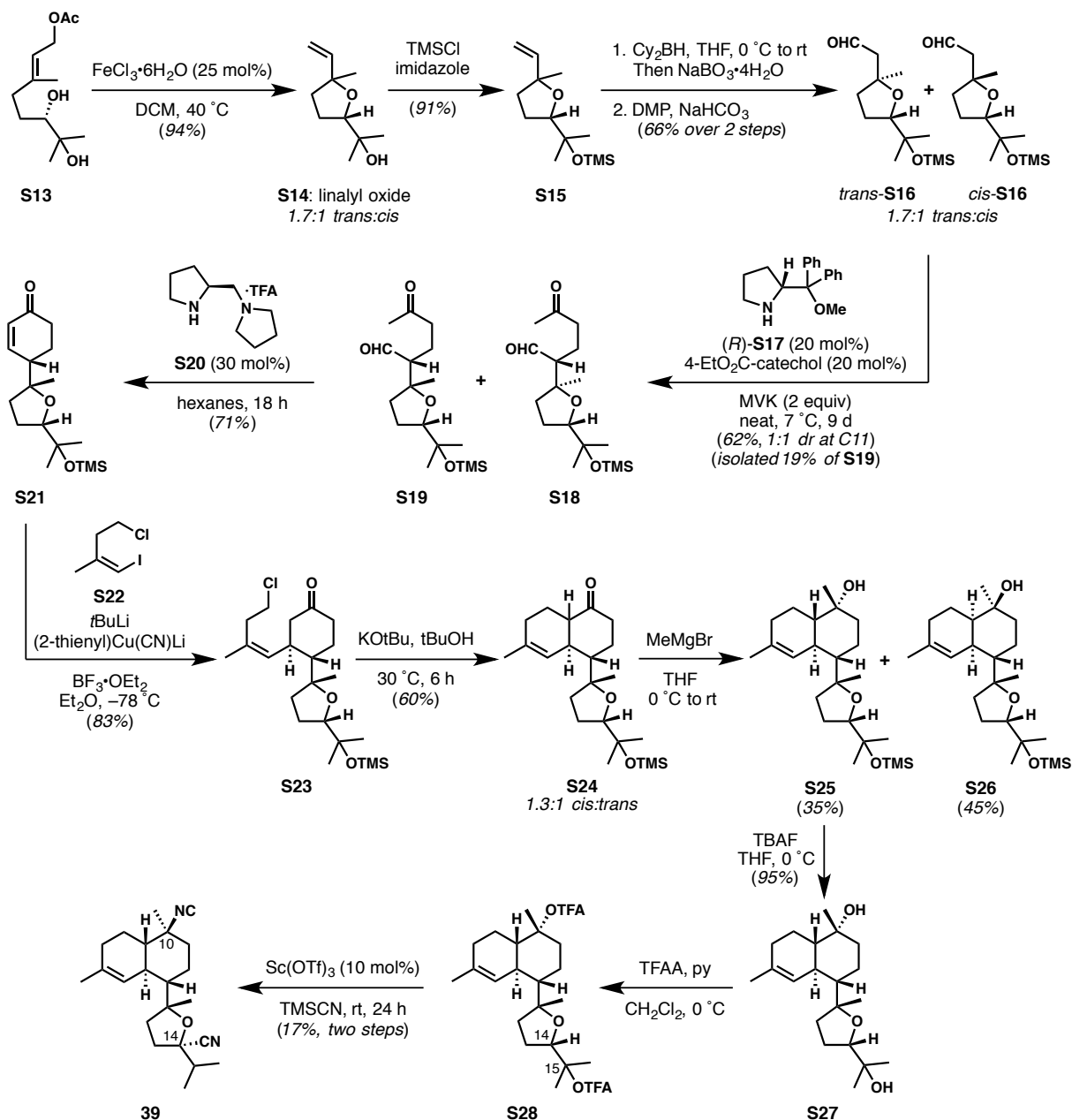


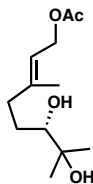
Isocyanide 38. To a solution of isocyanide **S12** (20.0 mg, 0.0456 mmol) in THF (0.5 mL) was added TBAF (0.10 mL, 1.0 M in THF, 0.100 mmol). After 1 h, the reaction was quenched with water (1 mL). The aqueous phase was extracted with ether (3 x 2 mL). The combined organic extracts were dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 2.5:55:45 EtOAc/CH₂Cl₂/hexanes) to afford a 3:1 mixture of **38** (8.3 mg, 50%) favoring the Δ^9 isomer as a thin film. The mixture of alkene isomers complicated the ¹H and ¹³C NMR spectra. $[\alpha]_D^{22} = +14.7$ (c = 0.83, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 5.35 (d, *J* = 4.0 Hz, 1H), 4.73 (s, 1H), 4.63 (s, 1H), 4.40 (s, 1H), 4.24 (s, 1H), 4.09 (dd, *J* = 9.0, 4.5 Hz, 1H), 4.07 (dd, *J* = 9.0, 4.0 Hz, 1H), 2.33 (dt, *J* = 12.8, 3.3 Hz, 1H), 2.20 (t, *J* = 11.2 Hz, 1H), 2.12 – 1.50 (complex), 1.65 (s, 6H), 1.62 (s, 3H), 1.57 (s, 3H), 1.53 (s, 3H), 1.43 (s, 3H), 1.42 (s, 3H), 1.34 – 1.13 (complex), 1.06 (m, 3H), 1.02 (m, 3H); ¹³C NMR (125 MHz,

¹⁴ Daub, M. E.; Prudhomme, J.; Le Roch, K.; Vanderwal, C. D. *J. Am. Chem. Soc.* **2015**, *137*, 4912–4915.

CDCl₃) δ 157.1 (br t), 151.1, 134.9, 120.7, 105.8, 87.8, 87.5, 84.9, 84.8, 71.7, 71.5, 71.1, 70.8, 63.7 (t, *J* = 5.8 Hz), 63.4 (t, *J* = 5.6 Hz), 47.6, 43.4, 42.0, 38.9, 38.8, 38.2, 37.8, 36.9, 36.0, 34.2, 32.8, 31.2, 30.7, 30.3, 29.8, 28.79, 28.76, 28.7, 26.0, 25.9, 25.7, 25.6, 24.1, 20.9, 17.9, 17.1; IR (thin film) ν 3418 (br), 2971, 2933, 2892, 2852, 2156, 2139, 1457, 1382, 1122, 1025, 758 cm⁻¹; HRMS (ESI) *m/z* calcd for C₂₁H₃₂ClNO₂Na (M + Na)⁺ 388.2019, found 388.2006.

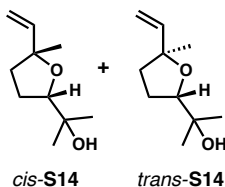
Scheme S1. Synthesis of C14-nitrile **39**.





S13

(*S,E*)-6,7-Dihydroxy-3,7-dimethyloct-2-enyl acetate (S13). (*S,E*)-6,7-Dihydroxy-3,7-dimethyloct-2-enyl acetate (**S13**) was prepared according to the literature procedure using a Sharpless dihydroxylation of geranyl acetate.¹⁵ The spectral data for this compound are consistent with those reported in the literature.¹⁵ $[\alpha]_D^{24} = -22.5$ ($c = 1.0$, CHCl_3), lit.¹⁵ $[\alpha]_D^{23} = +26.8$ ($c = 1.0$, CHCl_3) for the (*R*)-form; ¹H NMR (CDCl_3 , 500 MHz) δ 5.39 (t, $J = 7.0$ Hz, 1H), 4.58 (d, $J = 7.5$ Hz, 2H), 3.34 (ddd, $J = 10.5, 4.5, 1.5$ Hz, 1H), 2.37 – 2.28 (m, 1H), 2.25 (d, $J = 4.5$ Hz, 1H), 2.15 – 2.06 (m, 1H), 2.05 (s, 3H), 1.98 (s, 1H), 1.72 (s, 3H), 1.64 – 1.56 (m, 1H), 1.49 – 1.39 (m, 1H), 1.21 (s, 3H), 1.16 (s, 3H); ¹³C NMR (125 MHz, CDCl_3) δ 171.3, 142.2, 118.9, 78.2, 73.2, 61.5, 36.7, 29.6, 26.6, 23.4, 21.2, 16.6. The enantiomeric excess of **S13** was determined as 90% by ¹H NMR analysis of its corresponding mono-(*R*)-MTPA ester. ¹H NMR (500 MHz, CDCl_3) δ 5.30 ppm (t, 1H) for the *S* enantiomer; δ 5.23 ppm (t, 1H) for the *R* enantiomer, corresponding to the methyl group on the methoxy group of the ester.



***cis*- and *trans*-Linalyl oxides (S14).** The following procedure was adapted from Guérinot *et al.*¹⁶ To a solution of diol **S13** (4.23 g, 18.37 mmol) in CH_2Cl_2 (184 mL) was added $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ (1.24 g, 4.59 mmol) in one portion. After stirring at 40 °C for 5 h, the reaction mixture was cooled to

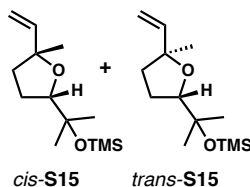
¹⁵ Surendra, K.; Corey, E. J. *J. Am. Chem. Soc.* **2008**, *130*, 8865–8869.

¹⁶ Guérinot, A.; Serra-Muns, A.; Gnamm, C.; Bensoussan, C.; Reymond, S.; Cossy, J. *Org. Lett.* **2010**, *12*, 1808–1811.

room temperature and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 25 → 30% Et₂O/pentane) to afford a 1:1.7 mixture of *cis*- and *trans*-linalyl oxides (3.06 g, 97%) as a colorless oil. The *cis*- and *trans*-THFs were separated for analytical purposes, and the spectral data for these compounds are consistent with those reported in the literature.^{17,18,19,20}

***trans*-S14:** $[\alpha]_D^{23} = +5.7$ (c = 0.95, CHCl₃), lit.¹⁷ $[\alpha]_D^{25} = +4.73$ (c = 2.07, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 5.88 (dd, *J* = 17.5, 10.5 Hz, 1H), 5.19 (dd, *J* = 17.0, 1.5 Hz, 1H), 5.00 (dd, *J* = 10.5, 1.5 Hz, 1H), 3.79 (t, *J* = 7.3 Hz, 1H), 2.16 (s, 1H), 1.94 – 1.80 (m, 3H), 1.76 – 1.69 (m, 1H), 1.31 (s, 3H), 1.23 (s, 3H), 1.13 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 143.8, 111.5, 85.7, 83.2, 71.3, 37.6, 27.4, 27.0, 26.4, 24.3.

***cis*-S14:** $[\alpha]_D^{23} = -3.8$ (c = 0.72, CHCl₃), lit.¹⁷ $[\alpha]_D^{25} = -2.94$ (c = 2.14, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 5.97 (dd, *J* = 17.3, 10.8 Hz, 1H), 5.20 (dd, *J* = 17.3, 1.3 Hz, 1H), 5.01 (dd, *J* = 10.5, 1.0 Hz, 1H), 3.86 (t, *J* = 7.3 Hz, 1H), 2.07 (s, 1H), 1.96 – 1.74 (m, 4H), 1.32 (s, 3H), 1.23 (s, 3H), 1.13 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 144.5, 111.7, 85.7, 82.9, 71.3, 38.1, 27.6, 26.6, 26.2, 24.5.



***cis*- and *trans*-Silyl ethers S15.** Imidazole (1.21 g, 17.8 mmol) was added in one portion to a solution of *cis*- and *trans*-linalyl oxides (S14) (1.01 g, 5.92 mmol) in CH₂Cl₂ (12 mL). Chlorotrimethylsilane (1.13 mL, 8.88 mmol) was added dropwise *via* syringe. After 30 min,

¹⁷ Méou, A.; Bouanah, N.; Archelas, A.; Zhang, X. M.; Guglielmetti, R.; Furstoss, R. *Synthesis* **1990**, 752–753.

¹⁸ Howell, A. R.; Pattenden, G. *J. Chem. Soc. Perkin Trans. 1* **1990**, 2715–2720.

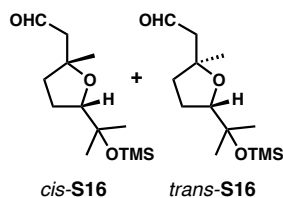
¹⁹ Fournier-Ngufack, C.; Lhoste, P.; Sinou, D. *Tetrahedron* **1997**, 53, 4353–4362.

²⁰ Wan, K. K.; Litz, J. P.; Vosburg, D. A. *Tetrahedron: Asymmetry* **2010**, 21, 2425–2428.

saturated NaHCO₃ solution (15 mL) was added to the reaction mixture. The reaction mixture was diluted with CH₂Cl₂ (10 mL) and water (10 mL). The biphasic mixture was separated, and the aqueous phase was extracted with CH₂Cl₂ (3 x 15 mL). The combined organic phases were washed with water (30 mL), dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 2.5% Et₂O/pentane) to afford a 1:1.7 mixture of *cis*-**S15** and *trans*-**S15** (1.30 g, 91%) as a colorless oil. The diastereomers were separated for analytical purposes.

trans-**S15**: $[\alpha]_D^{23} = +7.1$ (c = 1.0, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 5.86 (dd, *J* = 17.3, 10.8 Hz, 1H), 5.16 (dd, *J* = 17.3, 1.8 Hz, 1H), 4.97 (dd, *J* = 10.8, 1.8 Hz, 1H), 3.74 (t, *J* = 6.5 Hz, 1H), 1.94 – 1.75 (m, 3H), 1.71 – 1.62 (m, 1H), 1.30 (s, 3H), 1.21 (s, 3H), 1.20 (s, 3H), 0.11 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 144.3, 111.2, 86.4, 83.4, 75.2, 37.4, 27.9, 26.7, 26.4, 25.5, 2.7; IR (thin film) ν 2972, 2873, 1465, 1365, 1249, 1171, 1044, 839 cm⁻¹; HRMS (ESI) *m/z* calcd for C₁₃H₂₆O₂SiNa (M + Na)⁺ 265.1600, found 265.1604.

cis-**S15**: $[\alpha]_D^{23} = -0.27$ (c = 1.0, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 5.97 (dd, *J* = 17.5, 11.0 Hz, 1H), 5.20 (dd, *J* = 17.3, 1.8 Hz, 1H), 4.95 (dd, *J* = 11.0, 1.5 Hz, 1H), 3.78 (t, *J* = 7.0 Hz, 1H), 1.94 – 1.86 (m, 2H), 1.85 – 1.78 (m, 1H), 1.74 – 1.65 (m, 1H), 1.28 (s, 3H), 1.21 (s, 3H), 1.20 (s, 3H), 0.11 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 144.8, 111.3, 86.3, 83.1, 75.2, 38.0, 27.9, 26.6, 25.73, 25.70, 2.7; IR (thin film) ν 2972, 2872, 1459, 1365, 1250, 1172, 1045, 840 cm⁻¹; HRMS (ESI) *m/z* calcd for C₁₃H₂₆O₂SiNa (M + Na)⁺ 265.1600, found 265.1597.

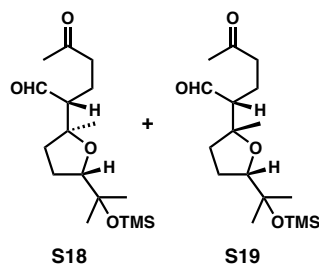


***cis*- and *trans*-Aldehydes S16.** A solution of $\text{BH}_3 \cdot \text{DMS}$ (0.60 mL, 6.31 mmol) in THF (10.5 mL) was cooled to 0 °C. Cyclohexene (1.28 mL, 12.62 mmol) was added dropwise *via* syringe over 2 min. After 20 min, the reaction mixture was allowed to warm to room temperature. After 2 h, the reaction mixture was cooled to 0 °C and a 1:1.7 mixture of *cis*- and *trans*-S15 (1.02 g, 4.21 mmol) was added as a solution in THF (10.5 mL). The transfer was completed with additional portions of THF (2 x 2 mL). The reaction mixture was allowed to stir at 0 °C for one hour before allowing to warm to room temperature. After 3 h, the reaction was quenched with water (30 mL) and $\text{NaBO}_3 \cdot 4\text{H}_2\text{O}$ (3.24 g, 21.04 mmol) was added in one portion. The reaction mixture was allowed to stir vigorously for 16 h. The reaction mixture was diluted with water (30 mL) and extracted with Et_2O (3 x 60 mL). The combined organic extracts were washed with brine (50 mL), dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 20% EtOAc /hexanes) to afford a mixture of *cis*- and *trans*-THFs contaminated with cyclohexanol. ^1H NMR and ^{13}C NMR spectra were complicated by the mixture of tetrahydrofuran diastereomers and cyclohexanol contaminant. Sodium bicarbonate (2.12 g, 25.18 mmol) was added to a solution of DMP (4.27 g, 10.07 mmol) in CH_2Cl_2 (40 mL) in one portion. A solution of *cis*- and *trans*-THFs contaminated with cyclohexanol (1.312 g, 5.036 mmol) in CH_2Cl_2 (5 mL) was added dropwise over 2 min. The transfer was completed with additional portions of CH_2Cl_2 (2 x 2.5 mL). After 2 h, the reaction was quenched with a 1:1 mixture of saturated NaHCO_3 solution and saturated $\text{Na}_2\text{S}_2\text{O}_3$ solution (50 mL), and allowed to stir until bubbling ceased (20 min). The phases were separated, and the

aqueous phase was extracted with CH₂Cl₂ (3 x 50 mL). The combined organic extracts were washed with 1 M NaOH (1 x 50 mL) and water (1 x 50 mL), dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 8.5% Et₂O/pentane) to afford a 1:1.7 mixture of *cis*-**S16** and *trans*-**S16** (0.714 g, 66% over two steps) as a colorless oil. The diastereomers were separated for analytical purposes using flash chromatography (SiO₂, 100% CH₂Cl₂).

trans-**S16**: $[\alpha]_D^{22} = +11.2$ (c = 0.90, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 9.82 (t, *J* = 3.0 Hz, 1H), 3.73 (t, *J* = 7.3 Hz, 1H), 2.58 (dd, *J* = 14.8, 2.8 Hz, 1H), 2.52 (dd, *J* = 14.5, 3.0 Hz, 1H), 2.00 – 1.72 (m, 4H), 1.31 (s, 3H), 1.21 (s, 3H), 1.17 (s, 3H), 0.11 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 203.0, 87.0, 81.4, 74.8, 54.4, 38.0, 27.7, 27.5, 26.4, 25.9, 2.7; IR (thin film) ν 2972, 2898, 2873, 2736, 1724, 1458, 1380, 1250, 1175, 1068, 1044, 840 cm⁻¹; HRMS (ESI) *m* / *z* calcd for C₁₃H₂₆O₃SiNa (M + Na)⁺ 281.1549, found 281.1554.

cis-**S16**: $[\alpha]_D^{22} = -49.4$ (c = 1.0, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 9.86 (dd, *J* = 3.5, 2.5 Hz, 1H), 3.77 (t, *J* = 7.3 Hz, 1H), 2.65 (dd, *J* = 15.0, 2.5 Hz, 1H), 2.46 (dd, *J* = 15.0, 3.5 Hz, 1H), 2.02 – 1.88 (m, 2H), 1.87 – 1.80 (m, 1H), 1.79 – 1.72 (m, 1H), 1.29 (s, 3H), 1.24 (s, 3H), 1.16 (s, 3H), 0.10 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 203.9, 86.8, 81.4, 74.9, 54.4, 38.0, 27.3, 27.0, 26.6, 26.2, 2.6; IR (thin film) ν 2970, 2873, 2736, 1723, 1456, 1380, 1250, 1177, 1040, 840 cm⁻¹; HRMS (ESI) *m* / *z* calcd for C₁₃H₂₆O₃SiNa (M + Na)⁺ 281.1549, found 281.1539.



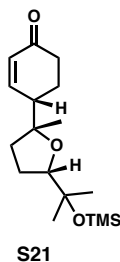
Keto-aldehydes S18 and S19. The following procedure was adapted from Peelen *et al.*²¹ A 2-dram vial was charged with a mixture of *cis*- and *trans*-aldehydes **S16** (0.714 g, 2.76 mmol), prolinol catalyst **S17**²² (0.148 g, 0.553 mmol), and 4-EtO₂C-catechol (0.101 g, 0.553 mmol). The vial was sealed with a septum, the headspace was purged with argon, and the reaction mixture was cooled to 0 °C. Methyl vinyl ketone (0.45 mL, 5.53 mmol) was added *via* syringe. The reaction mixture was allowed to stir at 0 °C until the solution became homogenous (30 min) before being placed in a 7 °C refrigerator for 9 days. The reaction mixture was purified directly using flash chromatography (SiO₂, 12.5% EtOAc/hexanes) to afford keto-aldehyde **S19** (0.177 g, 19%) as a colorless oil and keto-aldehyde **S18** (0.318 g, 35%) containing ca. 25% other diastereomers.

Keto-aldehyde **S18**: $[\alpha]_D^{22} = -48.6$ ($c = 1.0$, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 9.70 (d, $J = 3.0$ Hz, 1H), 3.59 (dd, $J = 8.5, 6.5$ Hz, 1H), 2.51 (ddd, $J = 18.0, 8.5, 5.8$ Hz, 1H), 2.45 – 2.34 (m, 2H), 2.11 (s, 3H), 1.97 – 1.77 (m, 6H), 1.74 – 1.59 (m, 1H), 1.20 (s, 5H), 1.14 (s, 3H), 0.09 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 208.3, 205.1, 87.3, 83.8, 74.4, 60.6, 41.7, 35.9, 30.1, 27.5, 26.2, 26.0, 25.7, 19.0, 2.7; IR (thin film) ν 2972, 2896, 2873, 2730, 1718, 1455, 1363, 1249, 1175, 1041, 839 cm⁻¹; HRMS (CI) m / z calcd for C₁₇H₃₂O₄SiH (M + H)⁺ 329.2148, found 329.2133.

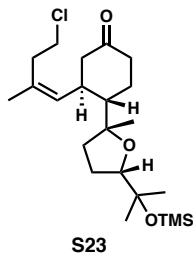
²¹ Peelen, T. J.; Chi, Y.; Gellman, S. H. *J. Am. Chem. Soc.* **2005**, *127*, 11598–11599.

²² Enders, D.; Kipphardt, H.; Gerdes, P.; Breña Valle, L. J.; Bhushan, V. *Bull. Soc. Chim. Belg.* **1988**, *97*, 691–704.

Keto-aldehyde **S19**: $[\alpha]_D^{22} = -92.7$ ($c = 1.0$, CHCl_3); $^1\text{H NMR}$ (CDCl_3 , 500 MHz) δ 9.90 (br s, 1H), 3.77 (t, $J = 7.3$ Hz, 1H), 2.56 (ddd, $J = 18.0, 8.0, 5.5$ Hz, 1H), 2.50 (dt, $J = 11.0, 1.8$ Hz, 1H), 2.33 (dt, $J = 18.0, 7.5$ Hz, 1H), 2.09 (s, 3H), 2.06 – 1.77 (m, 4H), 1.70 – 1.63 (m, 1H), 1.61 – 1.52 (m, 1H), 1.23 (s, 3H), 1.14 (s, 3H), 1.12 (s, 3H), 0.07 (s, 9H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 208.3, 207.1, 87.0, 85.0, 74.9, 59.5, 41.9, 37.2, 30.1, 27.2, 26.8, 25.8, 23.4, 19.5, 2.6; IR (thin film) ν 2970, 2873, 2730, 1718, 1449, 1364, 1250, 1176, 1038, 840 cm^{-1} ; HRMS (CI) m/z calcd for $\text{C}_{17}\text{H}_{32}\text{O}_4\text{SiH}$ ($\text{M} + \text{H}$) $^+$ 329.2148, found 329.2138.

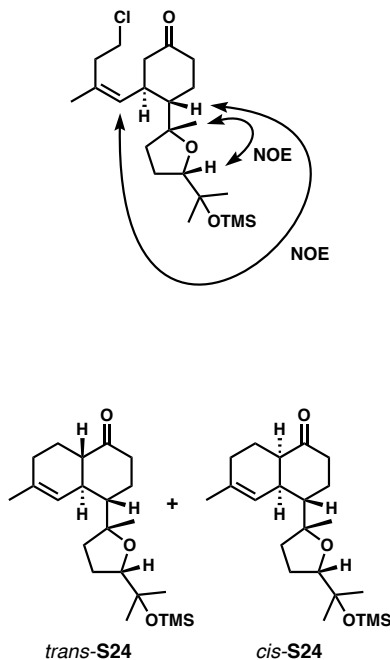


Enone S21. A solution of keto-aldehyde **S19** (0.177 g, 0.539 mmol) and catalyst **S20**¹³ (43.4 mg, 0.162 mmol) in hexanes (5.4 mL, previously sparged with argon gas for 20 min) was stirred at room temperature for 21 h. The reaction mixture was concentrated *in vacuo*, and the resulting crude residue was purified using flash chromatography (SiO_2 , 7.5% EtOAc/hexanes) to afford the title compound (0.119 g, 71%) as a colorless oil. $[\alpha]_D^{23} = -54.0$ ($c = 1.0$, CHCl_3); $^1\text{H NMR}$ (CDCl_3 , 500 MHz) δ 7.22 (dt, $J = 10.5, 1.5$ Hz, 1H), 6.06 (dd, $J = 10.3, 2.8$ Hz, 1H), 3.77 (t, $J = 7.0$ Hz, 1H), 2.67 – 2.60 (m, 1H), 2.50 (dt, $J = 16.3, 3.8$ Hz, 1H), 2.33 (td, $J = 15.5, 5.0$ Hz, 1H), 2.05 – 1.98 (m, 1H), 1.97 – 1.81 (m, 3H), 1.75 – 1.58 (m, 2H), 1.23 (s, 3H), 1.15 (s, 3H), 1.11 (s, 3H), 0.08 (s, 9H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 200.1, 154.2, 129.9, 86.4, 84.3, 74.9, 46.4, 37.8, 35.9, 27.3, 26.6, 26.3, 25.6, 22.7, 2.6; IR (thin film) ν 2969, 2872, 1683, 1452, 1380, 1249, 1178, 1042, 839 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{17}\text{H}_{30}\text{O}_3\text{SiNa}$ ($\text{M} + \text{Na}$) $^+$ 333.1862, found 333.1850.



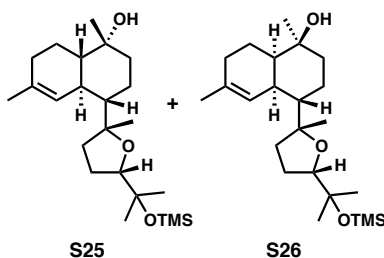
Ketone S23. To a solution of (*Z*)-4-chloro-1-iodo-2-methylbut-1-ene (**S22**)³ (0.154 g, 0.669 mmol) in Et₂O (1.1 mL, previously sparged with argon gas for 20 min) at $-78\text{ }^{\circ}\text{C}$ was added *tert*-butyllithium (0.84 mL, 1.59 M in pentane, 1.34 mmol) dropwise *via* syringe. After 20 min, lithium 2-thienylcyanocuprate solution (2.70 mL, 0.25 M in THF, 0.669 mmol) was added dropwise *via* syringe. The reaction mixture was allowed to stir for 1 h at $-78\text{ }^{\circ}\text{C}$. A solution of enone **S21** (0.139 g, 0.446 mmol) in Et₂O (0.5 mL) was added dropwise *via* syringe. The transfer was completed with additional portions of Et₂O (2 x 0.25 mL). Immediately following this addition, BF₃•OEt₂ (75 μL , 0.580 mmol) was added dropwise *via* syringe. After allowing the reaction mixture to stir for 3 h at $-78\text{ }^{\circ}\text{C}$, the reaction was quenched with 9:1 saturated NH₄Cl solution/NH₄OH (12 mL). After warming to room temperature, the solution turned a deep blue color. The aqueous layer was extracted with Et₂O (3 x 15 mL). The combined organic extracts were washed with 9:1 saturated NH₄Cl solution/NH₄OH (2 x 15 mL), washed with brine (15 mL), dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 7.5% EtOAc/hexanes) to afford the title compound (0.150 g, 81%) as a white solid (mp 56–58 $^{\circ}\text{C}$). $[\alpha]_{\text{D}}^{23} = -4.3$ ($c = 1.0$, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 5.37 (d, $J = 9.5$ Hz, 1H), 3.69 (t, $J = 6.8$ Hz, 1H), 3.61 – 3.48 (m, 2H), 2.76 (ddd, $J = 18.5, 9.8, 4.5$ Hz, 1H), 2.58 (quin, $J = 5.9$ Hz, 1H), 2.47 – 2.35 (m, 3H), 2.29 (td, $J = 15.0, 5.5$ Hz, 1H), 2.15 (dd, $J = 14.5, 10.0$ Hz, 1H), 2.11 – 2.05 (m, 1H), 1.86 (dt, $J = 9.5, 6.5$ Hz, 1H), 1.79 (ddd, $J = 11.8, 8.5, 3.8$ Hz, 1H), 1.72 – 1.62 (m, 1H), 1.68 (s, 3H), 1.62 – 1.51 (m, 3H), 1.19

(s, 3H), 1.17 (s, 3H), 1.07 (s, 3H), 0.10 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 211.9, 133.9, 127.4, 85.6, 85.1, 75.3, 50.7, 47.7, 42.5, 40.4, 38.4, 36.7, 35.2, 28.1, 27.2, 25.5, 23.0, 21.8, 2.8; IR (thin film) ν 2964, 2873, 1718, 1454, 1378, 1249, 1174, 1043, 839 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{22}\text{H}_{39}\text{ClO}_3\text{SiNa}$ ($\text{M} + \text{Na}$) $^+$ 437.2255, found 437.2252. ^1H -NOESY-2D (500 MHz, CDCl_3) spectra were obtained for ketone **S23** and selected NOE interactions are shown.



***cis*- and *trans*-Decalones S24.** To a solution of alkyl chloride **S23** (0.150 g, 0.362 mmol) in *tert*-butyl alcohol (1.81 mL) at 30 °C was added potassium *tert*-butoxide (0.27 mL, 1.6 M in THF, 0.434 mmol) dropwise *via* syringe. After stirring for 6 h at 30 °C, the reaction was quenched with saturated NH_4Cl solution (12 mL). The aqueous layer was extracted with pentane (4 x 10 mL), and the combined organic extracts were washed with water (3 x 10 mL) and brine (10 mL). The organic layer was dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 5% EtOAc/hexanes) to afford a 1.3:1 mixture of *cis*- and *trans*-decalones (99.7 mg, 73%) as a colorless oil, which was characterized as a mixture. $[\alpha]_{\text{D}}^{22} = -79.2$ ($c = 1.0$, CHCl_3); ^1H NMR (CDCl_3 , 500 MHz) δ 6.27

(s, 1H), 5.38 (s, 1H), 3.80 (t, $J = 6.3$ Hz, 1H), 3.73 (dd, $J = 7.9, 5.5$ Hz, 1H), 2.80 – 2.72 (m, 1H), 2.68 – 2.61 (m, 1H), 2.44 – 2.34 (m, 3 H), 2.26 – 2.15 (m, 4H), 2.13 – 1.38 (m, 21H), 1.64 (s, 3H), 1.58 (s, 3H), 1.20 (s, 3H), 1.18 (s, 9H), 1.17 (s, 3H), 1.07 (s, 3H), 0.11 (s, 9H), 0.10 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 215.2, 213.1, 133.1, 133.0, 127.7, 125.5, 86.4, 86.1, 85.4, 85.2, 75.6, 75.4, 51.4, 50.8, 50.0, 45.6, 44.4, 41.0, 38.8, 38.6, 36.5, 36.4, 29.9, 29.6, 28.0, 27.9, 27.2, 26.0, 25.7, 25.1, 24.8, 24.2, 23.84, 23.77, 22.7, 22.2, 22.0, 19.4, 2.77, 2.74; IR (thin film) ν 2963, 2875, 1714, 1454, 1378, 1249, 1172, 1041, 839 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{22}\text{H}_{38}\text{O}_3\text{SiNa}$ ($\text{M} + \text{Na}$) $^+$ 401.2488, found 401.2474.

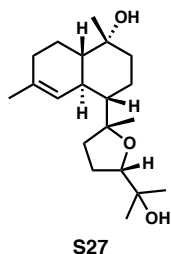


Decalins S25 and S26. Methylmagnesium bromide (0.44 mL, 3.0 M in Et_2O , 1.32 mmol) was added dropwise to a solution of a mixture of *cis*- and *trans*-decalones **S24** (99.7 mg, 0.263 mmol) in THF (2.63 mL) cooled to 0 °C. The reaction mixture was allowed to warm to room temperature for 30 min, and the reaction was quenched with saturated aqueous NH_4Cl solution (6 mL) at 0 °C. The aqueous layer was extracted with Et_2O (3 x 6 mL). The combined organic extracts were washed with brine (10 mL), dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 10% EtOAc /hexanes), yielding *trans*-decalin **S25** (36.5 mg, 35%) as a colorless oil and *cis*-decalin **S26** (47.1 mg, 45%) as a white solid (mp 82–84 °C).

cis-Decalin **S26**: $[\alpha]_{\text{D}}^{22} = +62.4$ ($c = 1.0$, CHCl_3); ^1H NMR (CDCl_3 , 500 MHz) δ 5.74, (d, $J = 5.0$ Hz, 1 H), 3.71 (t, $J = 6.3$ Hz, 1H), 2.24 – 2.15 (m, 1H), 2.00 – 1.80 (m, 5H), 1.66 – 1.44 (m, 9H),

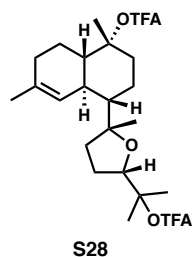
1.61 (s, 3H), 1.31 (s, 3H), 1.28 – 1.10 (m, 1H), 1.13 (s, 3 H), 1.11 (s, 3H), 1.08 (s, 3H), 0.07 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 130.2, 128.1, 86.2, 85.4, 75.7, 72.5, 49.1, 46.2, 38.2, 36.9, 35.3, 31.0, 28.2, 28.0, 26.8, 25.10, 25.07, 23.6, 19.9, 18.7, 2.7; IR (thin film) ν 3370 (br), 2963, 2940, 2894, 2872, 1458, 1376, 1249, 1043, 839 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{23}\text{H}_{42}\text{O}_3\text{SiNa}$ ($\text{M} + \text{Na}$) $^+$ 417.2801, found 417.2801.

trans-Decalin **S25**: $[\alpha]_{\text{D}}^{22} = -22.3$ ($c = 1.0$, CHCl_3); ^1H NMR (CDCl_3 , 500 MHz) δ 6.31 (s, 1H), 3.79 (dd, $J = 7.0, 5.5$ Hz, 1H), 2.17 (ap t, $J = 10.3$ Hz, 1H), 2.06 – 1.89 (m, 3H), 1.87 – 1.78 (m, 2H), 1.73 – 1.67 (m, 1H), 1.66 – 1.11 (m, 9H), 1.62 (s, 3H), 1.21 (s, 3H), 1.19 (s, 3H), 1.14 (s, 3H), 1.09 (s, 3H), 0.09 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 131.7, 126.6, 87.1, 85.1, 75.7, 70.7, 52.3, 47.8, 40.7, 38.7, 38.5, 30.9, 28.6, 28.1, 25.6, 24.8, 24.7, 23.8, 22.9, 19.1, 2.8; IR (thin film) ν 3464 (br), 2962, 2871, 1453, 1377, 1249, 1171, 1083, 1043, 839 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{23}\text{H}_{42}\text{O}_3\text{SiNa}$ ($\text{M} + \text{Na}$) $^+$ 417.2801, found 417.2801.



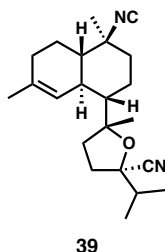
Diol S27. To a solution of *trans*-decalin **S25** (36.5 mg, 0.0925 mmol) in THF (0.93 mL) cooled to 0 °C was added TBAF (0.19 mL, 1.0 M in THF, 0.185 mmol). After 1 h, added additional TBAF (0.19 mL, 1.0 M in THF, 0.185). After 20 min, the reaction was quenched with water (5 mL). The reaction mixture was extracted with ether (4 x 5 mL). The combined organic extracts were dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 15% EtOAc/hexanes) to afford diol **S27** (28.2 mg, 95%) as a white solid (mp 98–100 °C). $[\alpha]_{\text{D}}^{23} = -21.1$ ($c = 0.77$, CHCl_3); ^1H NMR (CDCl_3 , 500 MHz) δ

6.25, (s, 1H), 3.85 (dd, $J = 7.8, 6.8$ Hz, 1H), 2.27 – 2.18 (m, 1H), 2.17 – 1.88 (m, 4H), 1.85 – 1.60 (m, 6H), 1.64 (s, 3H), 1.52 – 1.40 (m, 3H), 1.35 (ddd, $J = 24.5, 12.3, 5.8$ Hz, 1H), 1.28 – 1.12 (m, 2H), 1.24 (s, 3H), 1.22 (s, 3H), 1.15 (s, 3H), 1.10 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 132.8, 125.9, 87.1, 84.4, 71.4, 70.7, 52.3, 47.9, 40.7, 38.7, 38.5, 30.7, 28.7, 27.7, 25.8, 24.5, 24.3, 24.0, 22.8, 19.2; IR (thin film) ν 3436 (br), 2965, 2936, 2870, 1453, 1375, 1147, 1078, 892, 756 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{20}\text{H}_{34}\text{O}_3\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 345.2406, found 345.2415.

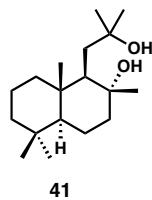


Bis(trifluoroacetate) S28. A mixture of diol **S27** (10.2 mg, 0.0316 mmol) and pyridine (30 μL , 0.371 mmol) in CH_2Cl_2 (0.32 mL) at $0\text{ }^\circ\text{C}$ was treated with trifluoroacetic anhydride (20 μL , 0.144 mmol). After 20 min, added additional pyridine (20 μL , 0.247 mmol) and trifluoroacetic anhydride (20 μL , 0.144 mmol). After 20 min, the reaction was quenched with 1 M HCl (2 mL). The aqueous layer was extracted with CH_2Cl_2 (3 x 2 mL). The combined organic extracts were washed with water (5 mL), washed with saturated NaHCO_3 solution (5 mL), dried over MgSO_4 , filtered and concentrated *in vacuo*. The crude residue was used directly in the subsequent reaction. $[\alpha]_{\text{D}}^{23} = -30.0$ ($c = 1.0$, CHCl_3); ^1H NMR (CDCl_3 , 500 MHz) δ 6.20 (s, 1H), 4.07 (dd, $J = 9.3, 3.8$ Hz, 1H), 2.75 (dt, $J = 14.8, 3.3$ Hz, 1H), 2.33 (ap t, $J = 10.3$ Hz, 1H), 2.08 – 1.92 (m, 4H), 1.86 – 1.78 (m, 1H), 1.73 – 1.49 (m, 7H), 1.62 (s, 3H), 1.57 (s, 3H), 1.53 (s, 3H), 1.44 (td, $J = 14.3, 3.8$ Hz, 1H), 1.33 (td, $J = 12.5, 2.8$ Hz, 1H), 1.19 (t, $J = 11.8$ Hz, 1H), 1.16 – 1.06 (m, 1H), 1.08 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 156.3 (q, $J = 36.3$ Hz), 156.2 (q, $J = 40.8$ Hz), 132.5, 125.3, 114.6 (q, $J = 285.6$ Hz), 114.5 (q, $J = 285.3$ Hz), 89.9, 89.0, 87.6, 82.9, 51.2,

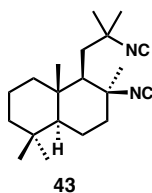
48.8, 38.2, 37.8, 34.9, 30.5, 25.1, 25.0, 23.72, 23.70, 22.6, 21.2, 21.3, 18.7; IR (thin film) ν 2962, 2931, 2877, 1779, 1456, 1375, 1220, 1156 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{24}\text{H}_{32}\text{F}_6\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$)⁺ 537.2051, found 537.2051.



C14-Nitrile 39. The following procedure was adopted from Pronin *et al.*⁶ Trifluoroacetate **S28** (10.2 mg, 0.0316 mmol) was dissolved in TMSCN (60 μL) and cooled to 0 $^{\circ}\text{C}$. A solution of scandium(III) trifluoromethanesulfonate (1.6 mg, 0.00316 mmol) in TMSCN (0.11 mL) was added to the reaction mixture dropwise *via* syringe. After 30 min, the reaction mixture was allowed to warm to room temperature. After 4 h, the reaction was quenched with TMEDA (20 μL) and the volatiles were removed under reduced pressure. The crude residue was dissolved in hexanes (6 mL), washed with saturated aqueous NaHCO_3 solution (6 mL), dried over MgSO_4 , filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO_2 , 3 \rightarrow 5% EtOAc/hexanes) to afford the title compound (2.0 mg, 19%) as a thin film. $[\alpha]_{\text{D}}^{22} = +32.1$ ($c = 0.20$, CHCl_3); ^1H NMR (CDCl_3 , 500 MHz) δ 6.02 (s, 1H), 2.28 – 2.20 (m, 1H), 2.20 – 2.02 (m, 4H), 2.02 – 1.77 (m, 7H), 1.72 – 1.48 (m, 3H), 1.65 (s, 3H), 1.39 – 1.27 (m, 1H), 1.31 (br s, 3H), 1.15 (d, $J = 6.5$ Hz, 3H), 1.08 (s, 3H), 1.05 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 152.6 (br t), 133.8, 123.8, 120.8, 88.9, 85.0, 60.6 (br t, $J = 5.1$ Hz), 51.9, 47.5, 40.6, 38.9, 38.7, 37.9, 35.5, 30.4, 25.3, 24.0, 21.8, 20.3, 18.2, 17.9; IR (thin film) ν 2965, 2933, 2875, 2128, 1453, 1384, 1063 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{22}\text{H}_{32}\text{N}_2\text{ONa}$ ($\text{M} + \text{Na}$)⁺ 363.2412, found 363.2417.

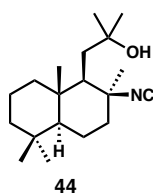


Diol 41. Methylmagnesium bromide (0.47 mL, 3.0 M in Et₂O, 1.40 mmol) was added dropwise to a solution of (3*aR*)-(+)-sclareolide (**40**) (70.0 mg, 0.280 mmol) in THF (2.8 mL) cooled to 0 °C. The reaction mixture was allowed to warm to room temperature for 30 min, and the reaction was quenched with saturated aqueous NH₄Cl solution (5 mL) at 0 °C. The aqueous layer was extracted with Et₂O (3 x 15 mL). The combined organic extracts were washed with brine (15 mL), dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 25% EtOAc/hexanes), yielding diol **41** (69.4 mg, 88%) as a white solid (mp 166–168 °C). $[\alpha]_D^{23} = +8.1$ (c = 1.0, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 4.06 (s, 2H), 1.88 (dt, *J* = 12.5, 3.0 Hz, 1H), 1.74 – 1.32 (m, 7H), 1.30 – 1.07 (m, 3H), 1.27 (s, 3H), 1.22 (s, 3H), 1.19 (s, 3H), 0.98 – 0.82 (m, 3H), 0.87 (s, 3H), 0.78 (s, 3H), 0.75 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 73.4, 69.8, 56.3, 54.8, 44.4, 42.0, 40.1, 38.9, 38.5, 33.6, 33.3, 28.5, 25.1, 21.8, 20.4, 18.7, 15.6; IR (thin film) ν 3230 (br), 2964, 2934, 2868, 1467, 2128, 1451, 1386, 1155, 1124, 940, 756 cm⁻¹; HRMS (ESI) *m/z* calcd for C₁₈H₃₄O₂Na (M + Na)⁺ 305.2456, found 305.2451.



Diisocyanide 43. The following procedure was adopted from Pronin *et al.*⁶ A mixture of diol (**41**) (25.8 mg, 0.0913 mmol) and pyridine (40 μL, 0.383 mmol) in CH₂Cl₂ (0.91 mL) at 0 °C was treated with trifluoroacetic anhydride (0.1 mL, 1.92 M in CH₂Cl₂, 0.192 mmol). After 15

minutes, the reaction was quenched with 1 M HCl (2 mL). The aqueous layer was extracted with hexanes (2 x 3 mL). The combined organic extracts were washed with water (5 mL), washed with saturated NaHCO₃ solution (5 mL), dried over MgSO₄, filtered and concentrated *in vacuo*. The crude trifluoroacetate was dissolved in TMSCN (0.1 mL) and cooled to 0 °C. A solution of scandium(III) trifluoromethanesulfonate (1.3 mg, 0.00274 mmol) in TMSCN (0.1 mL) was added. After 30 min, the reaction mixture was placed in a 3 °C refrigerator for 18 h. The reaction was quenched with TMEDA (20 μL) and the volatiles were removed under reduced pressure. The crude residue was suspended in hexanes (5 mL), washed with saturated aqueous NaHCO₃ solution (5 mL), dried over MgSO₄, filtered, and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 2.5% EtOAc/hexanes) to afford the title compound (6.3 mg, 23%) as a thin film. $[\alpha]_D^{24} = +17.1$ (c = 0.60, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 2.13 (dt, *J* = 14.0, 3.0 Hz, 1H), 1.87 – 1.31 (m, 7H), 1.53 (s, 3H), 1.50 (s, 3H), 1.48 (s, 3H), 1.29 – 1.11 (m, 3H), 1.09 – 1.75 (m, 3H), 1.04 (s, 3H), 0.90 (s, 3H), 0.85 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 157.0 (br t), 156.4 (br t), 61.4 (br t, *J* = 4.4 Hz), 57.4 (br t, *J* = 4.8 Hz), 55.4, 52.5, 42.0, 41.6, 40.9, 39.4, 37.9, 33.6, 33.5, 33.1, 32.6, 32.1, 21.9, 18.7, 18.1, 15.4; IR (thin film) ν 2929, 2841, 2133, 1465, 1444, 1392, 1367, 1156, 1131 cm⁻¹; HRMS (ESI) *m/z* calcd for C₂₀H₃₂N₂Na (M + Na)⁺ 323.2463, found 323.2456.



Isocyanide 44. The following procedure was adopted from Pronin *et al.*⁶ A mixture of diol (**41**) (69.4 mg, 0.246 mmol) and pyridine (80 μL, 1.03 mmol) in CH₂Cl₂ (2.5 mL) at 0 °C was treated with trifluoroacetic anhydride (70 μL, 0.516 mmol). After 15 minutes, the reaction was quenched

with 1 M HCl (3 mL). The aqueous layer was extracted with CH₂Cl₂ (3 x 5 mL). The combined organic extracts were washed with water (5 mL), washed with saturated NaHCO₃ solution (5 mL), dried over MgSO₄, filtered and concentrated *in vacuo*. The crude trifluoroacetate was dissolved in TMSCN (0.25 mL) and cooled to 0 °C. A solution of scandium(III) trifluoromethanesulfonate (2.4 mg, 0.00492 mmol) in TMSCN (0.16 mL) was added. After 2 h at 0 °C, the reaction was quenched with pyridine (50 µL) and the volatiles were removed under reduced pressure. The crude residue was dissolved in MeOH (2 mL) and cooled to 0 °C. Potassium carbonate (68.0 mg, 0.492 mmol) was added in one portion. After 15 min, the reaction mixture was filtered and concentrated *in vacuo*. The crude residue was purified using flash chromatography (SiO₂, 10% EtOAc/hexanes) to afford the title compound (14.8 mg, 21%) as a solid (mp 156–158 °C). $[\alpha]_D^{21} = +27.3$ (c = 1.0, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 2.10 (ap d, *J* = 14.0 Hz, 1H), 1.77 – 1.54 (m, 6H), 1.54 – 1.33 (m, 3H), 1.40 (s, 3H), 1.28 (s, 3H), 1.25 (s, 3H), 1.22 – 1.04 (m, 3H), 1.03 (s, 3H), 0.95 – 0.86 (m, 2H), 0.88 (s, 3H), 0.84 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 156.0 (br t, *J* = 4.9 Hz), 70.3, 61.7 (br t, *J* = 4.3 Hz), 55.6, 51.7, 42.3, 41.8, 40.6, 39.4, 38.9, 33.57, 33.55, 32.8, 32.3, 31.7, 21.9, 18.8, 18.2, 15.1; IR (thin film) ν 3486 (br), 2950, 2919, 2839, 2134, 1466, 1442, 1389, 1365, 1154, 1129 cm⁻¹; HRMS (ESI) *m* / *z* calcd for C₁₉H₃₃NONa (M + Na)⁺ 314.2460, found 314.2469.

IV. Evaluation of Inhibition of *P. falciparum* Growth

Method: Drug sensitive and resistant strains of *P. falciparum* malaria (BEI Resources, MR4/ATCC, Manassas, VA) were cultured in human type O+ erythrocytes in complete medium consisting of RPMI 1640 (Gibco), 0.043 mg/mL gentamicin (Gibco), 0.014 mg/mL hypoxanthine (Acros), 38.5 mM HEPES (Sigma), 0.18% sodium bicarbonate (Cellgro), 0.20% glucose (MP Biomedical), 0.003 mM NaOH (Sigma), 0.2% Albumax (Gibco), and 5% human serum as previously described.²³ Briefly, cultures were maintained in 25-cm² flasks (Corning) at a volume of 10 mL, gassed for 30 s with 3% CO₂, 1% O₂, and 96% N₂, and were finally incubated at 37 °C. The antiplasmodial activity was determined with an SYBR Green based parasite proliferation assay as previously described.²⁴ After 72 h of incubation in the presence of serial dilutions of compounds, the increase of parasite DNA contained in human red blood cells was evaluated. The relative fluorescence values were measured using a Molecular Devices SpectraMAX Gemini EM fluorimeter (excitation 495 nm, and emission 525 nm). Data were analyzed using Microsoft Excel and were plotted using SigmaPlot 10 (Systat).

²³ Le Roch, K. G.; Zhou, Y.; Blair, P. L.; Grainger, M.; Moch, J. K.; Haynes, J. D.; De La Vega, P.; Holder, A. A.; Batalov, S.; Carucci, D. J. *Science* **2003**, *301*, 1503–1508.

²⁴ Prudhomme, J.; McDaniel, E.; Ponts, N.; Bertani, S.; Fenical, W.; Jensen, P.; Le Roch, K. *PLoS One* **2008**, *3*, e2335.

Table S1. Activity of synthetic ICTs against wild-type (3D7) and chloroquine-resistant (Dd2) *P. falciparum*.

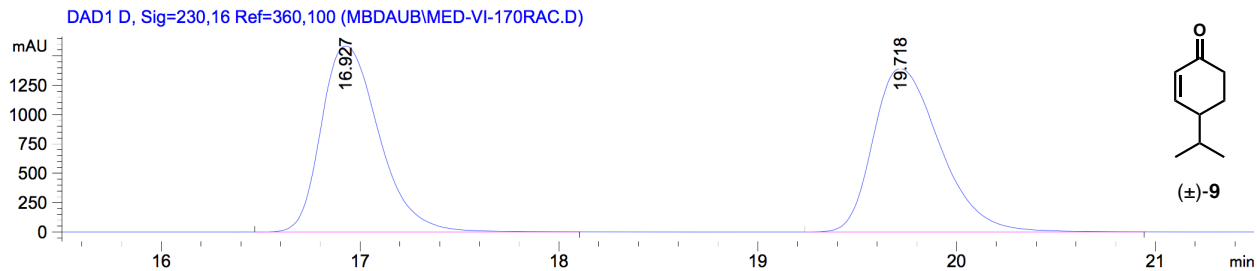
Compound	Pf strain ^a	IC50 [nM]	IC50 [μg/mL]	std error ^b	R	hillslope	Chloroquine ^c IC50 [μM]	std error
2	3D7	8.4	0.0033	0.0007	0.9914	1.342	0.0063	0.002
2	Dd2	4.6	0.0018	0.0002	0.9938	2.937	0.0399	0.0035
38	3D7	139	0.0508	0.0072	0.9952	2.3689	0.0052	0.0001
38	Dd2	144	0.0526	0.0093	0.9959	1.4242	0.0275	0.0086
39	3D7	175	0.0596	0.0117	0.9903	1.4691	0.0066	0.0004
39	Dd2	123	0.0418	0.0188	0.9808	0.6601	0.0413	0.0037
(±)-15	3D7	705	0.1631	0.0894	0.9899	0.5629	0.0059	0.0009
(±)-15	Dd2	247	0.057	0.0234	0.9961	0.5342	0.0322	0.0039
(±)-16^d	3D7	180	0.0417	0.0191	0.9748	0.8015	0.0091	0.0007
(±)-16^d	Dd2	45	0.0104	0.0073	0.9882	0.2933	0.0337	0.0031
22	3D7	12	0.0034	0.0006	0.9963	7.3863	0.0089	0.0018
22	Dd2	16	0.0043	0.0001	0.9979	4.6016	0.0497	0.0157
23	3D7	2.9	0.0008	0.0002	0.984	1.3984	0.0089	0.0018
23	Dd2	31	0.0084	0.0016	0.9917	1.5004	0.0497	0.0157
24	3D7	138	0.0342	0.0037	0.9935	2.6747	0.0089	0.0018
24	Dd2	200	0.0495	0.0089	0.9939	1.3615	0.0497	0.0157
28	3D7	15	0.0041	0.0006	0.9954	1.6148	0.0089	0.0018
28	Dd2	17	0.0047	0.0003	0.9983	3.1883	0.0497	0.0157
(±)-28^e	3D7	27	0.0074	0.0013	0.9927	--	0.0058	0.0005
(±)-28^e	Dd2	46	0.0127	0.0016	0.9918	--	0.0521	0.0274
29	3D7	1150	0.3155	0.0179	0.9989	2.0297	0.0089	0.0018
29	Dd2	958	0.2628	0.0427	0.9962	1.0929	0.0497	0.0157
30	3D7	312	0.0771	0.0032	0.9995	2.1804	0.0089	0.0018
30	Dd2	529	0.1308	0.0057	0.999	2.8612	0.0497	0.0157
(±)-36^f	3D7	302	0.0836	0.018	0.9773	4.7837	0.0091	0.0007
(±)-36^f	Dd2	205	0.0566	0.007	0.9833	3.2312	0.0337	0.0031
(±)-37^g	3D7	27	0.0075	0.0009	0.9831	2.043	0.0091	0.0007
(±)-37^g	Dd2	24	0.0065	0.0009	0.9879	1.2806	0.0337	0.0031
43	3D7	1.9	0.0006	0.0002	0.9833	1.7569	0.0059	0.0009
43	Dd2	1.6	0.0005	0.0002	0.9903	0.7528	0.0322	0.0039
44	3D7	244	0.0745	0.0083	0.997	1.4462	0.0061	0.0005
44	Dd2	416	0.1271	0.0438	1.2621	1.2721	0.0419	0.0905

^a3D7 is a chloroquine-sensitive strain of *P. falciparum*. Dd2 is a chloroquine-resistant strain of *P. falciparum*. ^bn = 3.

^cChloroquine was used as the antimalarial standard. ^dPrecipitate at 33.3 μg/mL. ^eAverage of three separate assays.

^fPrecipitate at 11.1 μg/mL. ^gPrecipitate at 3.7 μg/mL.

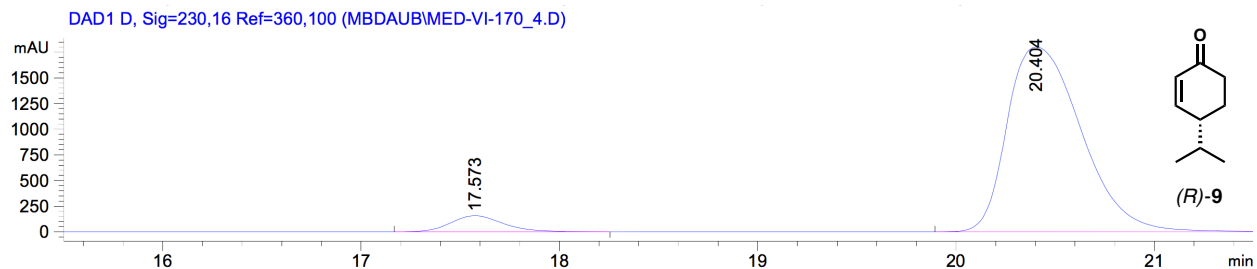
Determination of enantiomeric excess of (*R*)-cryptone (**9**)
 Chiralpak AD-H column, 2% *i*PrOH in hexanes, flow rate of 0.5 mL/min



Signal 3: DAD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.927	BB	0.3153	3.19316e4	1583.48645	49.5504
2	19.718	BB	0.3625	3.25110e4	1391.95129	50.4496

Totals : 6.44426e4 2975.43774

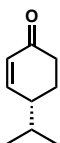


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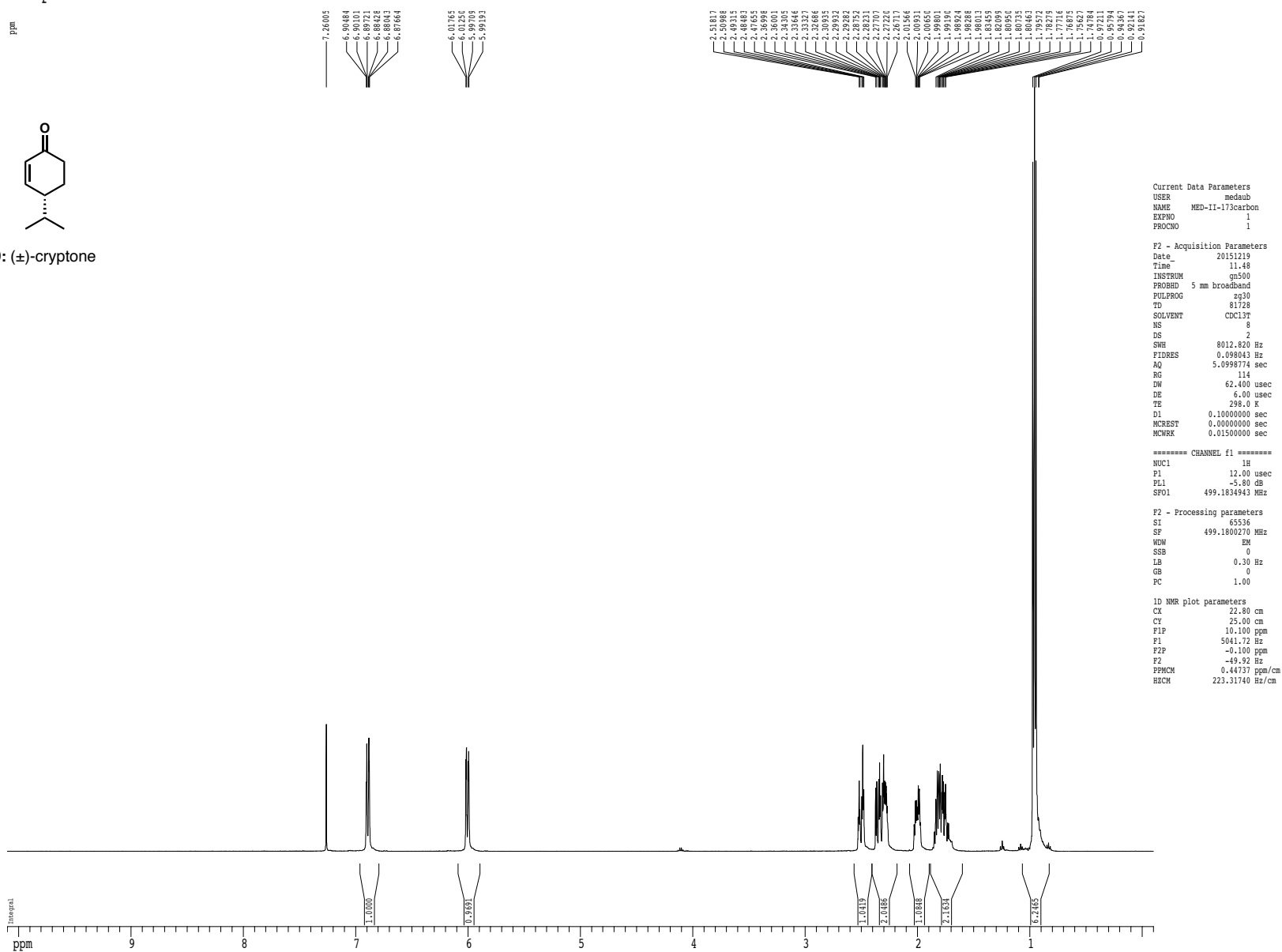
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Totals : 4.96805e4 1956.46533

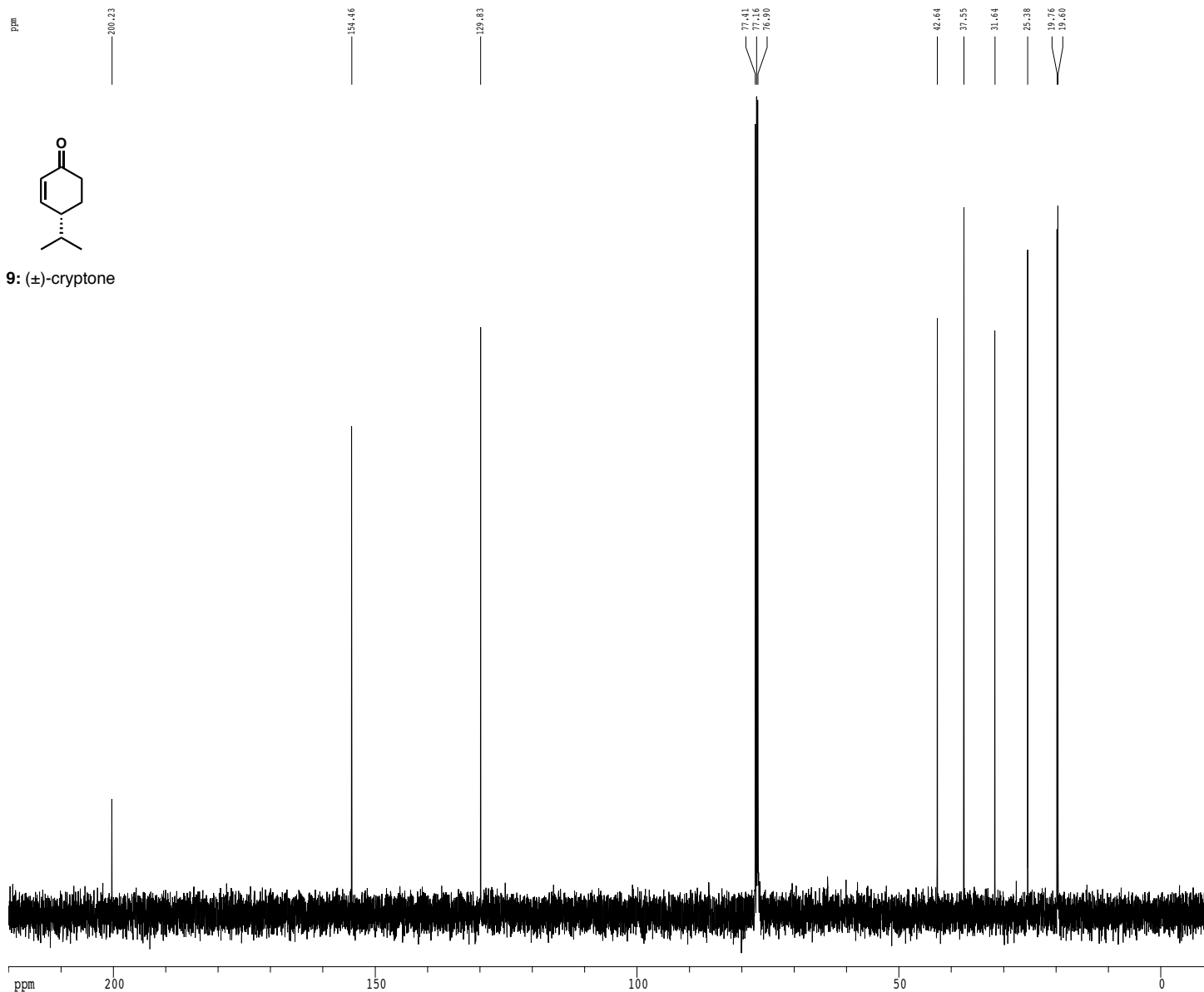
1H spectrum



9: (±)-cryptone



13C spectrum with 1H decoupling



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PROCNO    1

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DM         16.500 usec
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MCMK       0.01500000 sec

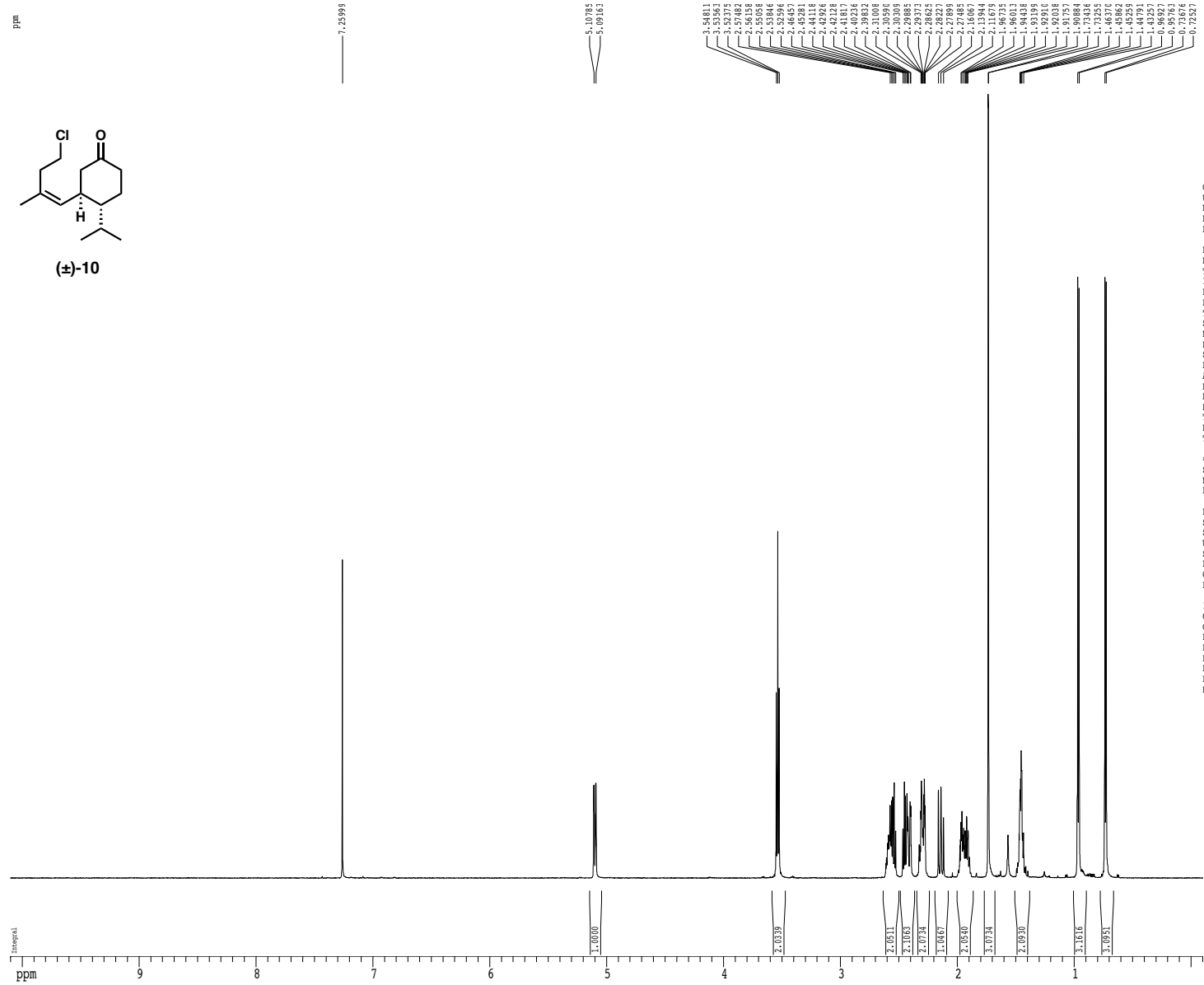
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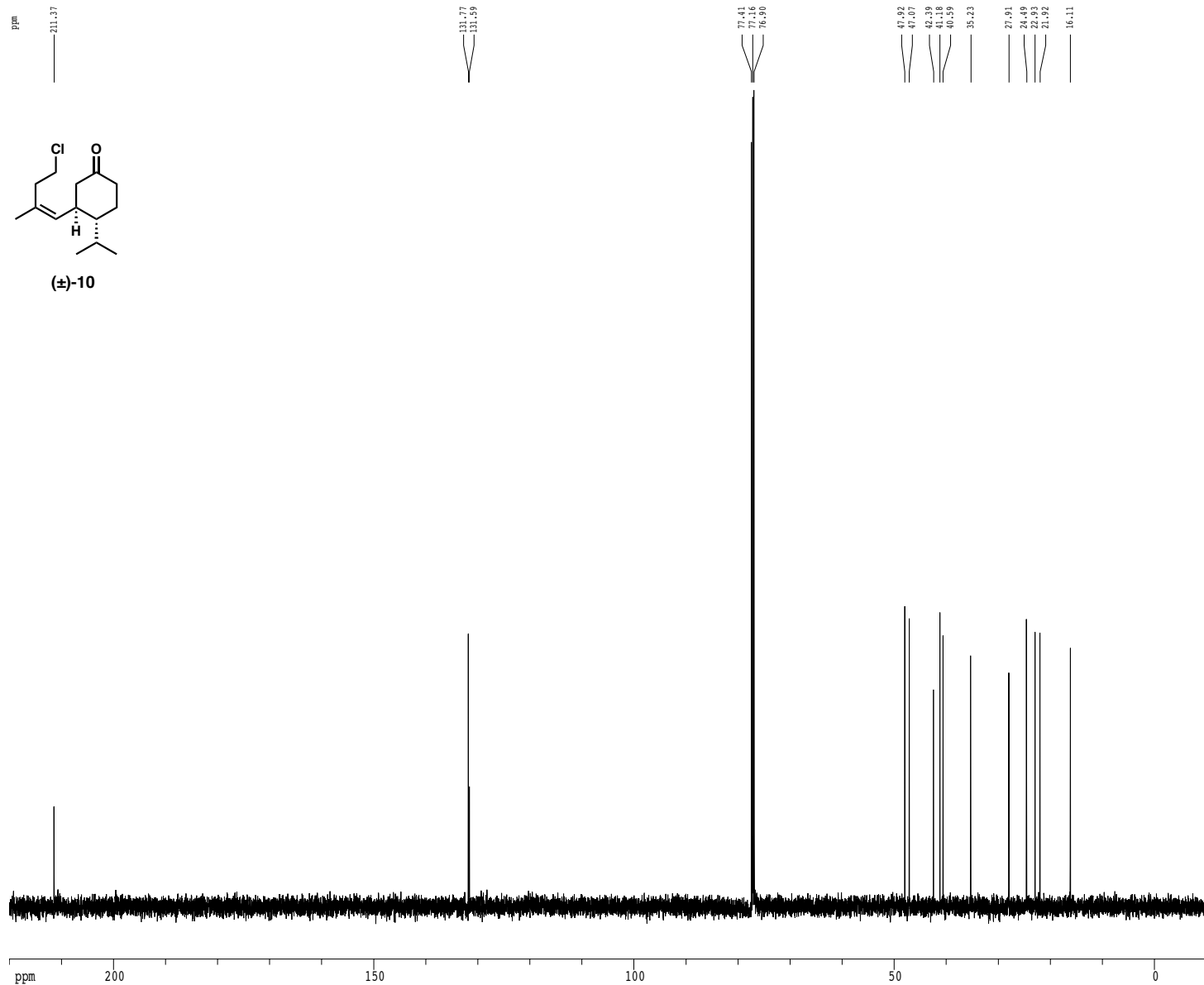
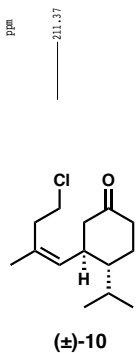
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HECM       1266.19946 Hz/cm
    
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1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling



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PROCNO    1

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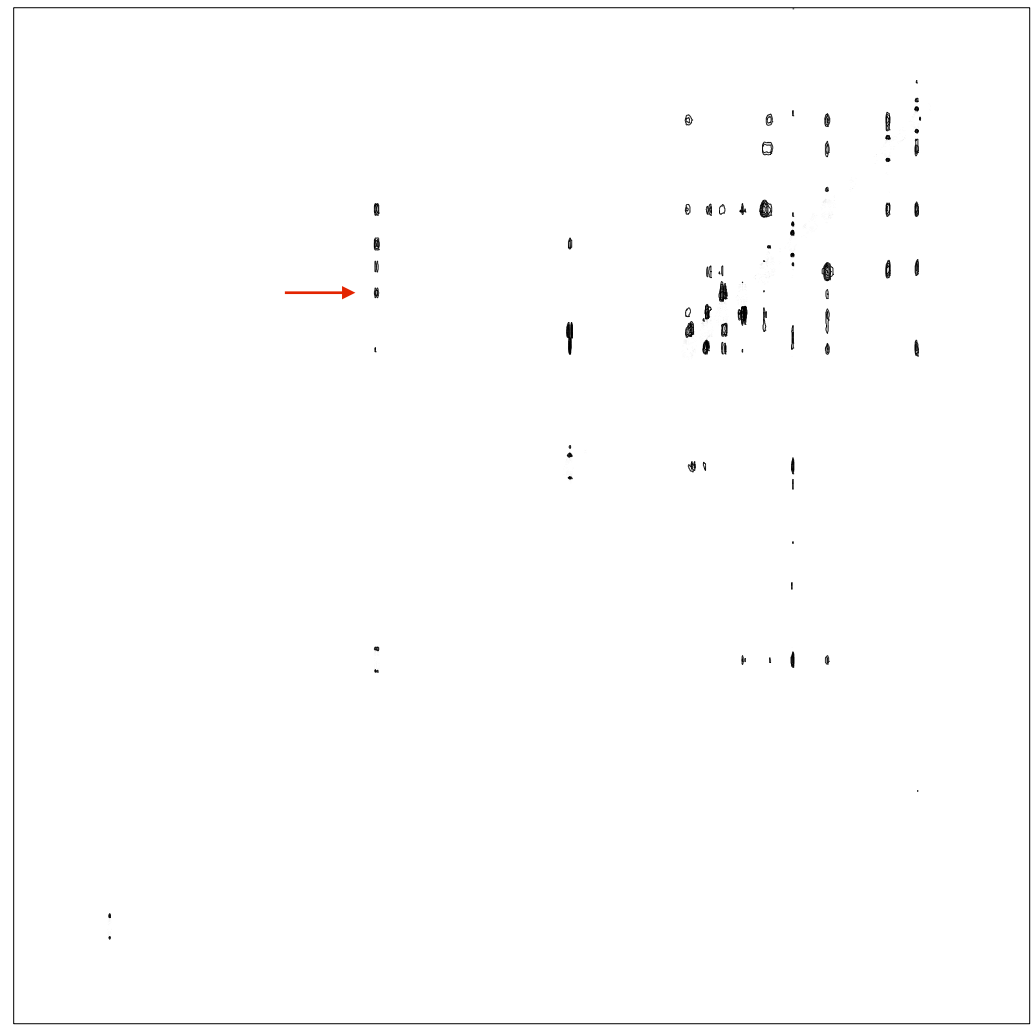
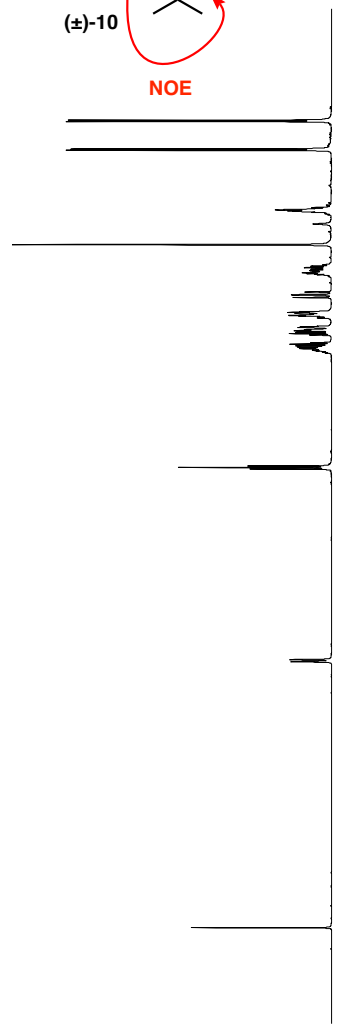
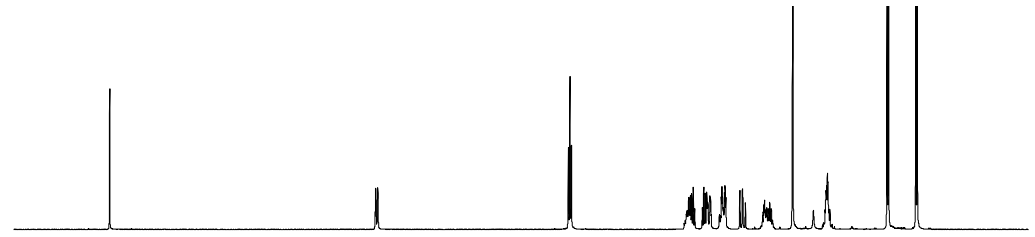
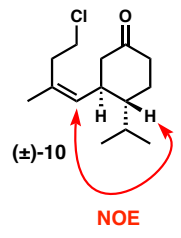
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GPMAM2     SINE.100
GPX1       0.00 t
GPF2       0.00 t
GPF1       0.00 t
GPF2       0.00 t
GPF1       30.00 t
GPF2       50.00 t
p15        500.00 usec
p16        1000.00 usec

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F2P        -10.000 ppm
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noesygp



ppm

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PROCNO 1

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S8 1.00000000 sec
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IN0 0.00020280 sec

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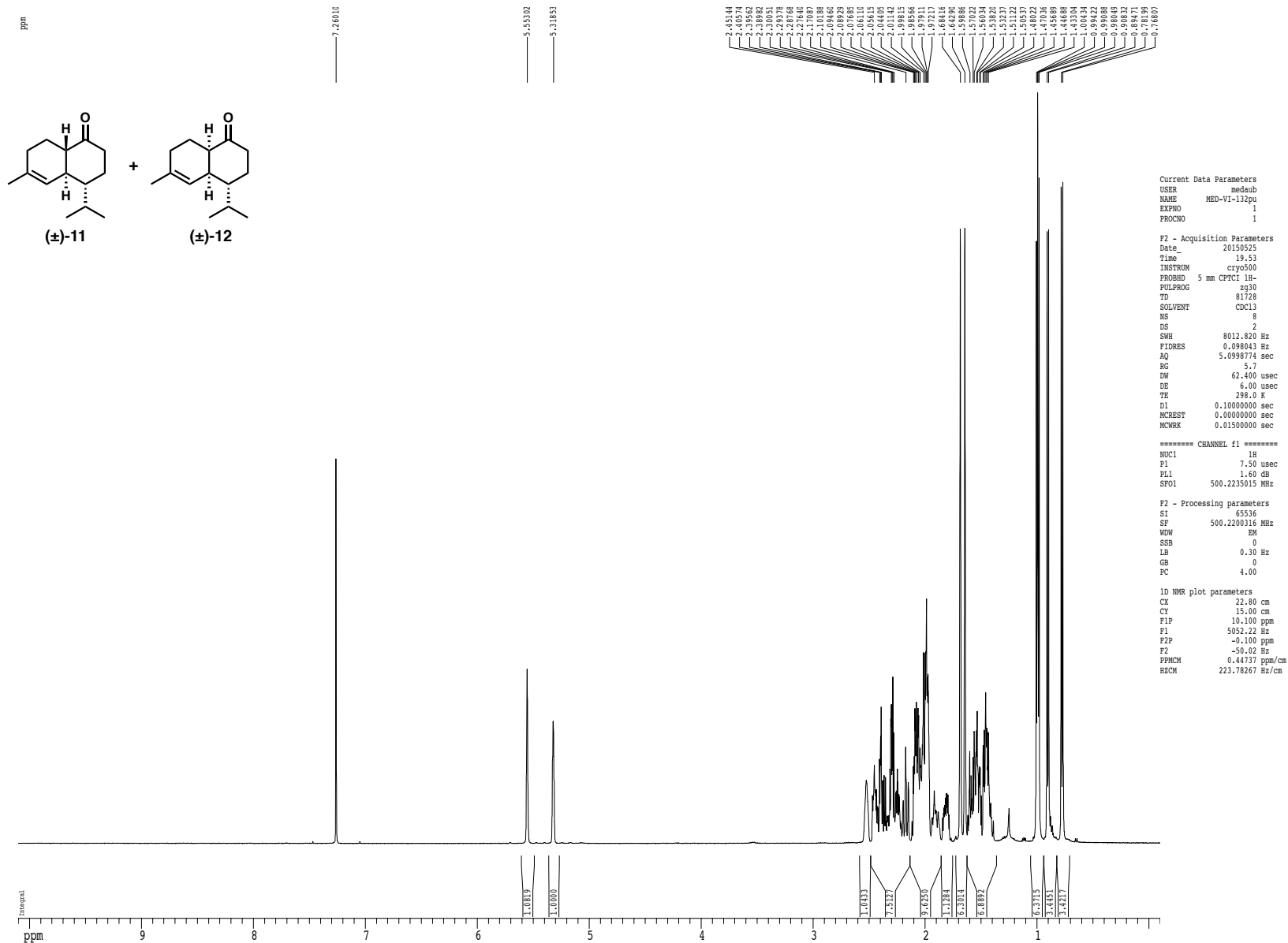
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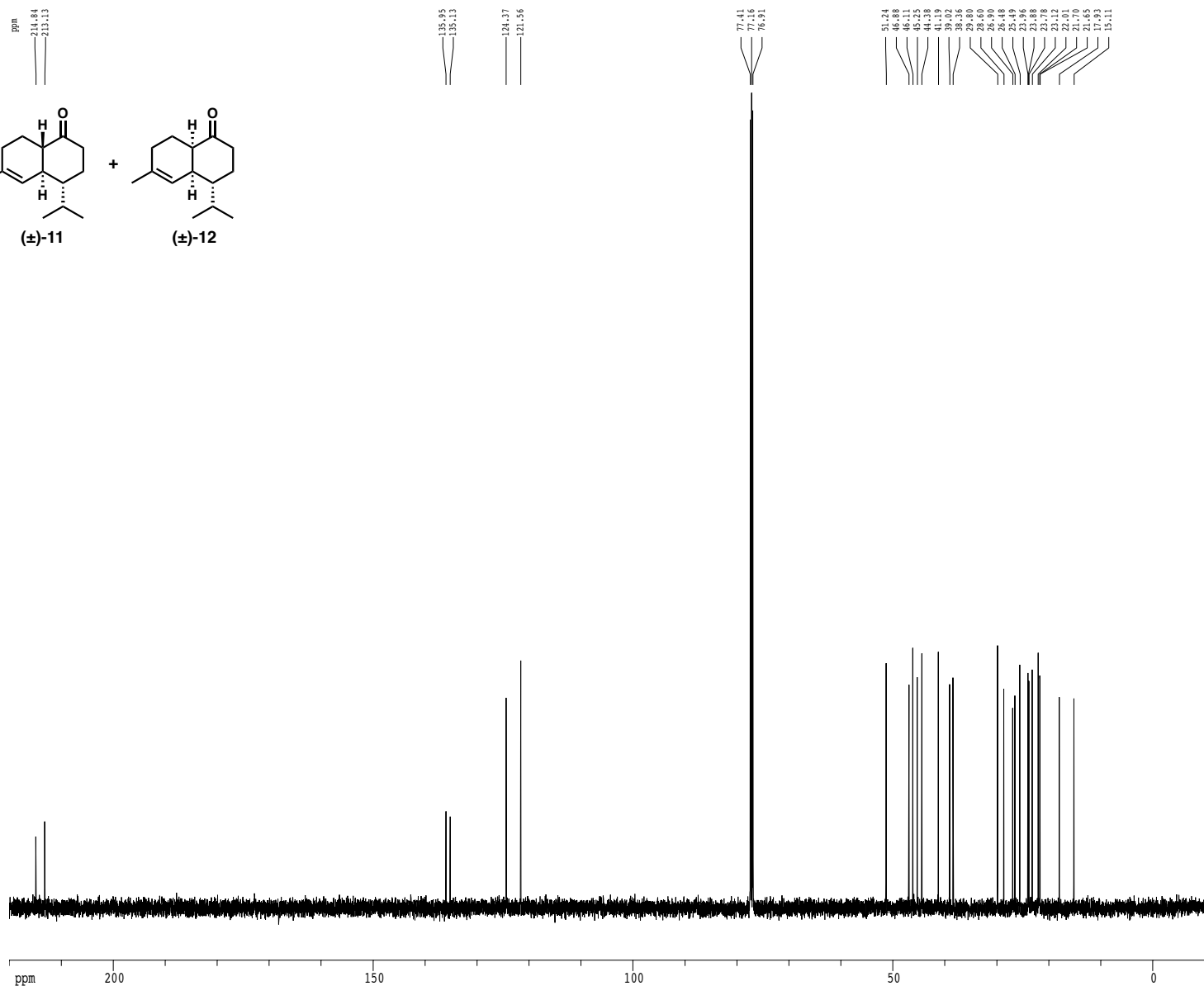
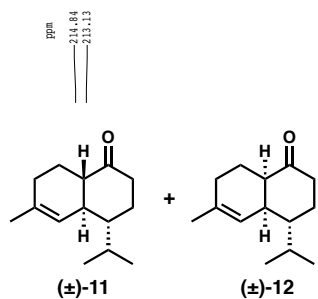
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LB 0 Hz
GB 0

ppm

1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling



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PROCNO       1

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FIDRES        0.462388 Hz
AQ            1.0813940 sec
RG            11585.2
DW            16.500 usec
DE            6.00 usec
TE            298.0 K
D1            0.25000000 sec
d11           0.03000000 sec
D16           0.00020000 sec
d17           0.00019600 sec
MCWREST       8.00000000 sec
MCWREK        0.01500000 sec
P2            33.10 usec

===== CHANNEL f1 =====
NUC1          13C
P1            16.55 usec
P11           500.00 usec
P12           2000.00 usec
PL0           120.00 dB
PL1           -1.00 dB
SFO1         125.7942548 MHz
SF1           2.70 dB
SP2           2.70 dB
SFOF1         Crp60, 0.5, 20.1
SFOF2         Crp60comp, 4
SFOFF1        0.00 Hz
SFOFF2        0.00 Hz

===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2          1H
PCPD2        100.00 usec
PL2           1.00 dB
PL12          24.50 dB
SFO2         500.2225011 MHz

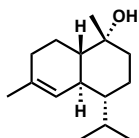
===== GRADIENT CHANNEL =====
GPMAM1       SINE.100
GPMAM2       SINE.100
GPY1         0.00 t
GPY2         0.00 t
GPY1         0.00 t
GPY2         0.00 t
GPY1         30.00 t
GPY2         50.00 t
p15          500.00 usec
p16          1000.00 usec

F2 - Processing parameters
SI            65536
SF            125.7804085 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            2.00

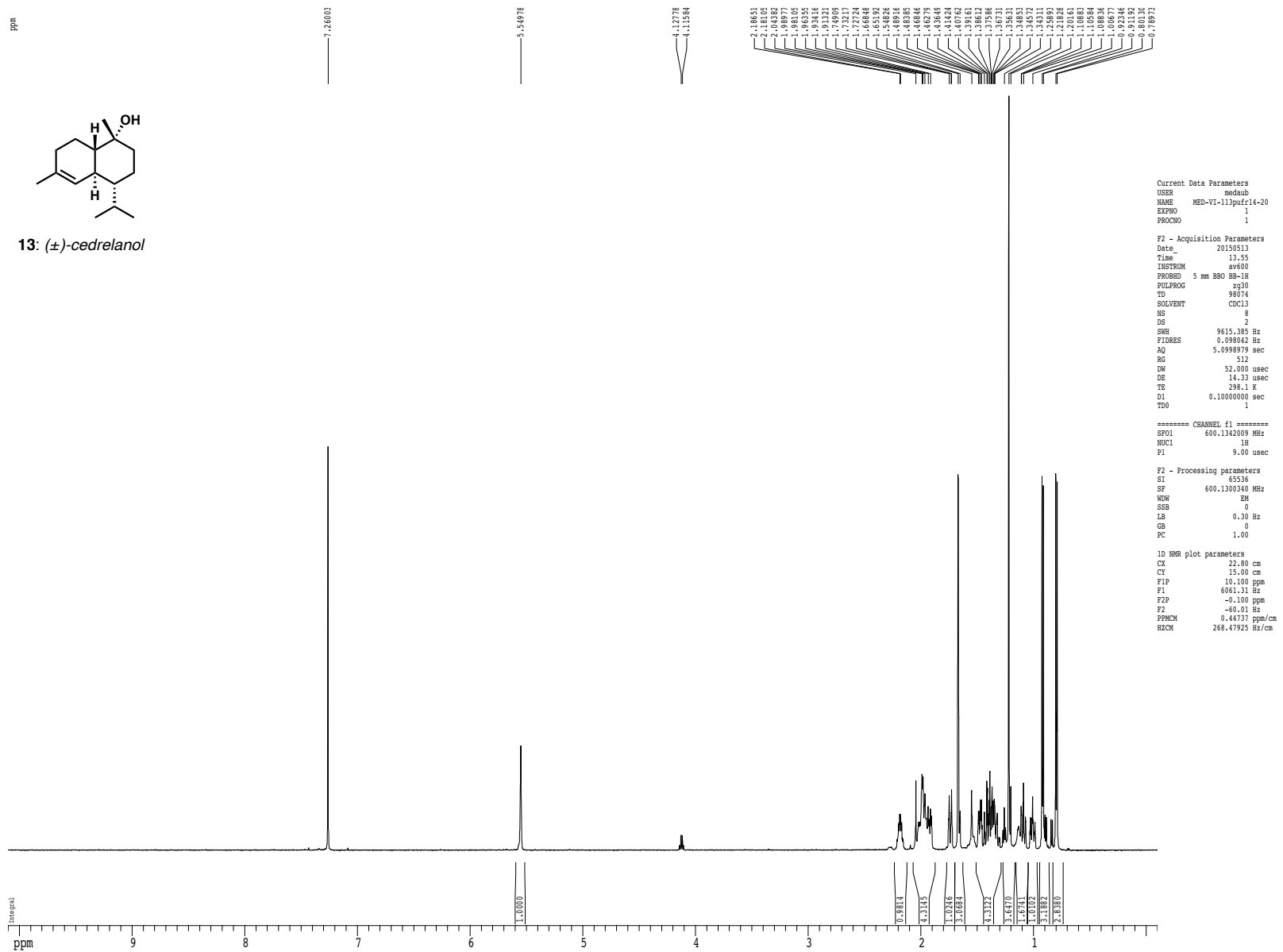
1D NMR plot parameters
CX            22.80 cm
CT            15.65 cm
PIF           220.000 ppm
F1            27671.69 Hz
F2P           -10.000 ppm
F2            -1257.80 Hz
FPMCM         10.88712 ppm/cm
BRCM          1268.83752 Hz/cm
    
```

1H spectrum

ppm



13: (±)-cedrelanol



```

Current Data Parameters
USER      medaub
NAME      MHD-VI-113puf14-20
EXPNO    1
PROCNO   1

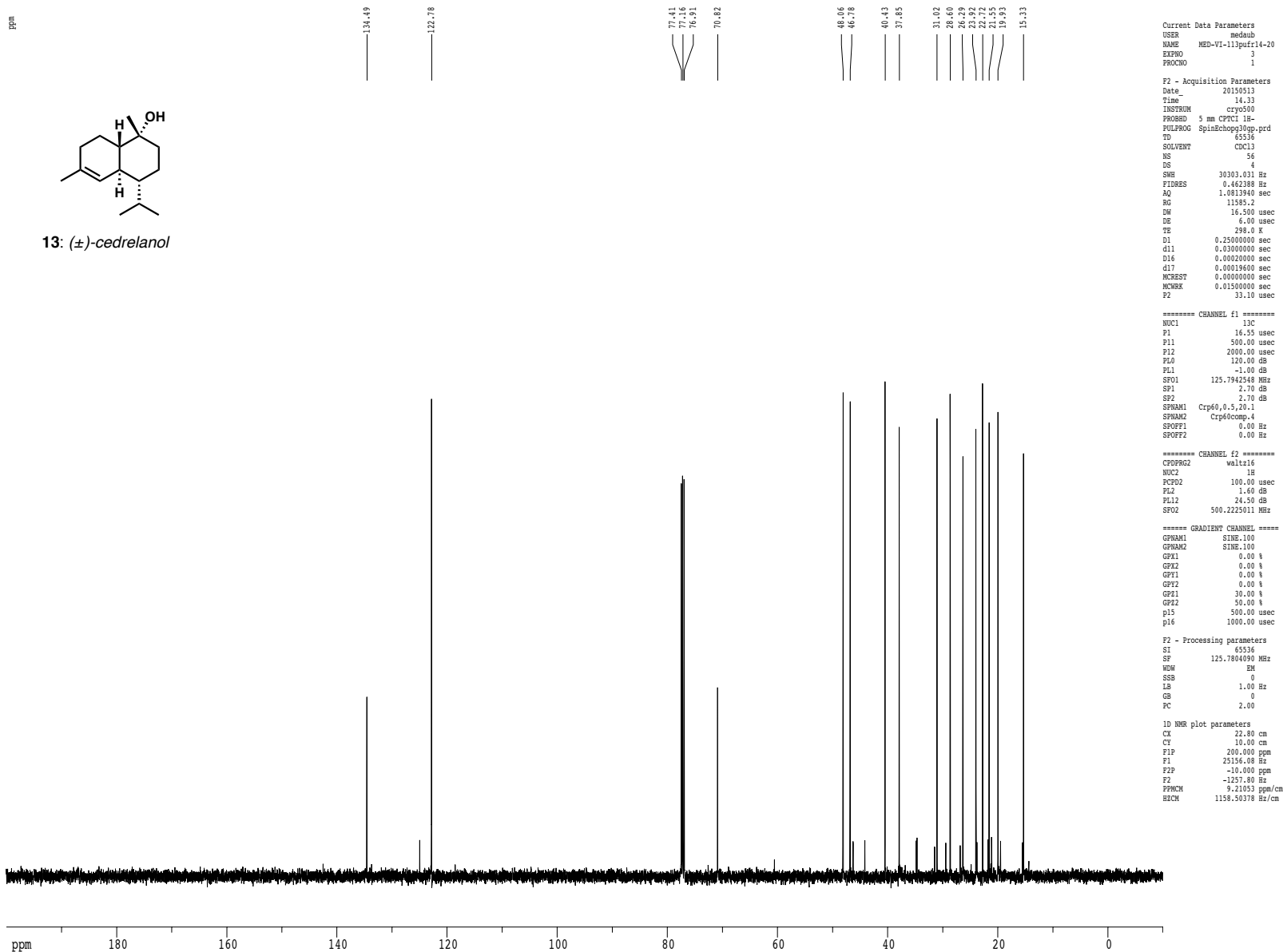
F2 - Acquisition Parameters
Date_    20130313
Time     13.55
INSTRUM  av600
PROBHD   5 mm BBO BB-1H
PULPROG  zg30
TD        98074
SOLVENT  CDCl3
NS        8
DS        2
SWH       9615.385 Hz
FIDRES    0.098042 Hz
AQ        5.098979 sec
RG        512
DW        52.000 usec
DE        14.33 usec
TE        298.1 K
D1        0.10000000 sec
TD0       1

***** CHANNEL f1 *****
SF01     600.1342009 MHz
NUC1     1H
P1       9.00 usec

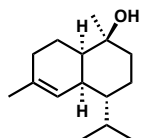
F2 - Processing parameters
SI       65536
SF       600.1300340 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00

1D NMR plot parameters
CX       22.80 cm
CY       15.00 cm
F1P      10.100 ppm
F1       6061.31 Hz
F2P      -0.100 ppm
F2       -60.01 Hz
FREQM    0.44137 ppm/cm
HZCM     268.47925 Hz/cm
    
```

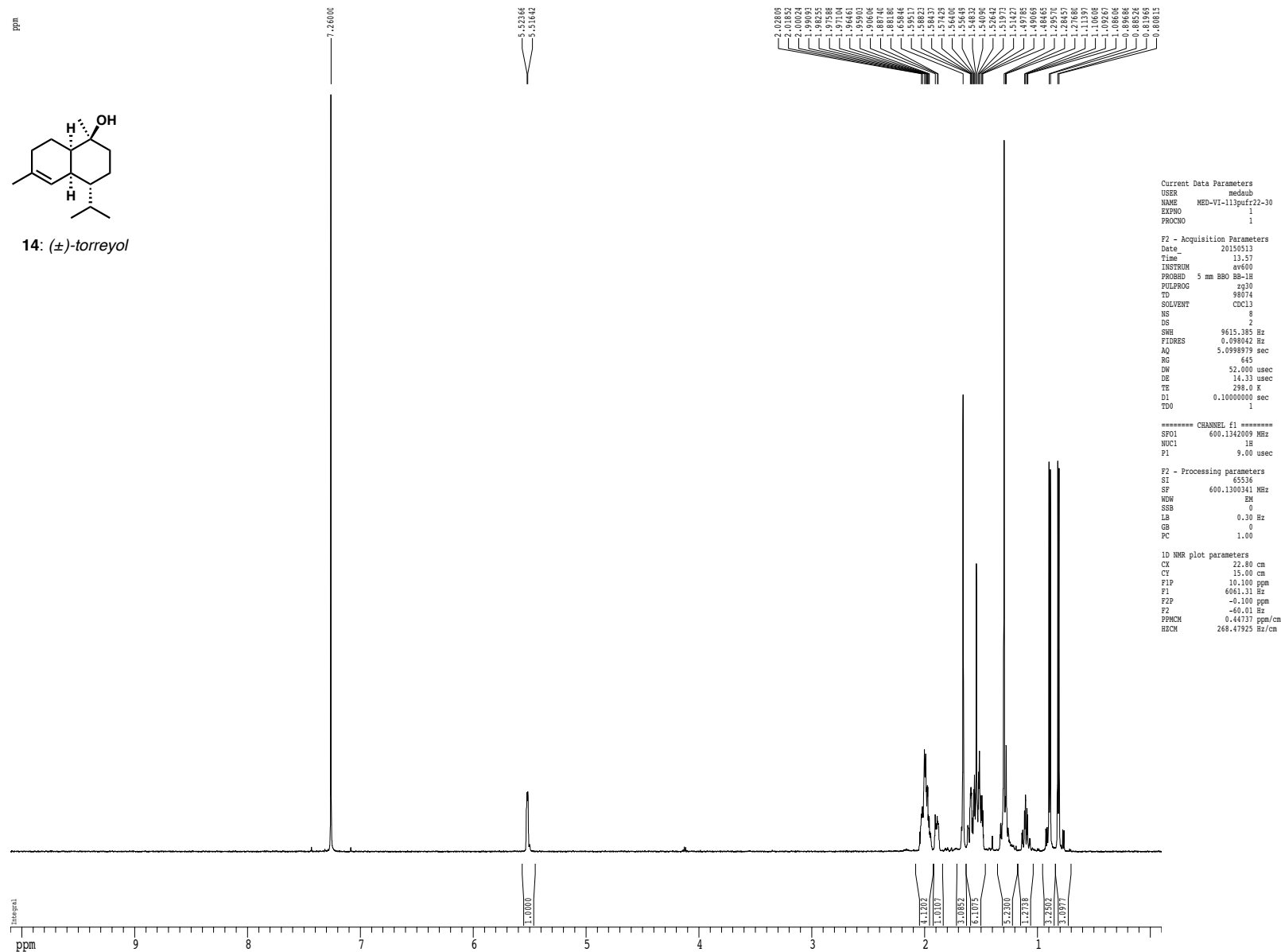
Z-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum

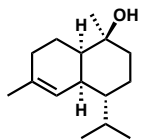


14: (±)-torreyol

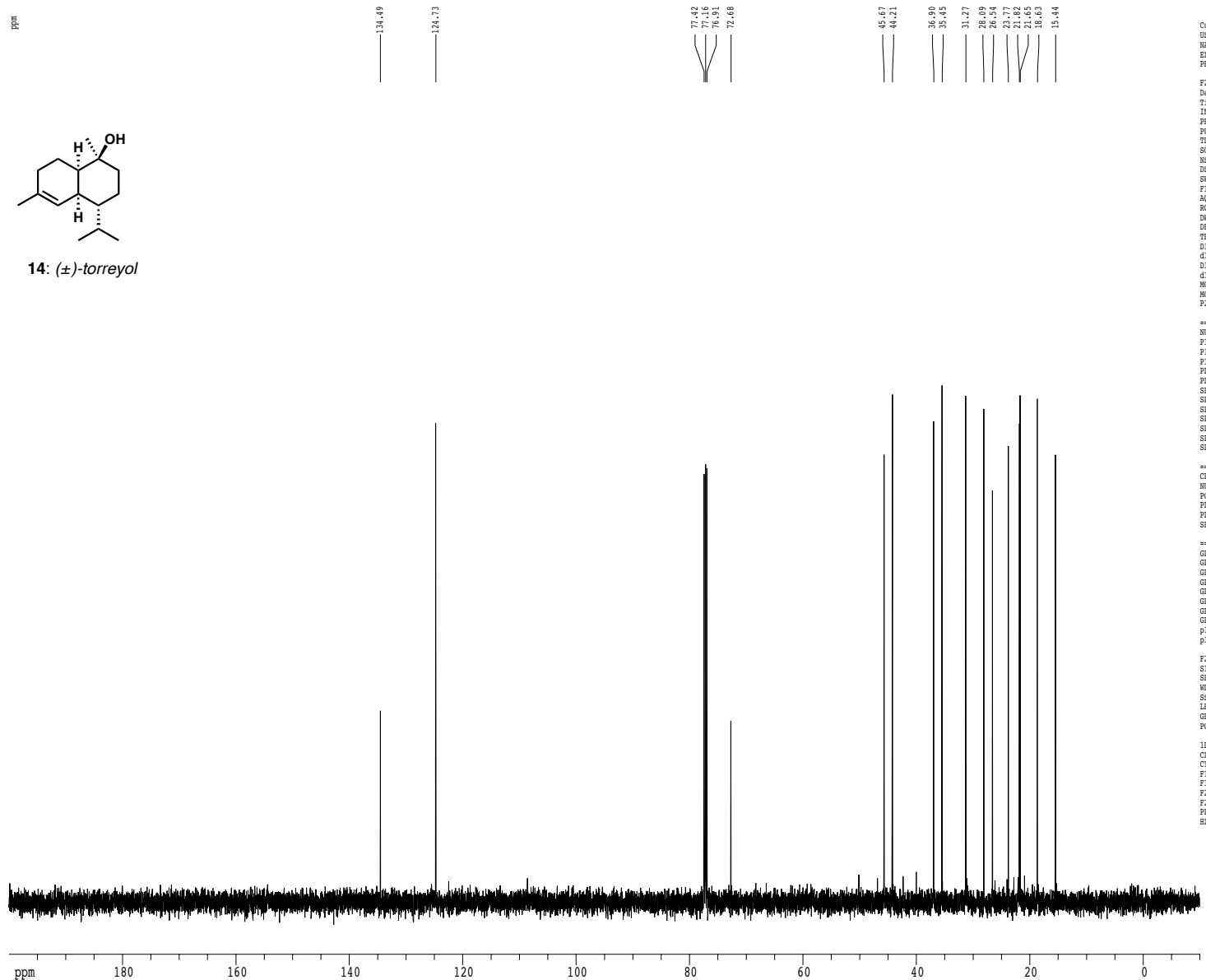


Z-restored spin-echo 13C spectrum with 1H decoupling

ppm



14: (\pm)-torreyol



```

Current Data Parameters
USER          medash
NAME          MED-VI-113pufz22-10
EXPNO         3
PROCNO        1

F2 - Acquisition Parameters
Date_         20150513
Time          14.37
INSTRUM       cryo500
PROBHD        5 mm CPTCI 1H-
PULPROG       SpinEchoq30gp.prd
TD            65536
SOLVENT       CDCl3
NS            32
DS            4
SWE           30303.031 Hz
FIDRES        0.462388 Hz
AQ            1.0813940 sec
RG            13004
DN            16.500 usec
DE            6.00 usec
TE            298.0 K
D1            0.2500000 sec
d11           0.0300000 sec
D16           0.0002000 sec
d17           0.0001900 sec
MCWREST       0.0000000 sec
MCWRK         0.0150000 sec
P2            33.10 usec

===== CHANNEL f1 =====
NUC1          13C
P1            16.50 usec
P11           500.00 usec
P12           2000.00 usec
PL0           120.00 dB
PL1           -1.00 dB
SFO1          125.7942548 MHz
SP1           2.70 dB
SP2           2.70 dB
SFOAM1        Crp60,0.5,20.1
SFOAM2        Crp60comp,4
SPOFF1        0.00 Hz
SPOFF2        0.00 Hz

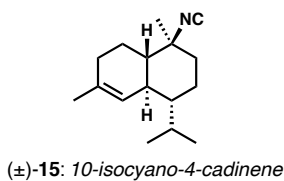
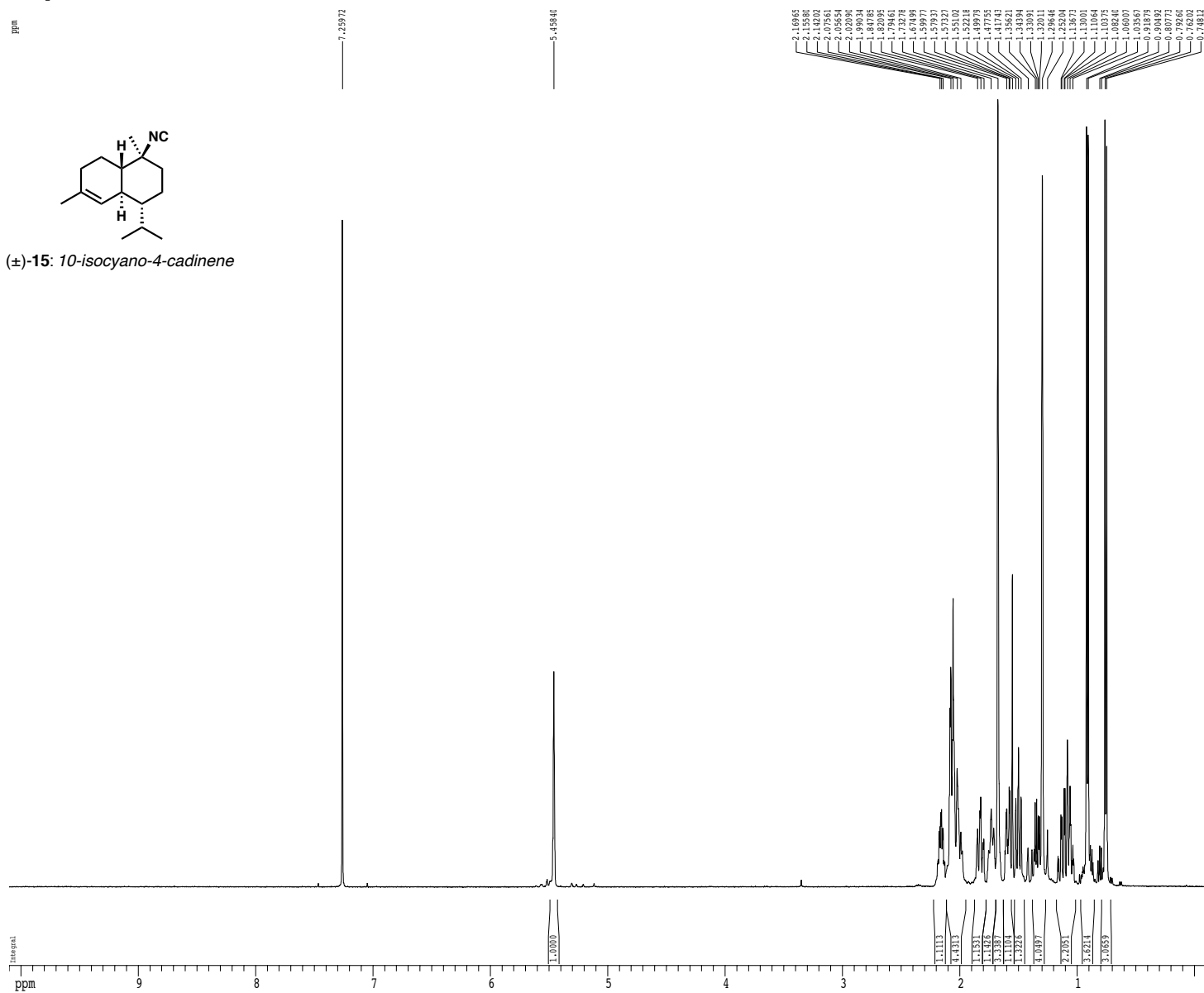
===== CHANNEL f2 =====
CPDPRG2       waltz16
NUC2          1H
PCPD2         100.00 usec
PL2           1.60 dB
PL12          24.50 dB
SFO2          500.2225011 MHz

===== GRADIENT CHANNEL =====
GPMAM1        SINE.100
GPMAM2        SINE.100
GPX1          0.00 %
GPX2          0.00 %
GPF1          0.00 %
GPF2          0.00 %
GPZ1          30.00 %
GPZ2          50.00 %
p15           500.00 usec
p16           1000.00 usec

F2 - Processing parameters
SI            65536
SF            125.7804085 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            2.00

1D NMR plot parameters
CX            22.80 cm
CY            10.00 cm
FIP           200.000 ppm
F1            25156.08 Hz
F2P          -10.000 ppm
F2            -1257.80 Hz
PPHMC         9.21053 ppm/cm
BRCM         1158.50378 Hz/cm
    
```

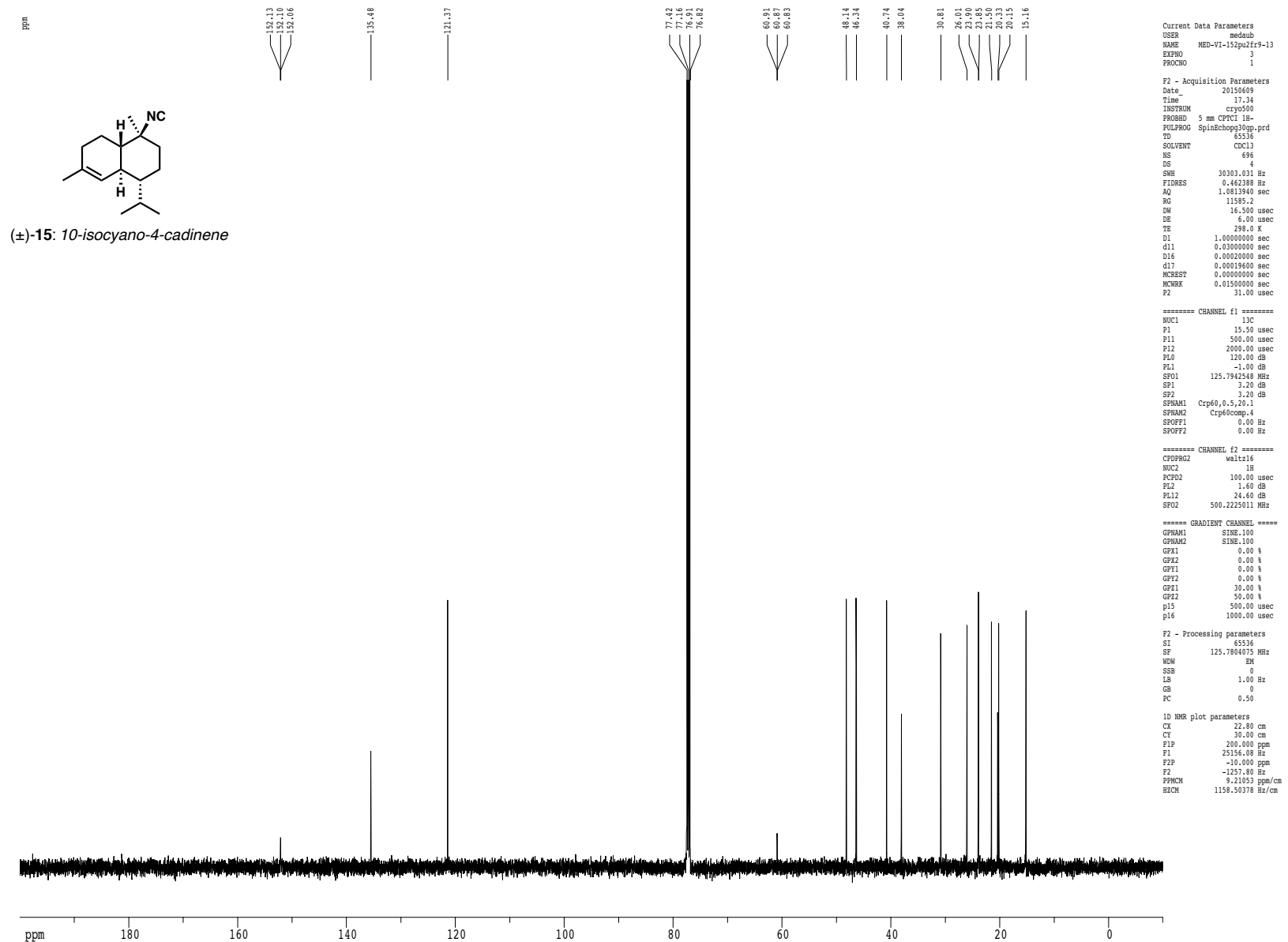
1H spectrum



2.16562
 2.15585
 2.14402
 2.07561
 2.05954
 1.99626
 1.99023
 1.84785
 1.82095
 1.79461
 1.77328
 1.65408
 1.59977
 1.57937
 1.57327
 1.55102
 1.49979
 1.47755
 1.41743
 1.38221
 1.37101
 1.33091
 1.32011
 1.28646
 1.25204
 1.25204
 1.19001
 1.18064
 1.10375
 1.06440
 1.03667
 0.91879
 0.90492
 0.80773
 0.76600
 0.75200
 0.74812

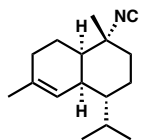
Current Data Parameters
 USER medaub
 NAME MED-VI-152puzif9-13
 EXPNO 2
 PROCNO 1
 F2 - Acquisition Parameters
 Date 20150609
 Time 17.31
 INSTRUM cryo300
 PROBHD 5 mm CPCTC 1B-
 PULPROG zg30
 TD 81728
 SOLVENT cdcl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 6.3
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCKEST 0.00000000 sec
 MCKRX 0.01500000 sec
 ===== CHANNEL f1 =====
 NU01 1H
 P1 7.50 usec
 PL1 1.60 dB
 SF01 500.2235015 MHz
 F2 - Processing parameters
 SI 65536
 SF 500.2200313 MHz
 NW 800
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00
 ID NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 FIP 10.100 ppm
 F1 5052.22 Hz
 F2P -0.100 ppm
 F2 -50.02 Hz
 PPMCM 0.44737 ppm/cm
 HCM 223.78267 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling

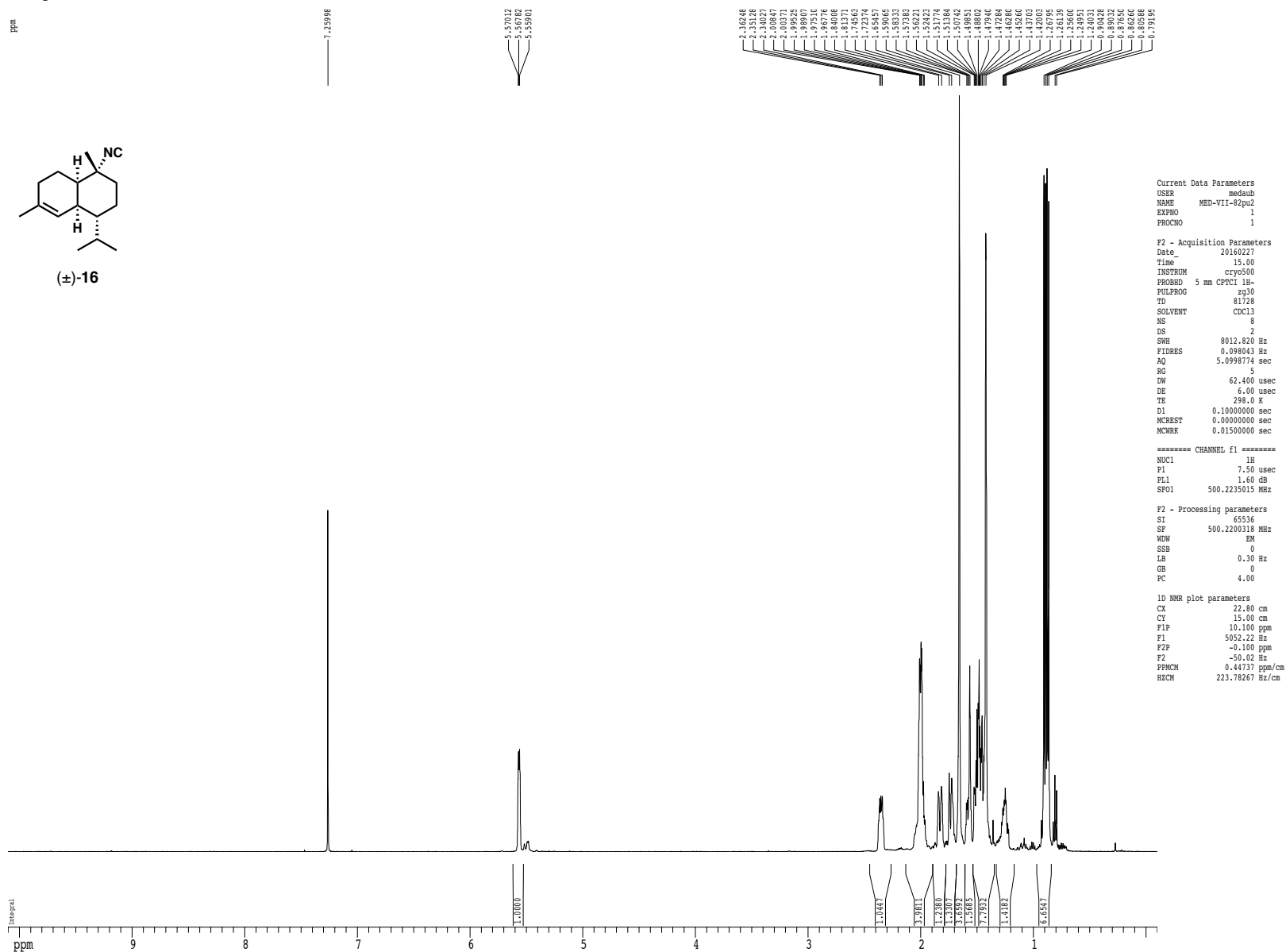


1H spectrum

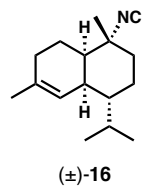
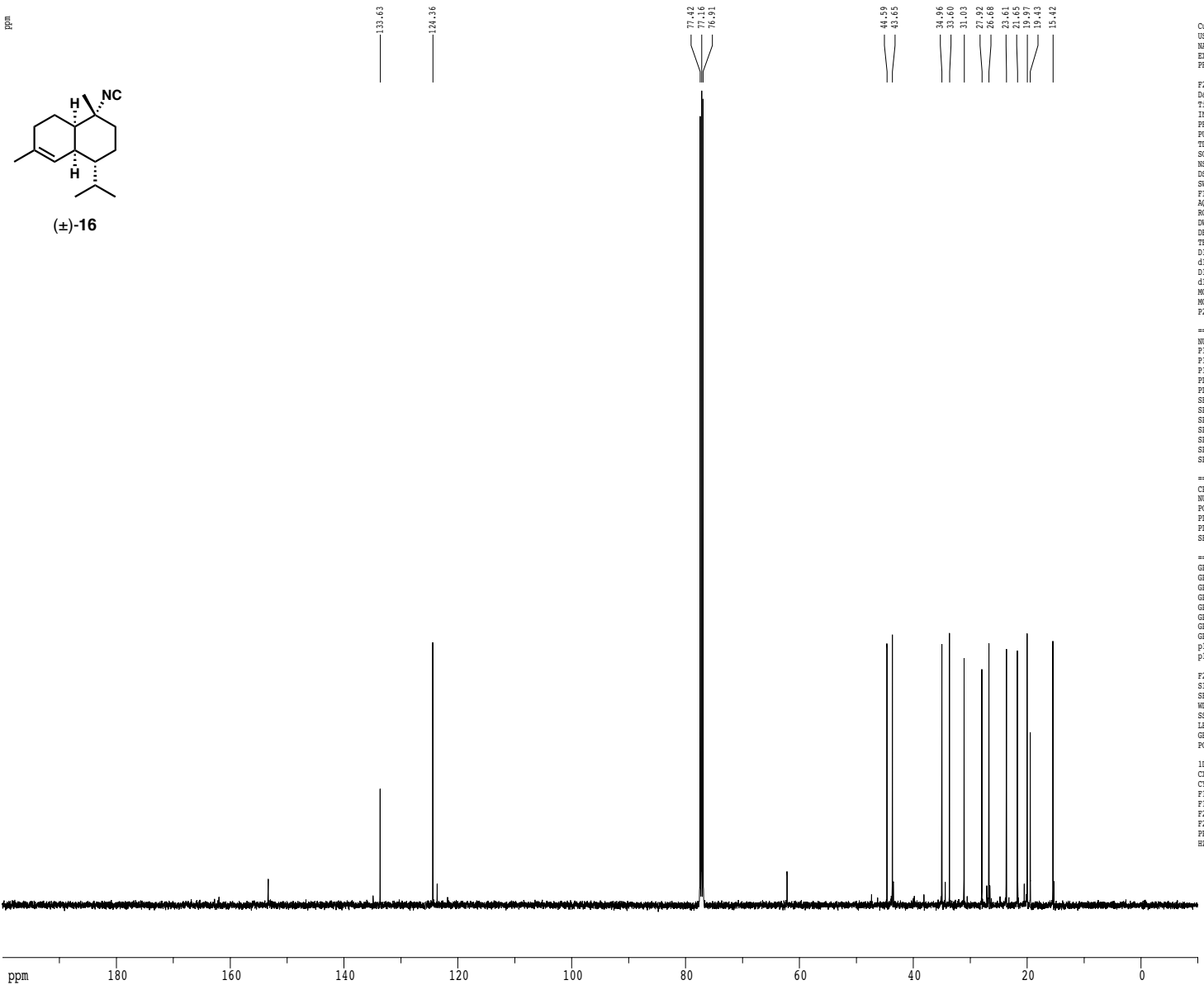
ppm



(±)-16



Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER      medaub
NAME      MED-VII-82pu2
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20160227
Time      15.03
INSTRUM   cryo500
PROBHD    5 mm CPCCI 1H-
PULPROG   SpinEcho30ap.prd
TD         65536
SOLVENT   CDCl3
NS         728
DS         4
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         13004
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         1.00000000 sec
d11        0.03000000 sec
D16        0.00020000 sec
d17        0.00019600 sec
MCWREST    0.00000000 sec
MCWFK      0.01500000 sec
P2         31.00 usec

===== CHANNEL f1 =====
NUC1       13C
P1         15.50 usec
P11        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1       125.7942548 MHz
SF1        3.20 dB
SP2        3.20 dB
SFOAM1     Crp60, 0.5, 20.1
SFOAM2     Crp60comp, 4
SFOFF1     0.00 Hz
SFOFF2     0.00 Hz

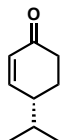
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2      100.00 usec
PL2        1.40 dB
PL12       24.40 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPMAM1     SINE.100
GPMAM2     SINE.100
GPX1       0.00 t
GPX2       0.00 t
GPY1       0.00 t
GPY2       0.00 t
GPZ1       30.00 t
GPZ2       50.00 t
p15        500.00 usec
p16        1000.00 usec

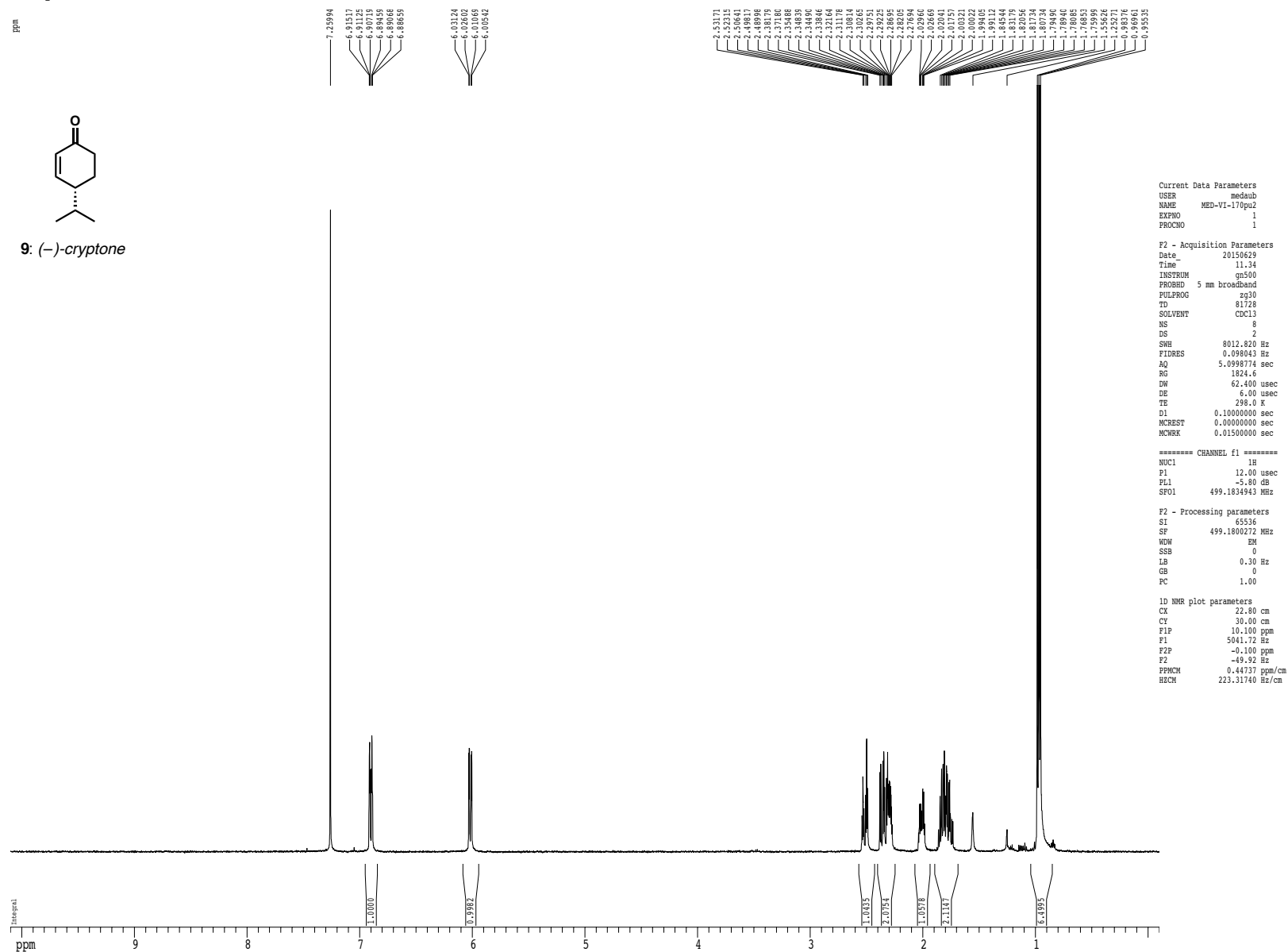
F2 - Processing parameters
SI         65536
SF         125.7804085 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

1D NMR plot parameters
CX         22.80 cm
CT         15.65 cm
PLP        200.000 ppm
F1         25156.08 Hz
F2P        -10.000 ppm
FZ         -1257.80 Hz
FPMCM      9.21053 ppm/cm
HZCM       1158.50378 Hz/cm
    
```

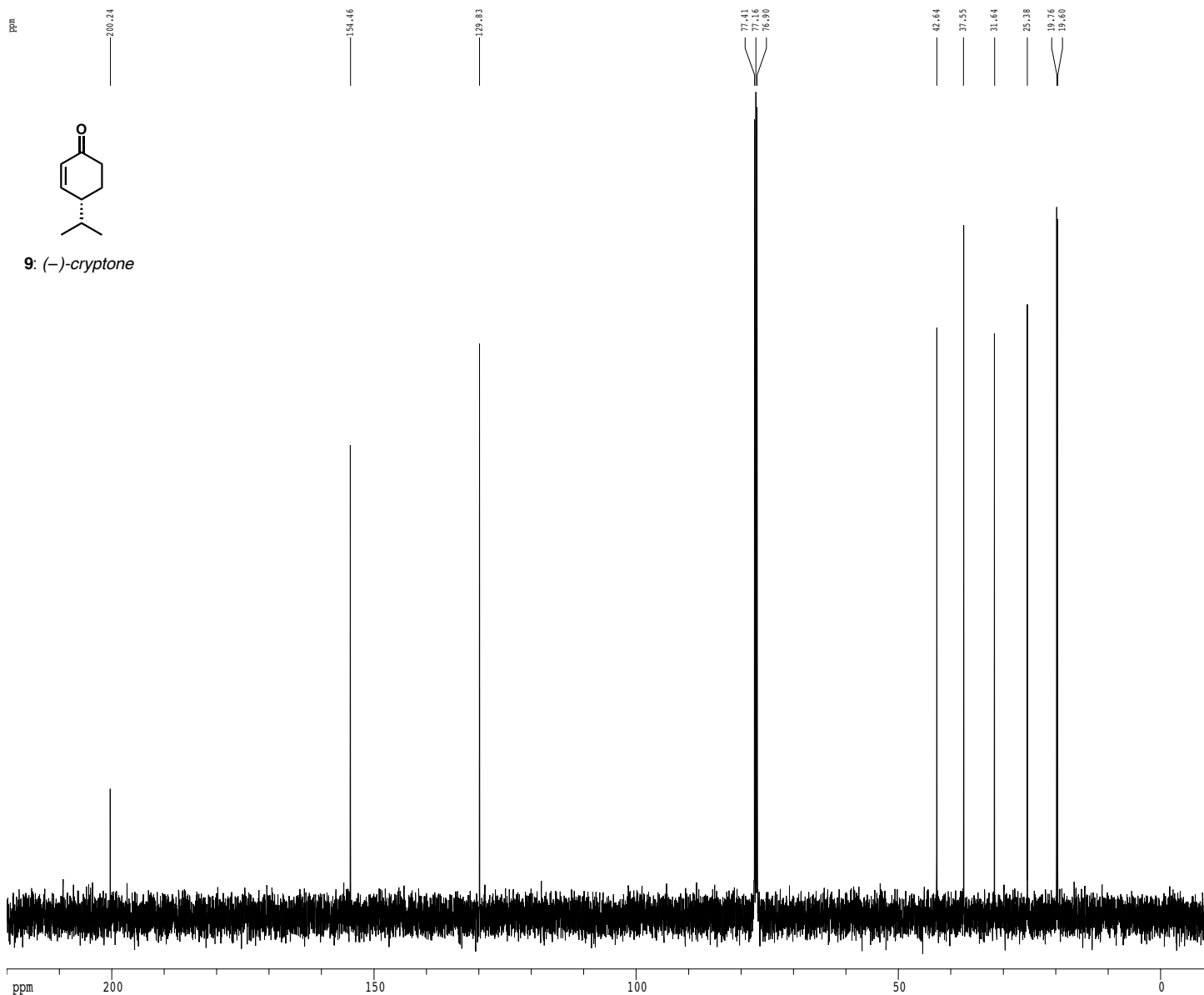
1H spectrum



9: (-)-cryptone



¹³C spectrum with ¹H decoupling



9: (-)-cryptone

```

Current Data Parameters
USER      medaub
NAME      MED-VI-150pu
EXPNO     3
PROCNO    1

F2 - Acquisition Parameters
Date_     20151219
Time      11.56
INSTRUM   gn500
PROBHD    5 mm broadband
PULPROG   zgpg30
TD        65536
SOLVENT   CDCl3
NS         88
DS         4
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         46341
DW         16.500 usec
DE         4.50 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
MCREST     0.00000000 sec
MCONRA     0.01500000 sec

===== CHANNEL f1 =====
NUC1       13C
P1         9.00 usec
PL1        -0.60 dB
SF01       125.5327181 MHz

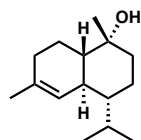
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2      80.00 usec
PL2        -3.00 dB
PL12       12.80 dB
SF02       499.1824959 MHz

F2 - Processing parameters
SI         65536
SF         125.5188998 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

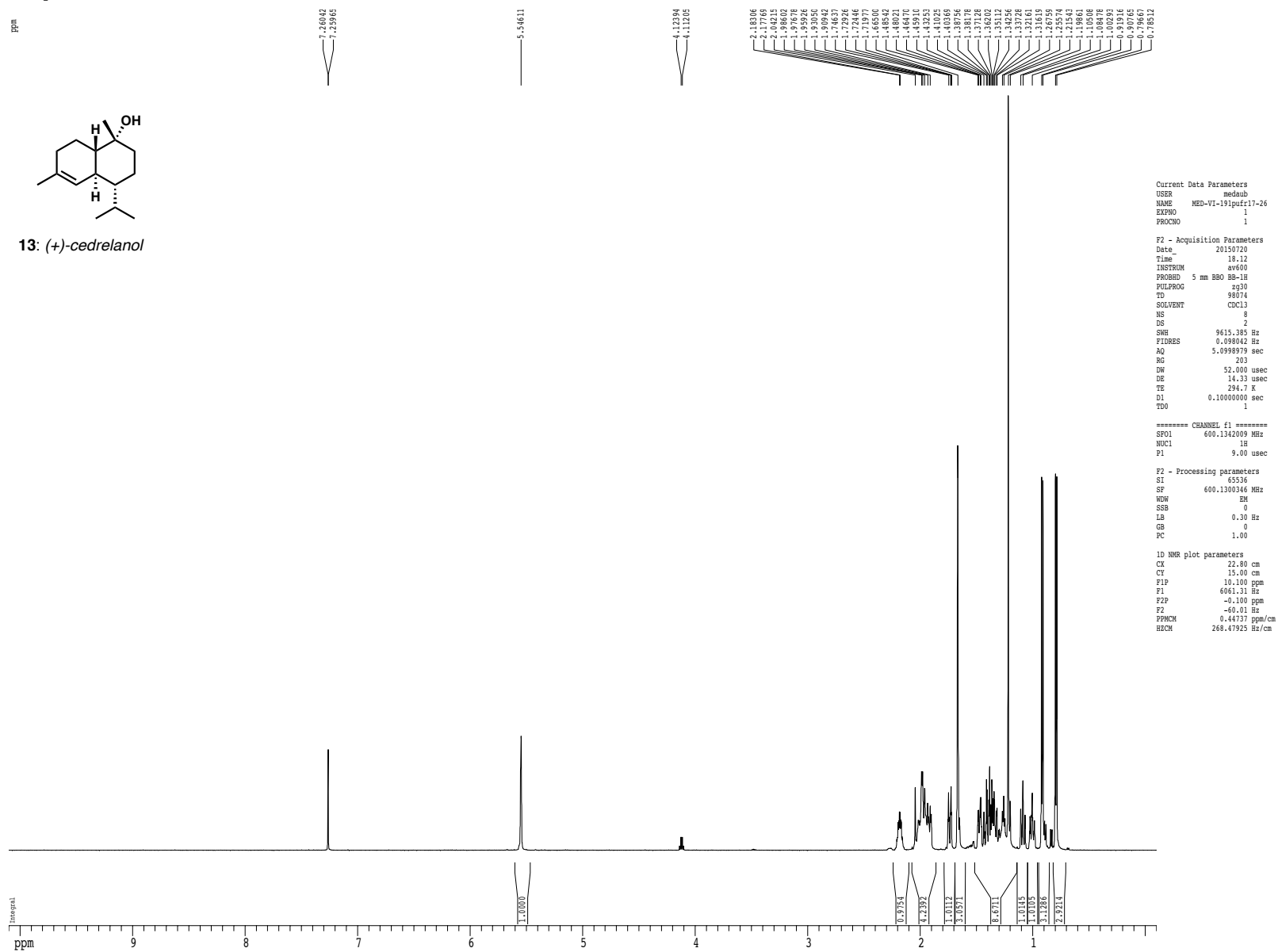
1D NMR plot parameters
CX         22.80 cm
CY         15.65 cm
F1P        220.000 ppm
F1         27614.16 Hz
F2P        -10.000 ppm
F2         -1255.19 Hz
PPMCM      10.08772 ppm/cm
HZCM       1266.19946 Hz/cm
    
```

1H spectrum

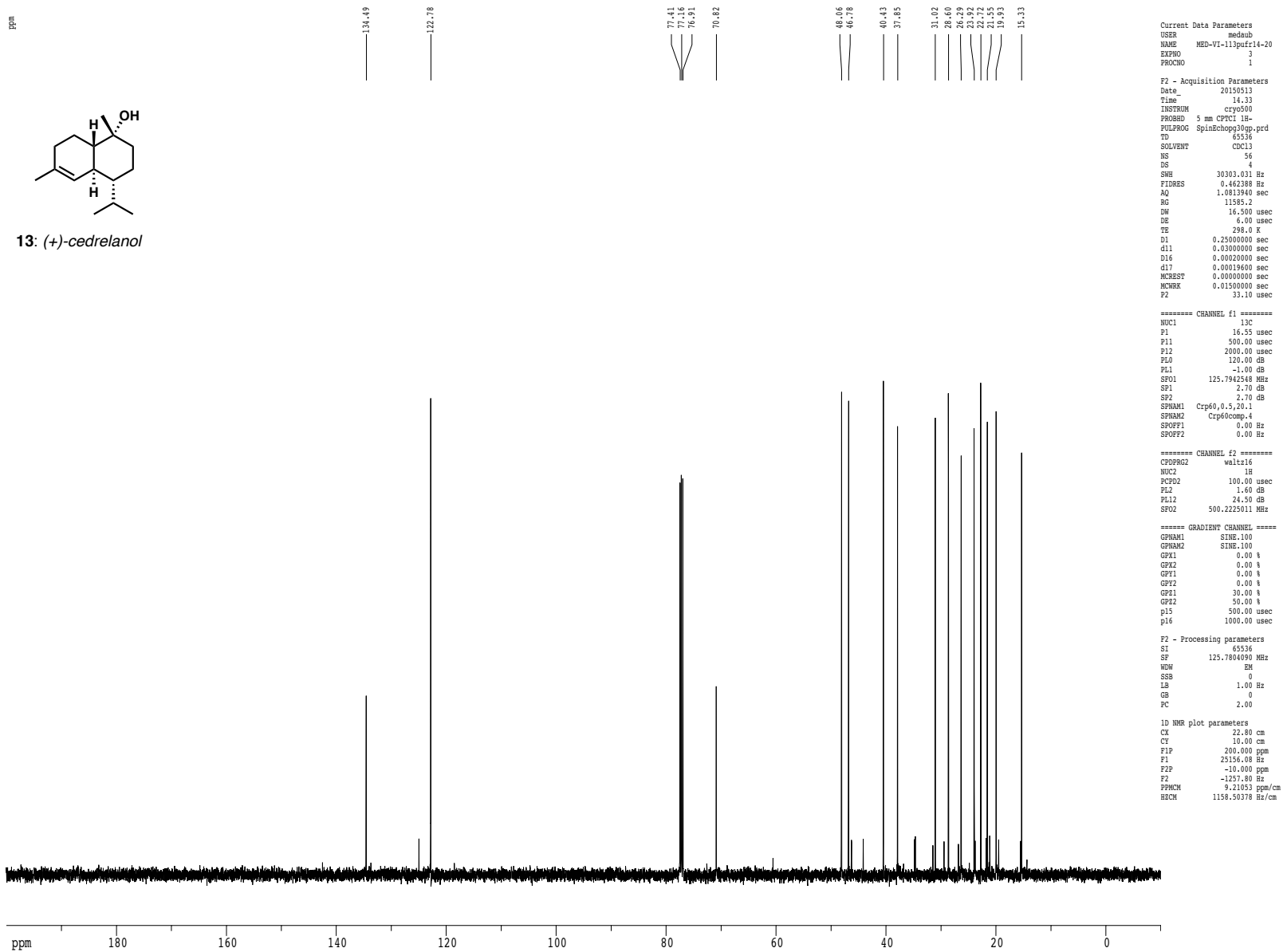
ppm



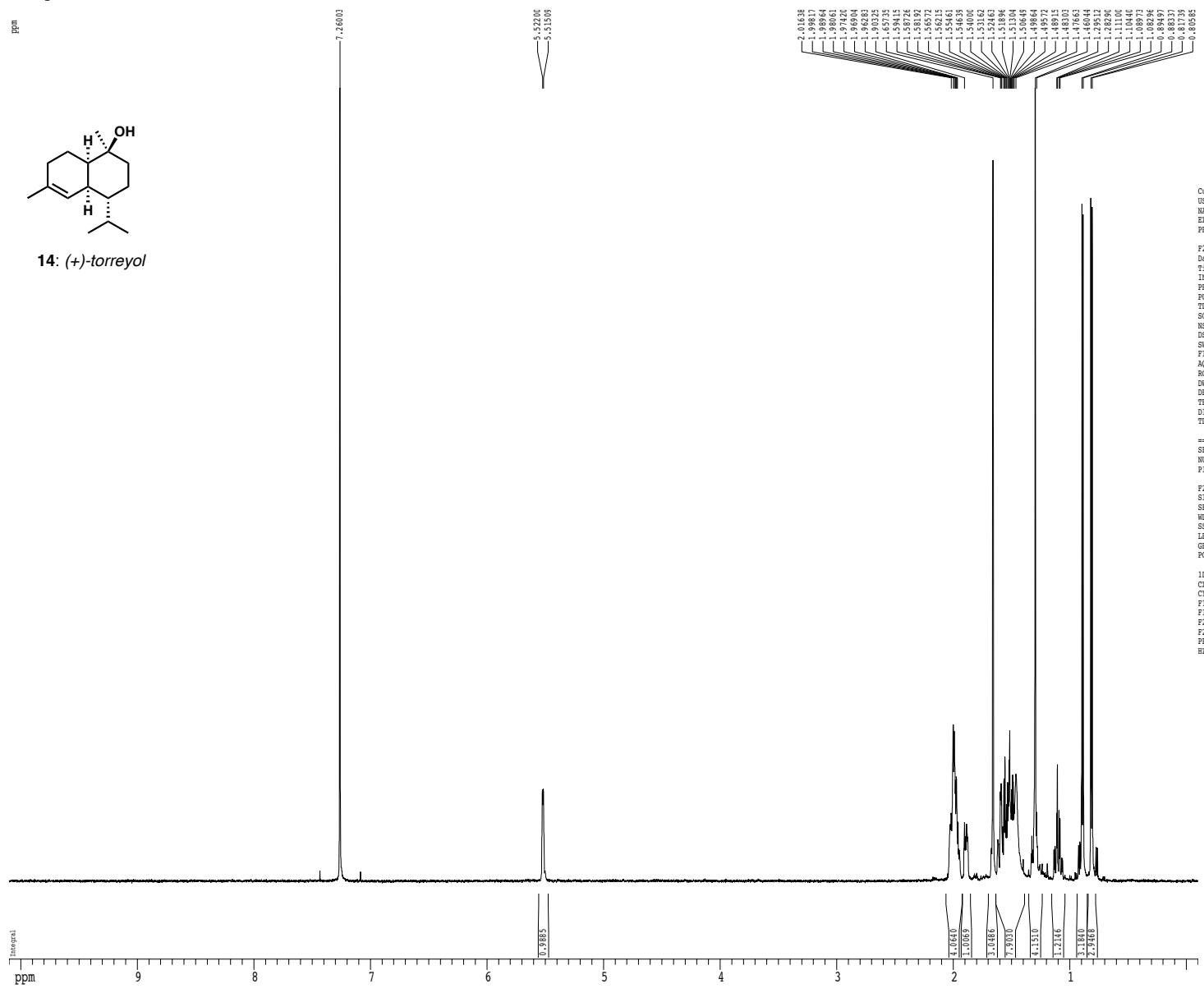
13: (+)-cedrelanol



Z-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum



```

Current Data Parameters
USER          medash
NAME          MED-VI-191pufz30-44
EXPNO        1
PROCNO       1

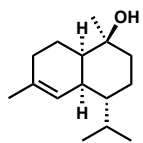
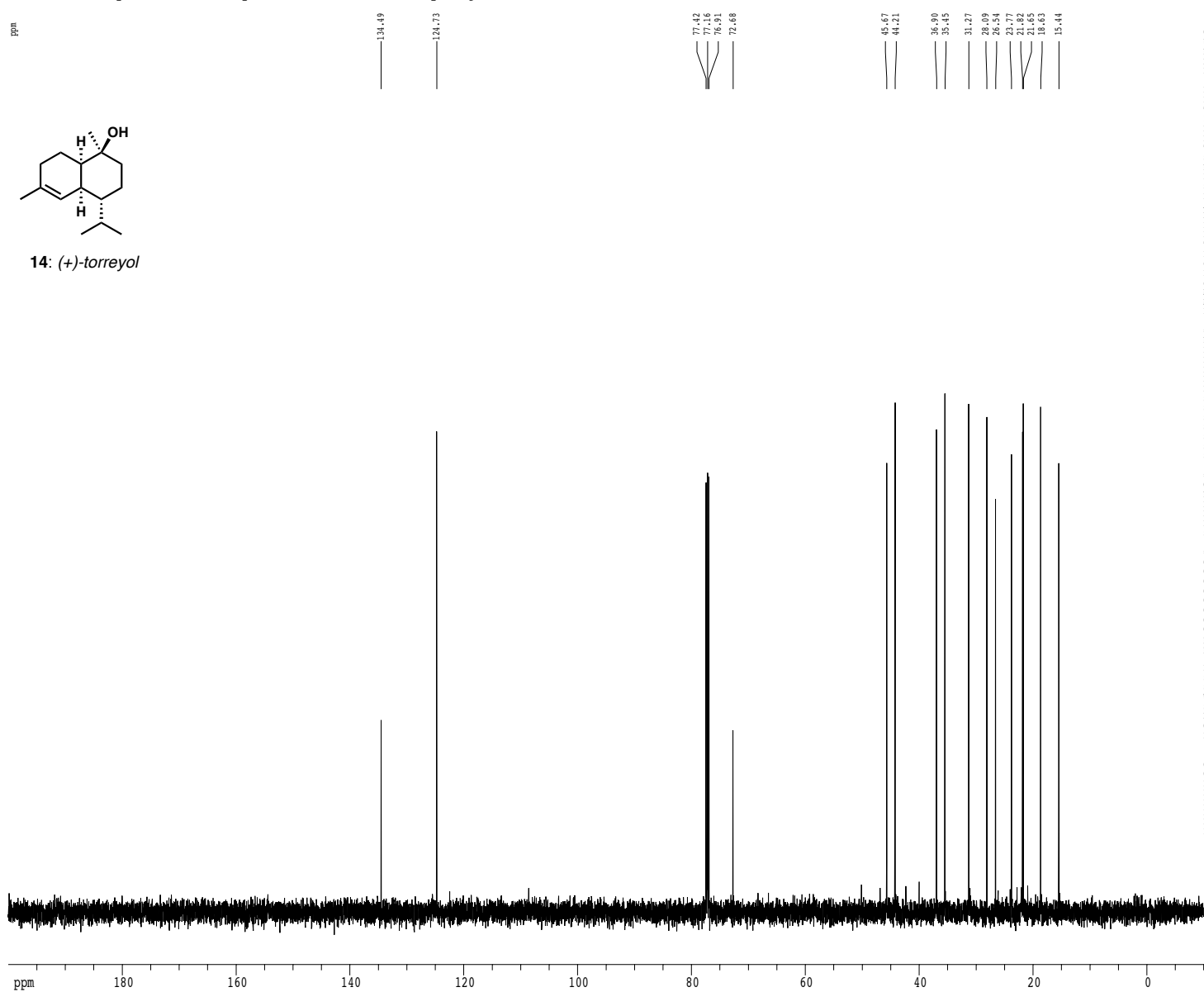
F2 - Acquisition Parameters
Date_        20150720
Time         18.15
INSTRUM      av600
PROBHD       5 mm BBO BB-1H
PULPROG      zg30
TD           98074
SOLVENT      CDCl3
NS           8
DS           2
SWH          9615.365 Hz
FIDRES       0.098042 Hz
AQ           5.0998979 sec
RG           724
DW           52.000 usec
DE           14.33 usec
TE           294.6 K
D1           0.1000000 sec
D0           1

===== CHANNEL f1 =====
SFO1         600.1342009 MHz
NUC1         1H
P1           9.00 usec

F2 - Processing parameters
SI           65536
SF           600.1300347 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB           0
PC           1.00

ID NMR plot parameters
CX           22.80 cm
CY           35.00 cm
F1P          10.100 ppm
F1           6061.31 Hz
F2P          -0.100 ppm
F2           -60.01 Hz
PPMCM        0.44737 ppm/cm
RECH         268.47925 Hz/cm
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



14: (+)-torreyol

```

Current Data Parameters
USER      medaub
NAME      MED-VI-113pufz22-30
EXPNO     3
PROCNO    1

F2 - Acquisition Parameters
Date_     20150513
Time      14.37
INSTRUM   cryo500
PROBHD    5 mm CPY11 H-
PULPROG   SpinEchoq30q.prd
TD         65536
SOLVENT   CDCl3
NS         32
DS         4
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0913940 sec
RG         13004
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
d16        0.00020000 sec
d17        0.00019600 sec
MCREST     0.00000000 sec
MCRM      0.01500000 sec
F2        33.10 usec

===== CHANNEL f1 =====
NUC1       13C
P1         16.55 usec
P11        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1       125.7942548 MHz
SF1        2.70 dB
SP2        2.70 dB
SFOF1      Crp60,0.5,20.1
SFOF2      Crp60comp,4
SFOFF1     0.00 Hz
SFOFF2     0.00 Hz

===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2      100.00 usec
PL2        1.60 dB
PL12       24.50 dB
SFO2       500.2225011 MHz

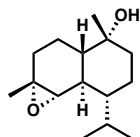
===== GRADIENT CHANNEL =====
GPMAM1     SINE.100
GPMAM2     SINE.100
GPF1       0.00 %
GPF2       0.00 %
GPF1       0.00 %
GPF2       0.00 %
GPF1       30.00 %
GPF2       50.00 %
p15        500.00 usec
p16        1000.00 usec

F2 - Processing parameters
SI         65536
SF         125.7804085 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

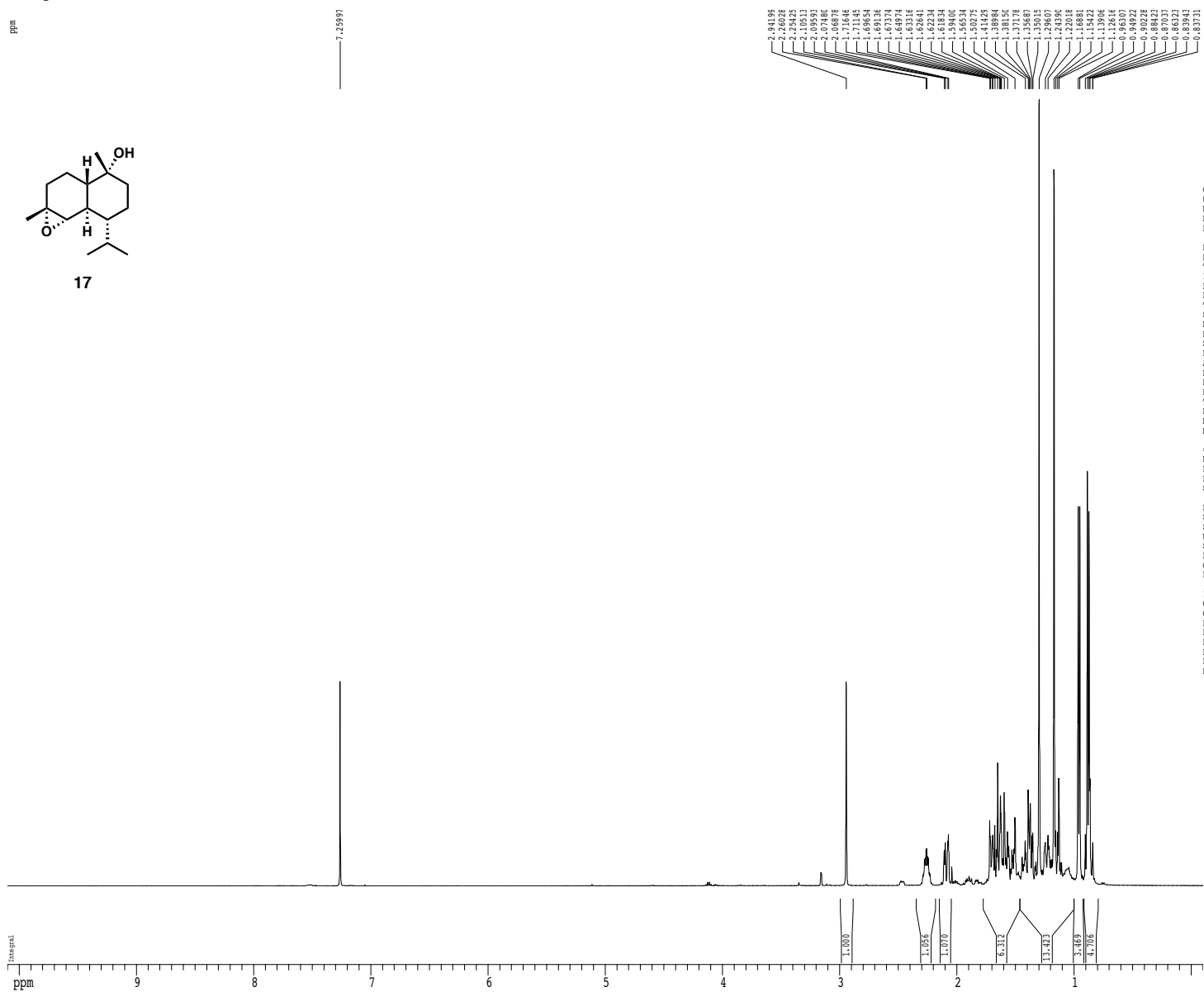
1D NMR plot parameters
CA         22.80 cm
CY         10.00 cm
F1P        200.000 ppm
F1         25156.08 Hz
F1P        -10.000 ppm
F2         -1257.80 Hz
PPHCH      9.21053 ppm/cm
H2CH       1158.50378 Hz/cm
    
```


1H spectrum

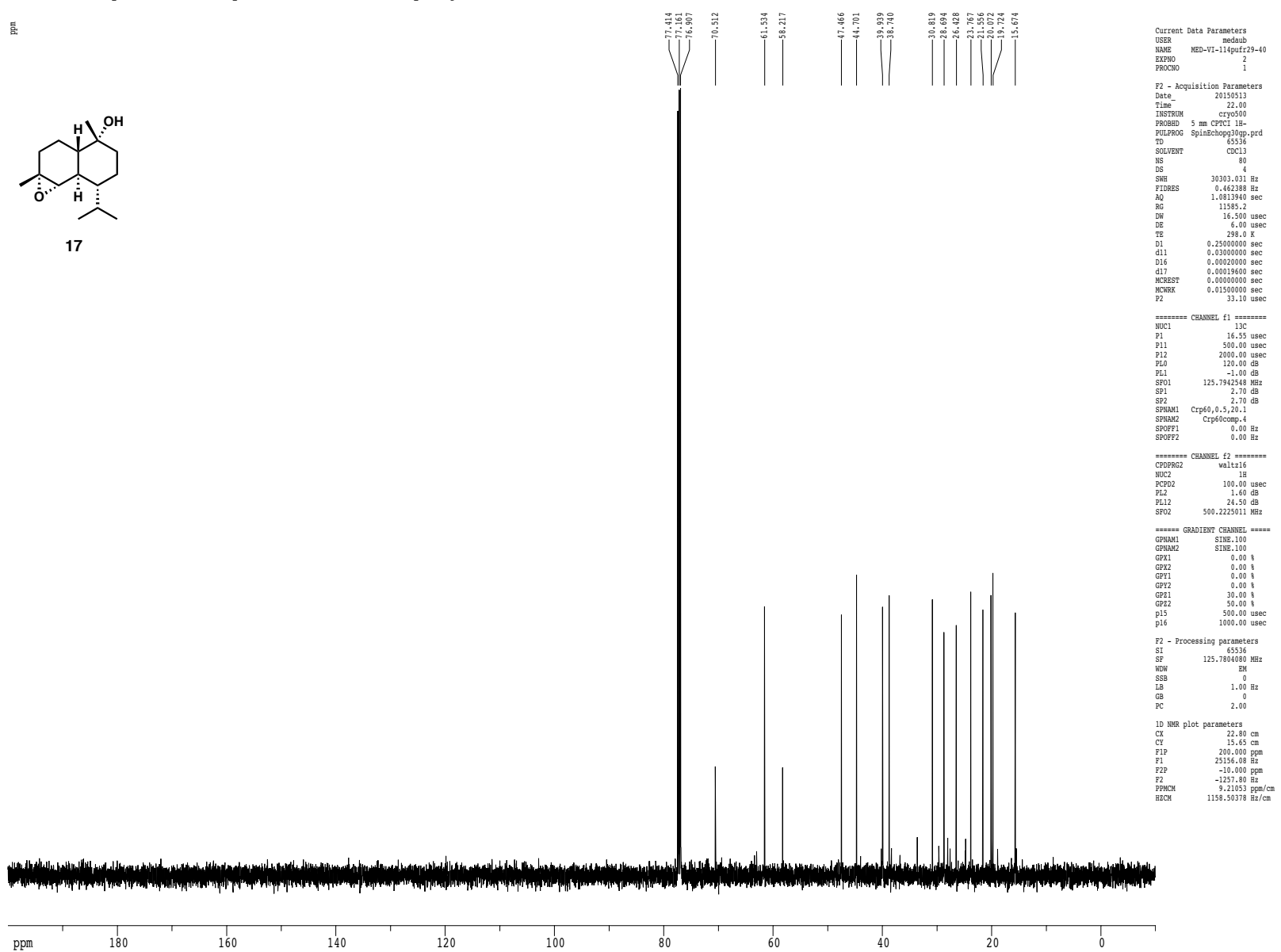
ppm



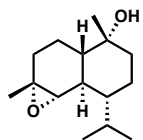
17



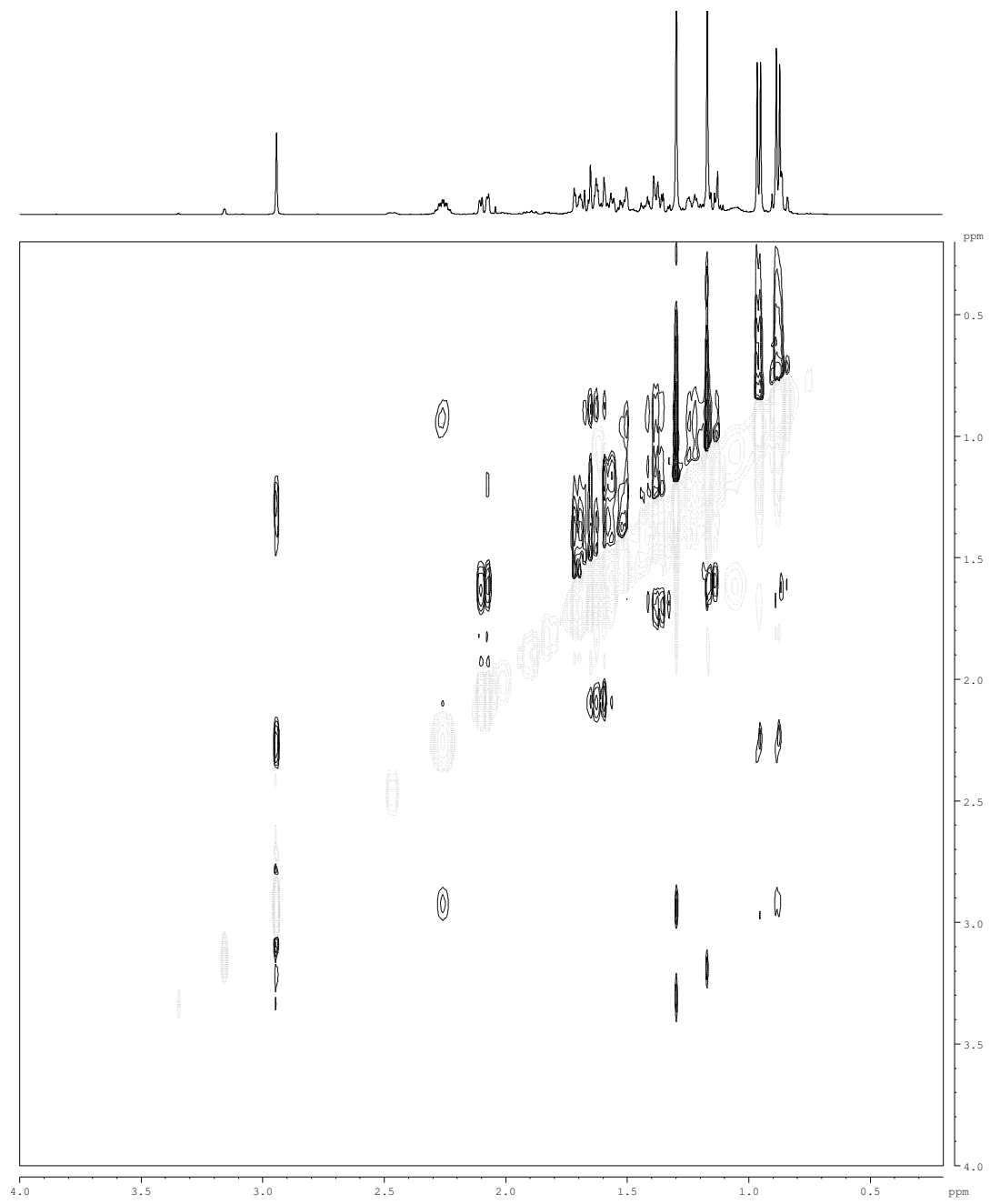
Z-restored spin-echo 13C spectrum with 1H decoupling



gnoesy



17



```
Current Data Parameters
NAME      MEO-VI-114purfz9-40
EXPNO    5
PROCNO   1

F2 - Acquisition Parameters
Date_    20150513
Time     22.08
INSTRUM  cryo400
PROBHD   5 mm CPTCI 1H-
PULPROG  noesygpp
TD        65536
SOLVENT  CDCl3
NS        2
DS        4
SWH       4006.410 Hz
FIDRES    1.956255 Hz
AQ        0.2555904 sec
RG        128
DW        124.800 usec
DE        6.00 usec
TE        298.0 K
D0        0.00000300 sec
D1        2.00000000 sec
D8        0.80000001 sec
D16       0.00020000 sec
d20       0.39880002 sec
IN0       0.00012480 sec

----- CHANNEL f1 -----
NUC1     1H
P1       7.50 usec
P2       15.00 usec
PL1      1.60 dB
SFO1    500.2219426 MHz

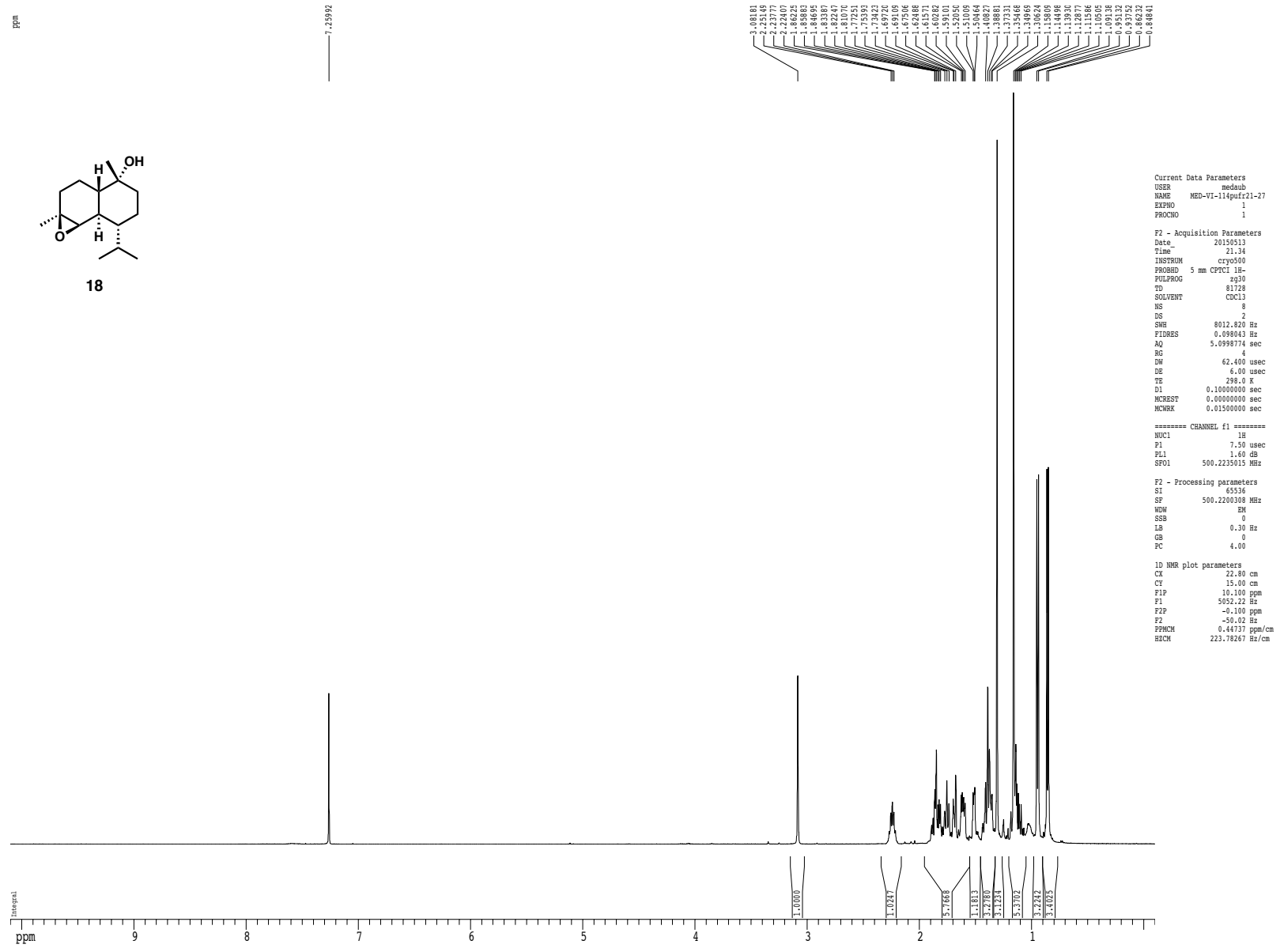
----- GRADIENT CHANNEL -----
GPNAM[1] sine.100
GPNAM[2] sine.100
GFX1     0 %
GFX2     0 %
GTY1     0 %
GTY2     0 %
GPZ1     40.00 %
GPZ2    -40.00 %
F16      1000.00 usec

F1 - Acquisition parameters
TD        63
SFO1     500.2219 MHz
FIDRES    63.593815 Hz
SW        8.009 ppm
FAMODE    undefined

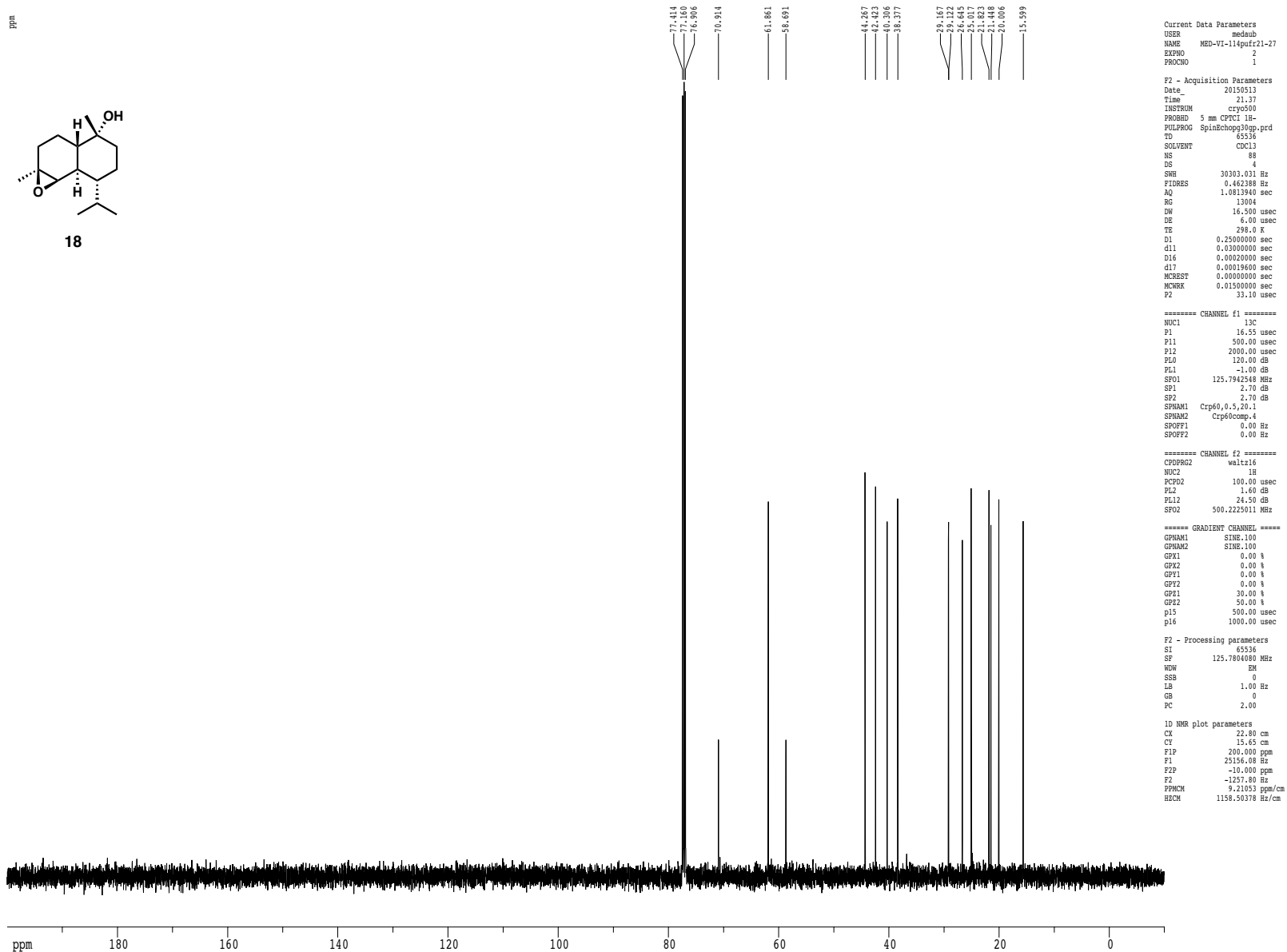
F2 - Processing parameters
SI        1024
SF        500.2200310 MHz
WDW       QSINE
SSB       2
LB        0 Hz
GB        0
PC        1.40

F1 - Processing parameters
SI        1024
MC2       TFE1
SF        500.2200310 MHz
WDW       QSINE
SSB       2
LB        0 Hz
GB        0
```

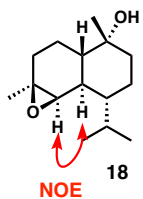
1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling



gnoesy



```
Current Data Parameters
NAME MED-VI-114puzr21-27
EXPNO 5
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150513
Time 21.45
INSTRUM cryo400
PROBHD 5 mm CPTCI 1H-
PULPROG zgpg30
TD 262144
SOLVENT cdcl3
NS 2
DS 4
SWH 4084.967 Hz
FIDRES 1.994613 Hz
AQ 0.2506752 sec
RG 128
DW 122.400 usec
DE 6.00 usec
TE 298.0 K
D0 0.0000300 sec
D1 2.0000000 sec
D8 0.8000001 sec
D16 0.0002000 sec
d20 0.3988002 sec
IN0 0.00012240 sec

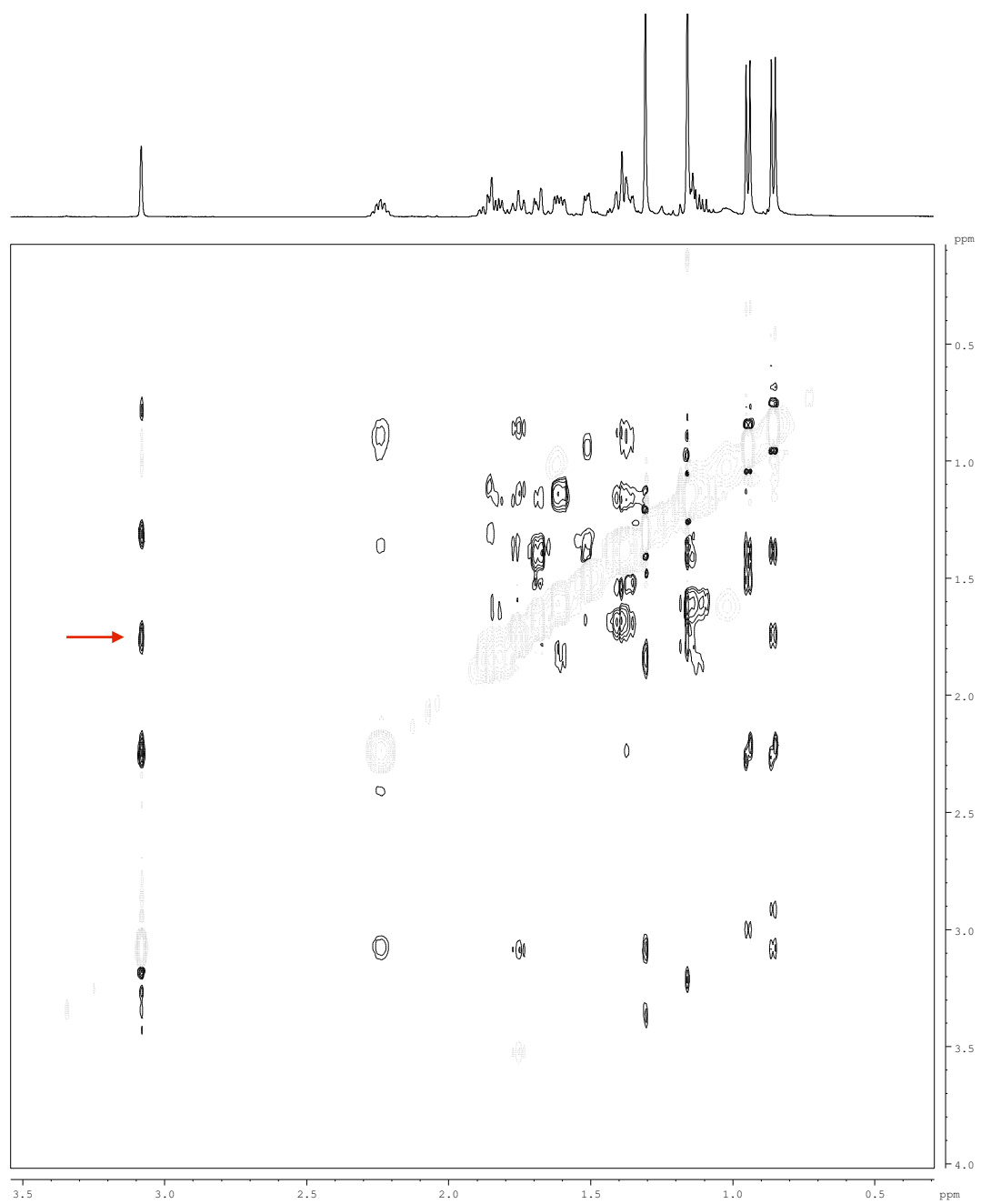
----- CHANNEL f1 -----
NUC1 1H
P1 7.50 usec
P2 15.00 usec
PL1 1.60 dB
SF01 500.2218797 MHz

----- GRADIENT CHANNEL -----
GPNAM[1] sine.100
GPNAM[2] sine.100
GPX1 0 %
GPX2 0 %
GPY1 0 %
GPY2 0 %
GPZ1 40.00 %
GPZ2 -40.00 %
P16 1000.00 usec

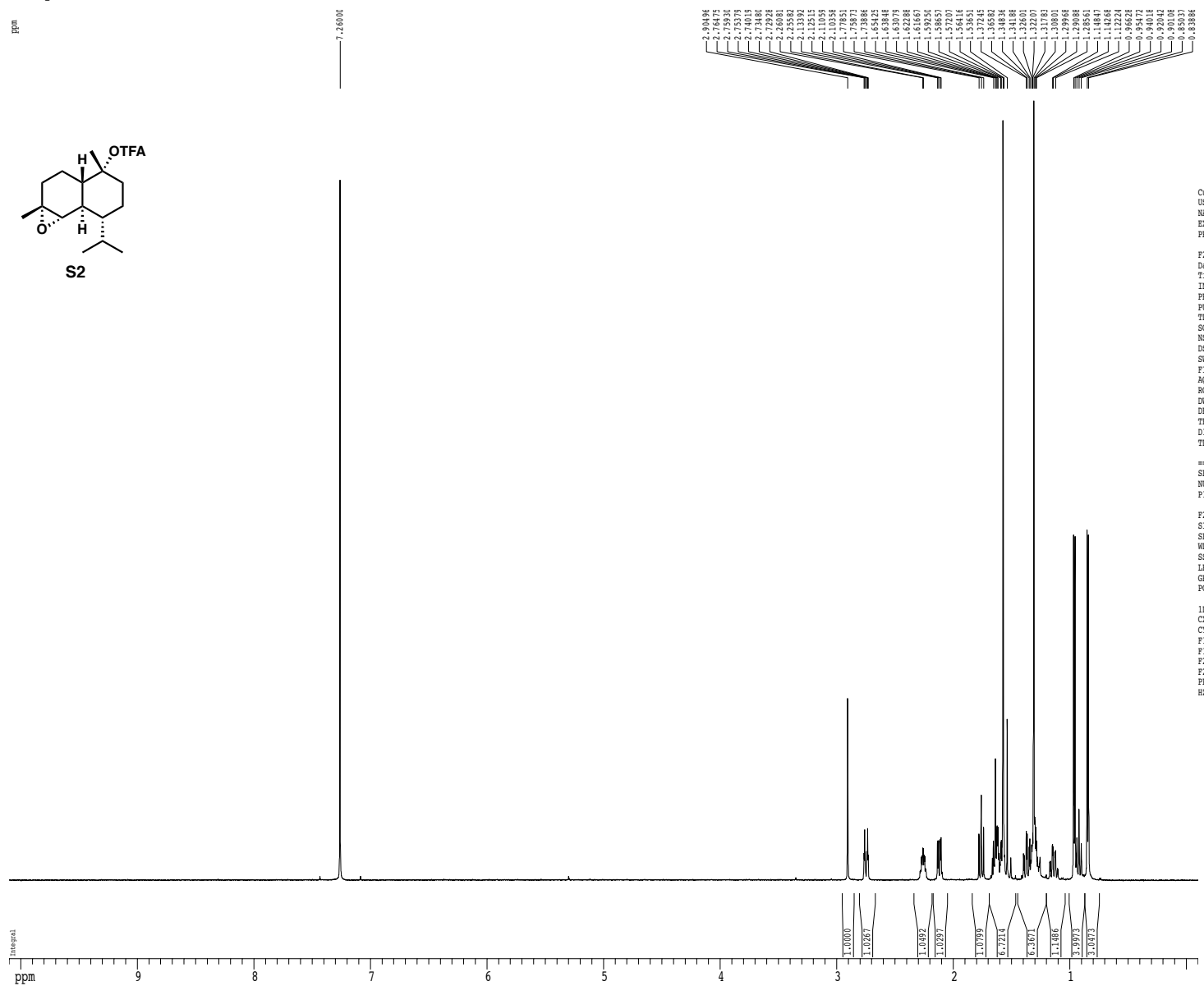
F1 - Acquisition parameters
TD 99
SF01 500.2219 MHz
FIDRES 41.262295 Hz
SW 8.166 ppm
FMODE undefined

F2 - Processing parameters
SI 1024
SF 500.2200309 MHz
WDW QSINE
SSB 2
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 TPF1
SF 500.2200309 MHz
WDW QSINE
SSB 2
LB 0 Hz
GB 0
```



1H spectrum



```

Current Data Parameters
USER          medaub
NAME          MED-VI-116pu
EXPNO        1
PROCNO       1

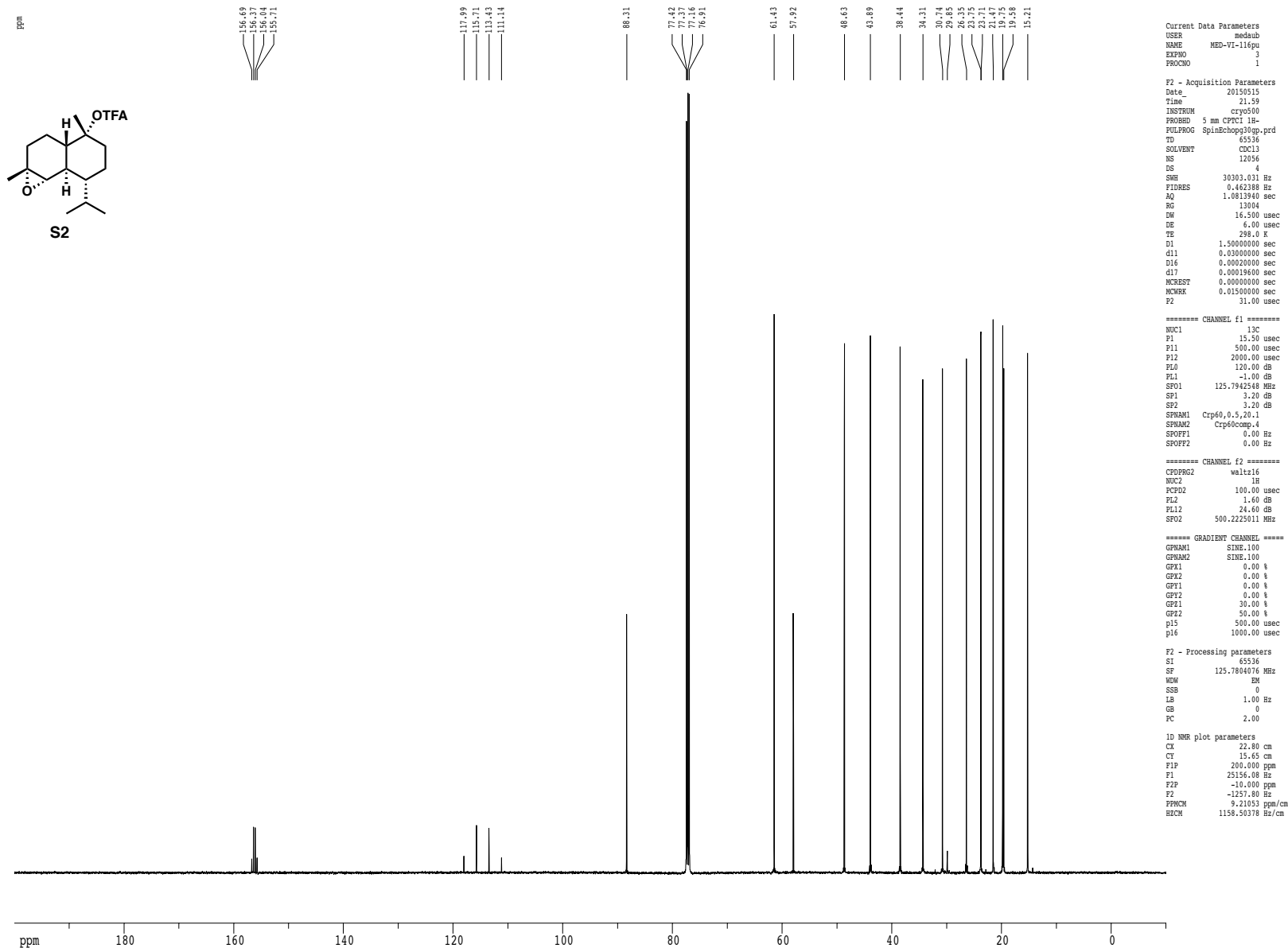
F2 - Acquisition Parameters
Date_        20150515
Time         18.02
INSTRUM      av600
PROBHD       5 mm BBO BB-1H
PULPROG      zg30
TD           98074
SOLVENT      CDCl3
NS           8
DS           2
SWH          9615.385 Hz
FIDRES       0.098042 Hz
AQ           5.0998979 sec
RG           724
DM           52.000 usec
DE           14.33 usec
TE           298.4 K
D1           0.10000000 sec
TD0          1

===== CHANNEL f1 =====
SF01         600.1342009 MHz
NUC1         1H
P1           9.00 usec

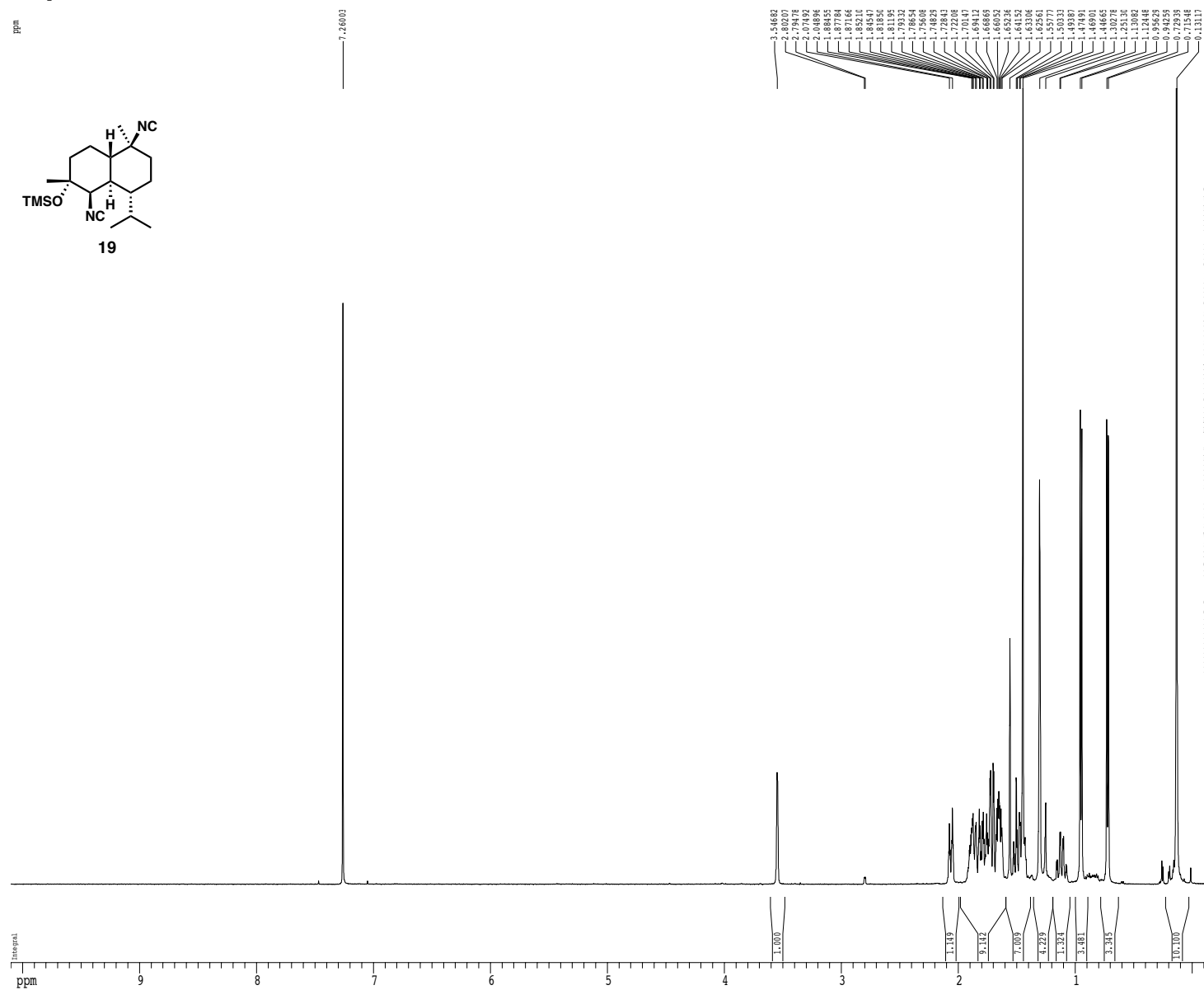
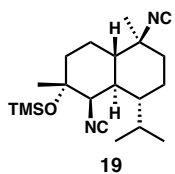
F2 - Processing parameters
SI           65536
SF           600.1300341 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB           0
PC           1.00

1D NMR plot parameters
CX           22.80 cm
CY           15.00 cm
F1P          10.100 ppm
F1           6061.31 Hz
F2P          -0.100 ppm
F2           -60.01 Hz
PPMCM       0.44737 ppm/cm
HZCM        268.47925 Hz/cm
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum



Current Data Parameters
USER medaub
NAME MED-VI-285pu2f8-14
EXPNO 2
PROCNO 1

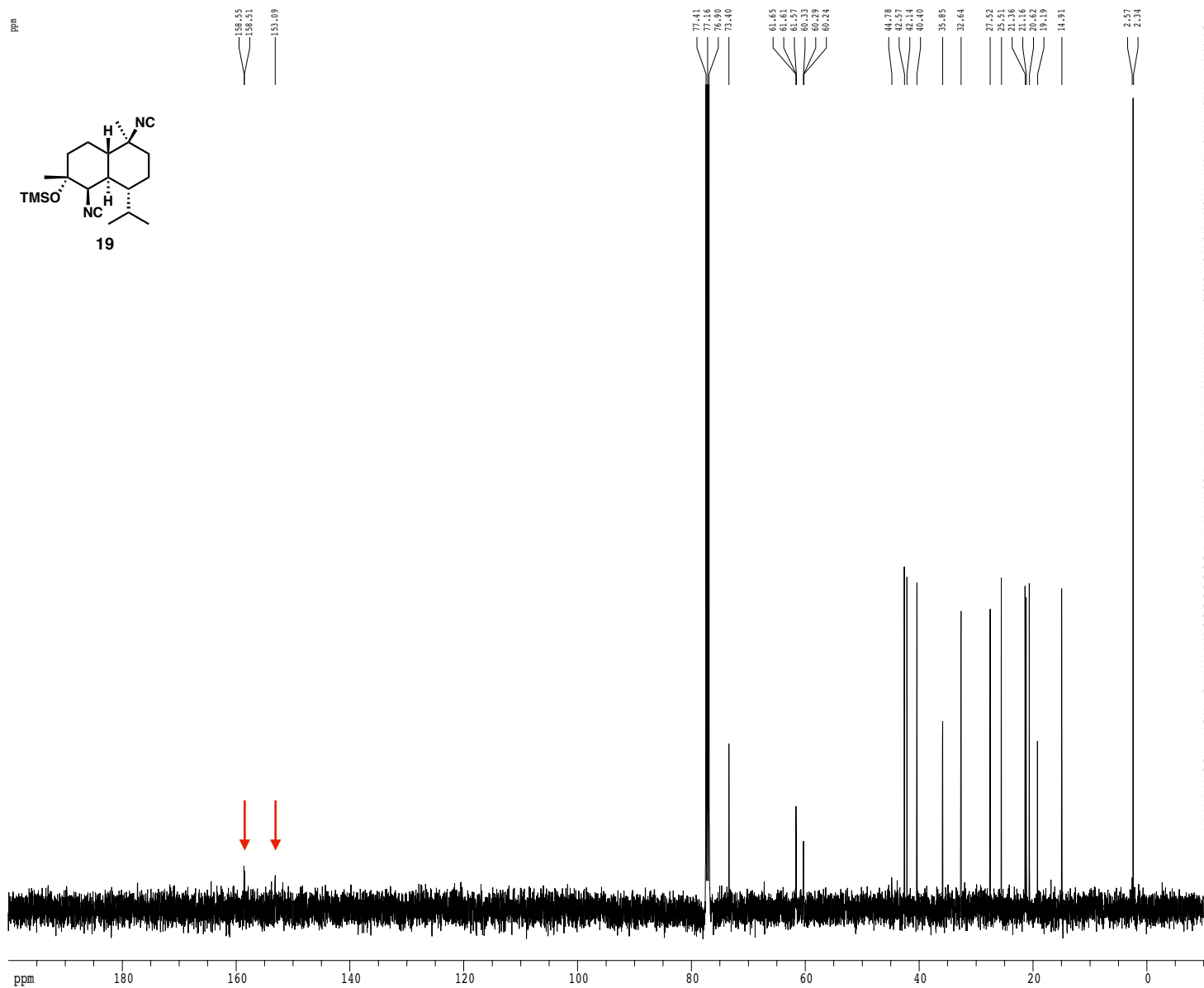
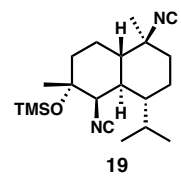
F2 - Acquisition Parameters
Date_ 20151223
Time 9.09
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG zg30
TD 81728
SOLVENT CDCl3
NS 8
DS 2
SWH 8012.820 Hz
FIDRES 0.098043 Hz
AQ 5.0988774 sec
RG 6.3
DW 62.400 usec
DE 6.00 usec
TE 298.0 K
D1 0.1000000 sec
MCREST 0.0000000 sec
MCWRX 0.0150000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 7.50 usec
PL1 1.60 dB
SFO1 500.225015 MHz

F2 - Processing parameters
SI 6536
SF 500.2200305 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 22.80 cm
CY 55.00 cm
FIP 10.100 ppm
F1 5052.22 Hz
FZP -0.100 ppm
FZ -50.02 Hz
FZCH 0.4437 ppm/cm
HZCH 223.78267 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER          medsub
NAME         MHD-VI-28pu2f8-14
EXPNO        3
PROCNO       1

F2 - Acquisition Parameters
Date_        20151223
Time         9.13
INSTRUM      cryo500
PROBHD       5 mm CPXI 1H-
PULPROG      SpinEcho30pp-prd
TD           65536
SOLVENT      CDCl3
NS           264
DS           4
SWH          30303.031 Hz
FIDRES       0.462388 Hz
AQ           1.0813940 sec
RG           13004
DW           16.500 usec
DE           6.00 usec
TE           298.0 K
D1           1.5000000 sec
d11          0.0300000 sec
d16          0.0002000 sec
d17          0.0001900 sec
MCKEST       0.0000000 sec
MCKRX        0.0150000 sec
F2           31.00 usec

===== CHANNEL f1 =====
NUC1          13C
P1           15.50 usec
PI1          500.00 usec
PI2          2000.00 usec
PL0          120.00 dB
PL1          -1.00 dB
SFO1         125.7804080 MHz
SP1          3.20 dB
SP2          3.20 dB
SFO1M1       Crp60,0.5,20.1
SFO1M2       Crp60comp,4
SPOFF1       0.00 Hz
SPOFF2       0.00 Hz

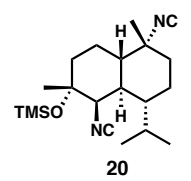
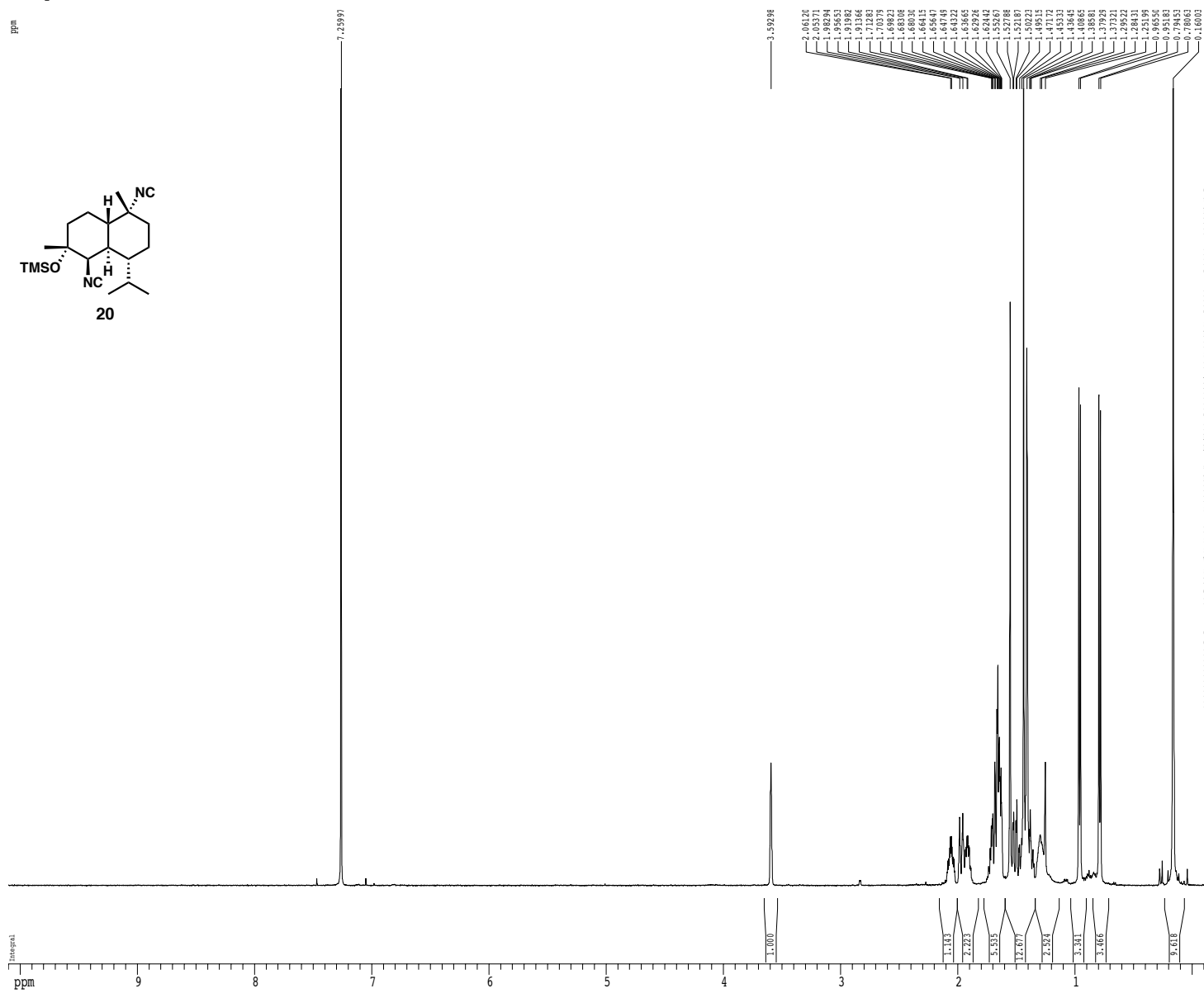
===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2          1H
P2           100.00 usec
PL2          1.60 dB
PL12         24.60 dB
SFO2         500.2225011 MHz

===== GRADIENT CHANNEL =====
GPRAM1       SINE.100
GPRAM2       SINE.100
GPK1         0.00 %
GPK2         0.00 %
GPI1         0.00 %
GPI2         0.00 %
GPE1         30.00 %
GPE2         50.00 %
p15          500.00 usec
p16          1000.00 usec

F2 - Processing parameters
SI           65536
SF           125.7804080 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           1.00

1D NMR plot parameters
CX           22.80 cm
CY           60.00 cm
FIP          200.000 ppm
F1           25156.08 Hz
F2P          -10.000 ppm
FZ           -1237.80 Hz
F2MCH        9.21053 ppm/cm
HSCM         1158.50378 Hz/cm
    
```

1H spectrum



7.25997
3.59298
2.06120
2.05371
1.98553
1.91982
1.81366
1.71283
1.69823
1.68308
1.68030
1.66415
1.65177
1.64745
1.63322
1.63665
1.62926
1.61741
1.55785
1.52788
1.52187
1.50223
1.47137
1.45333
1.43645
1.40865
1.39388
1.37321
1.29522
1.28431
1.25195
1.22195
0.95183
0.79453
0.78063
0.16001

```

Current Data Parameters
USER          medaub
NAME          MED-VI-285pu5
EXPNO         1
PROCNO        1

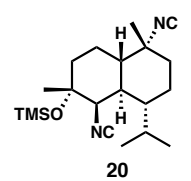
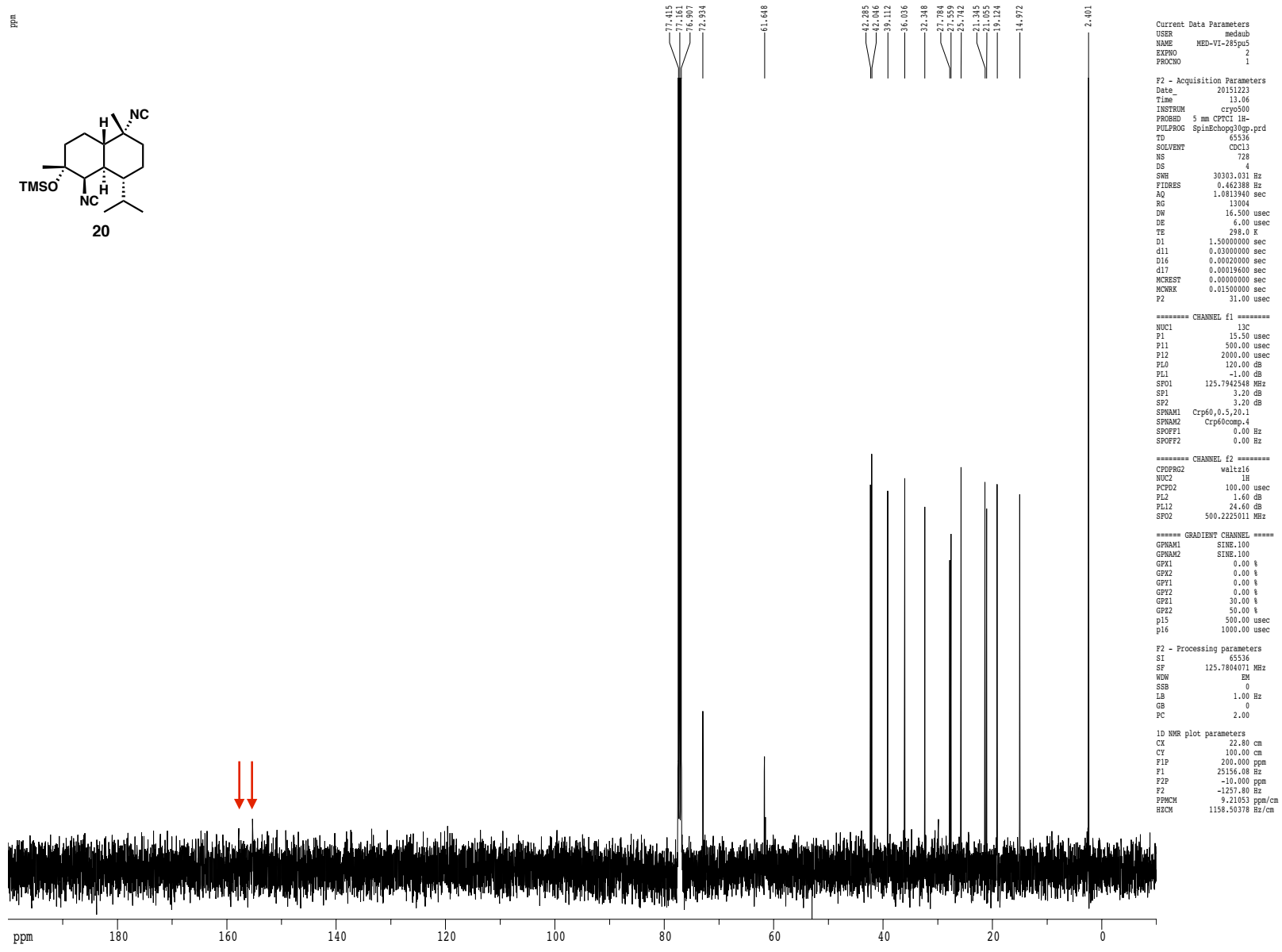
F2 - Acquisition Parameters
Date_         20151223
Time          13.02
INSTRUM       cryo500
PROBHD        5 mm CPTCT 1H
PULPROG       zg30
TD            81728
SOLVENT       CDCl3
NS            8
DS            2
SWH           8012.820 Hz
FIDRES        0.098043 Hz
AQ            5.0998774 sec
RG            71.1
DW            62.400 usec
DE            6.00 usec
TE            298.0 K
D1            0.10000000 sec
MCREST        0.00000000 sec
MCNKR         0.01500000 sec

===== CHANNEL f1 =====
NUC1           1H
P1             7.50 usec
PL1            1.60 dB
SF01          500.2235015 MHz

F2 - Processing parameters
SI            65536
SF            500.2200306 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            2.00

1D NMR plot parameters
CX            22.80 cm
CY            60.00 cm
FIP           10.100 ppm
F1            5052.22 Hz
F2P           -0.100 ppm
F2            -50.02 Hz
PPMCH         0.44737 ppm/cm
HSCX          223.78267 Hz/cm
    
```

Z-restored spin-echo ¹³C spectrum with ¹H decoupling



```

Current Data Parameters
USER          medaub
NAME          MED-VI-285pu5
EXPNO        2
PROCNO       1

F2 - Acquisition Parameters
Date_        20151223
Time         13.06
INSTRUM      cryo500
PROBHD       5 mm CPXI 1H-
PULPROG      SpinEchopg30pp.prd
TD           65536
SOLVENT      CDCl3
NS           728
DS           4
SMB          30303.031 Hz
FIDRES       0.462988 Hz
AQ           1.0813940 sec
RG           13004
DA           16.500 usec
DE           6.00 usec
TE           298.0 K
D1           1.5000000 sec
d11          0.0300000 sec
D16          0.0020000 sec
d17          0.00019600 sec
MCREST      0.0000000 sec
MORPK       0.01500000 sec
F2           31.00 usec

===== CHANNEL f1 =====
NUC1         13C
P1           15.50 usec
P11          500.00 usec
P12          2000.00 usec
PL0          120.00 dB
PL1          -1.00 dB
SFO1        125.7942548 MHz
SP1          3.20 dB
SP2          3.20 dB
SFO1M1      Crp60,0.5,20.1
SFO1M2      Crp60comp,4
SFOFF1      0.00 Hz
SFOFF2      0.00 Hz

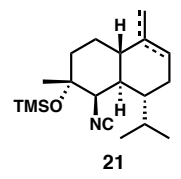
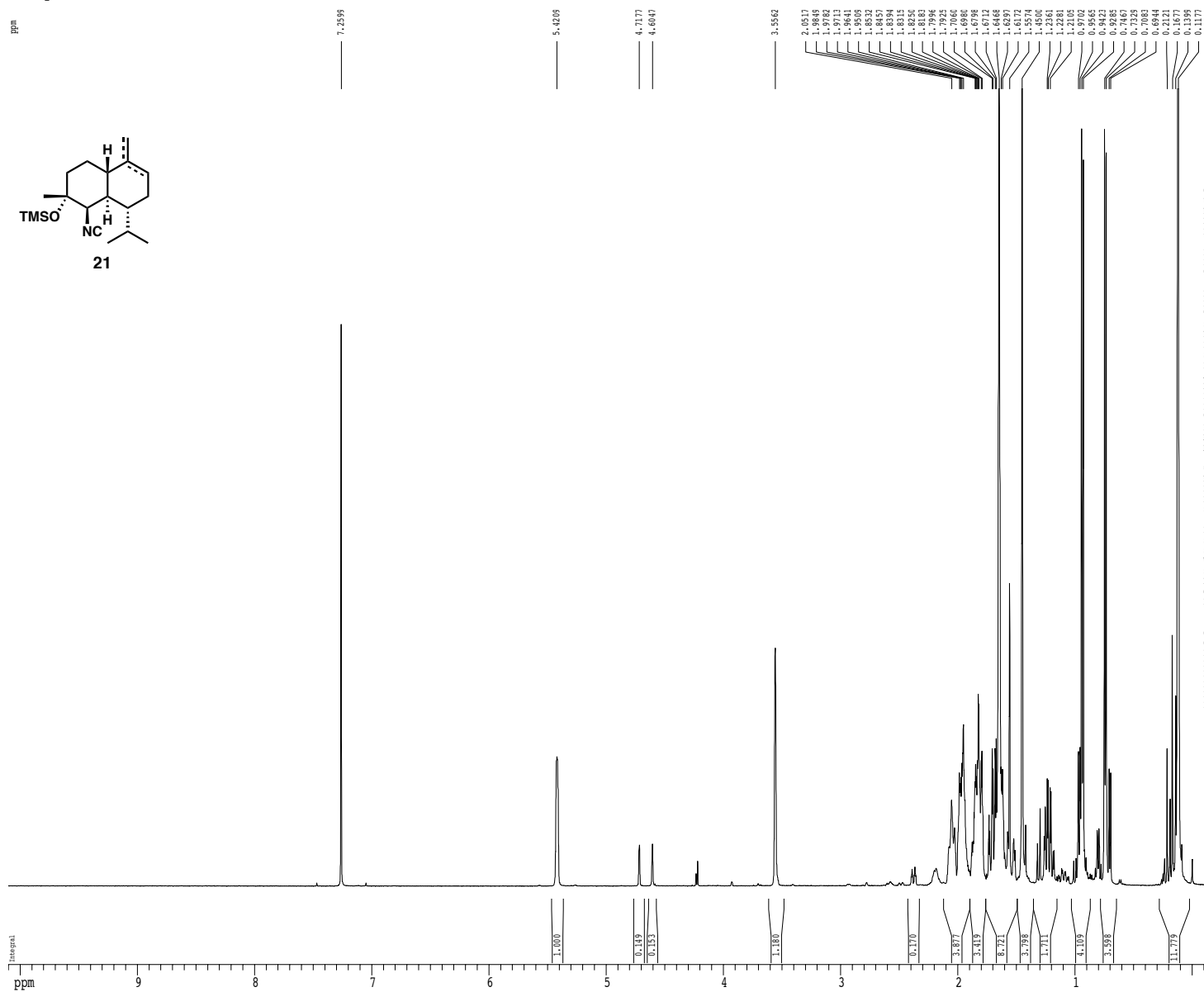
===== CHANNEL f2 =====
CPDPRG2     waltz16
NUC2         1H
PCPD2       100.00 usec
PL2         1.60 dB
PL12        24.60 dB
SFO2        500.2229011 MHz

===== GRADIENT CHANNEL =====
GPM1        SINE,100
GPM2        SINE,100
GPX1        0.00 %
GPX2        0.00 %
GPI1        0.00 %
GPI2        0.00 %
GPI3        30.00 %
GPI4        50.00 %
p15         500.00 usec
p16         1000.00 usec

F2 - Processing parameters
SI          65536
SF          125.7804071 MHz
WDW         EM
SSB         0
LB          1.00 Hz
GB          0
PC          2.00

1D NMR plot parameters
CY          22.80 cm
CY          100.00 cm
F1          25156.08 Hz
F2          -10.000 ppm
F2          -1257.80 Hz
FPMCH      9.21053 ppm/cm
HSCN       1158.50378 Hz/cm
    
```

1H spectrum



Current Data Parameters

USER medaub

NAME MED-VI-285pu4

EXPNO 1

PROCNO 1

F2 - Acquisition Parameters

Date_ 20151223

Time 9.24

INSTRUM cryo500

PROBHD 5 mm CPCTC 1H-

PULPROG zg30

TD 81728

SOLVENT CDCl3

NS 8

DS 2

SWH 8012.820 Hz

FIDRES 0.098043 Hz

AQ 5.0998774 sec

RG 9

DW 62.400 usec

DE 6.00 usec

TE 298.0 K

D1 0.1000000 sec

MCREST 0.0000000 sec

MCONK 0.0150000 sec

===== CHANNEL f1 =====

NUC1 1H

P1 7.50 usec

PL1 1.60 dB

SFO1 500.2235015 MHz

F2 - Processing parameters

SI 65536

SF 500.2200305 MHz

WDW EM

SSB 0

LB 0.30 Hz

GB 0

PC 15.00

1D NMR plot parameters

CX 22.80 cm

CY 80.00 cm

F1P 10.100 ppm

F1 5052.22 Hz

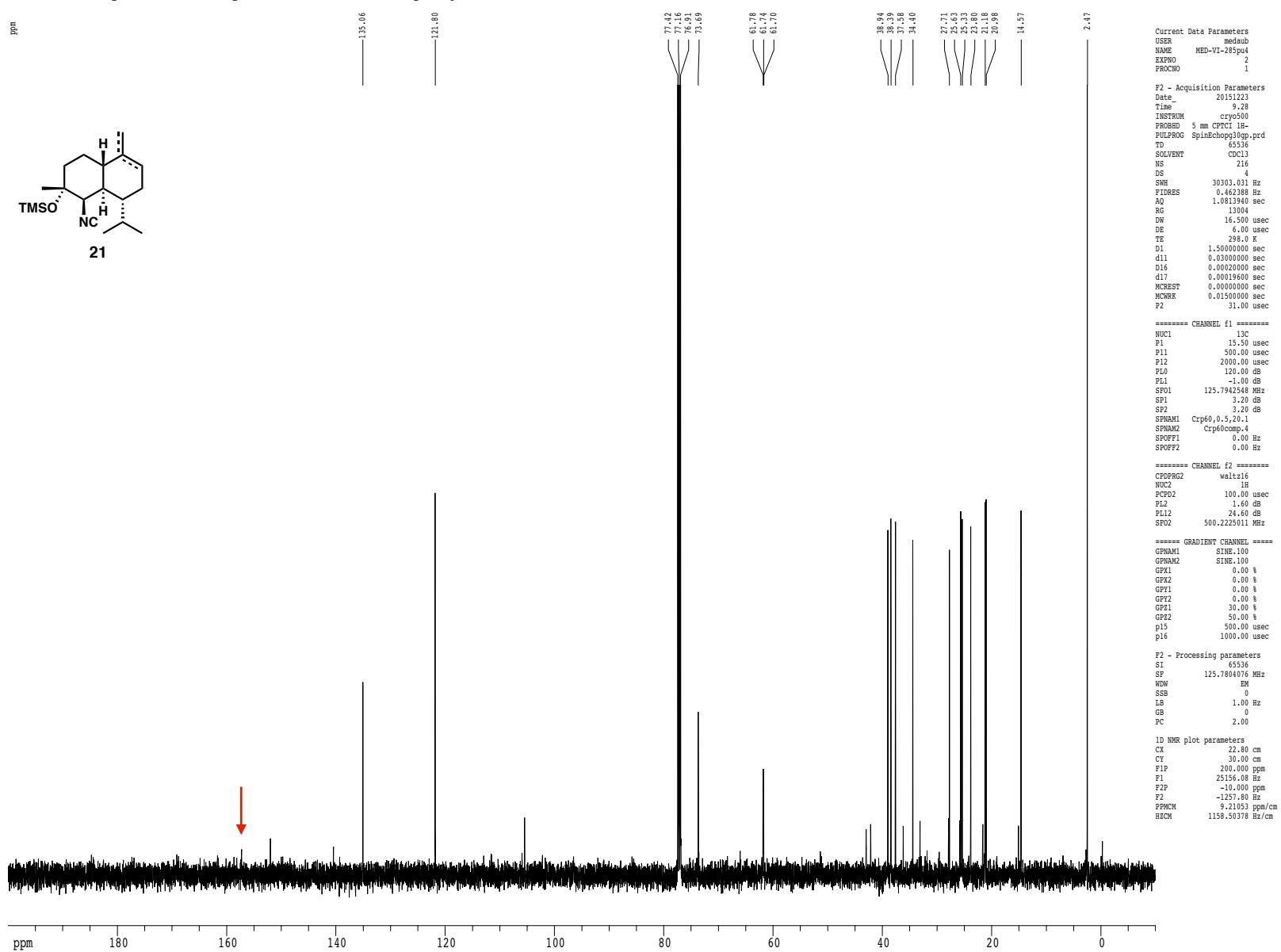
F2P -0.100 ppm

F2 -50.02 Hz

PPMCH 0.44737 ppm/cm

HZCH 223.78267 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER      medaub
NAME      MED-VI-285p4
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20151223
Time      9.28
INSTRUM   cryo500
PROBHD    5 mm CP1C1 1H-
FILEPROG  SpinEcho30p.prd
TD         65536
SOLVENT   CDCl3
NS         216
DS         4
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         13004
DW         16.500 usec
DE         6.000 usec
TE         298.0 K
D1         1.50000000 sec
d11        0.03000000 sec
d16        0.00020000 sec
d17        0.00019600 sec
MCREST     0.00000000 sec
MCMRFX     0.01500000 sec
P2         31.000 usec

===== CHANNEL f1 =====
NUC1       13C
P1         15.50 usec
PL1        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1       125.7842548 MHz
SP1        3.20 dB
SP2        3.20 dB
SFO1M1     Crp60,0.5,20.1
SFO1M2     Crp60cm0,4
SFOFF1     0.00 Hz
SFOFF2     0.00 Hz

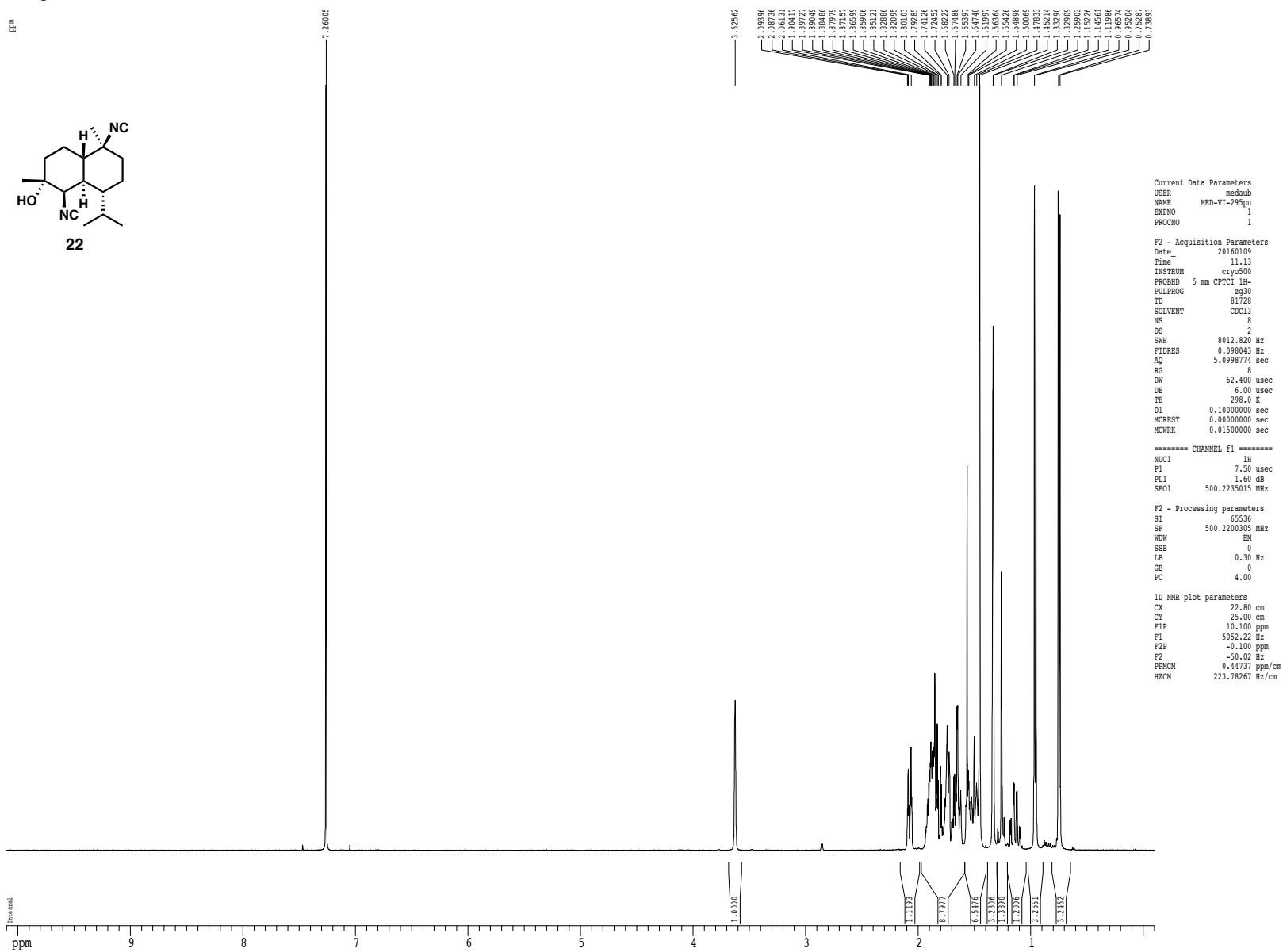
===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2       1H
PCPD2     100.00 usec
PL2        1.60 dB
PL12       24.60 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1     SINE.100
GPNAM2     SINE.100
GPF1       0.00 %
GPF2       0.00 %
GPF1       0.00 %
GPF2       0.00 %
GPF1       30.00 %
GPF2       50.00 %
p15        500.00 usec
p16        1000.00 usec

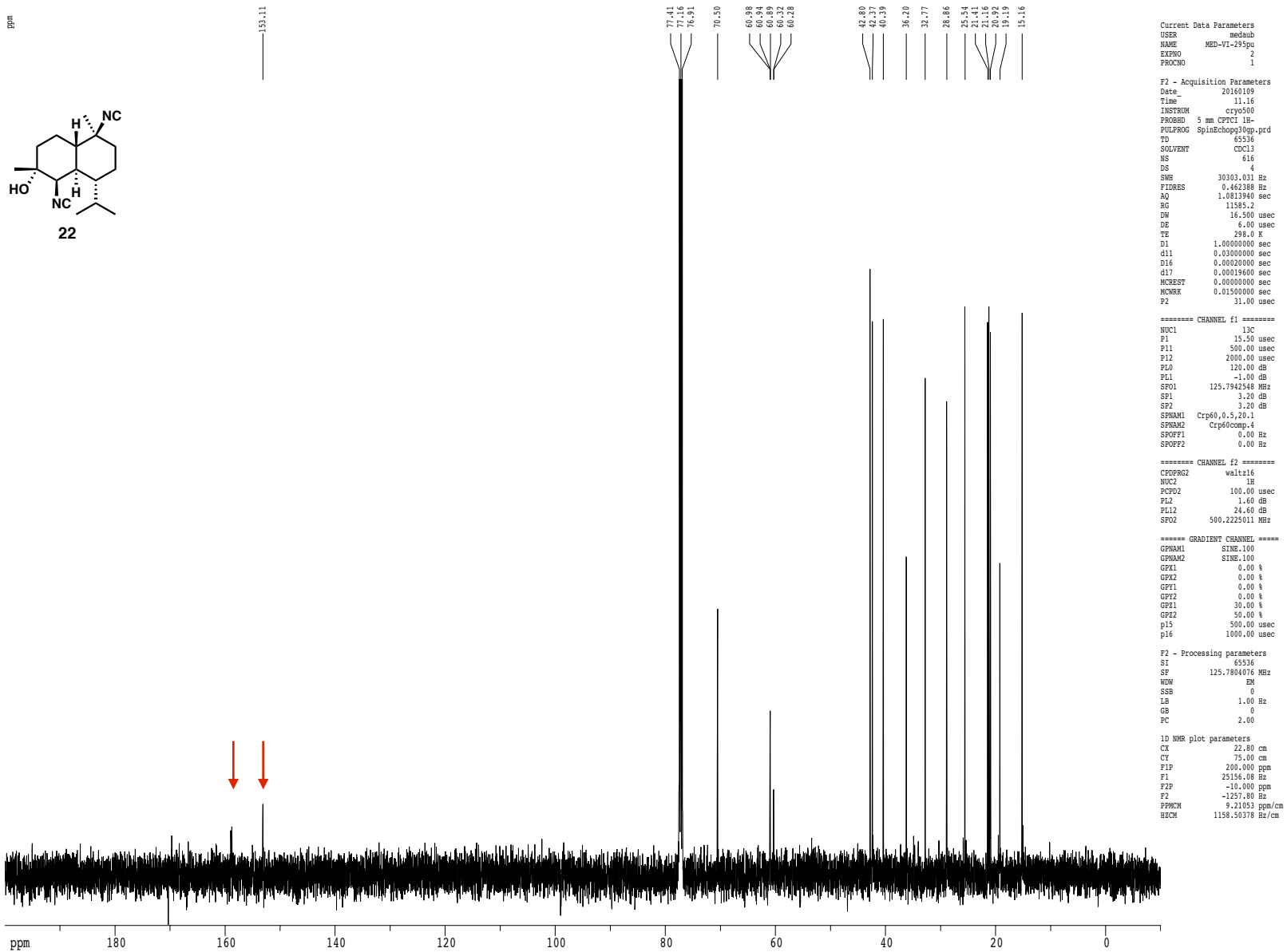
F2 - Processing parameters
SI         65536
SF         125.7804076 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

1D NMR plot parameters
CX         22.80 cm
CI         30.00 cm
FIP        200.000 ppm
F1         25156.08 Hz
F2P        -10.000 ppm
F2         -1257.80 Hz
F2MCH      9.21953 ppm/cm
HECM       1158.50378 Hz/cm
    
```

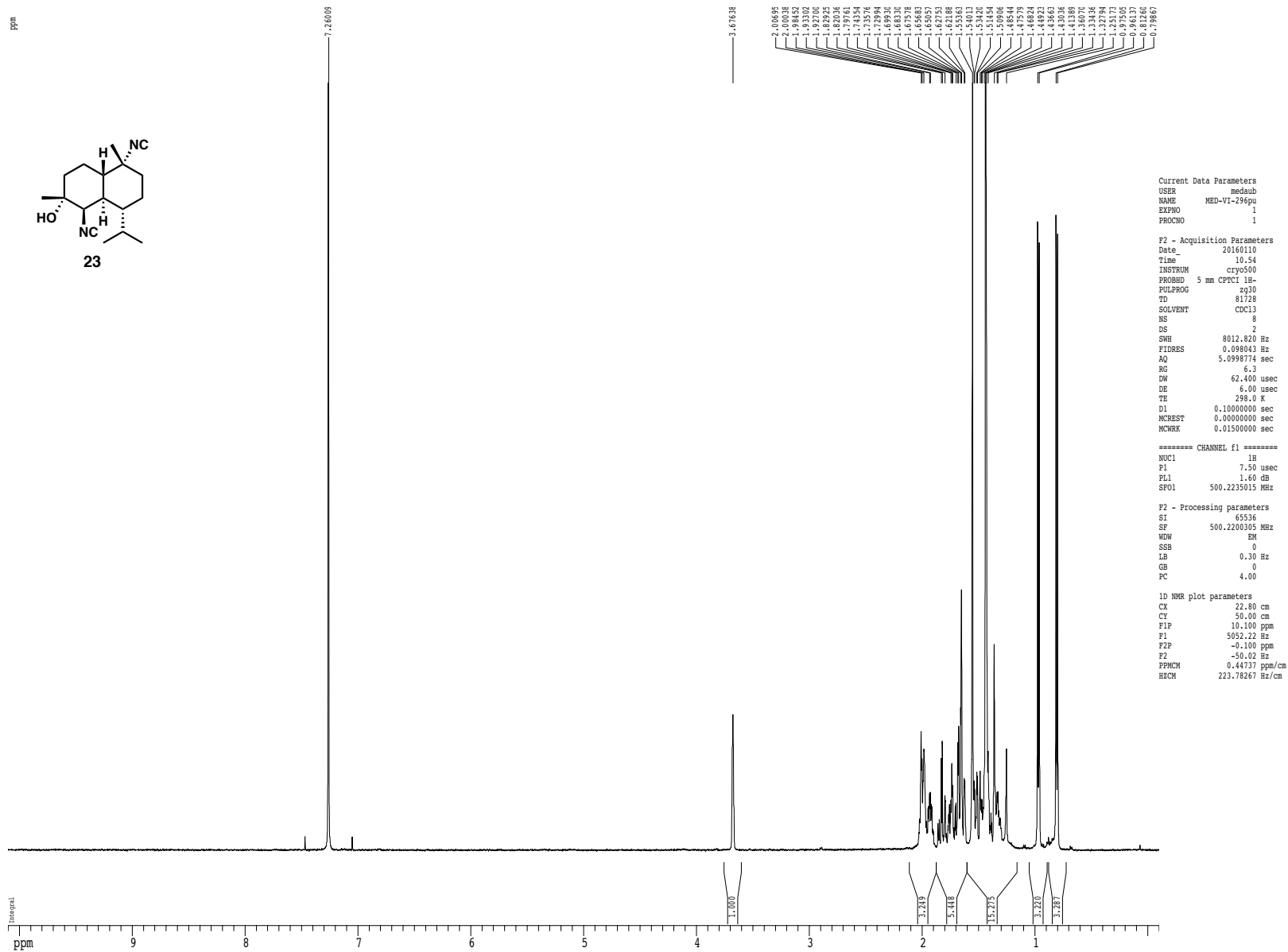
1H spectrum



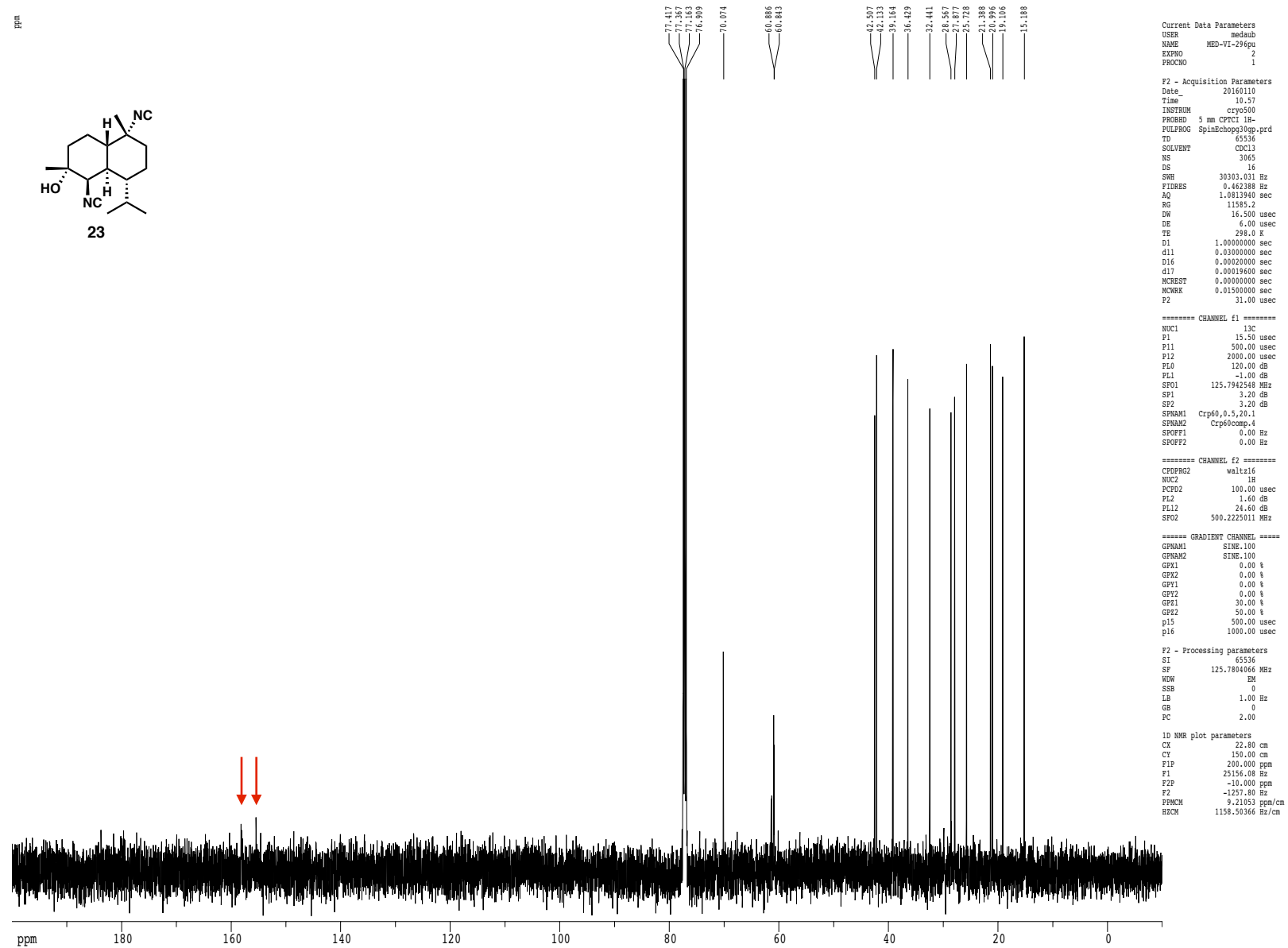
Z-restored spin-echo 13C spectrum with 1H decoupling



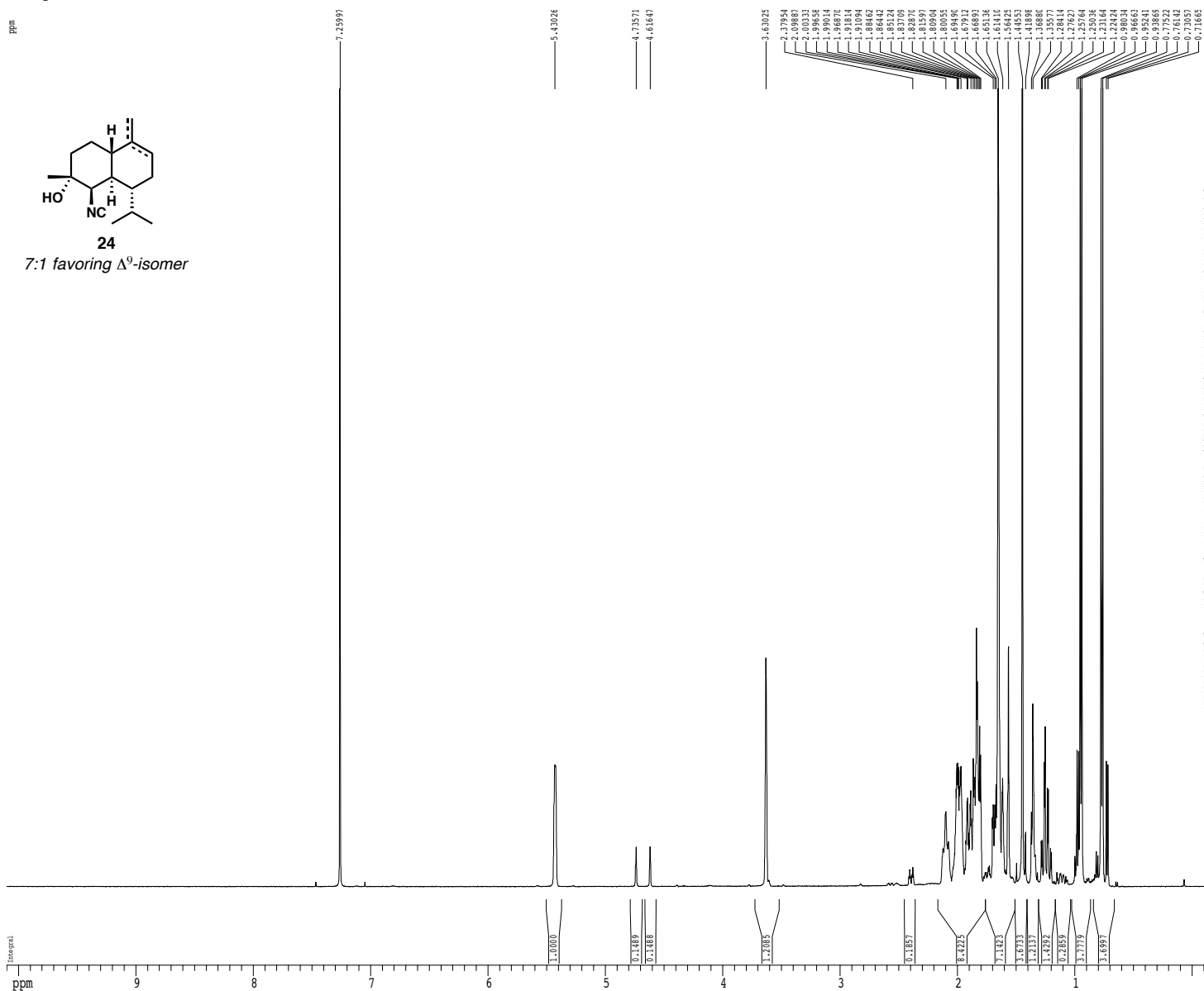
1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum



```

Current Data Parameters
USER      medaub
NAME      MED-VI-299pu
EXPNO     1
PROCNO    1

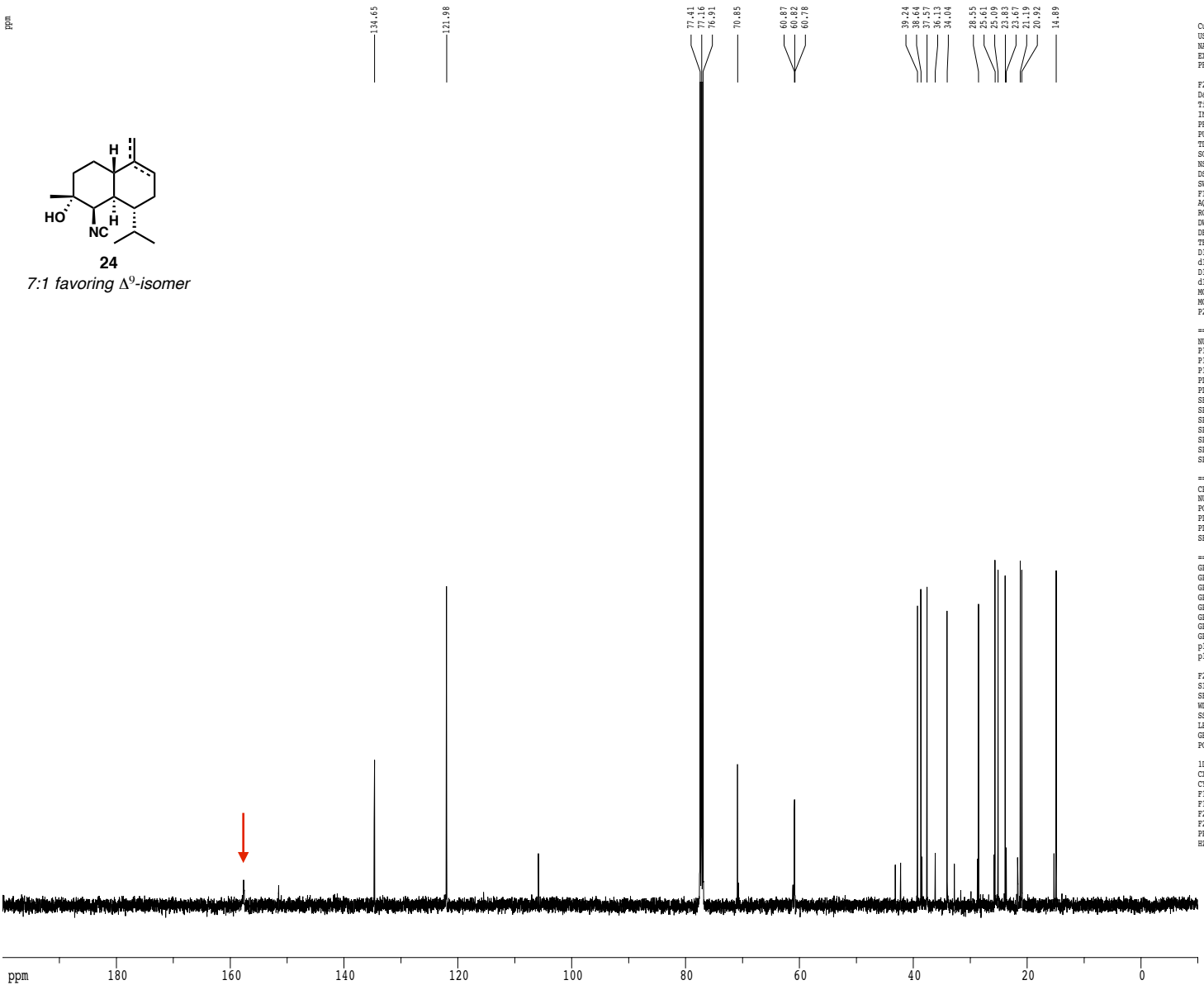
F2 - Acquisition Parameters
Date_     20161110
Time      13.30
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         81728
SOLVENT   CDCl3
NS         8
DS         2
SHH        8012.920 Hz
FIDRES    0.098043 Hz
AQ         5.0998774 sec
RG         9
DW         62.400 usec
DE         4.00 usec
TE         298.0 K
D1         0.10000000 sec
MCREST    0.00000000 sec
MCNRX     0.01500000 sec

===== CHANNEL f1 =====
NUC1      1H
P1        7.50 usec
PL1       1.60 db
SF01      500.2235015 MHz

F2 - Processing parameters
SI         65536
SF         500.2200306 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00

1D NMR plot parameters
CX         22.80 cm
CY         35.00 cm
F1P        10.100 ppm
F1         5052.22 Hz
F2P        -0.100 ppm
F2         -50.00 Hz
FPMCM     0.44737 ppm/cm
HZCM      223.78267 Hz/cm
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER      medaub
NAME      MED-VI-299pu
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20160110
Time      14.02
INSTRUM   cryo500
PROBHD    5 mm CPCCI 1H-
PULPROG   SpinEcho30ap.prd
TD         65536
SOLVENT   CDCl3
NS         904
DS         4
SNR        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         13004
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         1.00000000 sec
d11        0.03000000 sec
D16        0.00020000 sec
d17        0.00019600 sec
MCKWST     0.00000000 sec
MCKWRK     0.01500000 sec
P2         31.00 usec

===== CHANNEL f1 =====
NUC1       13C
P1         15.50 usec
P11        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1      125.7942548 MHz
SF1        3.20 dB
SP2        3.20 dB
SFOAM1     Crp60,0.5,20.1
SFOAM2     Crp60comp,4
SFOFF1     0.00 Hz
SFOFF2     0.00 Hz

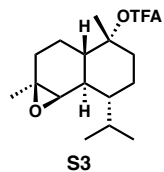
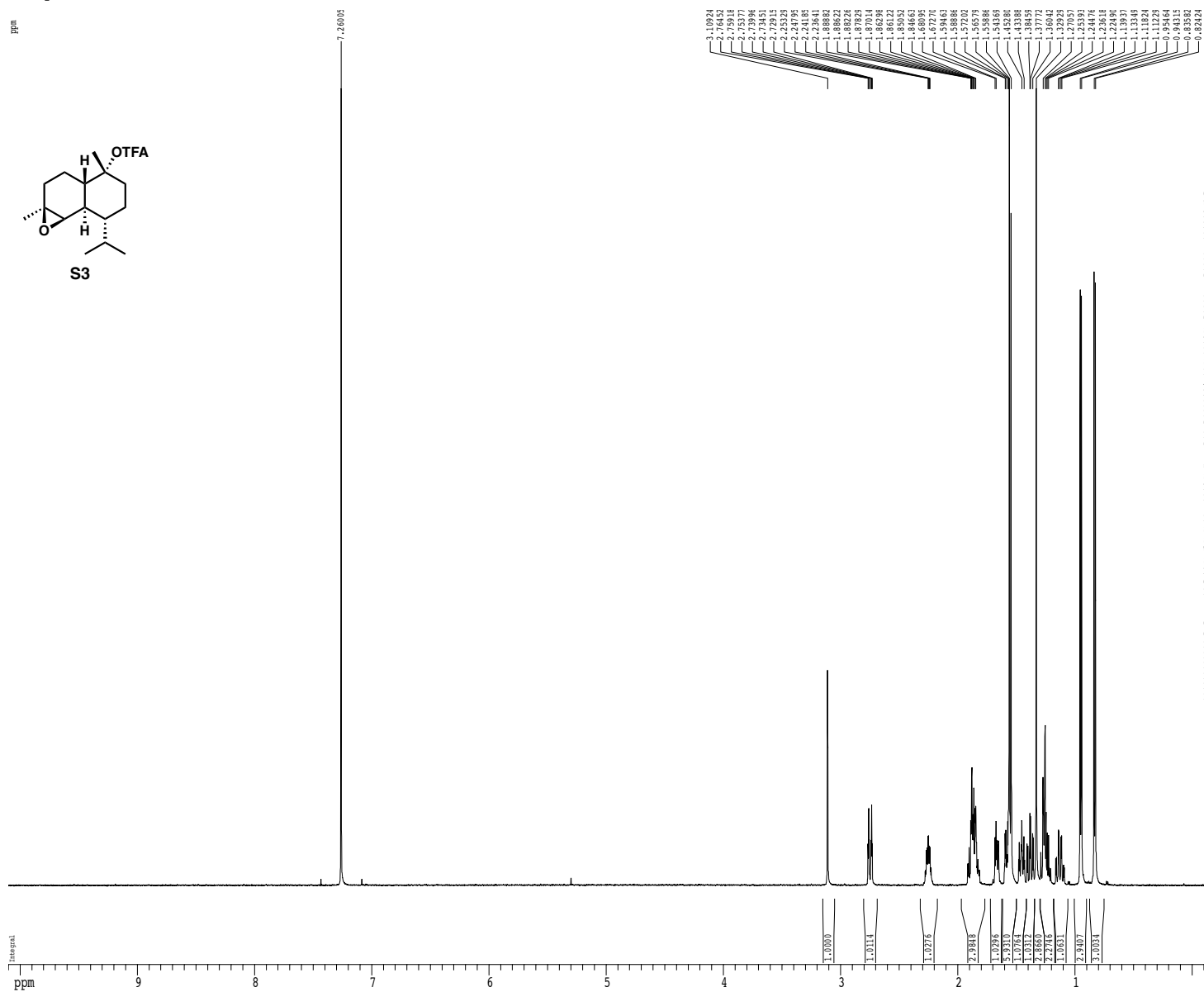
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2     100.00 usec
PL2        1.40 dB
PL12       24.40 dB
SFO2      500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1     SINE.100
GPNAM2     SINE.100
GPX1       0.00 t
GPX2       0.00 t
GPY1       0.00 t
GPY2       0.00 t
GPZ1       30.00 t
GPZ2       50.00 t
p15        500.00 usec
p16        1000.00 usec

F2 - Processing parameters
SI         65536
SF         125.7804076 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

1D NMR plot parameters
CX         22.80 cm
CT         30.00 cm
PLP        200.000 ppm
F1         25156.08 Hz
F2P        -10.000 ppm
F2         -1257.80 Hz
PPHMC      9.21053 ppm/cm
HRCM       1158.50378 Hz/cm
    
```

1H spectrum



Current Data Parameters
 USER medaub
 NAME MED-VI-199pu
 EXPNO 1
 PROCNO 1

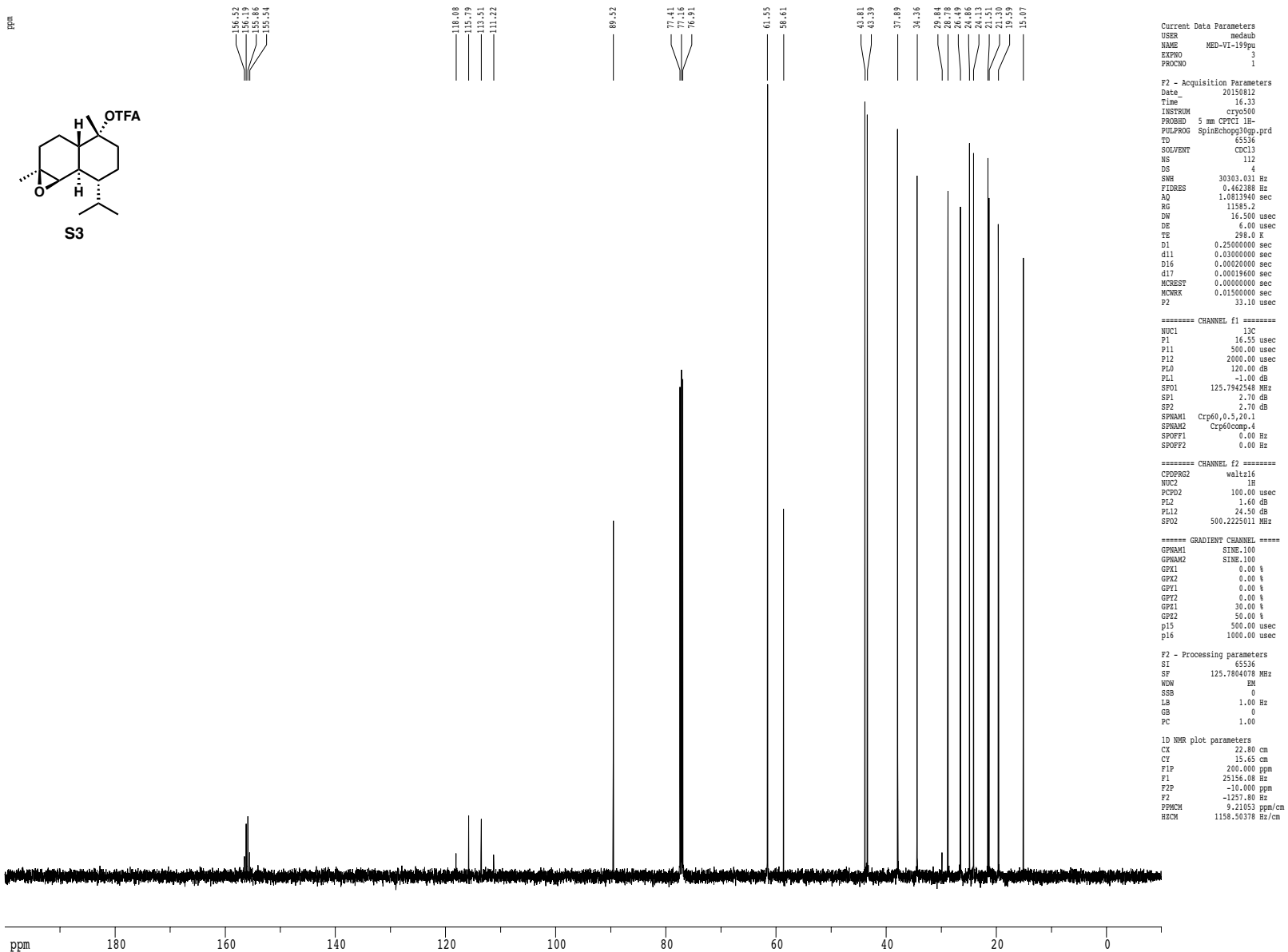
F2 - Acquisition Parameters
 Date_ 20150812
 Time 14.33
 INSTRUM av600
 PROBRD 5 mm BBO BB-1H
 PULPROG zg30
 TD 98074
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 9615.385 Hz
 FIDRES 0.098042 Hz
 AQ 5.0998979 sec
 RG 645
 DW 52.000 usec
 DE 14.33 usec
 TE 298.4 K
 D1 0.1000000 sec
 TDO 1

===== CHANNEL f1 =====
 SF01 600.1342009 MHz
 NUCL 1H
 P1 9.00 usec

F2 - Processing parameters
 S1 65536
 SF 600.1300344 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 22.80 cm
 CY 25.00 cm
 FIP 10.100 ppm
 F1 6061.31 Hz
 F2P -0.100 ppm
 F2 -60.01 Hz
 PPMCM 0.44737 ppm/cm
 HCM 266.47925 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER      medaub
NAME      MED-VI-199pu
EXPNO     3
PROCNO    1

F2 - Acquisition Parameters
Date_     20150812
Time      16.33
INSTRUM   cryo500
PROBHD    5 mm CP1C1 1H-
PULPROG   SpinEchoSgppg.prd
TD         65536
SOLVENT   CDCl3
NS         112
DS         4
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         11985.2
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
d16        0.00020000 sec
d17        0.00019600 sec
MCREST     0.00000000 sec
MCMRFX     0.01500000 sec
P2         33.10 usec

===== CHANNEL f1 =====
NUC1       13C
P1         16.55 usec
P11        500.00 usec
P12        2000.00 usec
PLO        120.00 dB
PL1        -1.00 dB
SFO1       125.7842548 MHz
SP1        2.70 dB
SP2        2.70 dB
SFOAM1     Crp60,0.5,20.1
SFOAM2     Crp60cmd,4
SFOFF1     0.00 Hz
SFOFF2     0.00 Hz

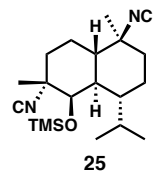
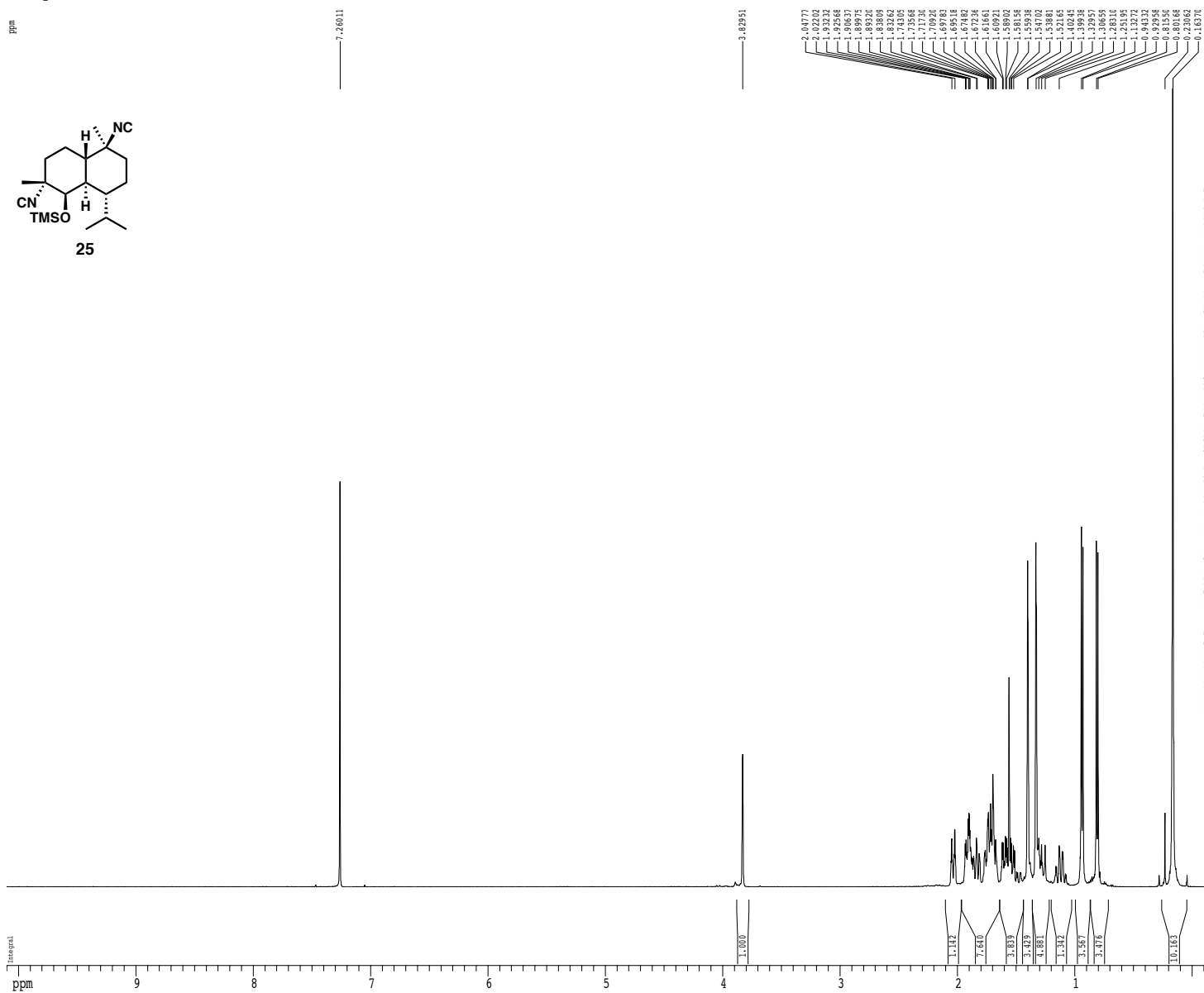
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2     100.00 usec
PL2        1.60 dB
PL12       24.50 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1     SINE.100
GPNAM2     SINE.100
GPF1       0.00 %
GPF2       0.00 %
GPI1       0.00 %
GPI2       0.00 %
GPI3       30.00 %
GPI4       50.00 %
p15        500.00 usec
p16        1000.00 usec

F2 - Processing parameters
SI         65536
SF         125.7804078 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.00

1D NMR plot parameters
CX         22.80 cm
CI         15.65 cm
FIP        200.000 ppm
F1         25156.08 Hz
F2P        -10.000 ppm
F2         -1257.80 Hz
FPMCH      9.21833 ppm/cm
HECM       1158.50378 Hz/cm
    
```

1H spectrum



Current Data Parameters
 USER medaub
 NAME MED-VI-283pub
 EXPNO 1
 PROCNO 1

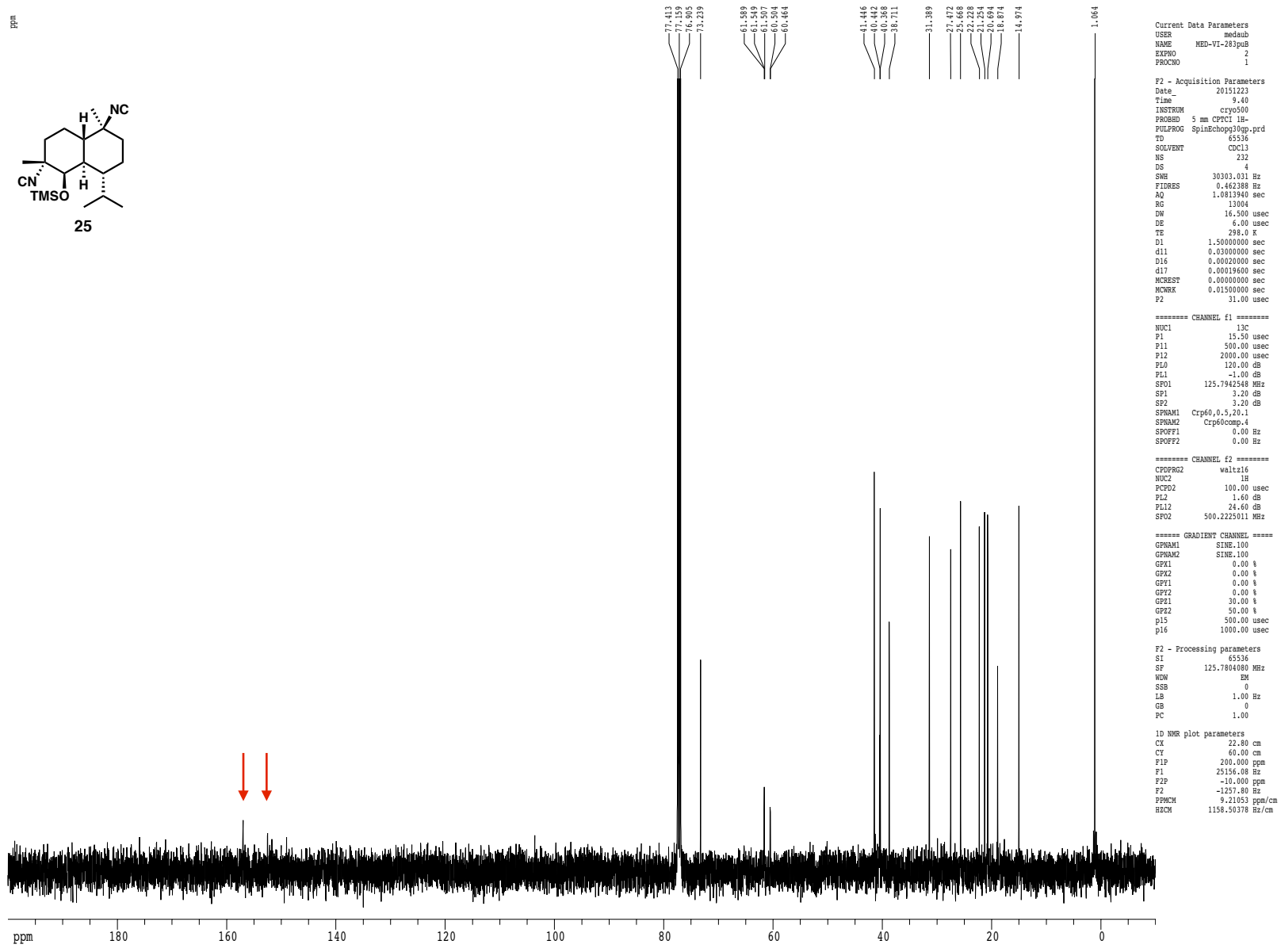
F2 - Acquisition Parameters
 Date_ 20151223
 Time 8.37
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.920 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 10.1
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCMRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.60 db
 SF01 500.2235015 MHz

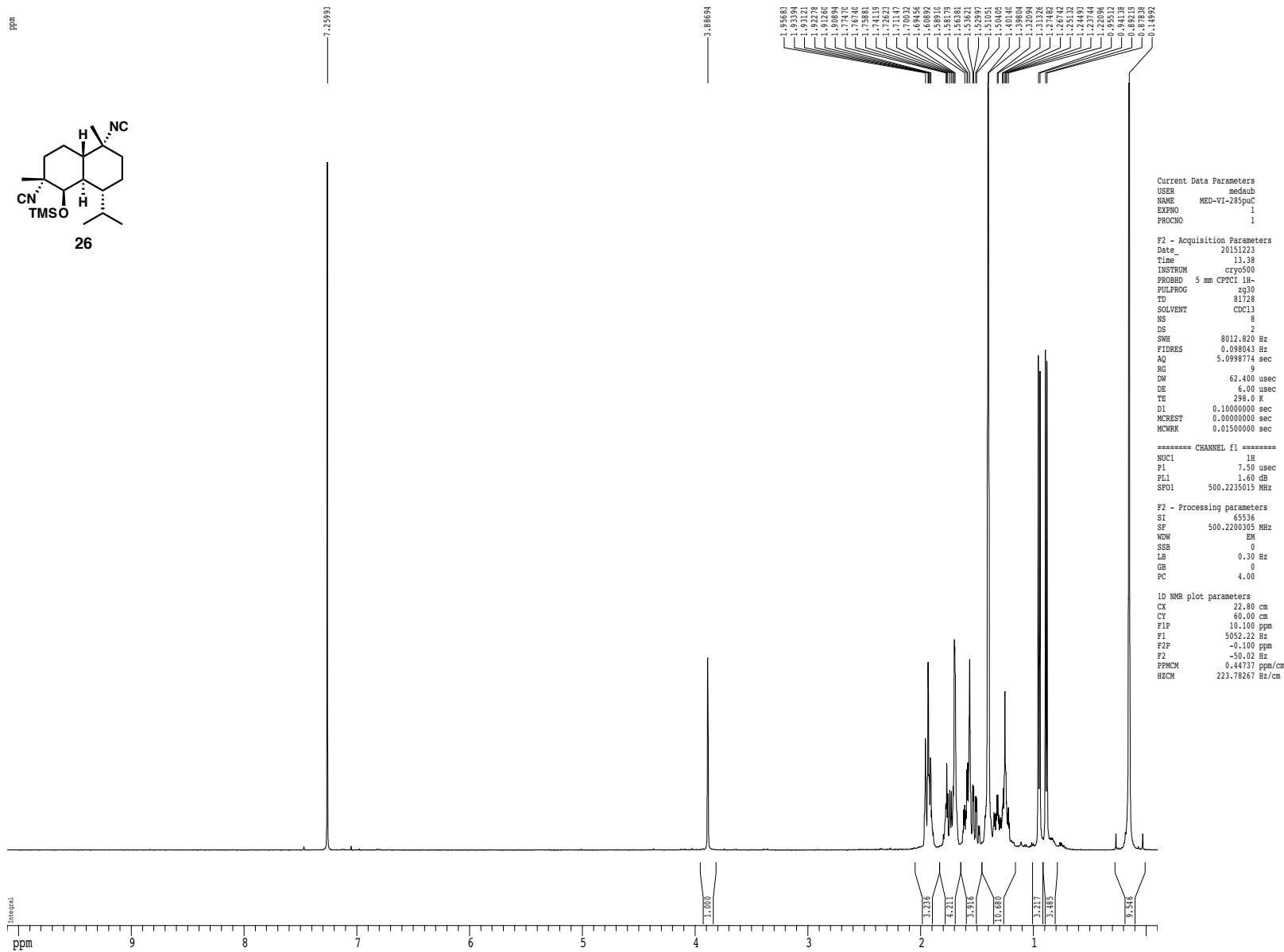
F2 - Processing parameters
 SI 65536
 SF 500.2200304 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 22.80 cm
 CY 40.00 cm
 F1P 10.100 ppm
 F1 5052.22 Hz
 F2P -0.100 ppm
 F2 -50.00 Hz
 FPMCM 0.44737 ppm/cm
 HCM 223.78267 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling

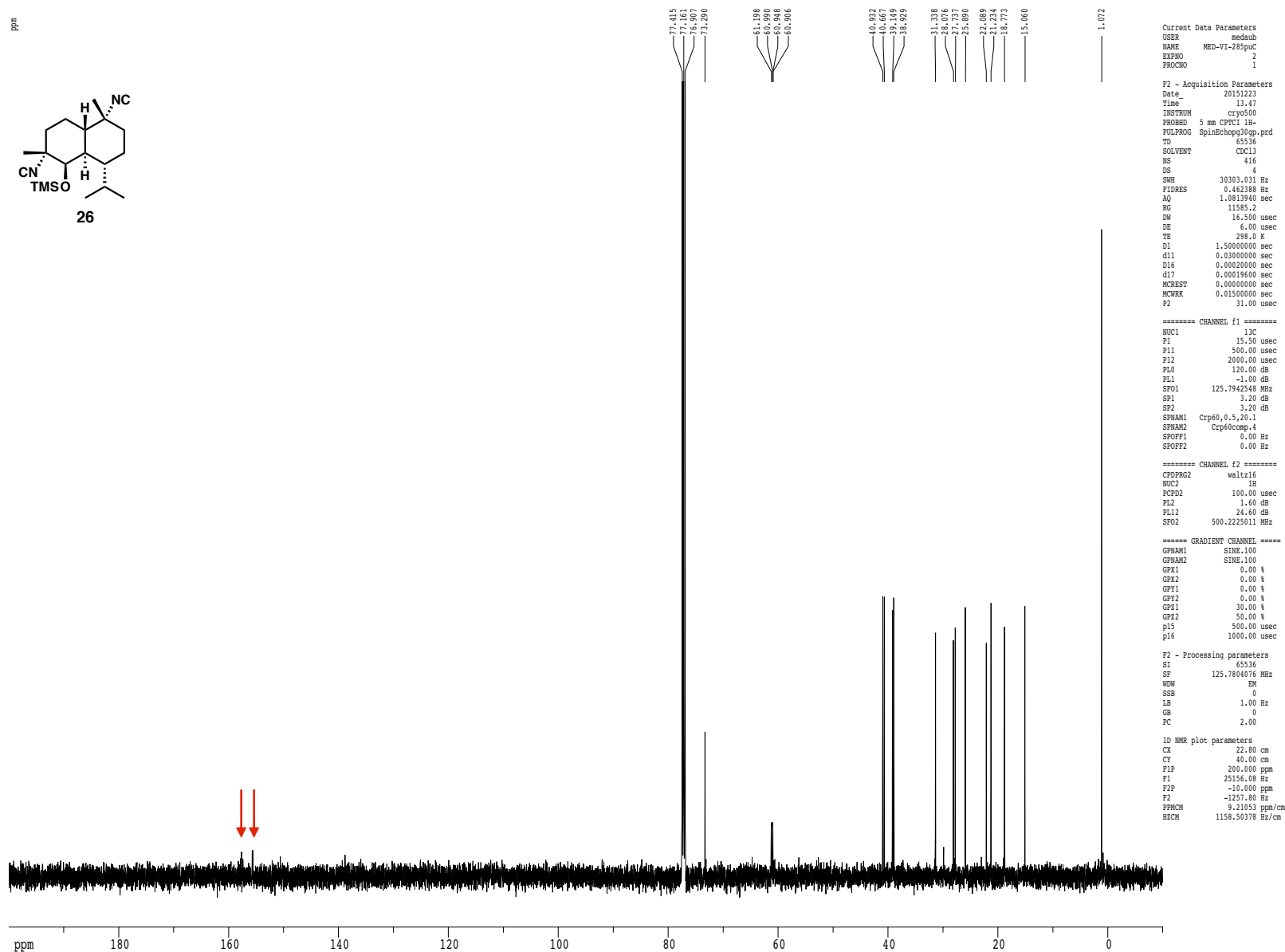
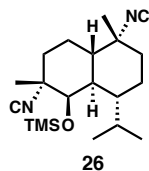


¹H spectrum

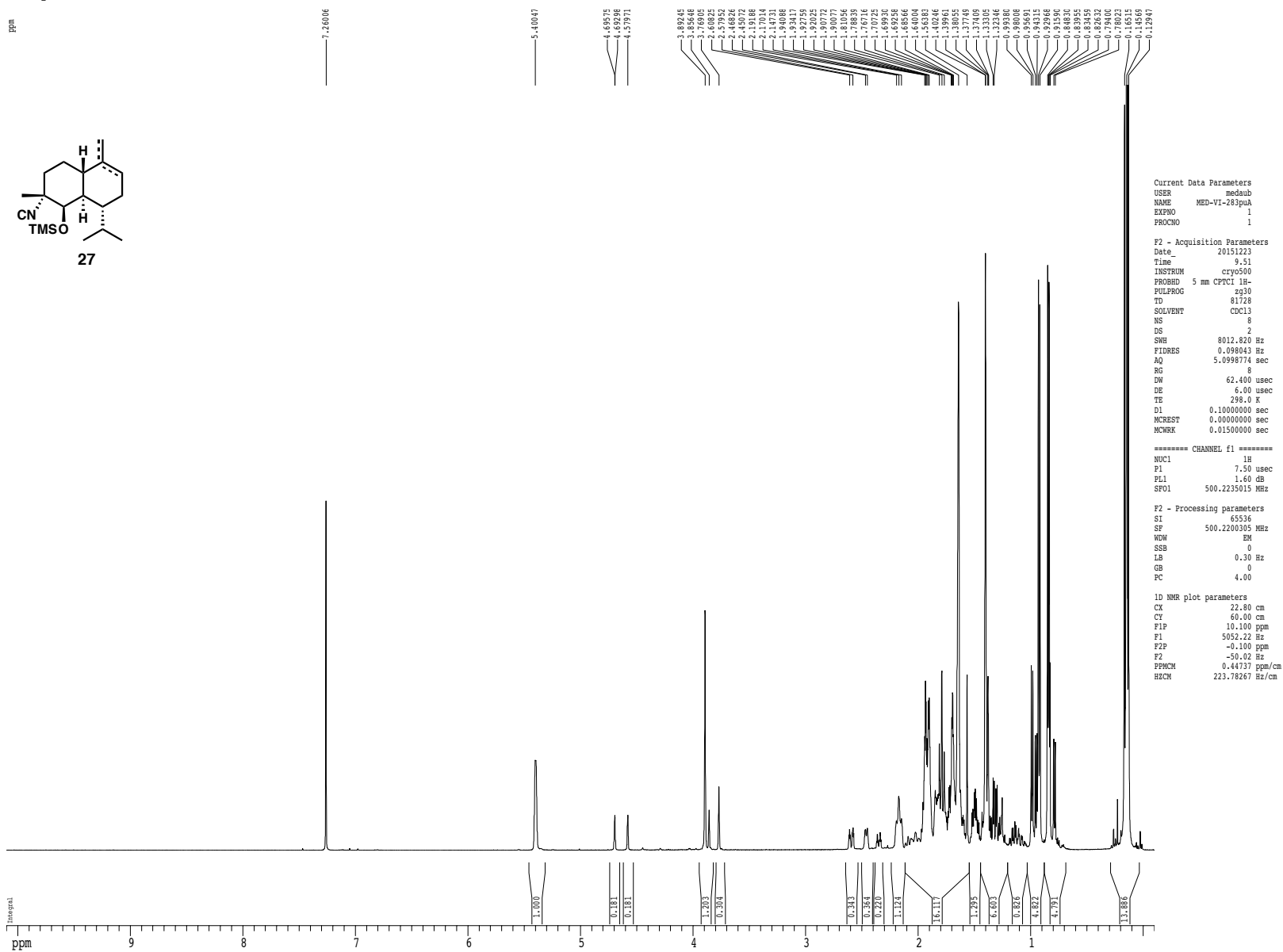


Z-restored spin-echo 13C spectrum with 1H decoupling

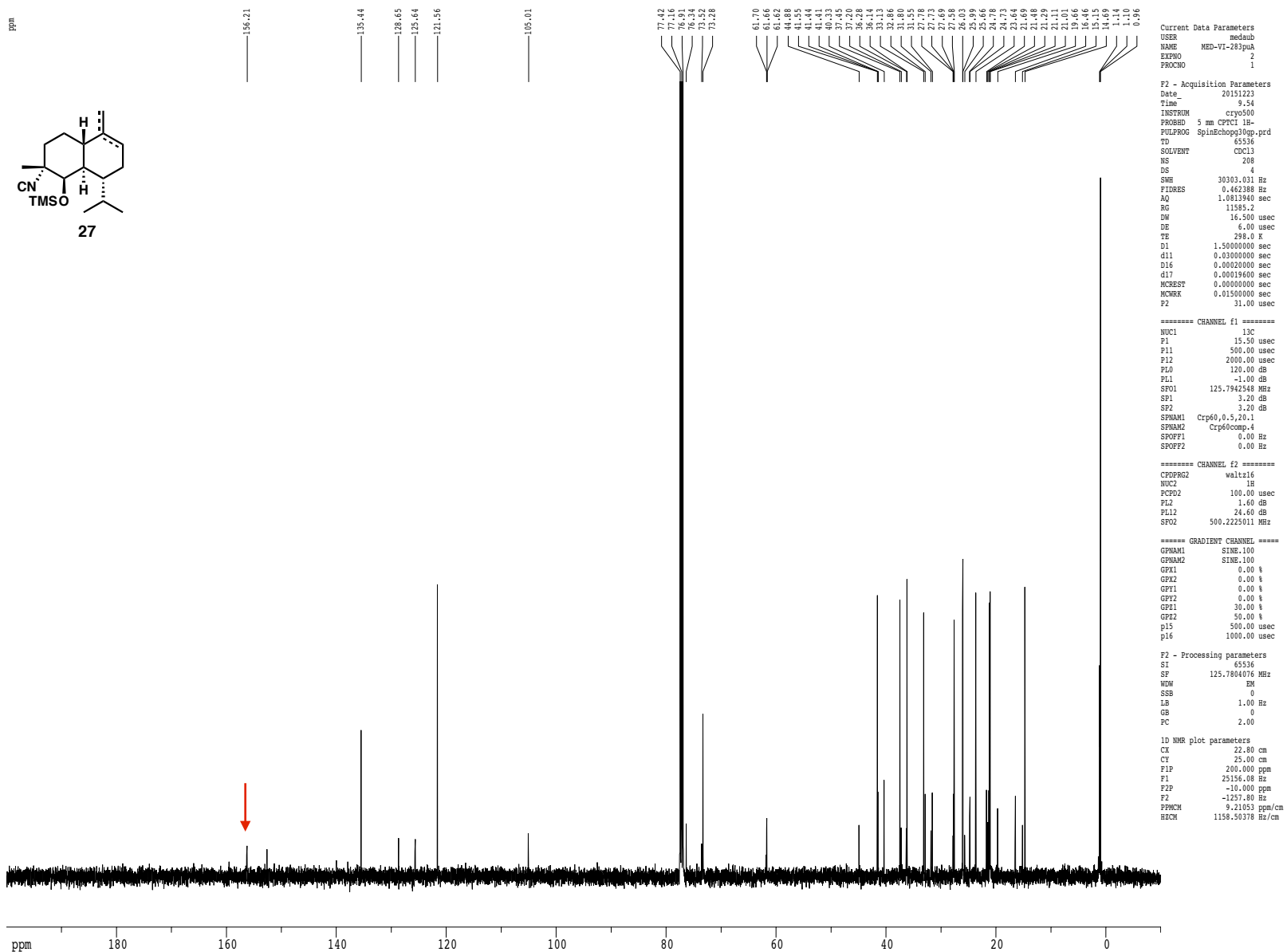
ppm



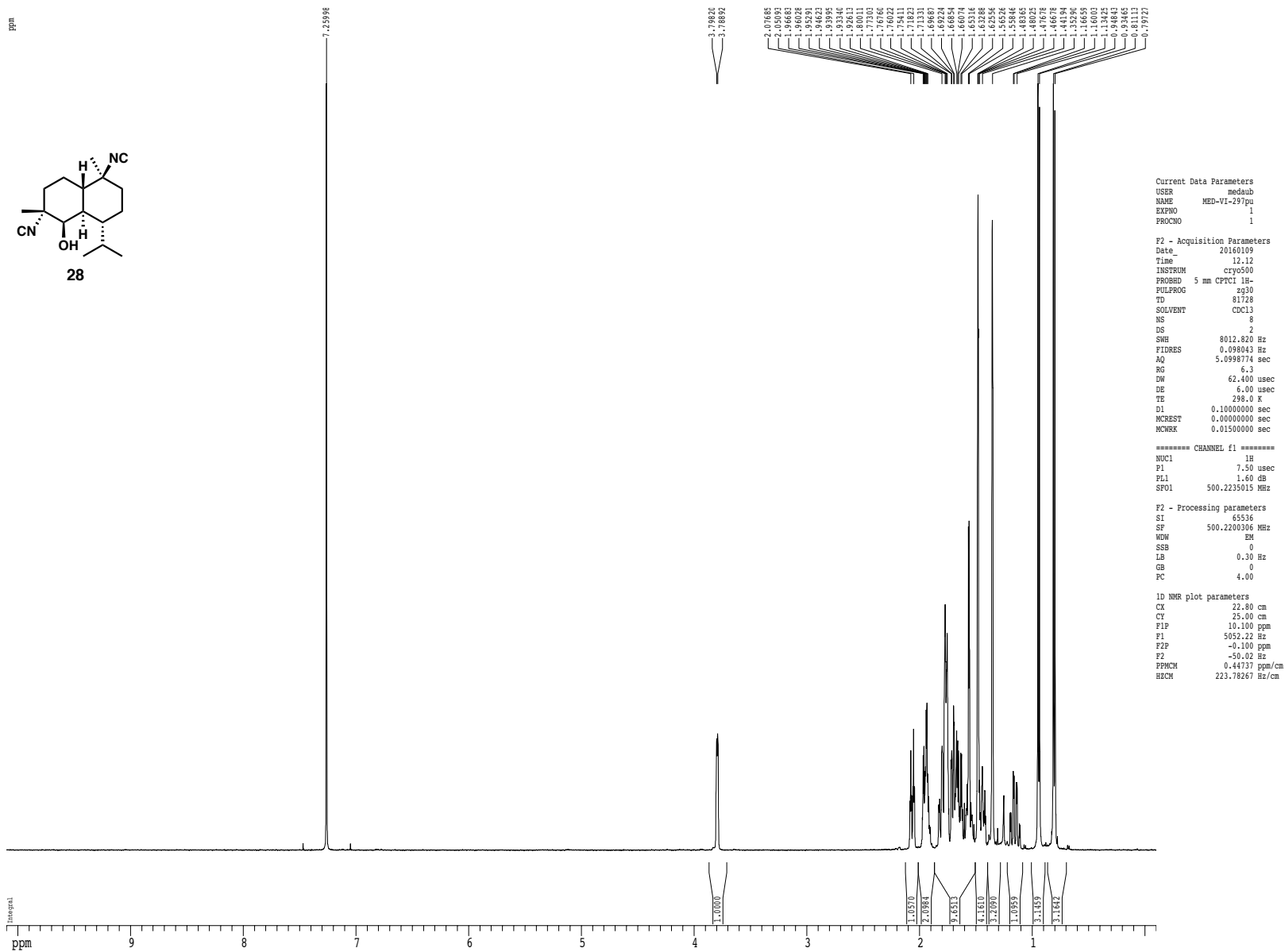
1H spectrum



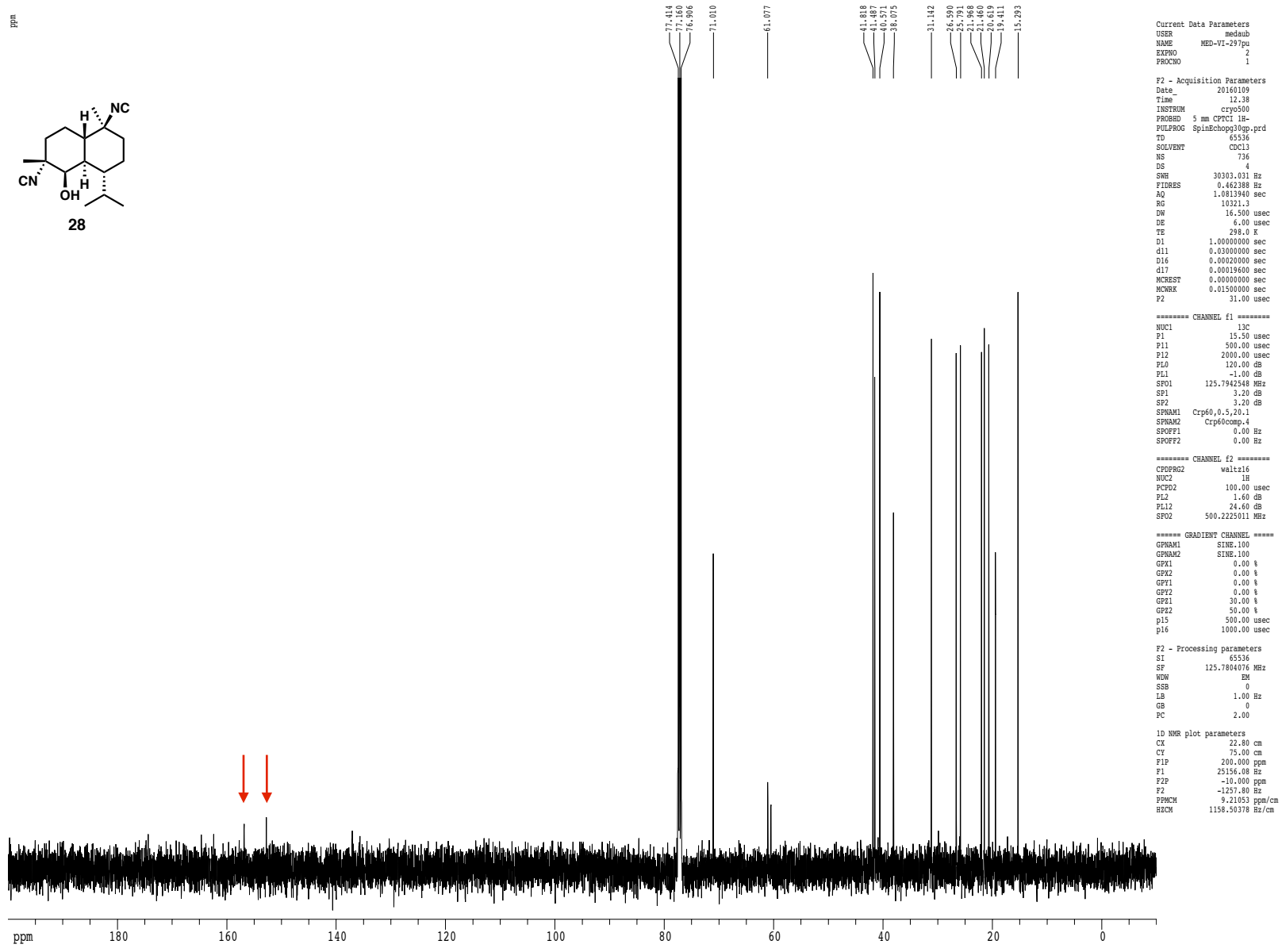
Z-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER          medaubb
NAME         MED-VI-297pu
EXPNO        2
PROCNO       1

F2 - Acquisition Parameters
Date_        20160109
Time         12.38
INSTRUM      cryo500
PROBHD       5 mm CPXI 1H-
PULPROG      SpinEcho30pp-prd
TD           65536
SOLVENT      CDCl3
NS           736
DS           4
SWH          30303.031 Hz
FIDRES       0.462388 Hz
AQ           1.0813940 sec
RG           10321.3
DW           16.500 usec
DE           6.00 usec
TE           298.0 K
D1           1.00000000 sec
d11          0.03000000 sec
D16          0.00200000 sec
d17          0.00019600 sec
MCREST      0.00000000 sec
MCRMK       0.01500000 sec
F2           31.00 usec

===== CHANNEL f1 =====
NUC1         13C
P1           15.50 usec
P11          500.00 usec
P12          2000.00 usec
PL0          120.00 dB
PL1          -1.00 dB
SFO1        125.7942548 MHz
SP1          3.20 dB
SP2          3.20 dB
SFOAM1      Crp60,0.5,20.1
SFOAM2      Crp60comp,4
SFOFF1      0.00 Hz
SFOFF2      0.00 Hz

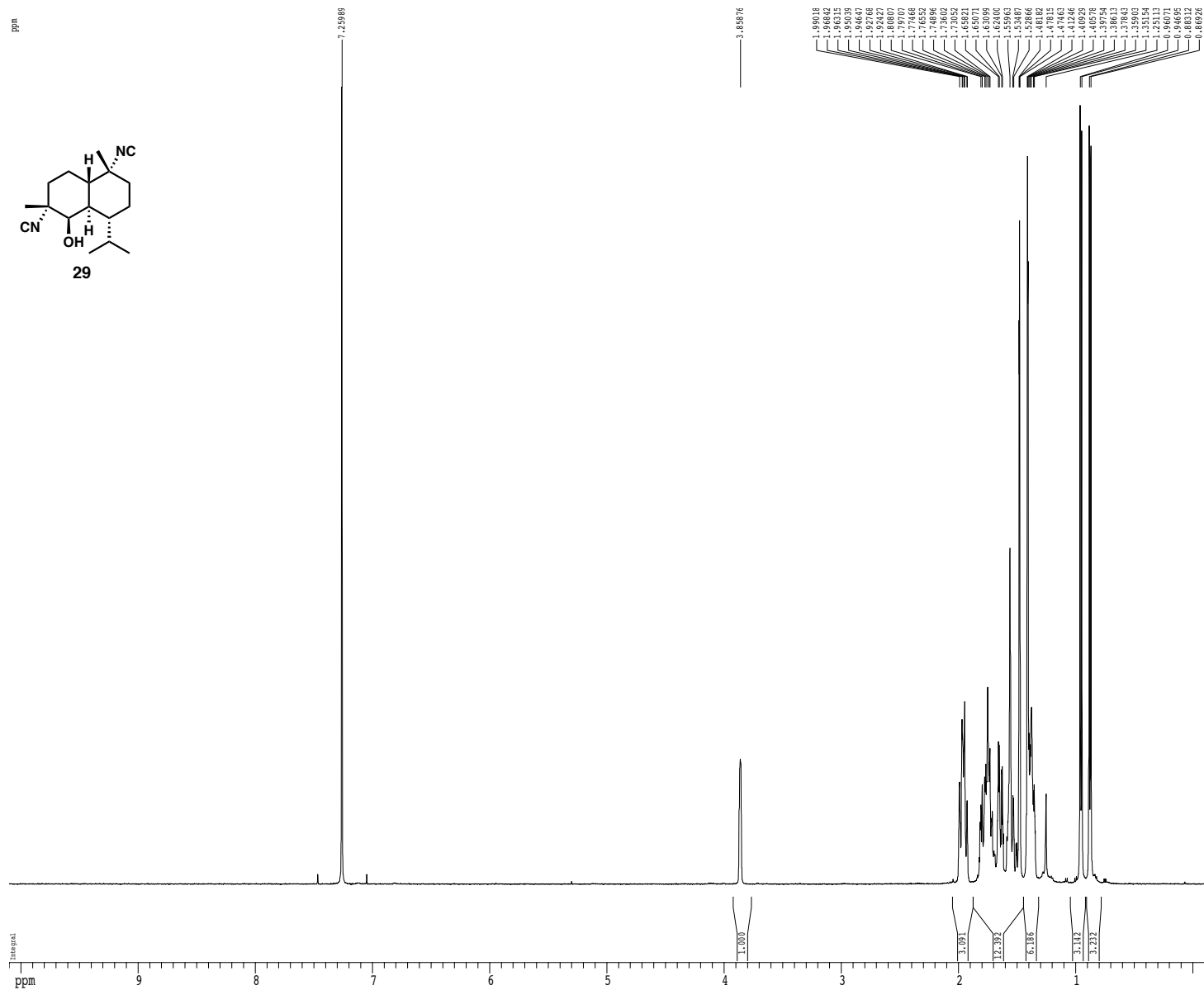
===== CHANNEL f2 =====
CPDPRG2     waltz16
NUC2         1H
PCPD2       100.00 usec
PL2         1.60 dB
PL12        24.60 dB
SFO2        500.2225011 MHz

===== GRADIENT CHANNEL =====
GPMAM1      SINE,100
GPMAM2      SINE,100
GPX1        0.00 %
GPX2        0.00 %
GPI1        0.00 %
GPI2        0.00 %
GPI3        30.00 %
GPI4        50.00 %
p15         500.00 usec
p16         1000.00 usec

F2 - Processing parameters
SI           65536
SF           125.7804076 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           2.00

1D NMR plot parameters
CY           22.80 cm
CY           75.00 cm
F1           200.000 ppm
F1           25156.08 Hz
F2P         -10.000 ppm
F2          -1257.80 Hz
FPMCH       9.21053 ppm/cm
H2CH        1158.50378 Hz/cm
    
```

¹H spectrum



Current Data Parameters
 USER medaub
 NAME MED-VI-298Pu
 EXNO 1
 PROCNO 1

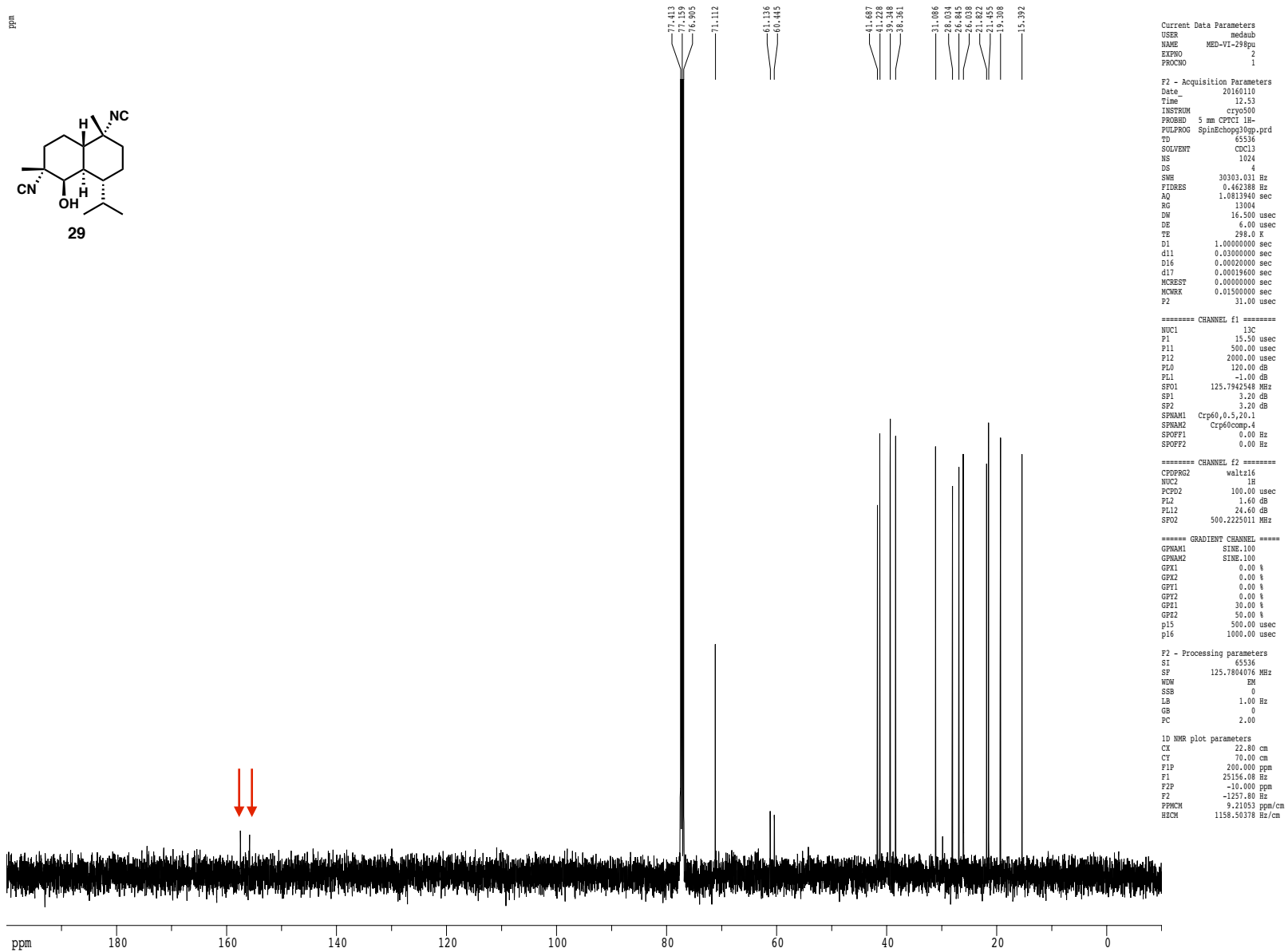
F2 - Acquisition Parameters
 Date_ 20160110
 Time 12.50
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.620 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 9
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 7.50 usec
 PL1 1.60 dB
 SF01 500.2235015 MHz

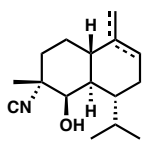
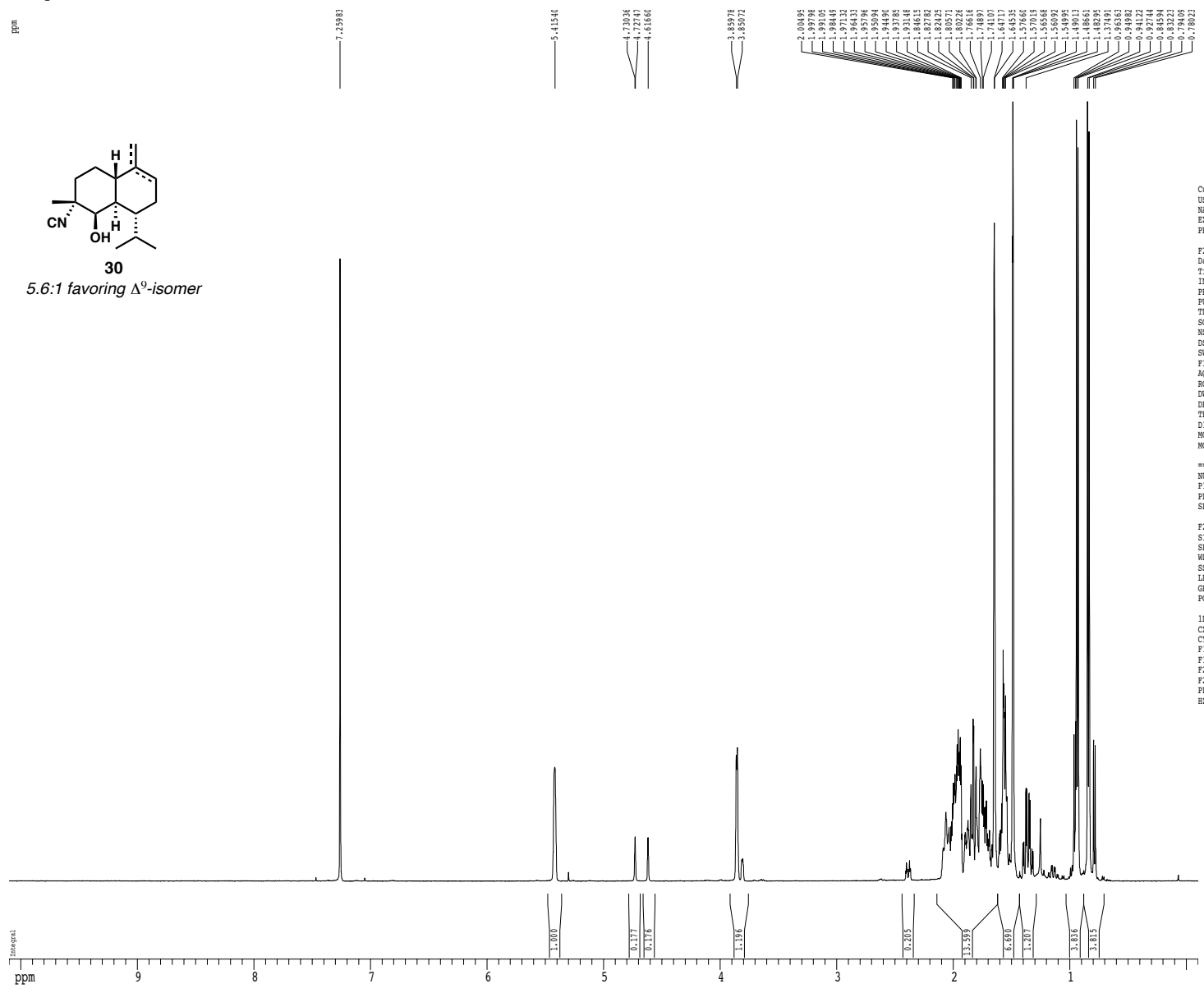
F2 - Processing parameters
 SI 65536
 SF 500.2200306 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.80 cm
 CY 35.00 cm
 P1P 10.100 ppm
 F1 5052.22 Hz
 F2P -0.100 ppm
 F2 -50.02 Hz
 FFCM 0.44737 ppm/cm
 HZCM 223.78267 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum



30
5.6:1 favoring Δ^9 -isomer

```

Current Data Parameters
USER      medaub
NAME      MED-VI-300pu
EXPNO     1
PROCNO    1

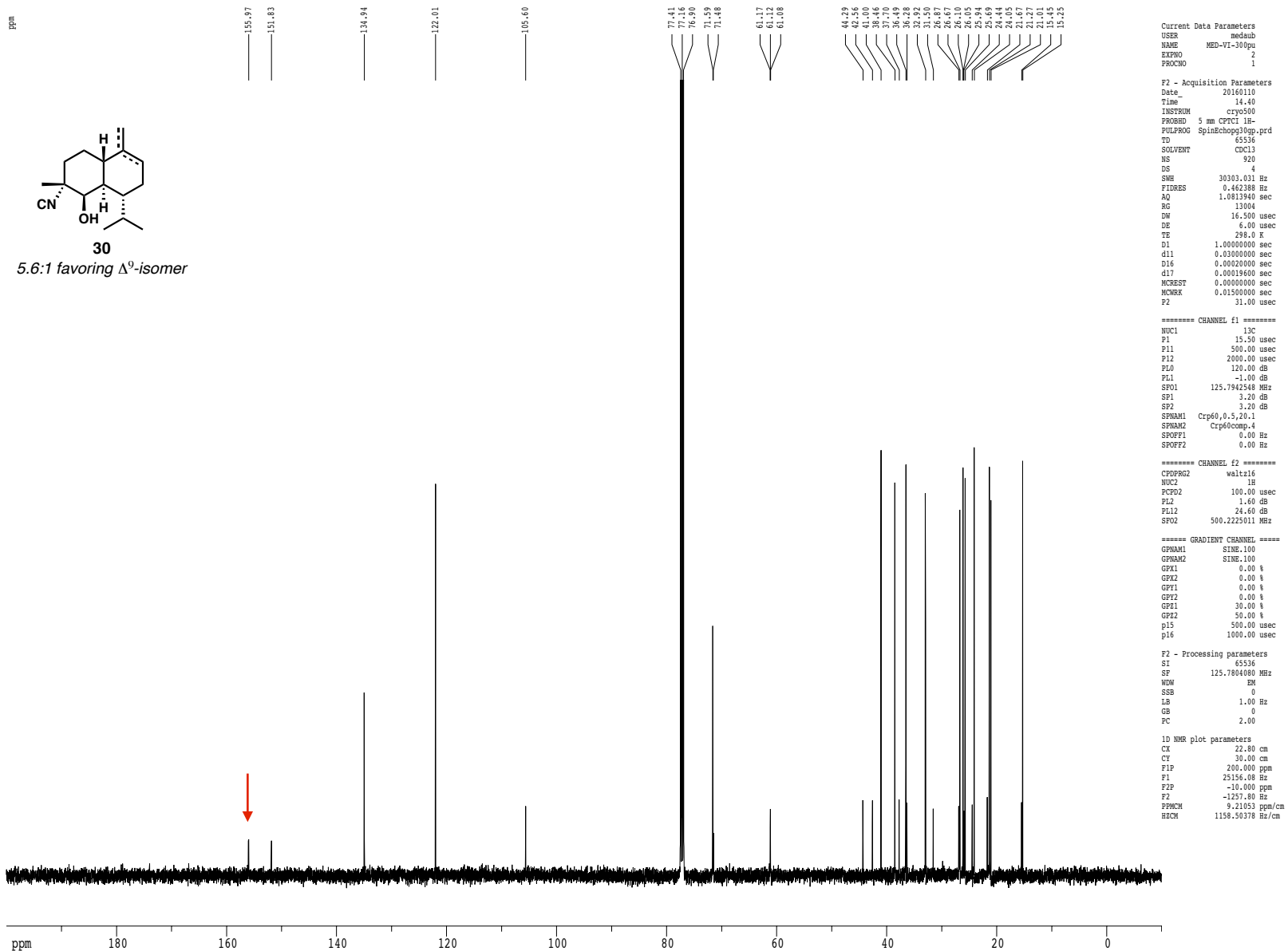
F2 - Acquisition Parameters
Date_     20160110
Time      14.07
INSTRUM   cryo500
PROBHD    5 mm CPCL1 H-
PULPROG   zg30
TD         81728
SOLVENT   CDCl3
NS         8
DS         2
SWH        8012.820 Hz
FIDRES     0.098043 Hz
AQ         5.0998774 sec
RG         8
DW         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
MCREST    0.00000000 sec
MCMRFX    0.01500000 sec

===== CHANNEL f1 =====
NUC1       1H
P1         7.50 usec
PL1        1.60 dB
SFO1       500.2235015 MHz

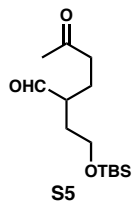
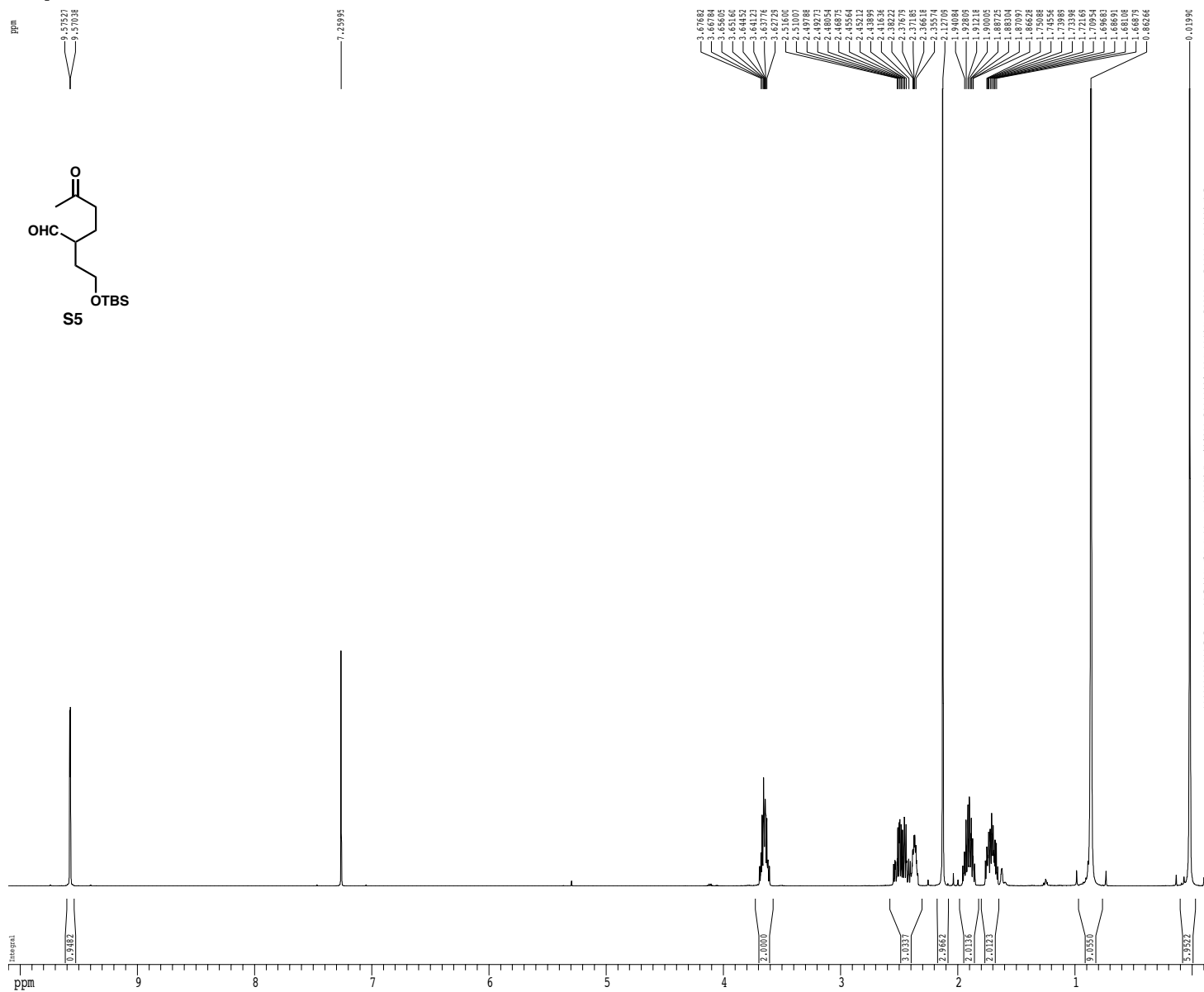
F2 - Processing parameters
SI         65536
SF         500.2200307 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00

ID NMR plot parameters
CX         22.80 cm
CY         15.00 cm
F1P        10.100 ppm
F1         5052.22 Hz
F2P        -0.100 ppm
F2         -50.02 Hz
PPMCM     0.44737 ppm/cm
HZCM      223.78267 Hz/cm
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum



Current Data Parameters
 USER medaub
 NAME MED-VI-287pu
 EXPNO 1
 PROCNO 1

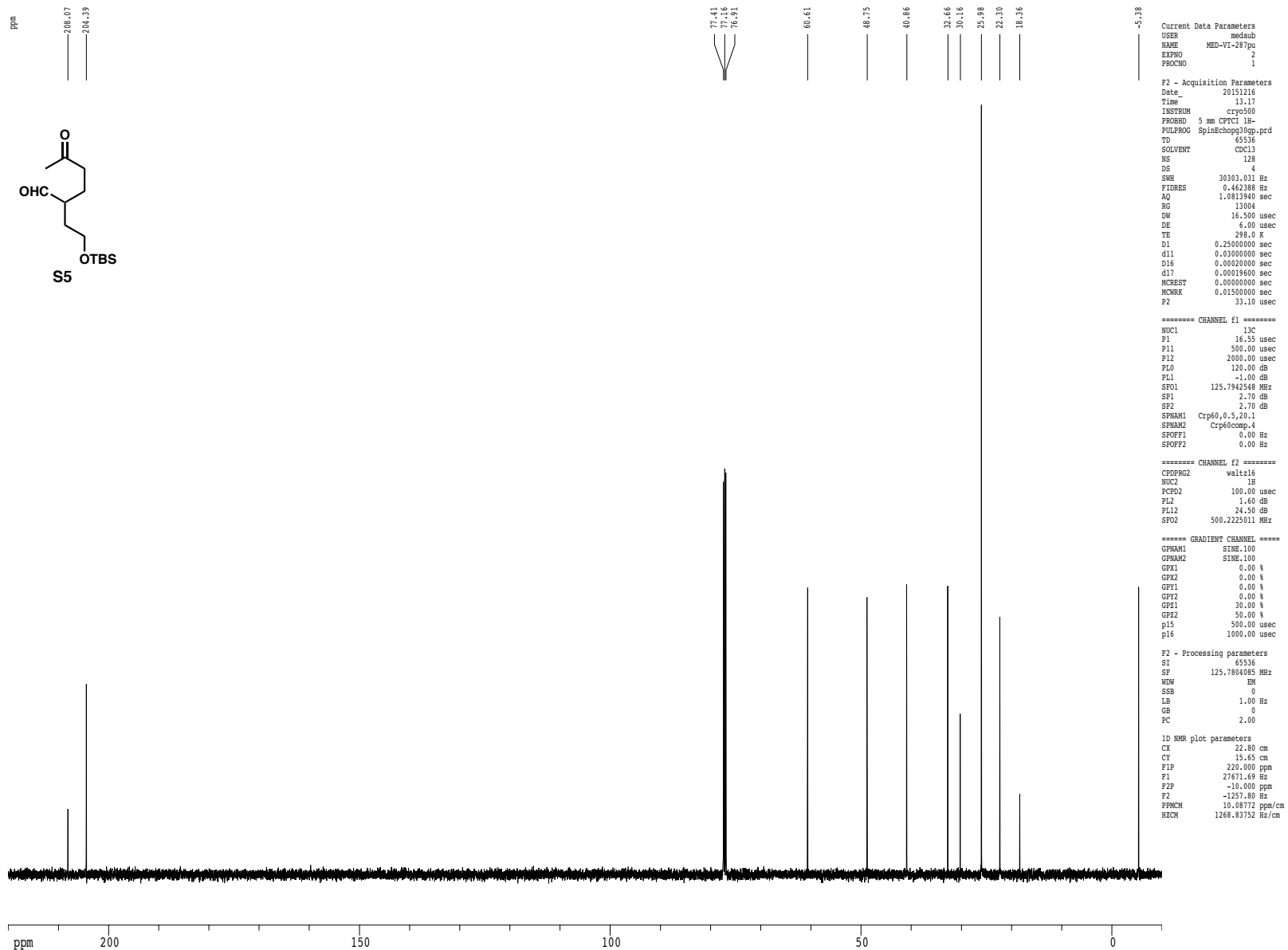
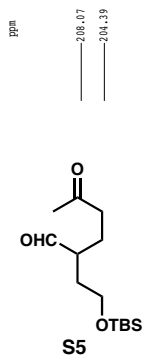
F2 - Acquisition Parameters
 Date_ 20151216
 Time 13.14
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3
 NS 8
 DS 2
 SHW 8012.920 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 7.1
 DW 62.400 usec
 DE 4.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.60 db
 SF01 500.2235015 MHz

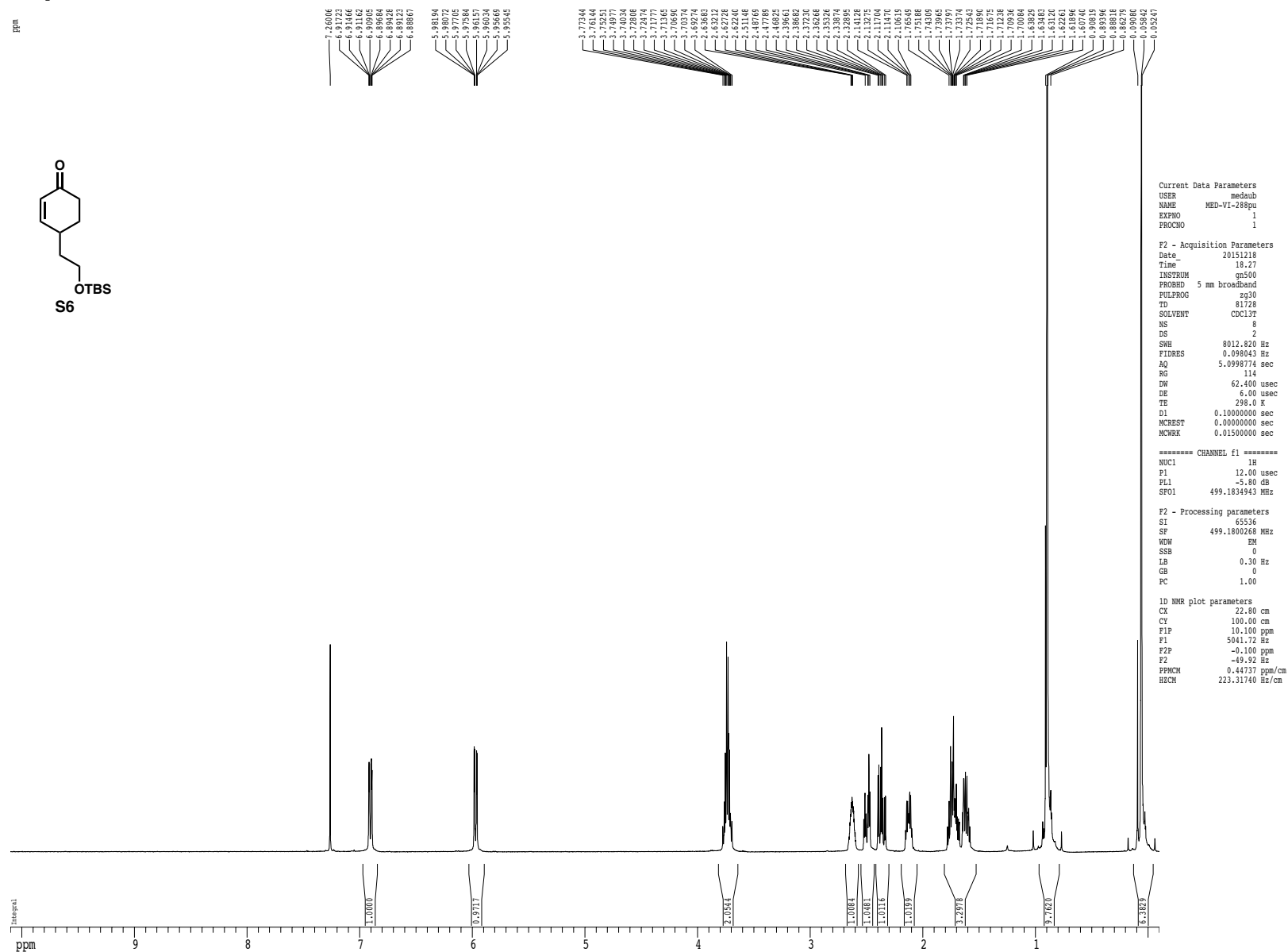
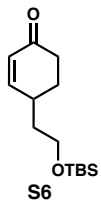
F2 - Processing parameters
 SI 65536
 SF 500.2209317 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.80 cm
 CY 40.00 cm
 F1P 10.100 ppm
 F1 5052.22 Hz
 F2P -0.100 ppm
 F2 -50.00 Hz
 FPMCM 0.44737 ppm/cm
 HCM 223.78267 Hz/cm

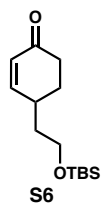
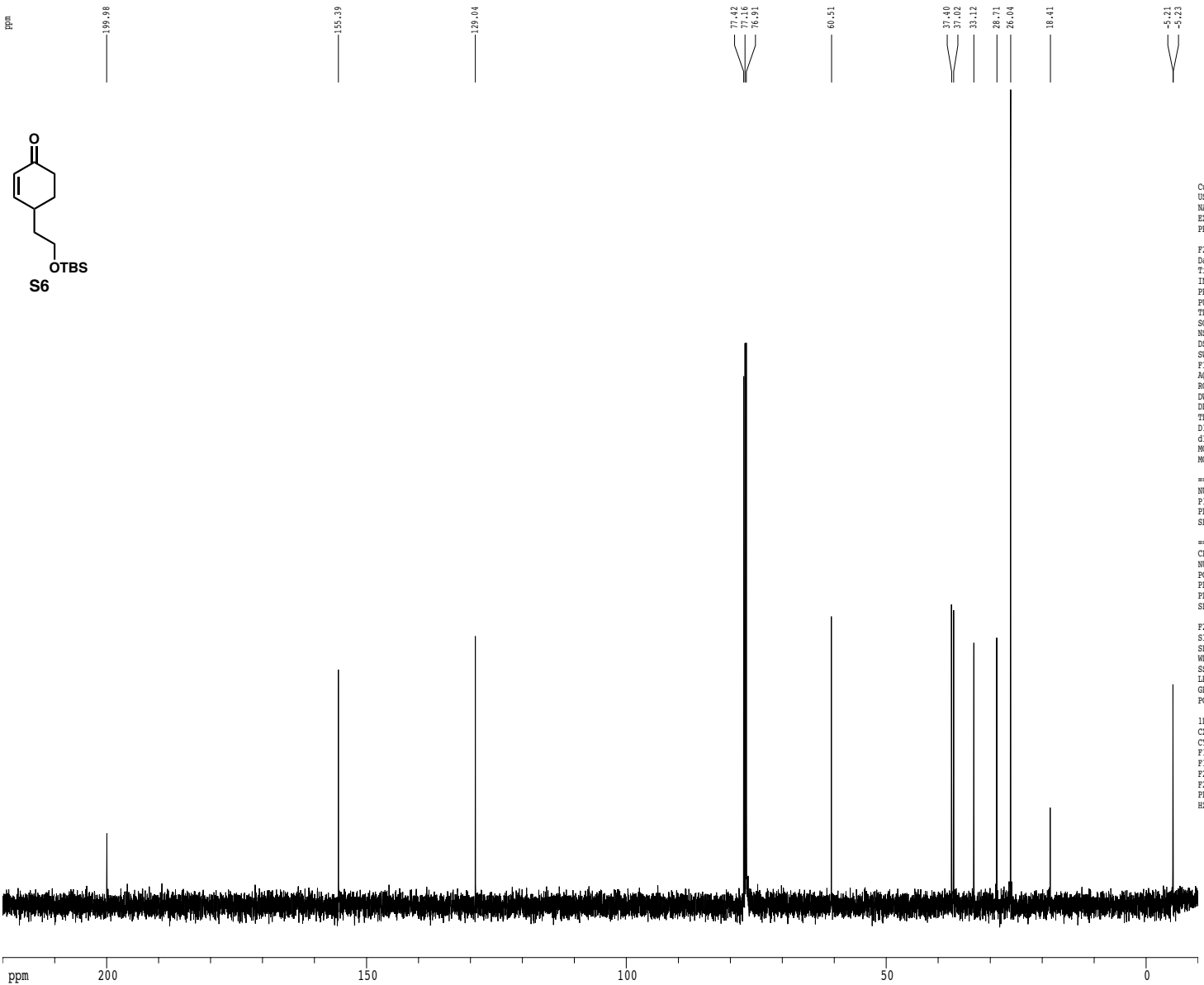
Z-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum



13C spectrum with 1H decoupling



```

Current Data Parameters
USER      medaub
NAME      MED-VI-288pu
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20151218
Time      18.30
INSTRUM   qm500
PROBHD    5 mm broadband
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         120
DS         4
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         46341
DM         16.500 usec
DE         4.50 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
MCREST     0.00000000 sec
MCWRK      0.01500000 sec

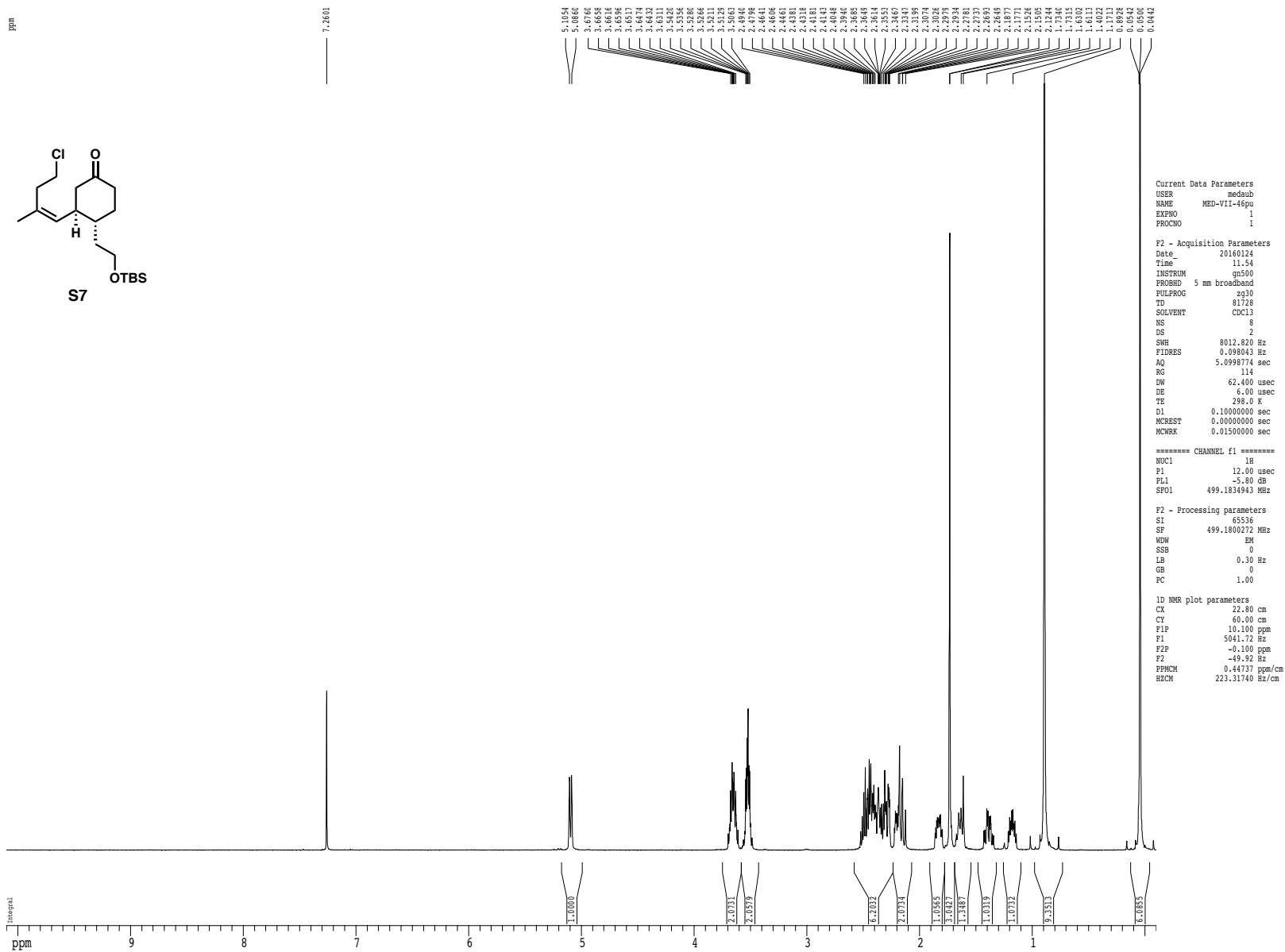
***** CHANNEL f1 *****
NUC1       13C
P1         9.00 usec
PL1        -0.60 dB
SFO1       125.5327181 MHz

***** CHANNEL f2 *****
CPDPRG2    waltz16
NUC2       1H
PCPD2      80.00 usec
PL2        -3.00 dB
PL12       12.80 dB
SFO2       499.1824959 MHz

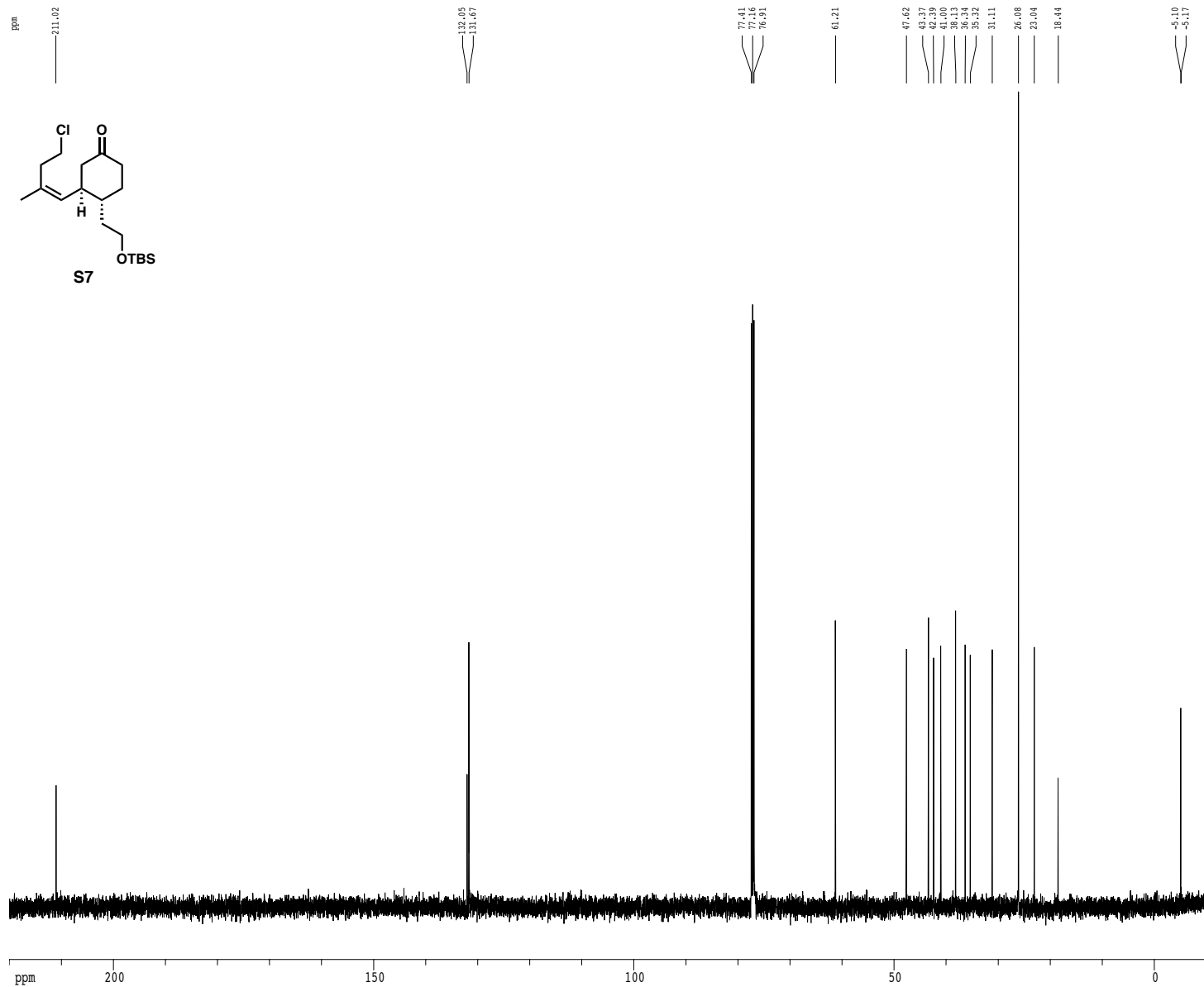
F2 - Processing parameters
SI         65536
SF         125.5188979 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

1D NMR plot parameters
CX         22.80 cm
CY         15.65 cm
F1P        220.000 ppm
F1         27614.16 Hz
F2P        -10.000 ppm
F2         -1255.19 Hz
PPMCM      10.0872 ppm/cm
HZCM       1266.19946 Hz/cm
    
```

1H spectrum



13C spectrum with 1H decoupling



```

Current Data Parameters
USER          medaub
NAME          MED-VII-46pu
EXPNO         2
PROCNO        1

F2 - Acquisition Parameters
Date_         20160124
Time          11:57
INSTRUM       gm500
PROBHD        5 mm broadband
PULPROG       zgpg30
TD            65536
SOLVENT       CDCl3
NS            200
DS            4
SWH           30303.031 Hz
FIDRES        0.462388 Hz
AQ            1.0813940 sec
RG            46341
DN            16.500 usec
DE            4.500 usec
TE            298.0 K
D1            0.25000000 sec
d11           0.03000000 sec
MCREST        0.00000000 sec
MCWRK         0.01500000 sec

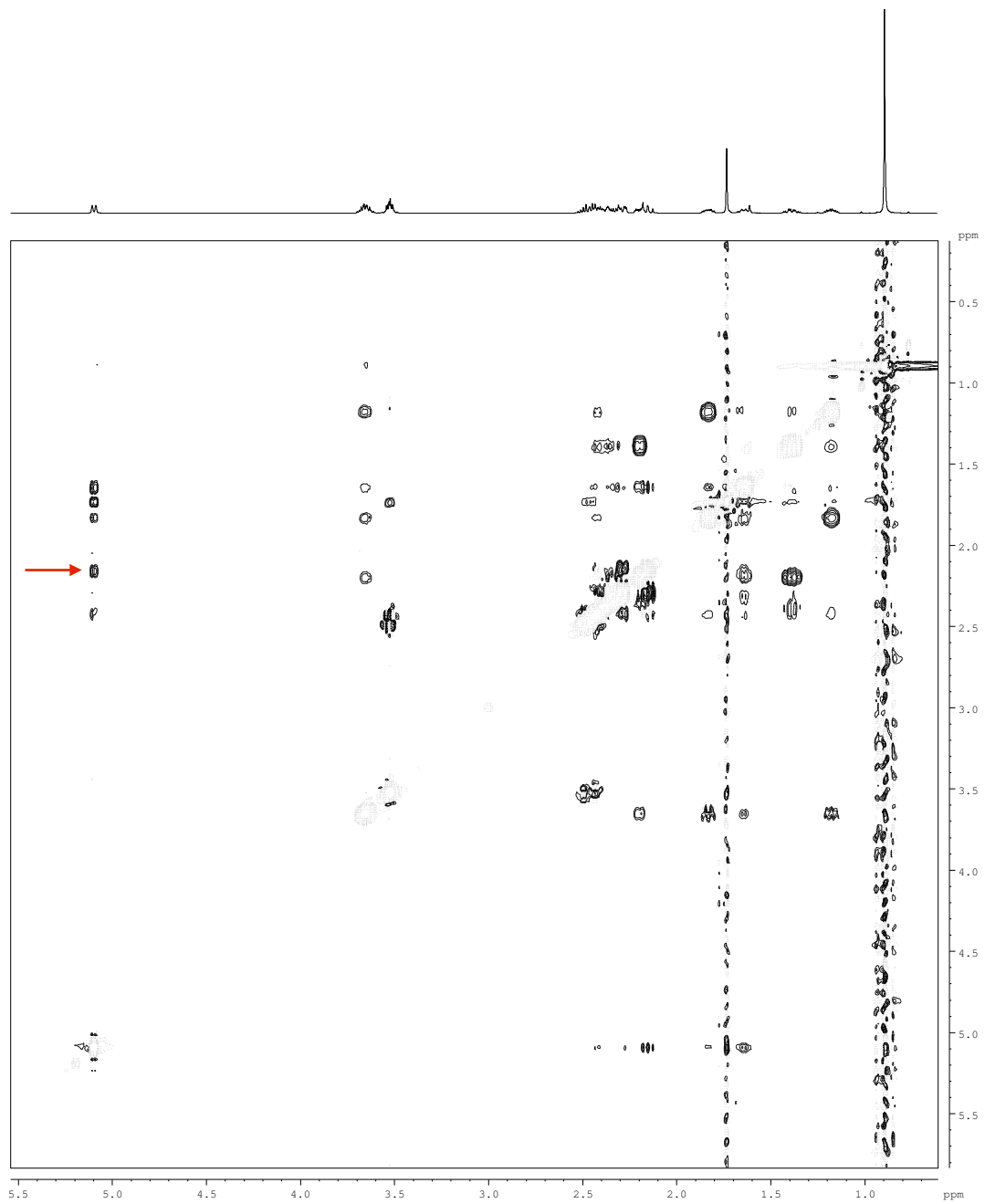
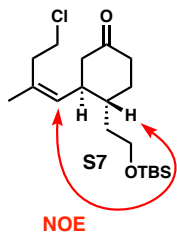
***** CHANNEL f1 *****
NUC1           13C
P1             9.00 usec
PL1           -0.60 dB
SFO1          125.5327181 MHz

***** CHANNEL f2 *****
CPDPRG2       waltz16
NUC2           1H
PCPD2         80.00 usec
PL2           -3.00 dB
PL12          12.80 dB
SFO2          499.1824959 MHz

F2 - Processing parameters
SI            65536
SF            125.5188984 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            2.00

1D NMR plot parameters
CX            22.80 cm
CY            15.65 cm
P1P          220.000 ppm
F1           27614.16 Hz
F2P          -10.000 ppm
F2           -1255.19 Hz
PPMCM        10.08772 ppm/cm
HZCM         1266.19946 Hz/cm
    
```


noesygtp



```
Current Data Parameters
NAME      MED-VII-46pu
EXPNO     4
PROCNO    1

F2 - Acquisition Parameters
Date_     20160124
Time      12.04
INSTRUM   gm400
PROBHD    5 mm broadband
PULPROG   noesygtp
TD         262144
SOLVENT   CDCl3
NS         2
DS         4
SWH        4496.403 Hz
FIDRES     2.195509 Hz
AQ         0.2277376 sec
RG         1290.2
DW         111.200 usec
DE         6.00 usec
TE         298.0 K
DO         0.0000300 sec
D1         2.0000000 sec
D8         1.0000000 sec
D16        0.0002500 sec
d20        0.4987500 sec
IN0        0.0001120 sec

----- CHANNEL f1 -----
NUC1       1H
P1         12.00 usec
P2         24.00 usec
PL1        -5.80 dB
SF01       499.1821199 MHz

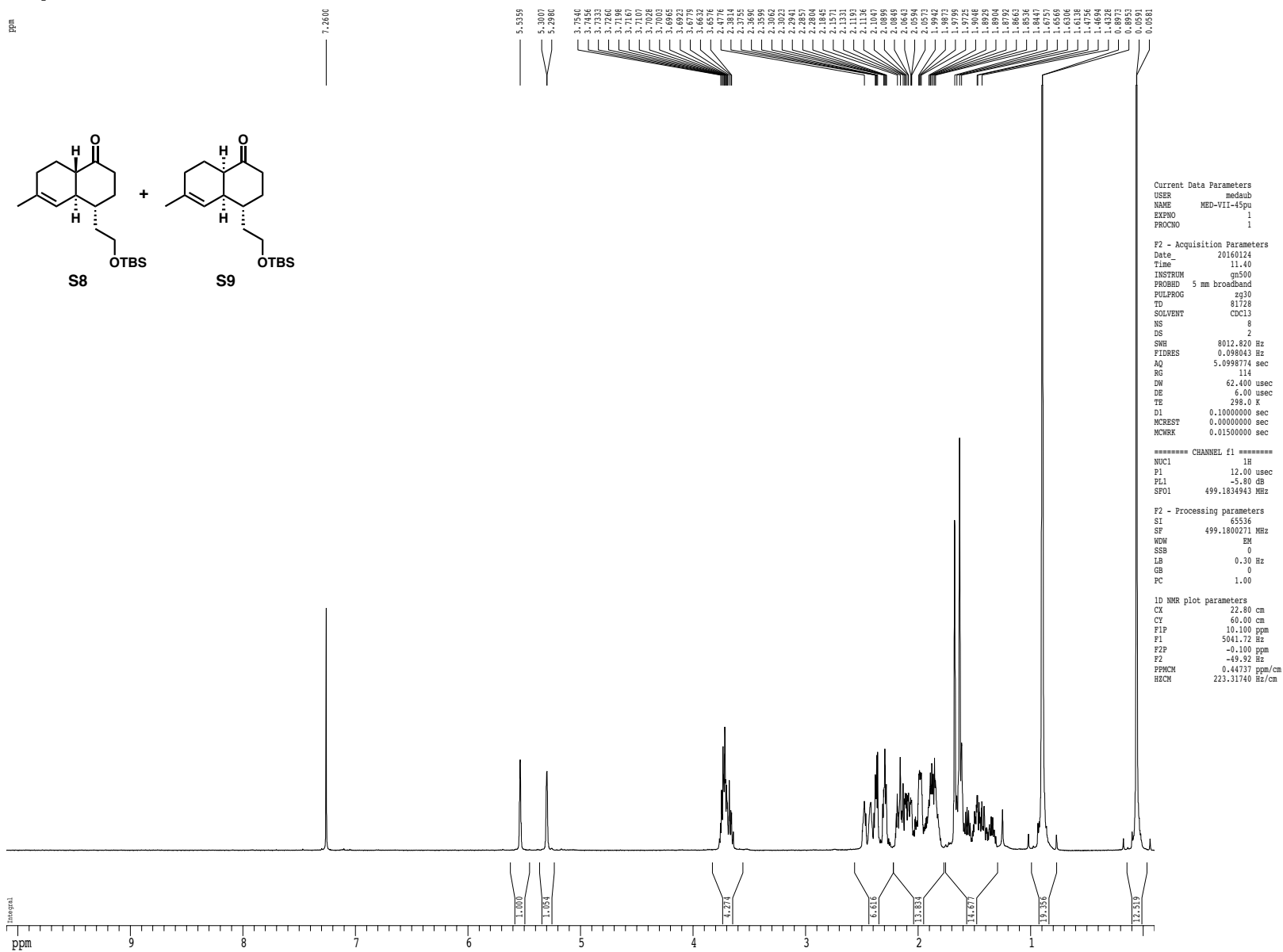
----- GRADIENT CHANNEL -----
GFNAM[1]   sine.100
GFNAM[2]   sine.100
GPX1       0 %
GPX2       0 %
GPY1       0 %
GPY2       0 %
GPZ1       40.00 %
GPZ2       -40.00 %
P16        1000.00 usec

F1 - Acquisition parameters
TD         256
SF01       499.1821 MHz
FIDRES     17.564074 Hz
SW         9.008 ppm
FAMODE     TPF1

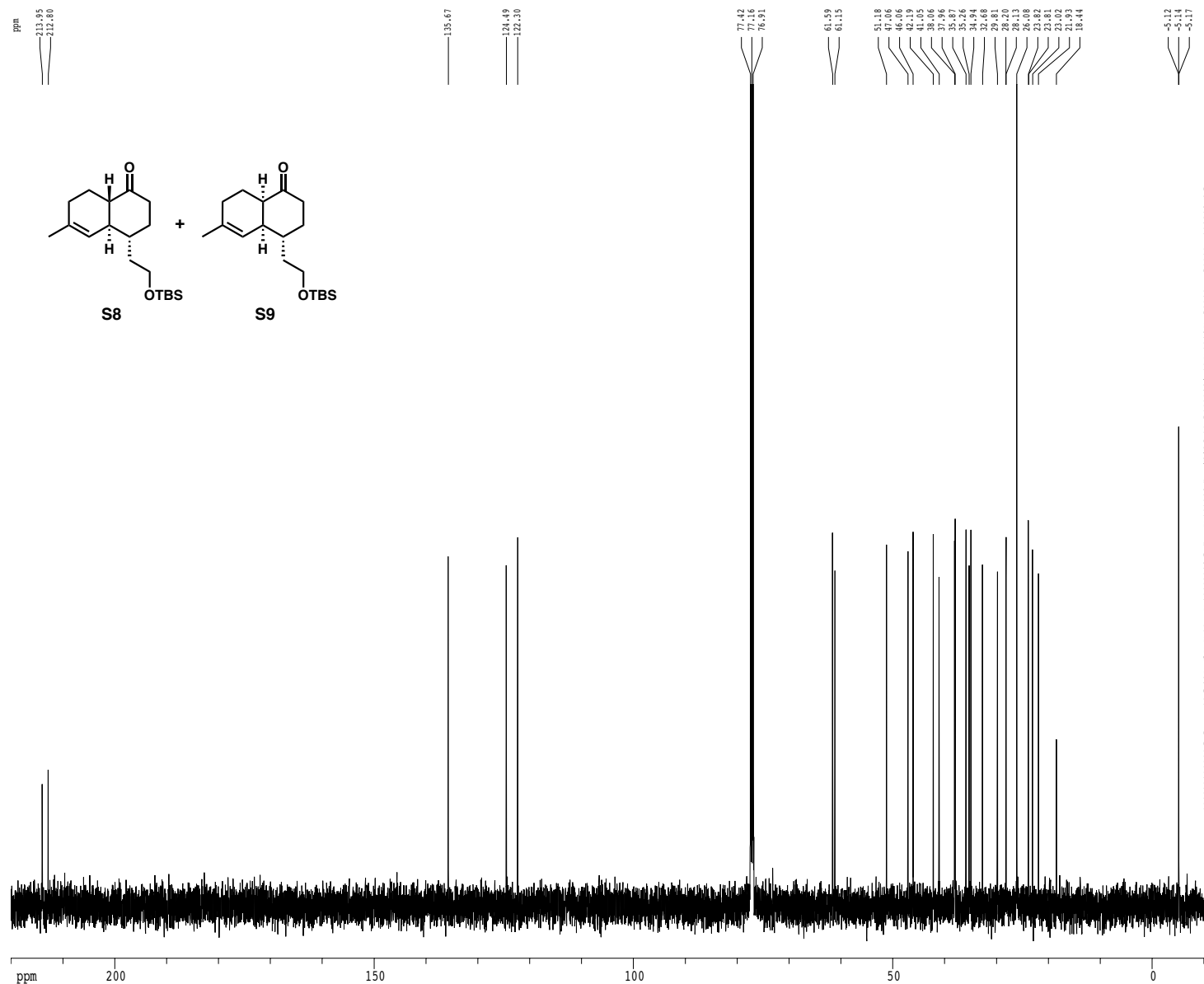
F2 - Processing parameters
SI         1024
SF         499.1800272 MHz
WDW        QSINE
SSB        2
LB         0 Hz
GB         0
PC         1.40

F1 - Processing parameters
SI         1024
MC2        TPF1
SF         499.1800272 MHz
WDW        QSINE
SSB        2
LB         0 Hz
GB         0
```

1H spectrum



13C spectrum with 1H decoupling



```

Current Data Parameters
USER      medaub
NAME      MED-VII-45pu
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20160124
Time      11.41
INSTRUM   gm500
PROBHD    5 mm broadband
PULPROG   zgdc30
TD         65536
SOLVENT   CDCl3
NS         408
DS         4
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         46241
DW         16.500 usec
DE         4.50 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
MCREST     0.00000000 sec
MCMRXX     0.01500000 sec

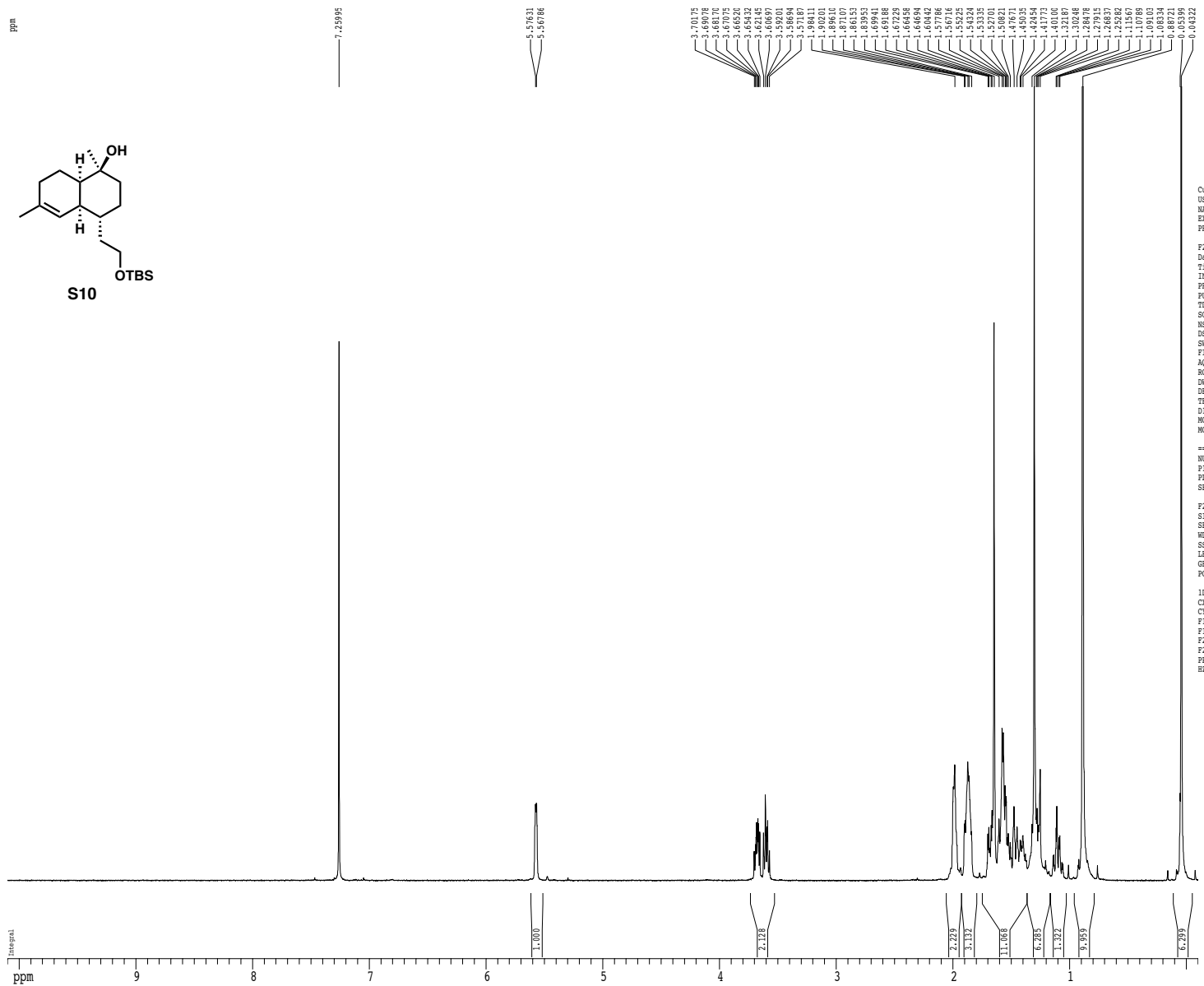
===== CHANNEL f1 =====
NUC1       13C
P1         9.00 usec
PL1        -0.60 dB
SFO1       125.5327181 MHz

===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2      80.00 usec
FL2        -3.00 dB
PL12       12.80 dB
SFO2       499.1824959 MHz

F2 - Processing parameters
SI         65536
SF         125.5188976 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

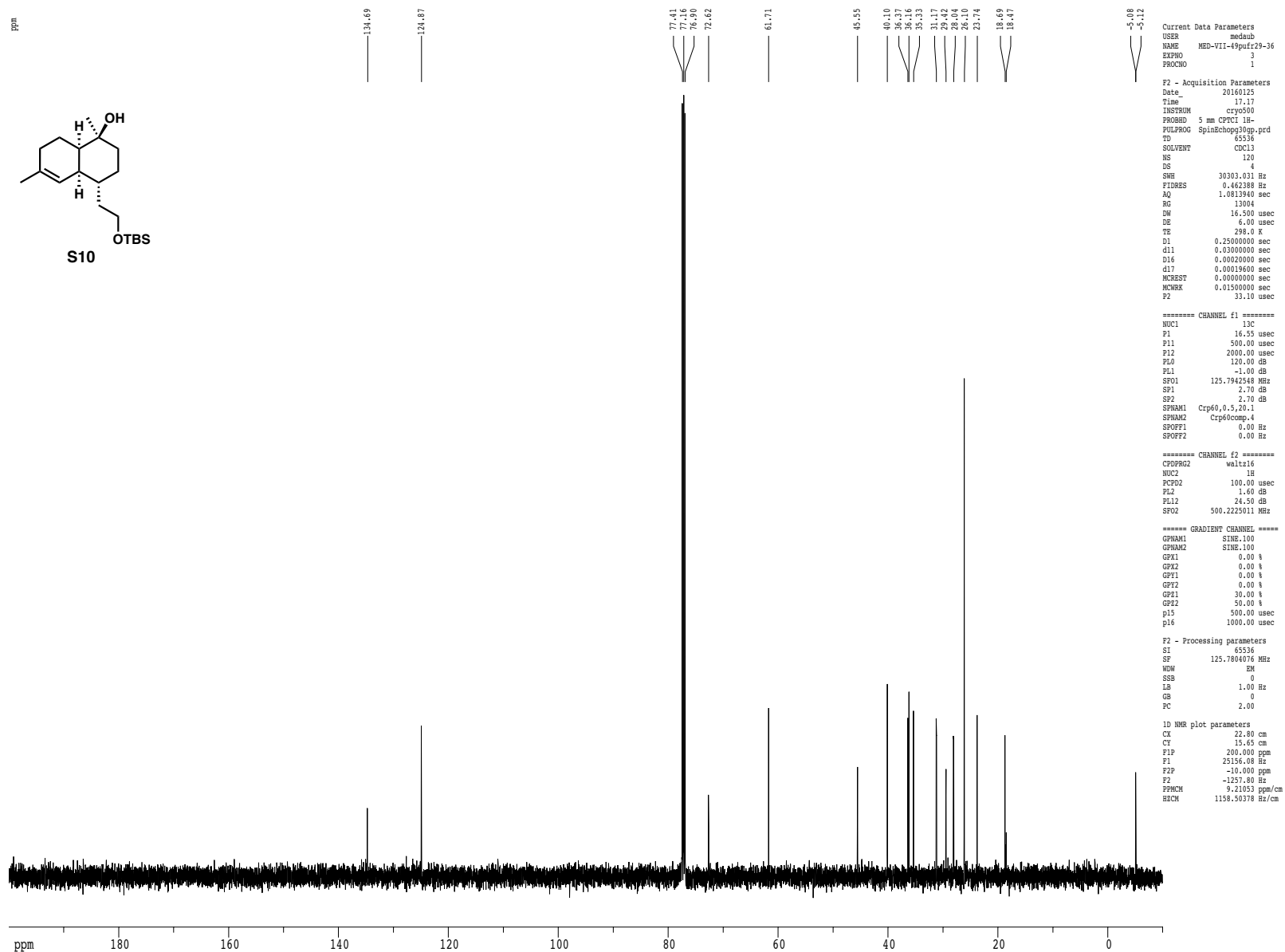
1D NMR plot parameters
CX         22.80 cm
CY         30.00 cm
FIP        220.000 ppm
F1         27614.16 Hz
F2P        -10.000 ppm
F2         -125.19 Hz
F2MCM      10.08712 ppm/cm
H2CM       1266.19946 Hz/cm
    
```

1H spectrum

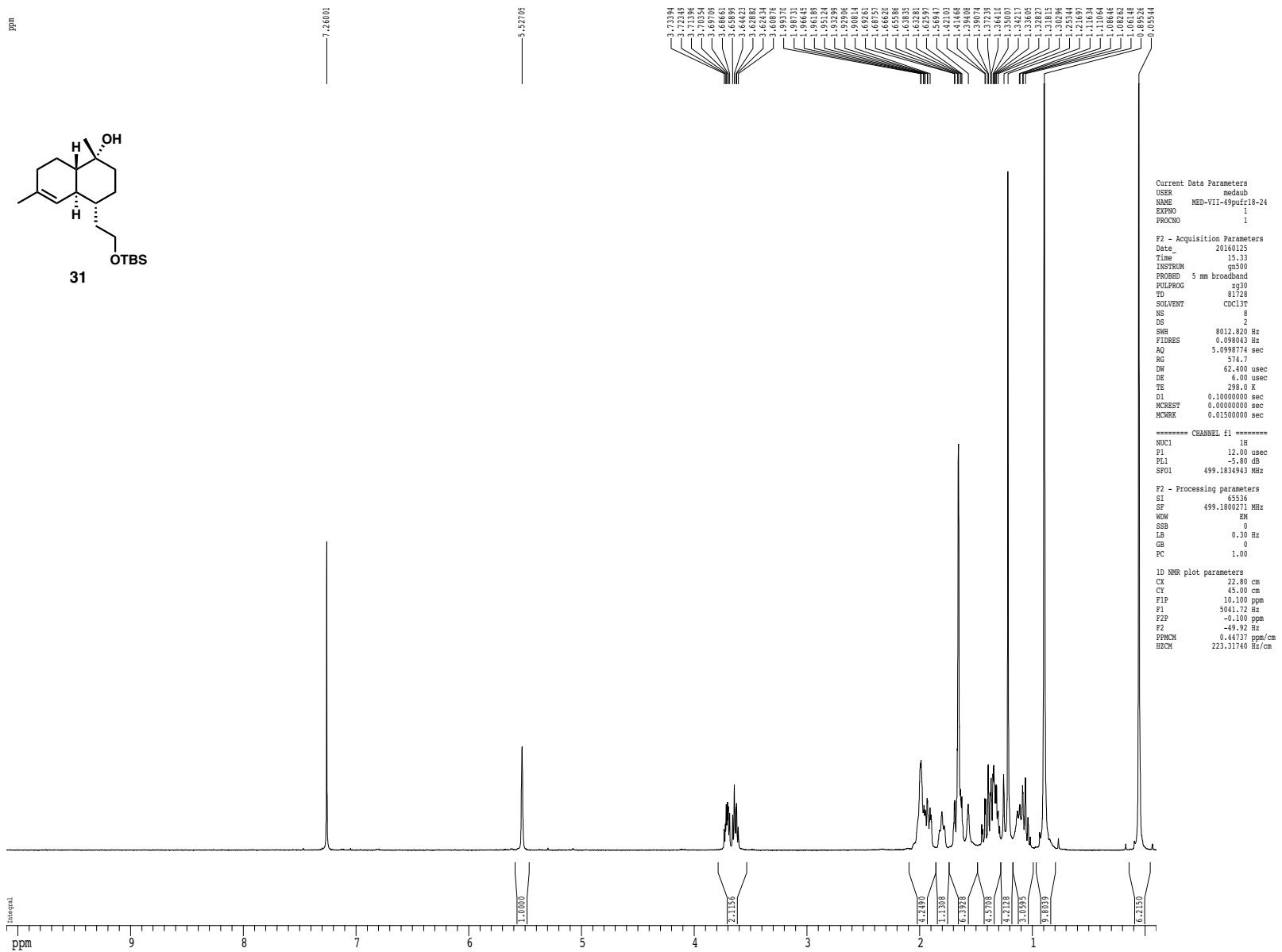


Current Data Parameters
USER medaub
NAME MED-VII-49pufz29-36
EXPNO 1
PROCNO 1
F2 - Acquisition Parameters
Date_ 20161225
Time 15.36
INSTRUM gn500
PROBHD 5 mm broadband
PULPROG zg30
TD 81728
SOLVENT CDCl3T
NS 8
DS 2
SWH 8012.820 Hz
FIDRES 0.098043 Hz
AQ 5.0998774 sec
RG 724.1
DW 62.400 usec
DE 6.00 usec
TE 298.0 K
D1 0.10000000 sec
MCREST 0.00000000 sec
MCWEX 0.01500000 sec
===== CHANNEL f1 =====
NUC1 1H
P1 12.00 usec
PL1 -5.80 dB
SFO1 499.1834943 MHz
F2 - Processing parameters
SI 65536
SF 499.1800272 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00
ID NMR plot parameters
CX 22.80 cm
CY 60.00 cm
FIP 10.100 ppm
F1 5041.72 Hz
F2P -0.100 ppm
F2 -49.50 Hz
PPMCM 0.44737 ppm/cm
HZCM 223.31740 Hz/cm

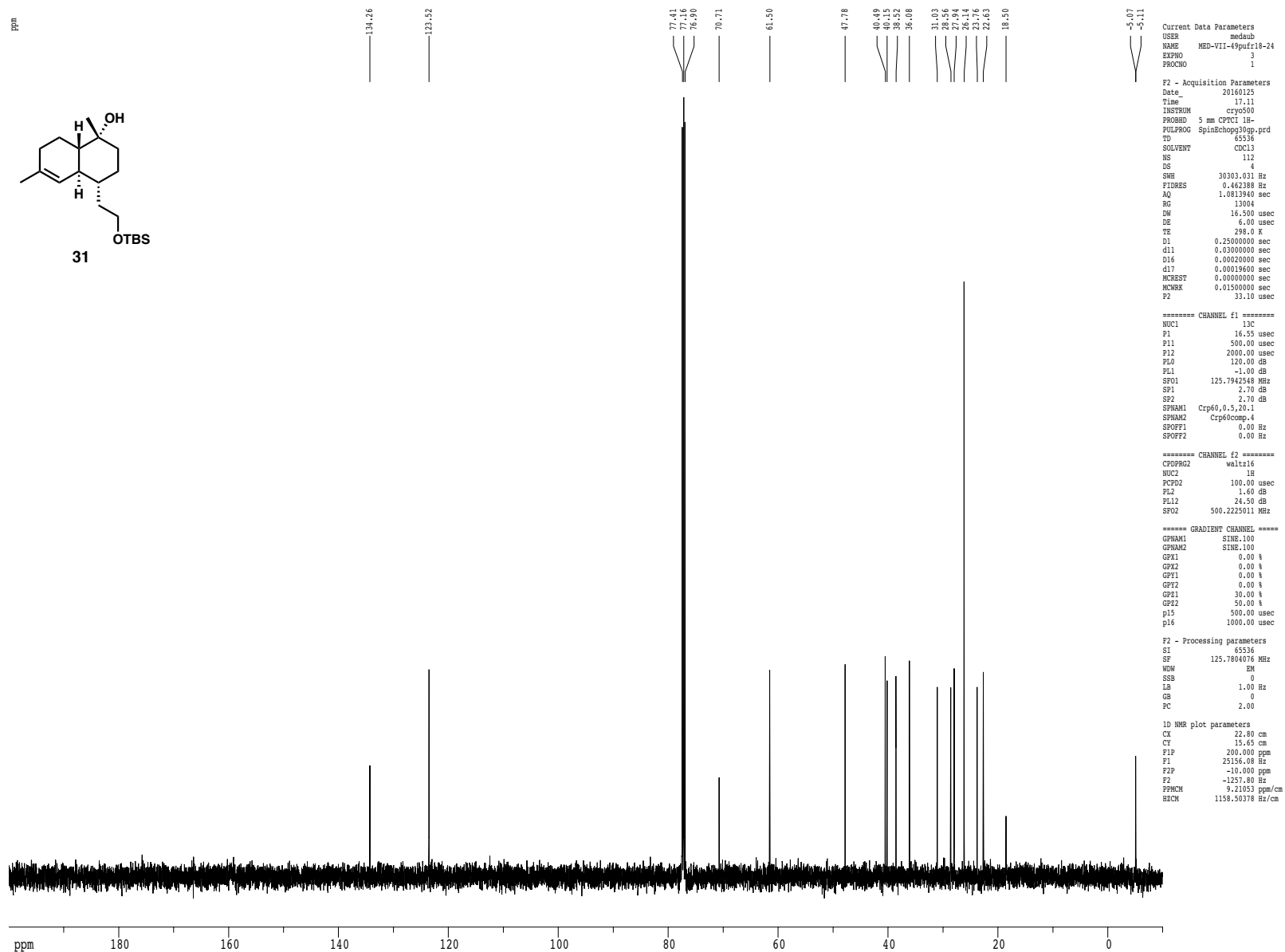
Z-restored spin-echo 13C spectrum with 1H decoupling



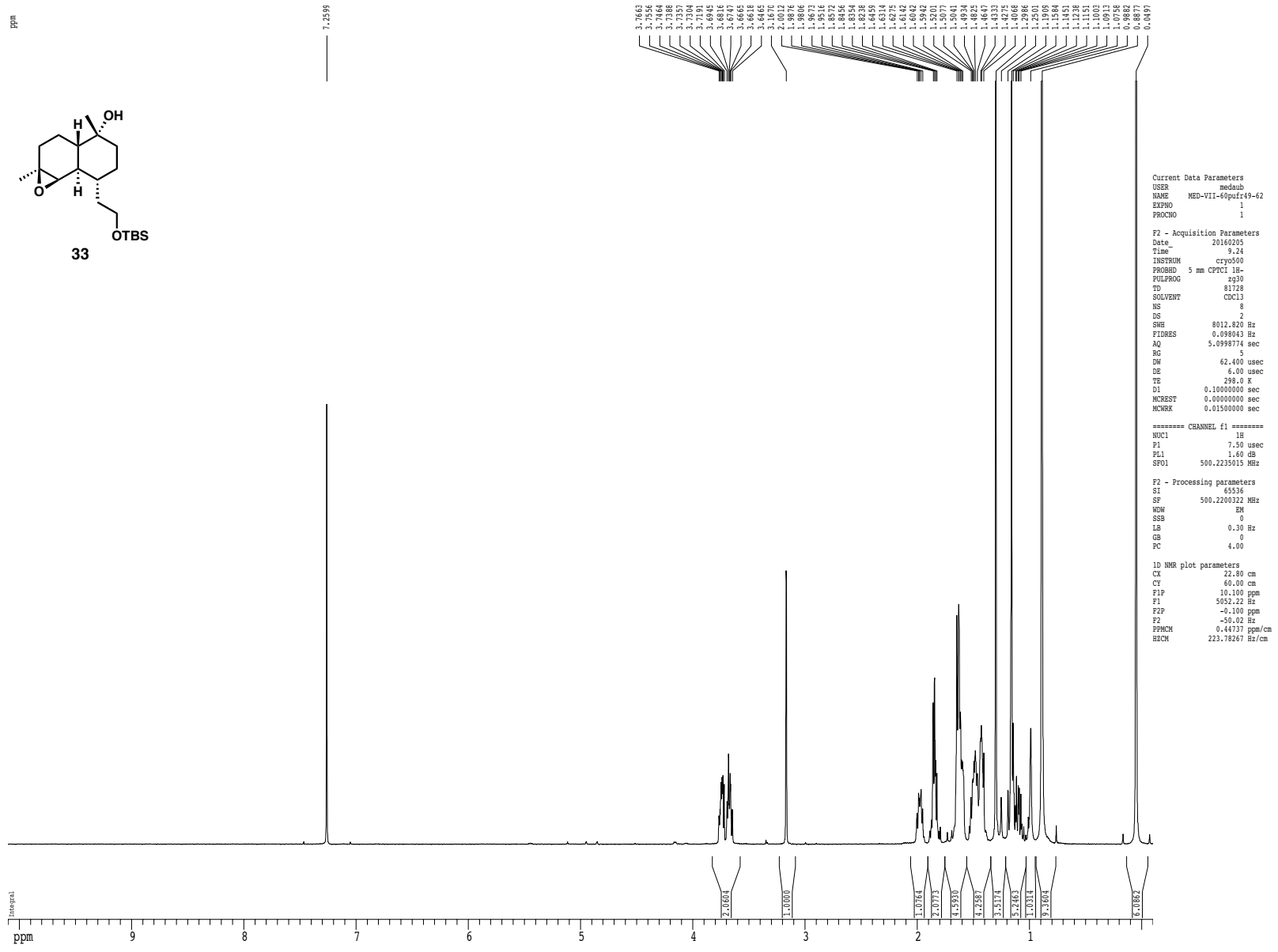
1H spectrum



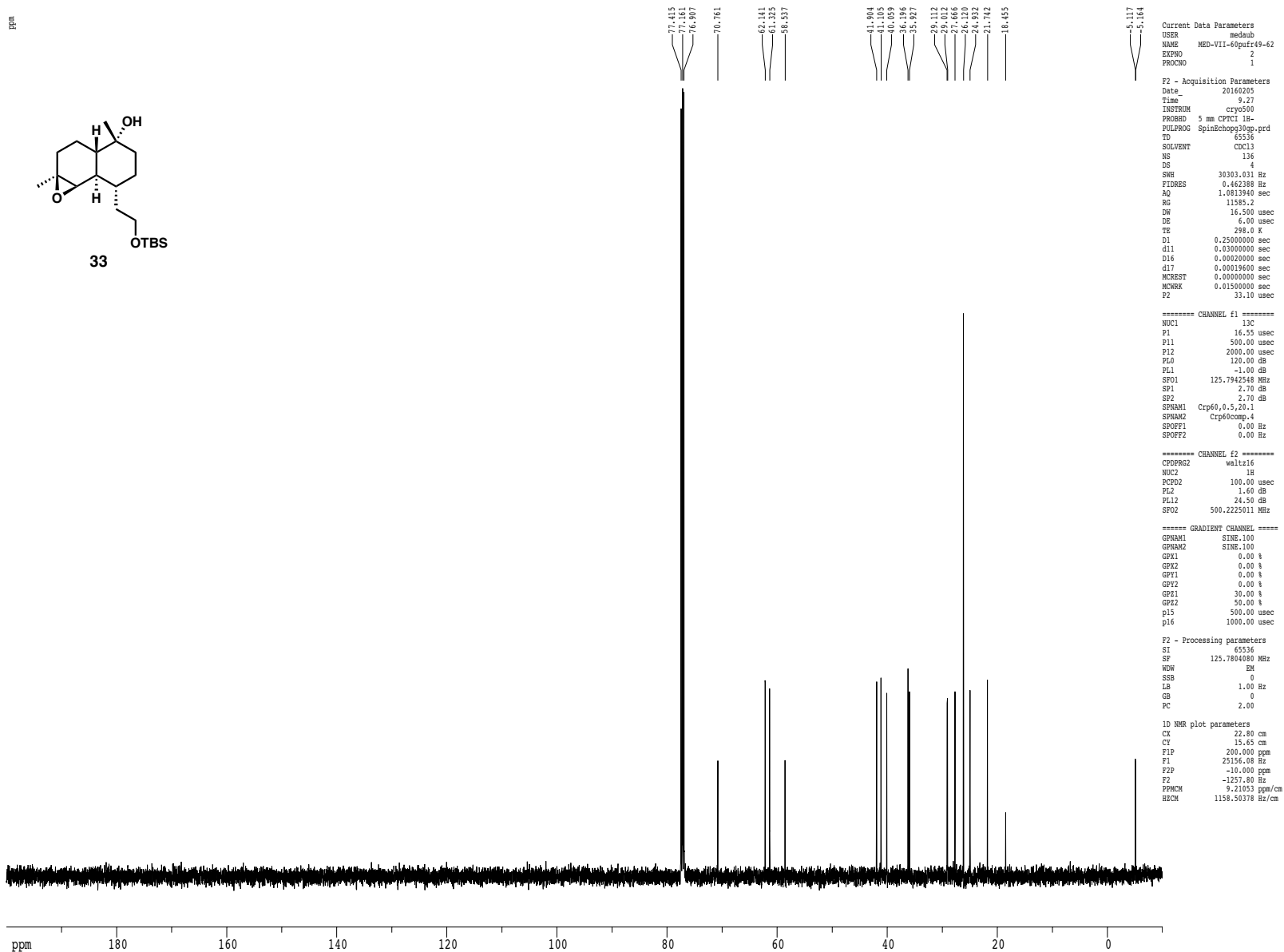
Z-restored spin-echo 13C spectrum with 1H decoupling



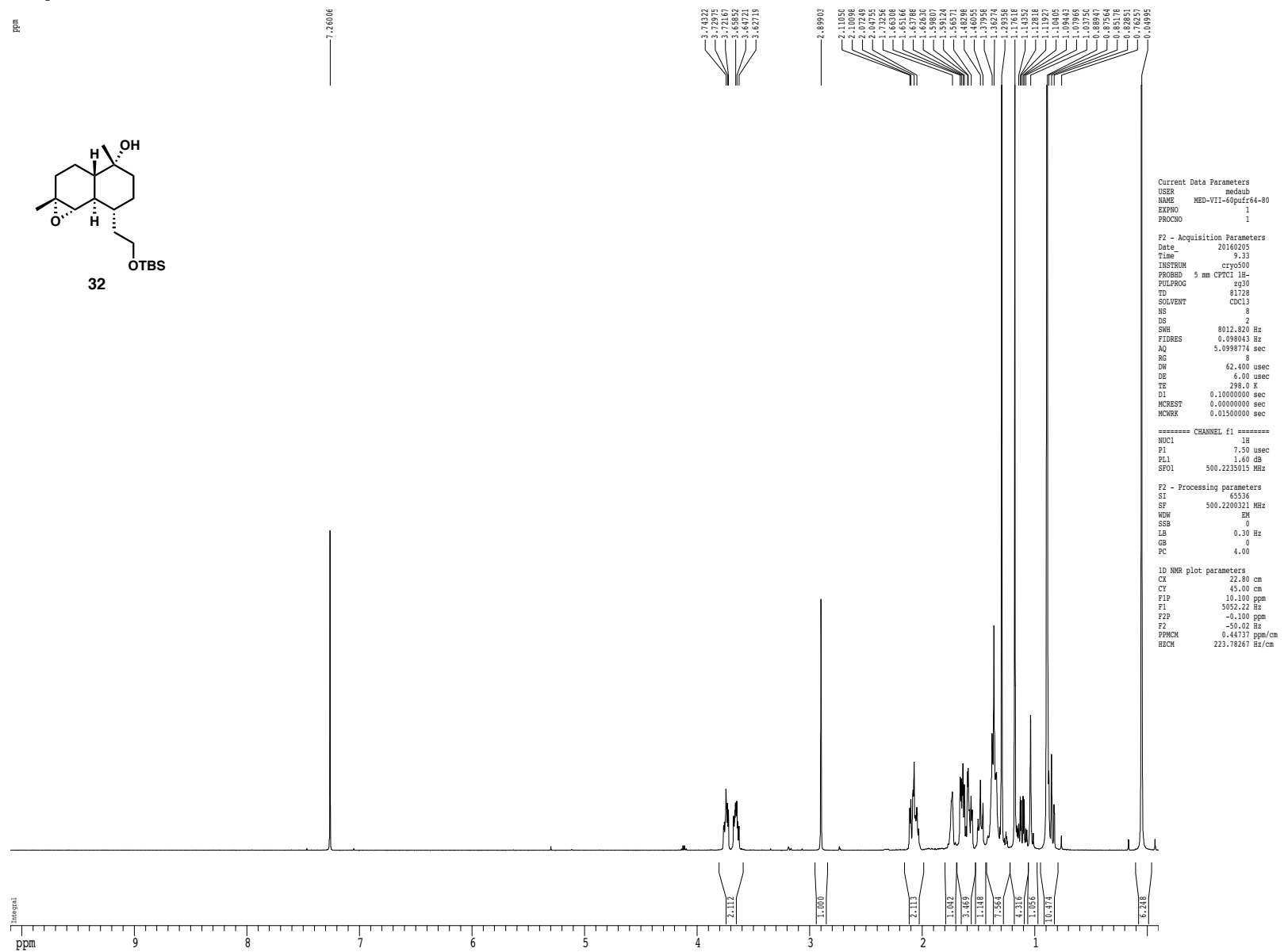
1H spectrum



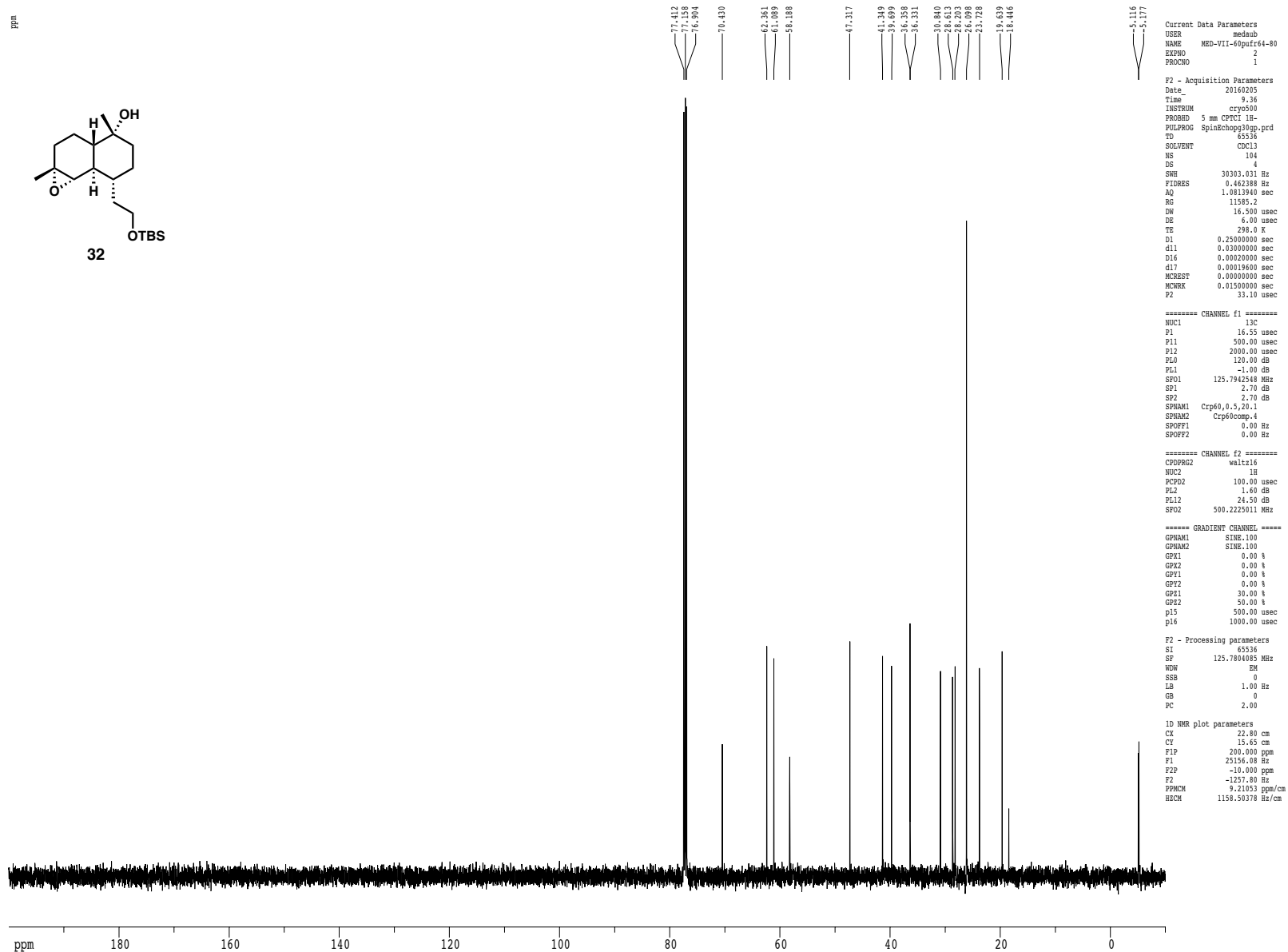
Z-restored spin-echo 13C spectrum with 1H decoupling



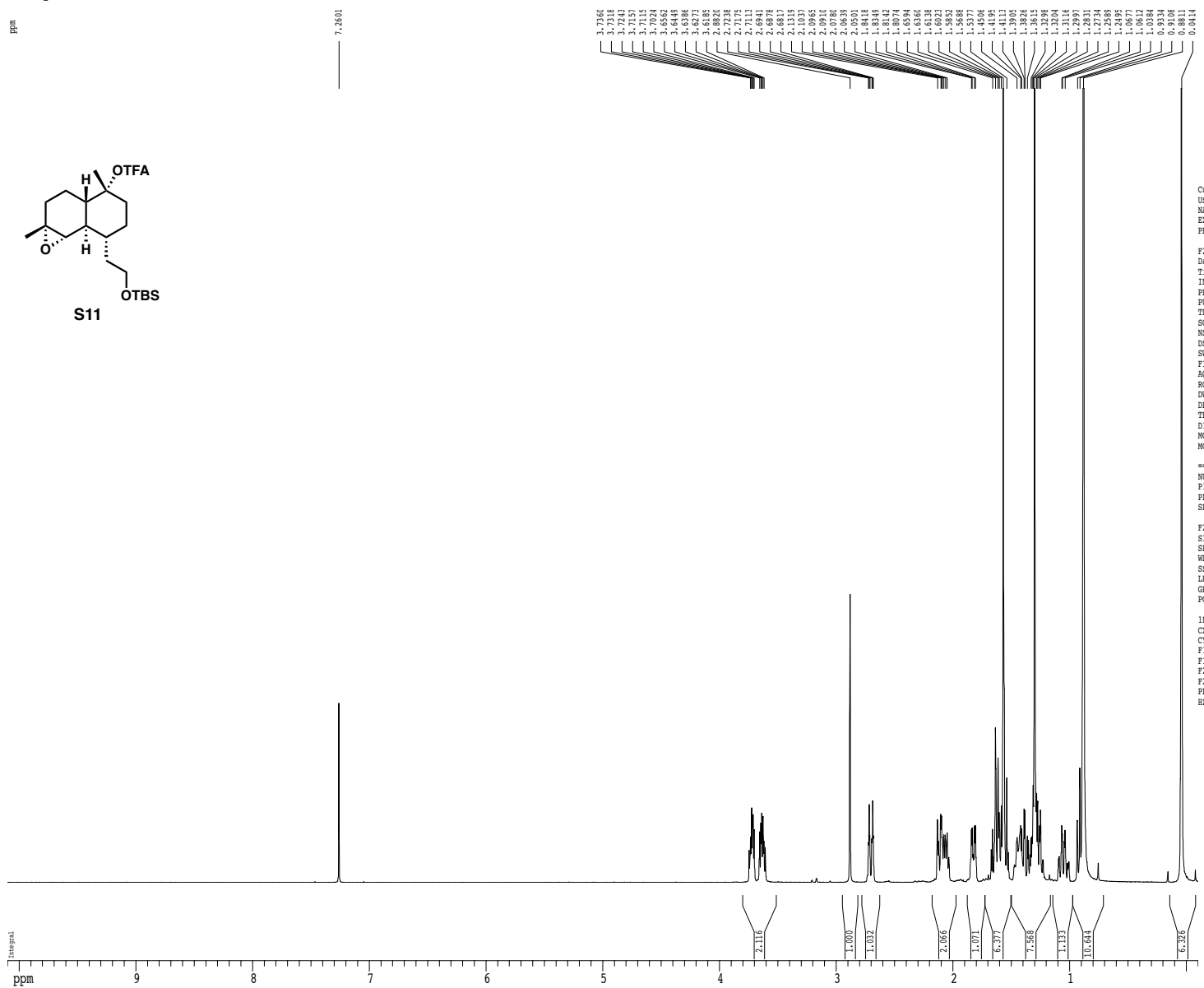
1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum



ppm

```

Current Data Parameters
USER      medaub
NAME      MED-VII-65pu
EXNO      1
PROCNO    1

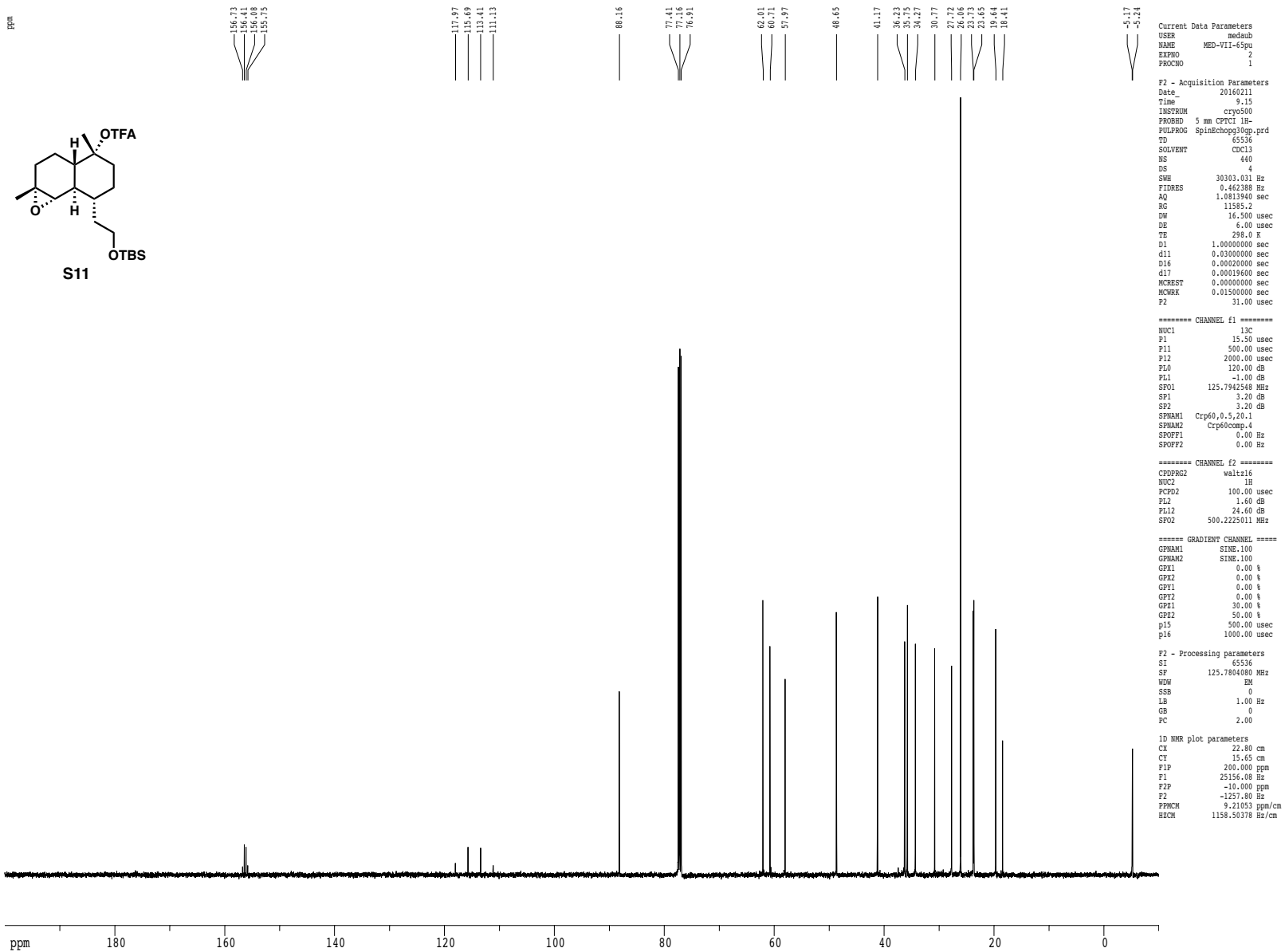
F2 - Acquisition Parameters
Date_     20160211
Time      9.12
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         81728
SOLVENT   CDCl3
NS         8
DS         2
SWH       8012.620 Hz
FIDRES    0.098043 Hz
AQ         5.0998774 sec
RG         5.7
DW         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
MCREST    0.00000000 sec
MWRK      0.01500000 sec

===== CHANNEL f1 =====
NUC1      1H
P1        7.50 usec
PI1       1.60 dB
SFO1      500.2235015 MHz

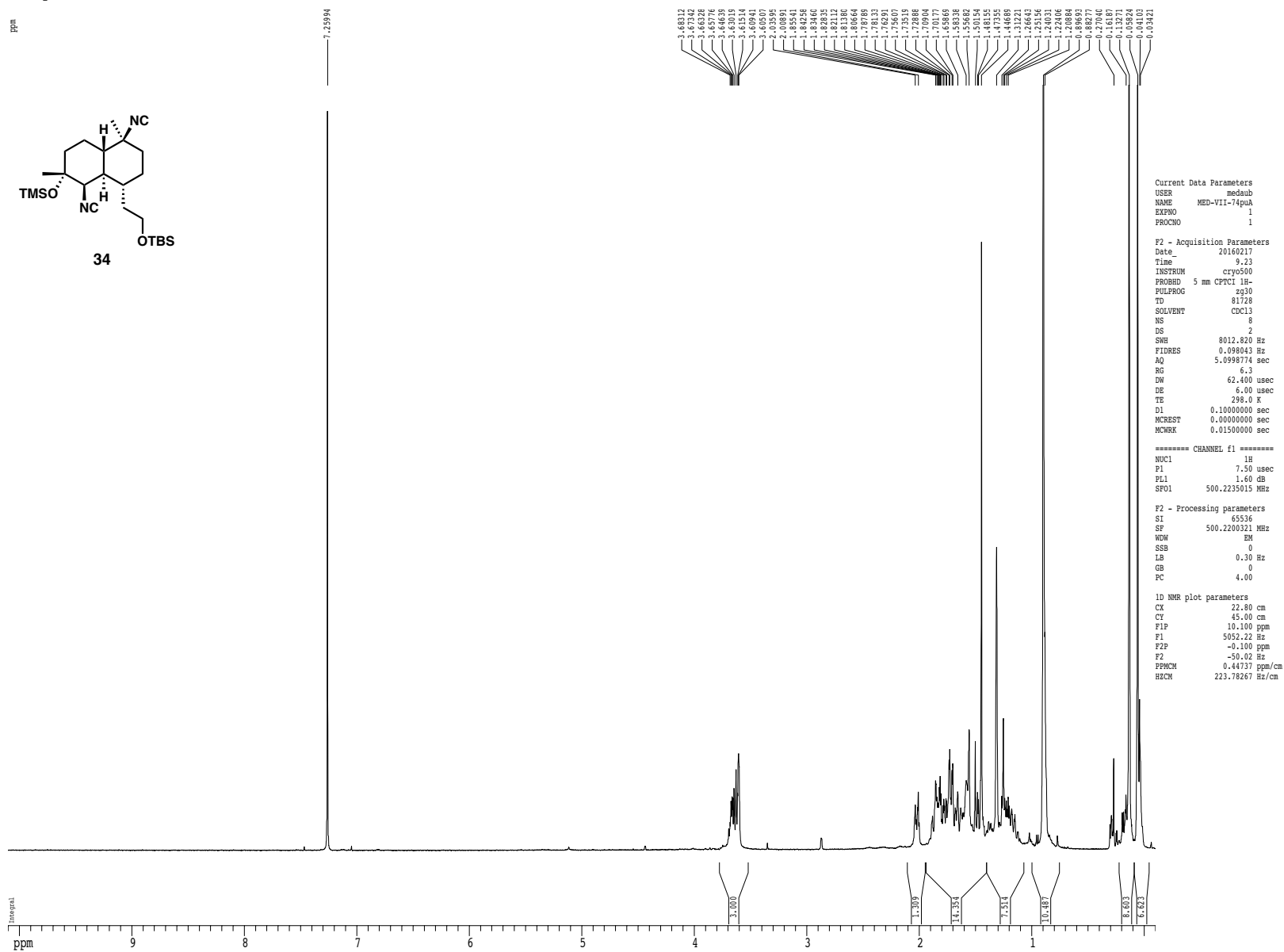
F2 - Processing parameters
SI        65536
SF        500.2200321 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB         0
PC         4.00

ID NMR plot parameters
CX        22.80 cm
CY        45.00 cm
PIP       10.100 ppm
F1        5052.22 Hz
F2P       -0.100 ppm
F2        -50.02 Hz
PPMCM     0.444737 ppm/cm
HZCM      223.78267 Hz/cm
    
```

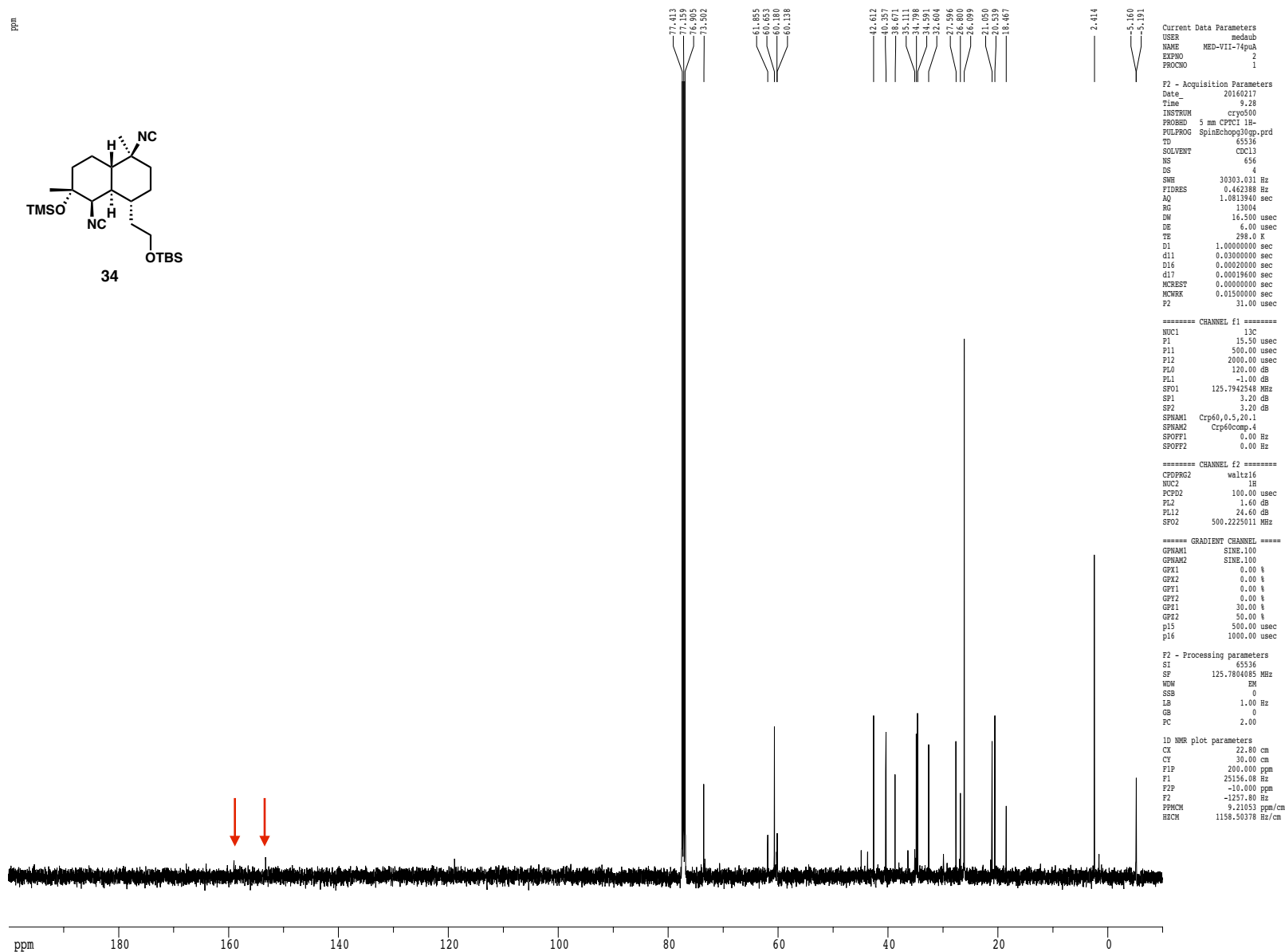
Z-restored spin-echo 13C spectrum with 1H decoupling



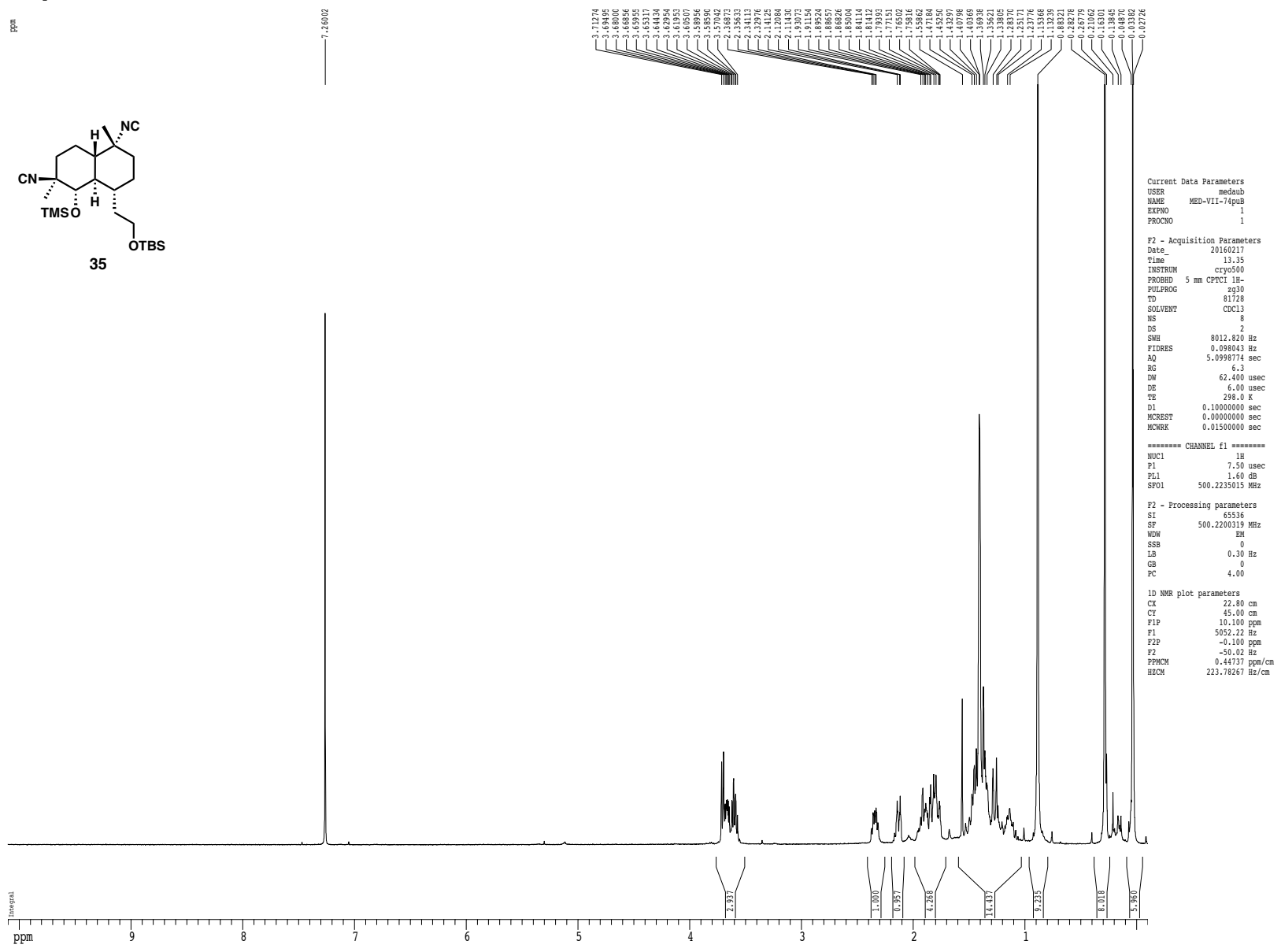
1H spectrum



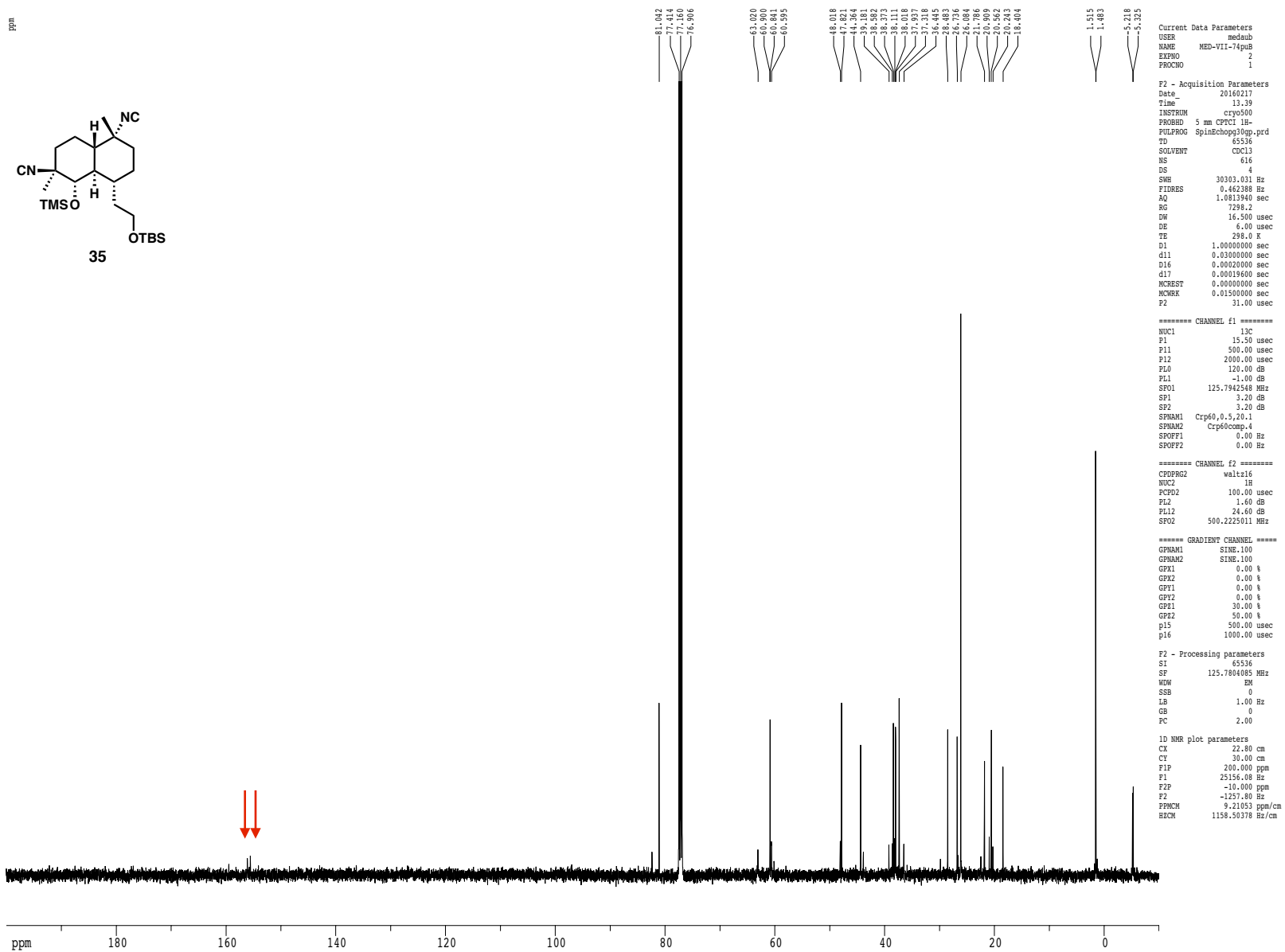
Z-restored spin-echo 13C spectrum with 1H decoupling



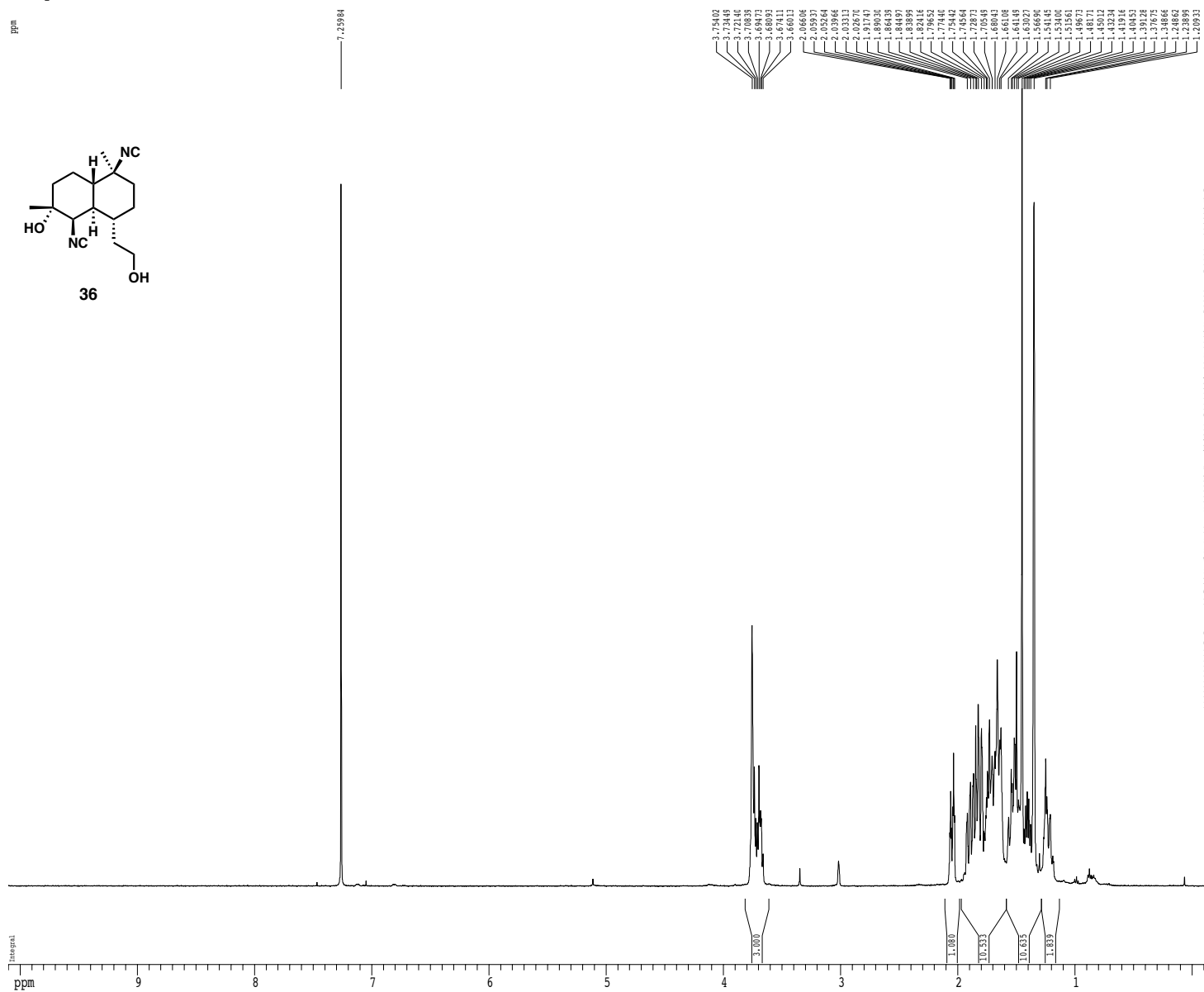
1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum



```

Current Data Parameters
USER      medaub
NAME      MED-VII-107pu
EXPNO     2
PROCNO    1

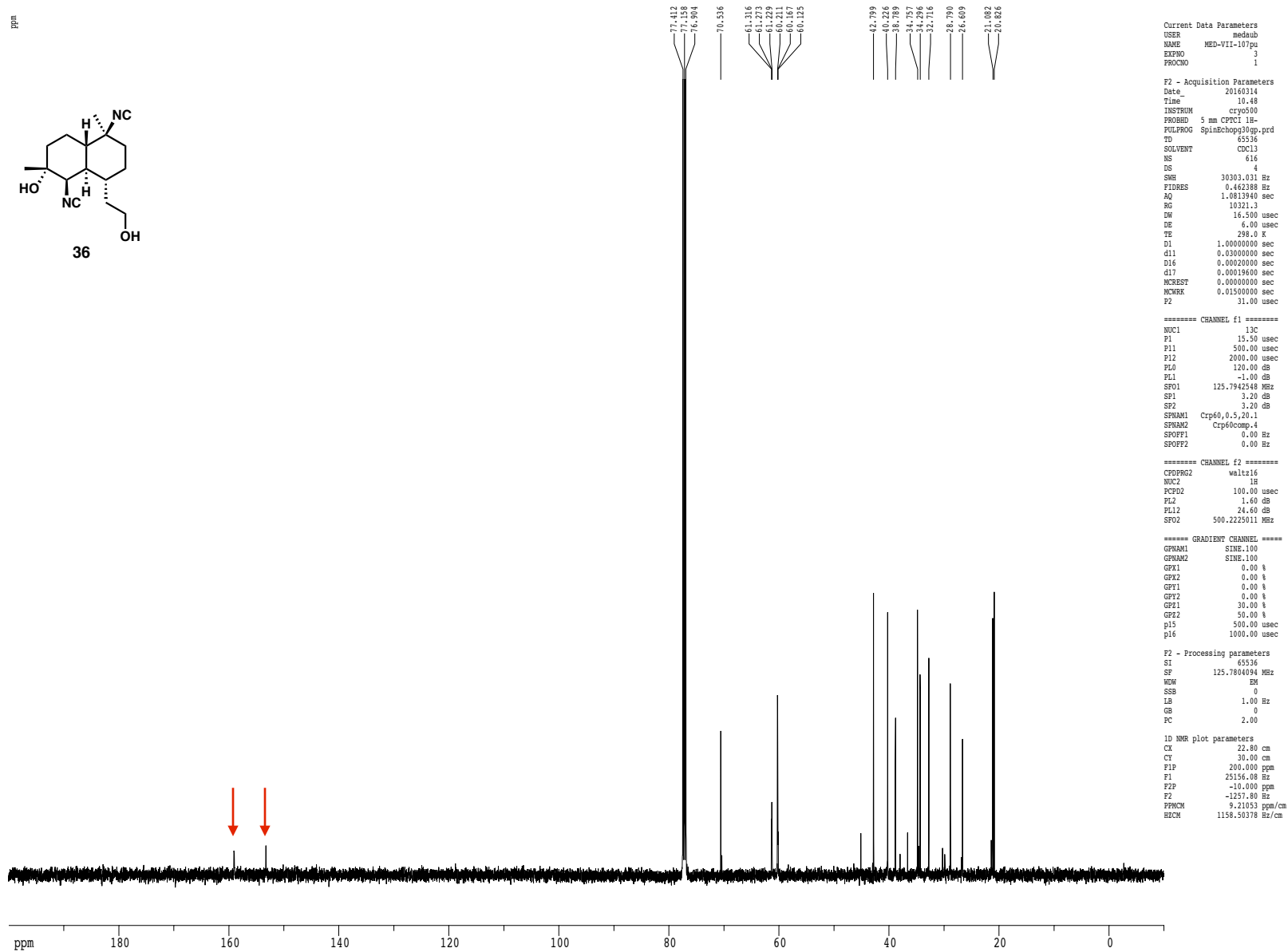
F2 - Acquisition Parameters
Date_     20160314
Time      10.44
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         81728
SOLVENT   CDCl3
NS         8
DS         2
SH         8012.920 Hz
FIDRES    0.098043 Hz
AQ         5.0998774 sec
RG         8
DW         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
MCREST    0.00000000 sec
MCONX     0.01500000 sec

===== CHANNEL f1 =====
NUC1      1H
P1        7.50 usec
PL1       1.60 db
SF01      500.2235015 MHz

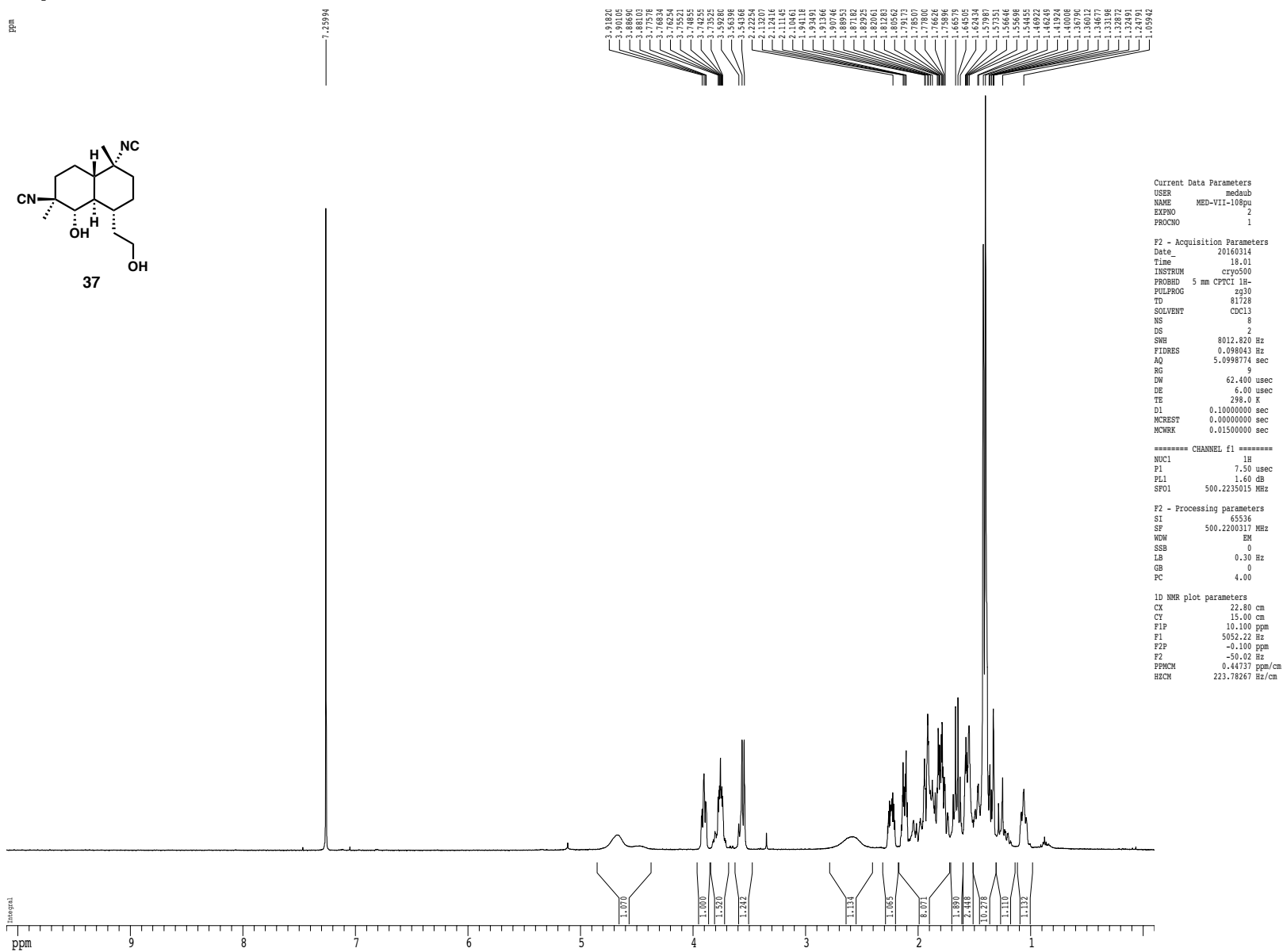
F2 - Processing parameters
SI         65536
SF         500.2209317 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

1D NMR plot parameters
CX         22.80 cm
CY         23.00 cm
F1P        10.100 ppm
F1         5052.22 Hz
F2P        -0.100 ppm
F2         -50.00 Hz
FPMCM     0.44737 ppm/cm
HCM        223.78267 Hz/cm
    
```

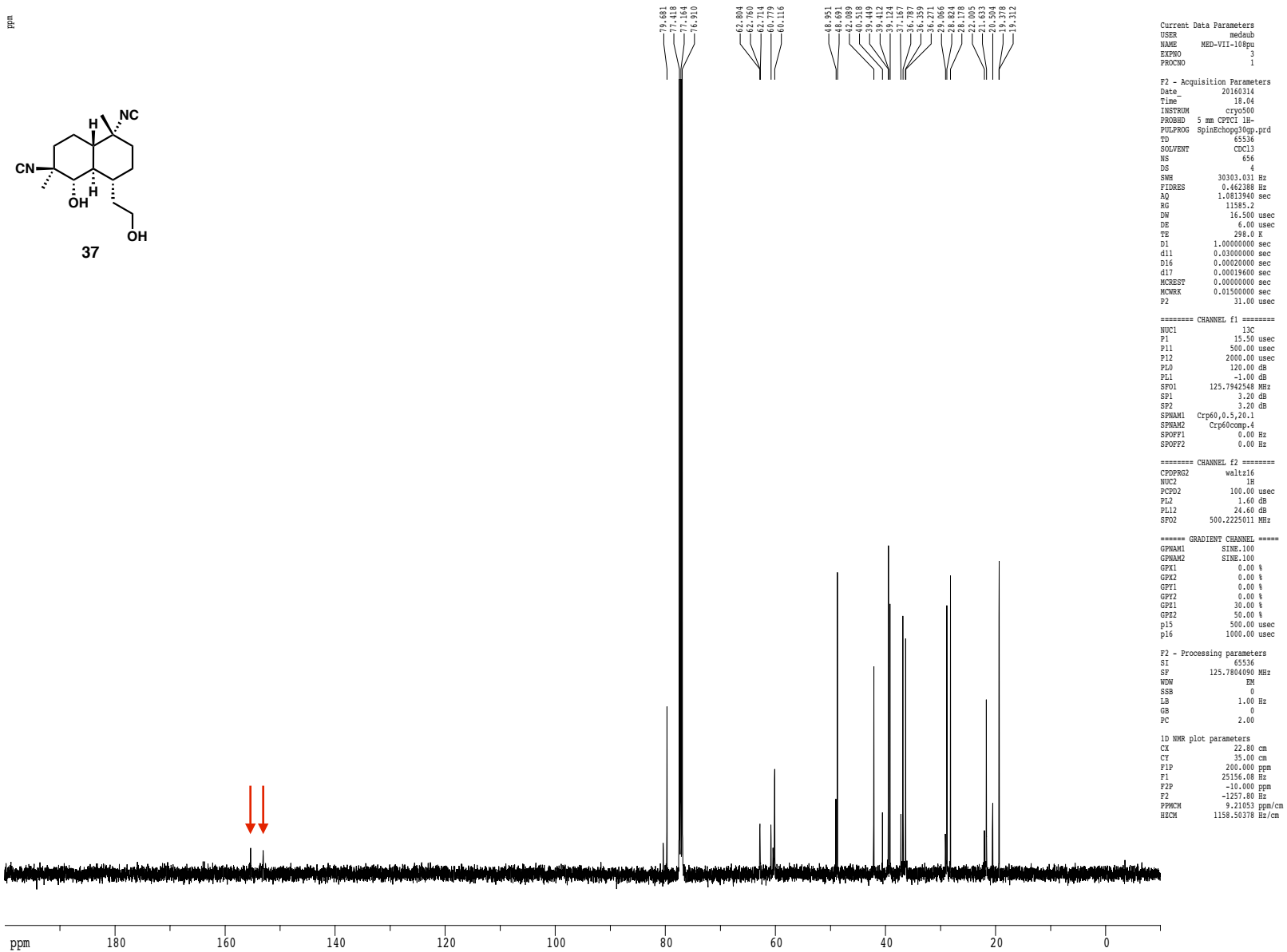
Z-restored spin-echo 13C spectrum with 1H decoupling



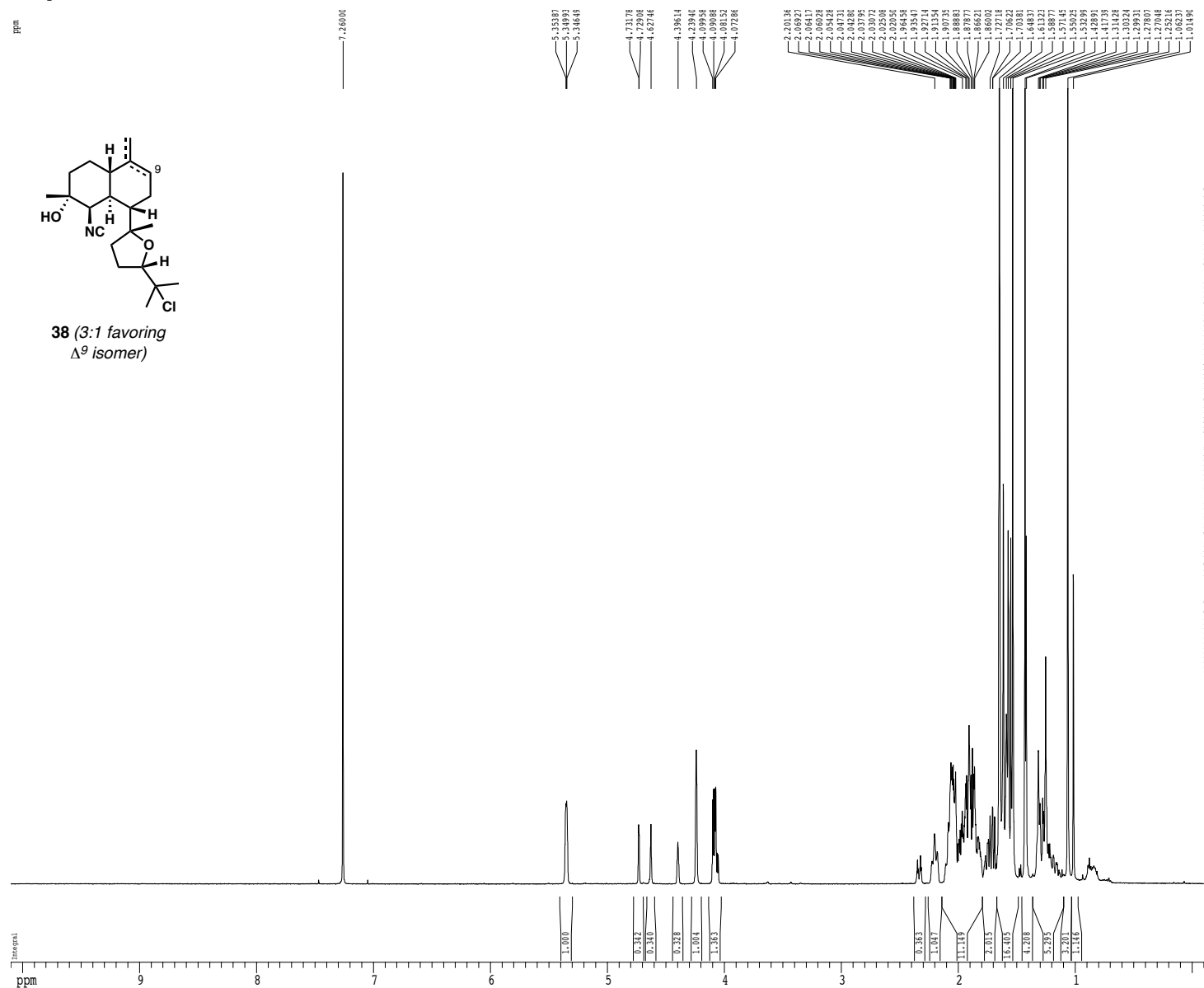
1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum



```

Current Data Parameters
USER          medaub
NAME          MED-VI-260pufr13-16
EXFNO        2
PROCNO        1

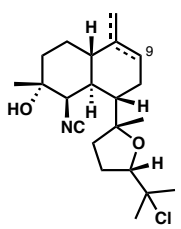
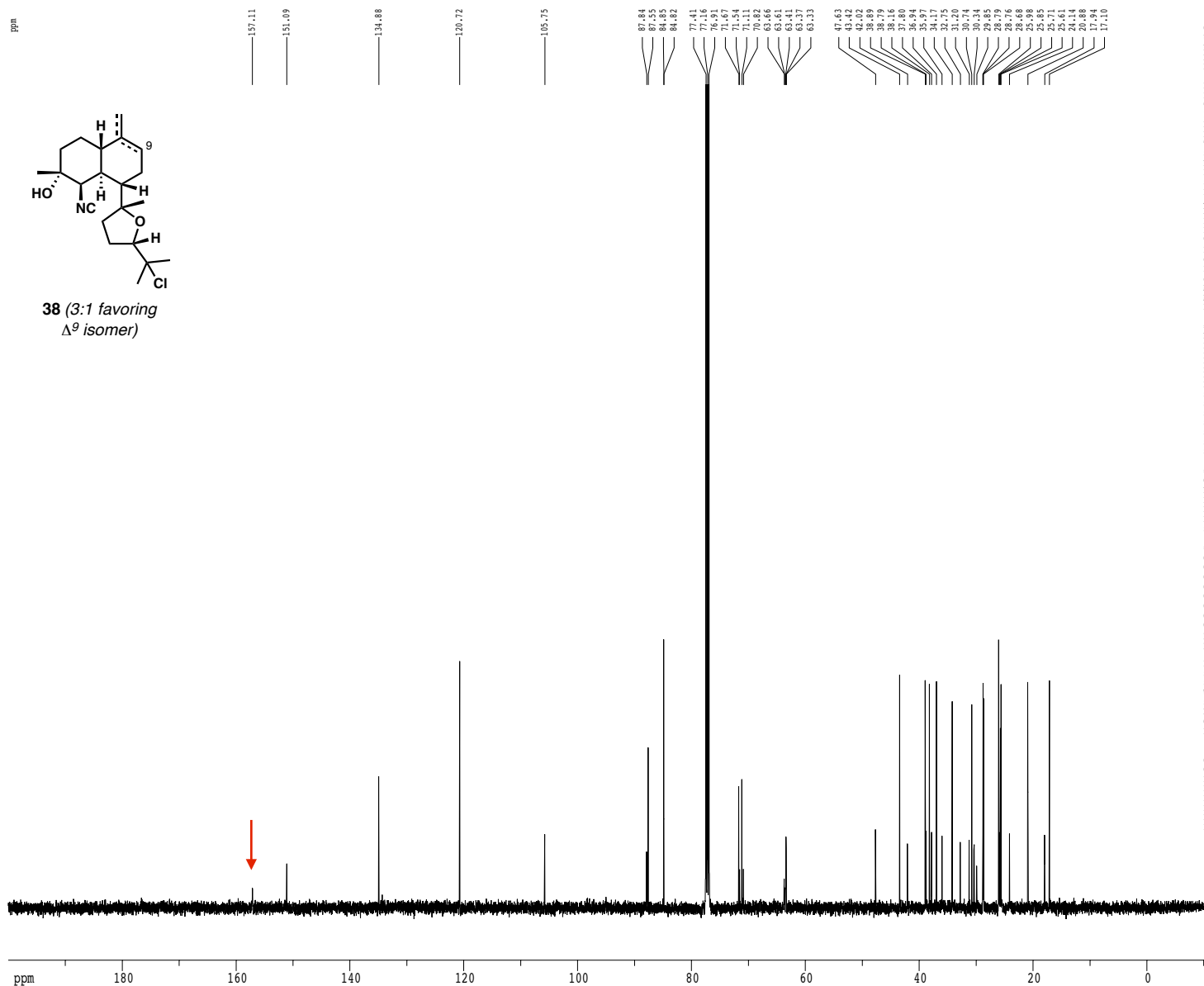
F2 - Acquisition Parameters
Date_         20151017
Time          9.52
INSTRUM       cryo500
PROBHD        5 mm CPTCI 1H-
PULPROG       zg30
TD            81728
SOLVENTY      CDCl3
NS            8
DS            2
SHE           8012.820 Hz
FIDRES        0.098043 Hz
AQ            5.0998774 sec
RG            6.3
DW            62.400 usec
DE            6.00 usec
TE            298.0 K
D1            0.10000000 sec
MCREST        0.00000000 sec
MCRXK         0.01500000 sec

===== CHANNEL f1 =====
NUC1          1H
P1            7.50 usec
PL1           1.60 dB
SFO1          500.225015 MHz

F2 - Processing parameters
SI            65536
SF            500.2200315 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            0.50

1D NMR plot parameters
CX            22.80 cm
CY            25.00 cm
FIP           10.100 ppm
F1            5052.22 Hz
F2P           -0.100 ppm
F2            -50.02 Hz
F2MCH         0.44337 ppm/cm
H2MCH         223.78267 Hz/cm
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



38 (3:1 favoring Δ^9 isomer)

```

Current Data Parameters
USER          medaub
NAME         MED-VI-26(pufr13-16
EXPNO        3
PROCNO       1

F2 - Acquisition Parameters
Date_        20151017
Time         9.54
INSTRUM      cryo500
PROBHD       5 mm CPXI 1H-
PULPROG      SpinEchoq30q.prd
TD           65536
SOLVENT      CDCl3
NS           752
DS           4
SWH          30303.031 Hz
FIDRES      0.462388 Hz
AQ          1.0813940 sec
RG          11585.2
DW          16.500 usec
DE          6.00 usec
TE          298.0 K
D1          1.00000000 sec
d11         0.03000000 sec
d16         0.00020000 sec
d17         0.00019600 sec
MCREST      0.00000000 sec
MCRMX       0.01500000 sec
F2          31.00 usec

===== CHANNEL f1 =====
NUC1         13C
P1           15.50 usec
P11          500.00 usec
P12          2000.00 usec
PL0          120.00 dB
PL1          -1.00 dB
SFO1        125.7942548 MHz
SF1          3.20 dB
SP2          3.20 dB
SFO2        Crp60,0.5,20.1
SF2          3.20 dB
SFOFF1       0.00 Hz
SFOFF2       0.00 Hz

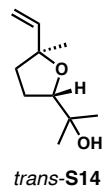
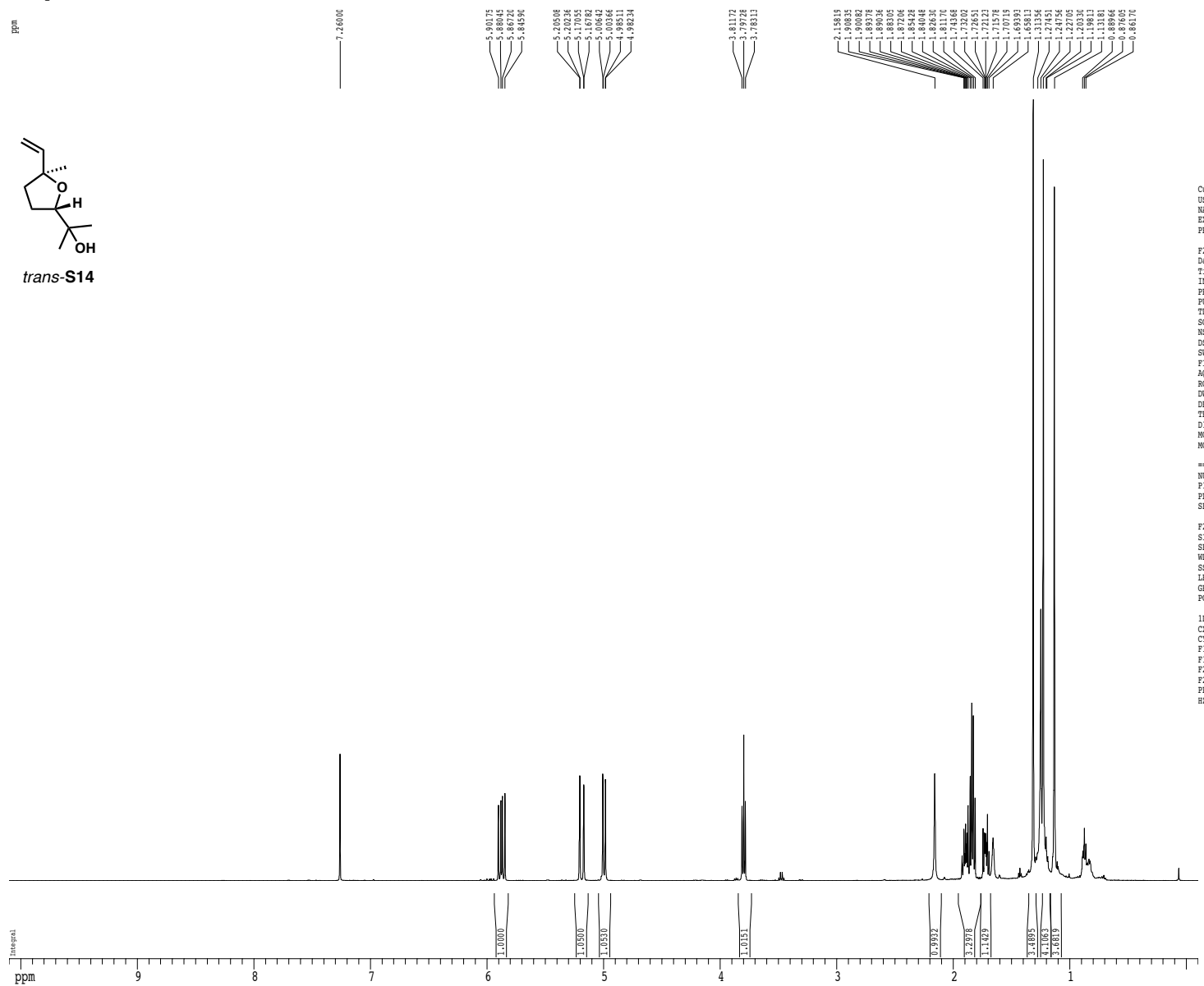
===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2         1H
PCPD2        100.00 usec
PL2          1.60 dB
PL12         24.60 dB
SFO2        500.2225011 MHz

===== GRADIENT CHANNEL =====
GPMAM1      SINE.100
GPMAM2      SINE.100
GPF1        0.00 %
GPF2        0.00 %
GPF1        0.00 %
GPF2        0.00 %
GPF1        30.00 %
GPF2        50.00 %
p15         500.00 usec
p16         1000.00 usec

F2 - Processing parameters
SI          65536
SF          125.7804082 MHz
WDW         EM
SSB         0
LB          1.00 Hz
GB          0
PC          2.00

1D NMR plot parameters
CA          22.80 cm
CY          30.00 cm
F1P         200.000 ppm
F1          25156.08 Hz
F1P         -10.000 ppm
F2          -1257.80 Hz
PPMCH      9.21053 ppm/cm
HSCM       1158.50378 Hz/cm
    
```

¹H spectrum



```

Current Data Parameters
USER      medaub
NAME      MED-VI-246pu
EXNO      1
PROCNO    1

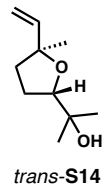
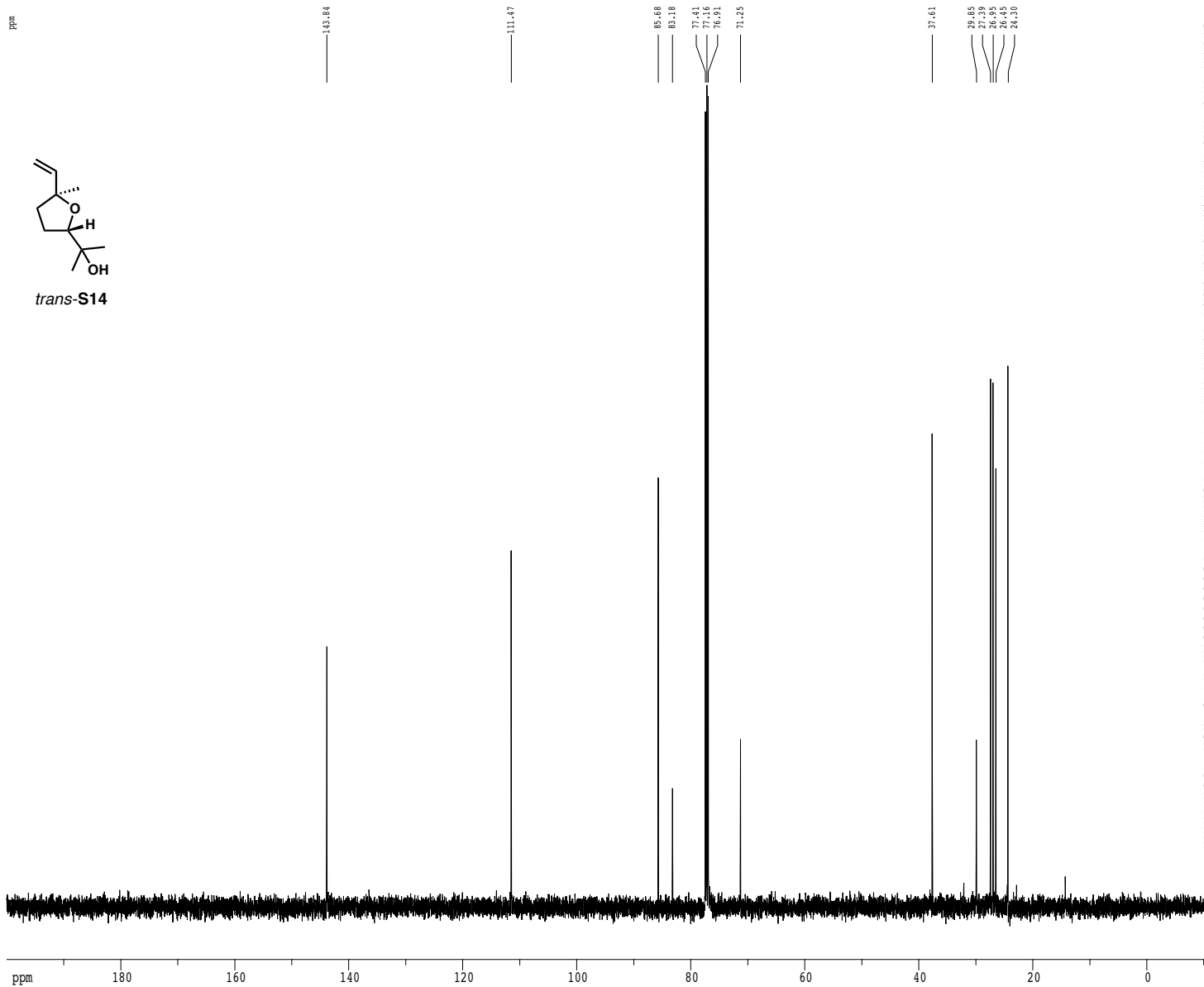
F2 - Acquisition Parameters
Date_     20150926
Time      15.16
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         81728
SOLVENT   CDCl3
NS         8
DS         2
SWH        8012.620 Hz
FIDRES     0.098043 Hz
AQ         5.0998774 sec
RG         6.3
DW         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
MCREST    0.00000000 sec
MCMR      0.01500000 sec

===== CHANNEL f1 =====
NUC1       1H
P1         7.50 usec
PI1        1.60 dB
SFO1       500.2235015 MHz

F2 - Processing parameters
SI         65536
SF         500.2200313 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00

ID NMR plot parameters
CX         22.80 cm
CY         15.00 cm
PI1        10.100 ppm
F1         5052.22 Hz
F2P        -0.100 ppm
F2         -50.02 Hz
PPHMC      0.444737 ppm/cm
HZCM       223.78267 Hz/cm
    
```


Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER      medaub
NAME      MED-VI-240pu
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20150926
Time      15.19
INSTRUM   cryo500
PROBHD    5 mm CP1C1 1H-
PULPROG   SpinEchoSgppg-prd
TD         65536
SOLVENT   CDCl3
NS         104
DS         4
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         11985.2
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
d16        0.00020000 sec
d17        0.00019600 sec
MCREST    0.00000000 sec
MCMRFX    0.01500000 sec
P2         33.10 usec

===== CHANNEL f1 =====
NUC1       13C
P1         16.55 usec
PL1        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1       125.7842548 MHz
SP1        2.70 dB
SP2        2.70 dB
SFOF1      Crp60,0.5,20.1
SFOF2      Crp60comp,4
SPOFF1     0.00 Hz
SPOFF2     0.00 Hz

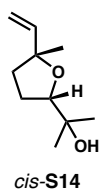
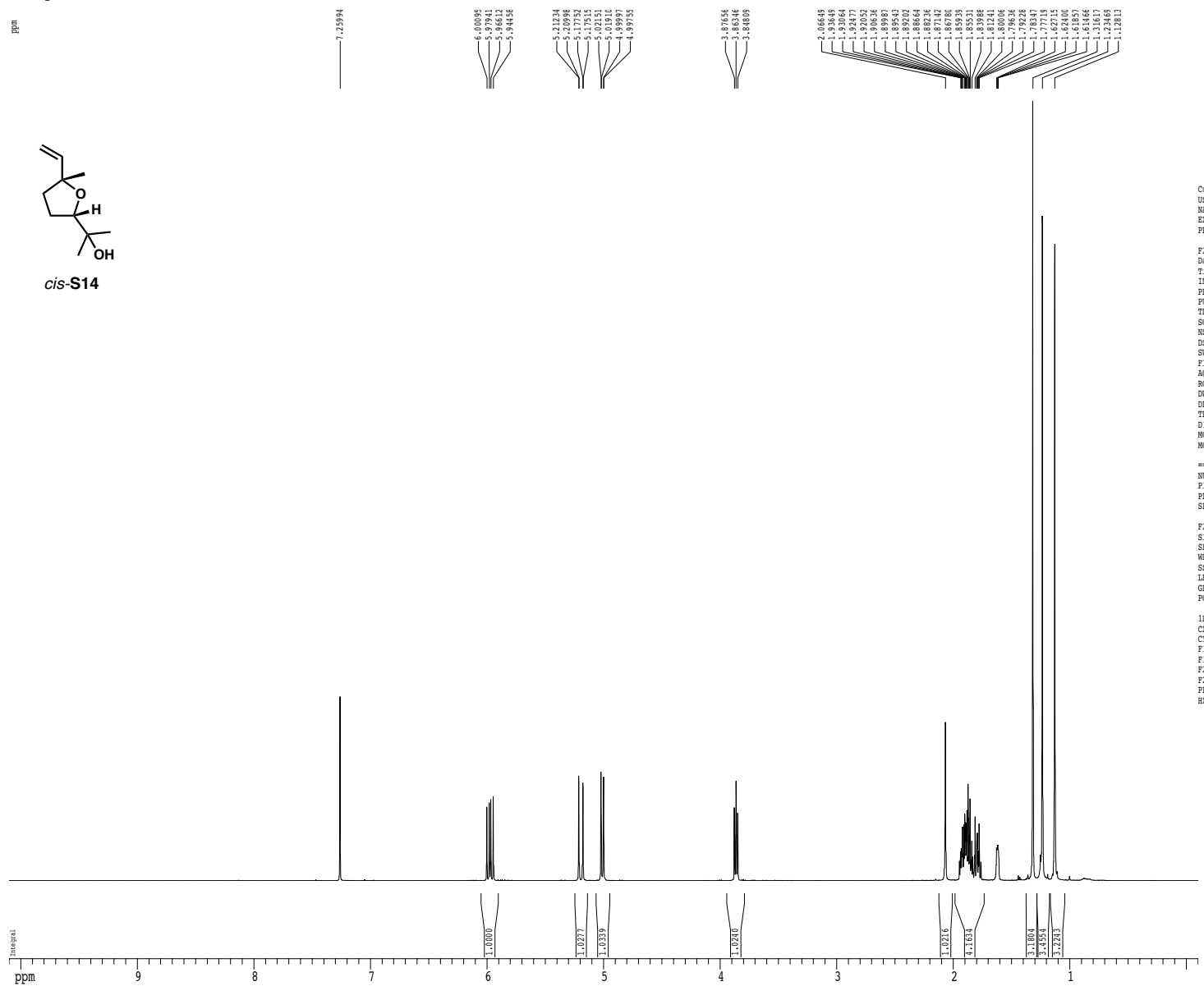
===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2       1H
PCPD2     100.00 usec
PL2        1.60 dB
PL12       24.50 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1    SINE.100
GPNAM2    SINE.100
GPF1      0.00 %
GPF2      0.00 %
GPF1      0.00 %
GPF2      0.00 %
GPF1      30.00 %
GPF2      50.00 %
p15       500.00 usec
p16       1000.00 usec

F2 - Processing parameters
SI         65536
SF         125.7804080 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

1D NMR plot parameters
CX         22.80 cm
CI         15.65 cm
FIP        200.000 ppm
F1         25156.08 Hz
F2         -10.000 ppm
F2         -1257.80 Hz
F2FCH      9.21953 ppm/cm
H2CM       1158.50378 Hz/cm
    
```

¹H spectrum



```

Current Data Parameters
USER      medaub
NAME      MED-VI-241pu
EXPNO     1
PROCNO    1

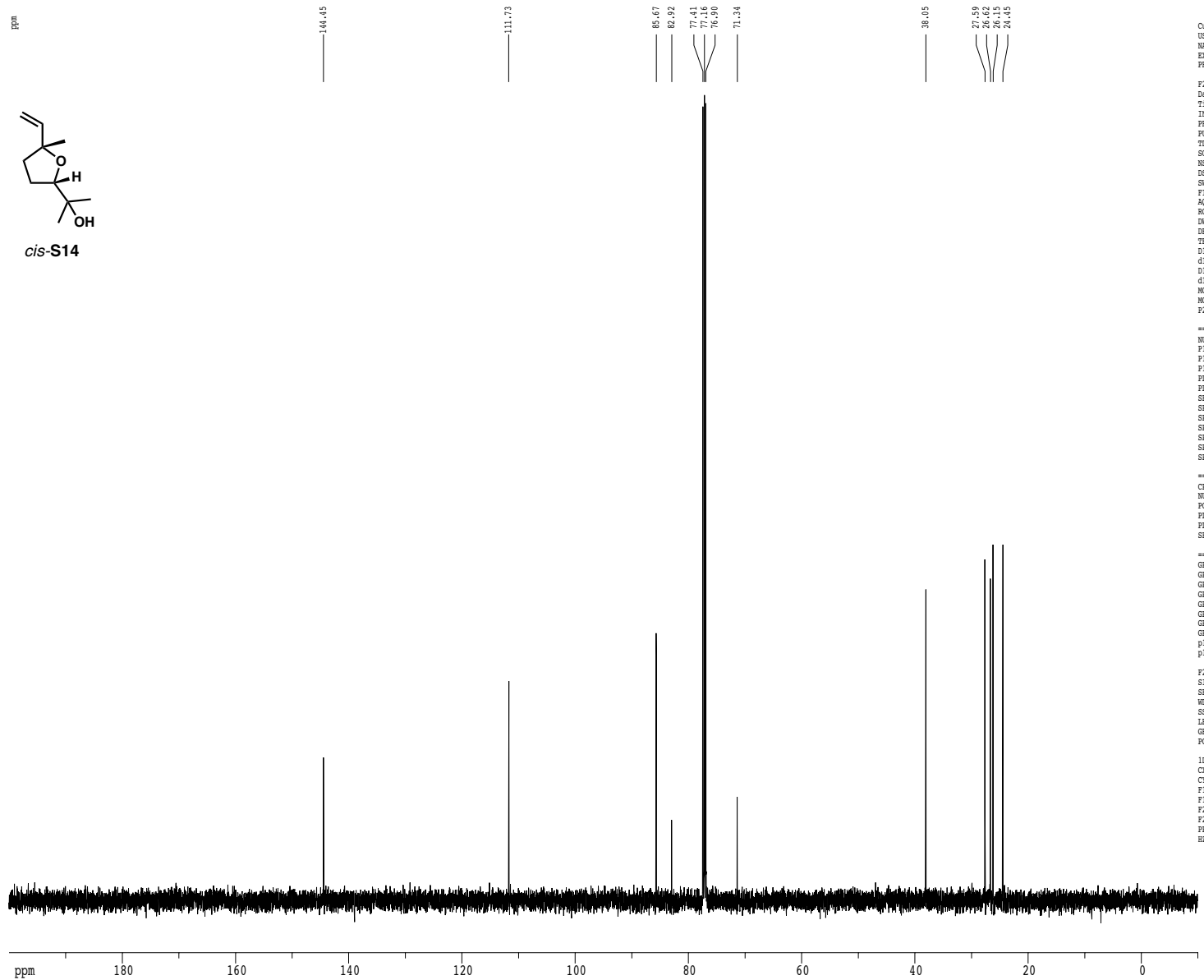
F2 - Acquisition Parameters
Date_     20150926
Time      15.25
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         81728
SOLVENT   CDCl3
NS         8
DS         2
SWH        8012.620 Hz
FIDRES     0.098043 Hz
AQ         5.0998774 sec
RG         7.1
DW         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
MCREST    0.00000000 sec
MCWRK     0.01500000 sec

===== CHANNEL f1 =====
NUC1       1H
P1         7.50 usec
PI1        1.60 dB
SFO1       500.2235015 MHz

F2 - Processing parameters
SI         65536
SF         500.2200313 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00

ID NMR plot parameters
CX         22.80 cm
CY         15.00 cm
PI1        10.100 ppm
F1         5052.22 Hz
F2P        -0.100 ppm
F2         -50.02 Hz
PPHNCM     0.444737 ppm/cm
HZCM       223.78267 Hz/cm
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER      medaub
NAME      MED-VI-241pu
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20150926
Time      15.27
INSTRUM   cryo500
PROBHD    5 mm CPCCI 1H-
PULPROG   SpinEcho30ap.prd
TD         65536
SOLVENT   CDCl3
NS         88
DS         4
SNR        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         11585.2
DM         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
D16        0.00020000 sec
d17        0.00019600 sec
MCKEST     0.00000000 sec
MCKWK      0.01500000 sec
P2         33.10 usec

===== CHANNEL f1 =====
NUC1       13C
P1         16.55 usec
P11        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1      125.7942548 MHz
SF1        2.70 dB
SP2        2.70 dB
SFOAM1     Crp60,0.5,20.1
SFOAM2     Crp60comp.4
SFOFF1     0.00 Hz
SFOFF2     0.00 Hz

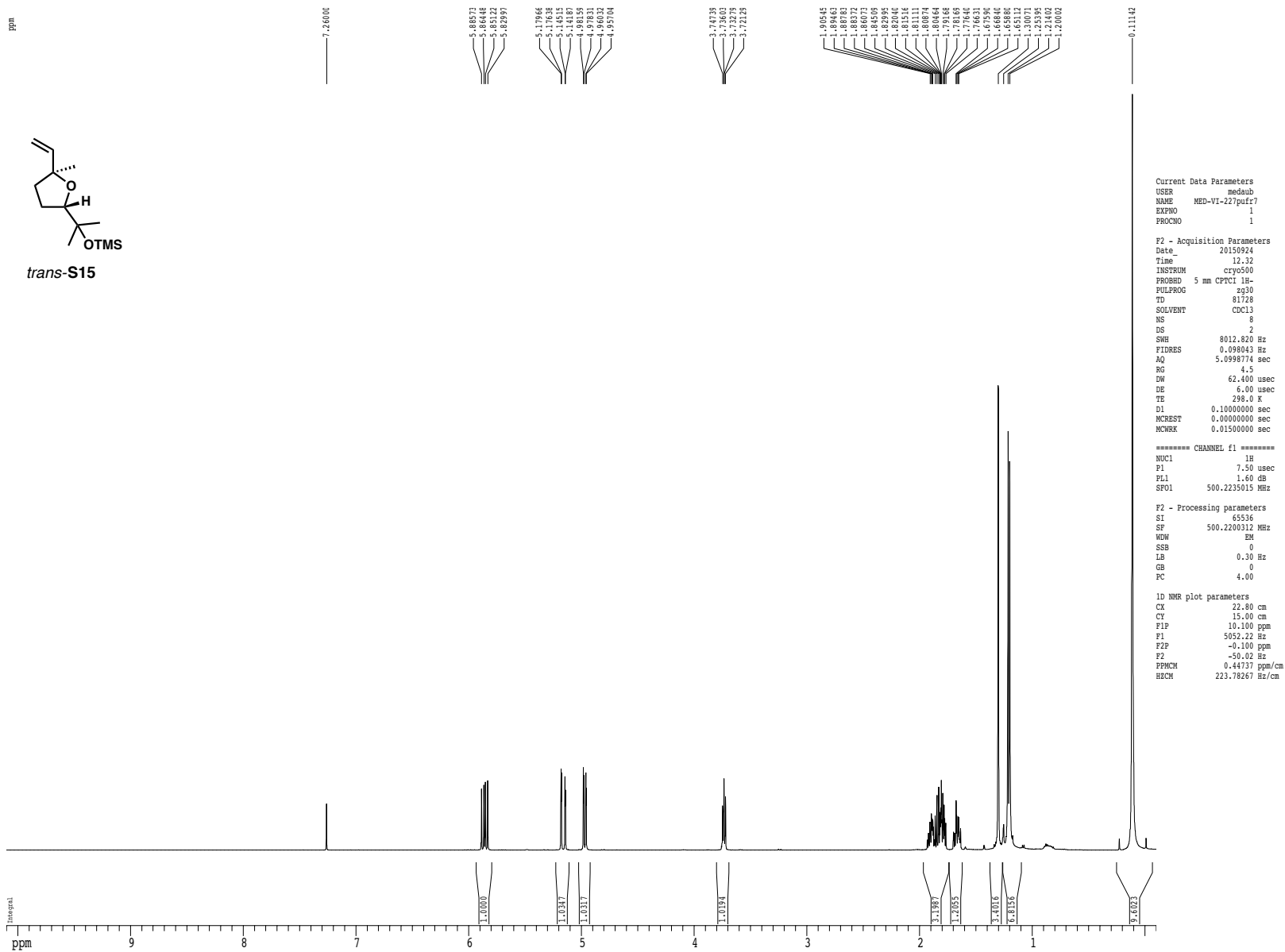
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2     100.00 usec
PL2        1.00 dB
PL12       24.50 dB
SFO2      500.2225011 MHz

===== GRADIENT CHANNEL =====
GPMAM1     SINE.100
GPMAM2     SINE.100
GPX1       0.00 t
GPX2       0.00 t
GPY1       0.00 t
GPY2       0.00 t
GPZ1       30.00 t
GPZ2       50.00 t
p15        500.00 usec
p16        1000.00 usec

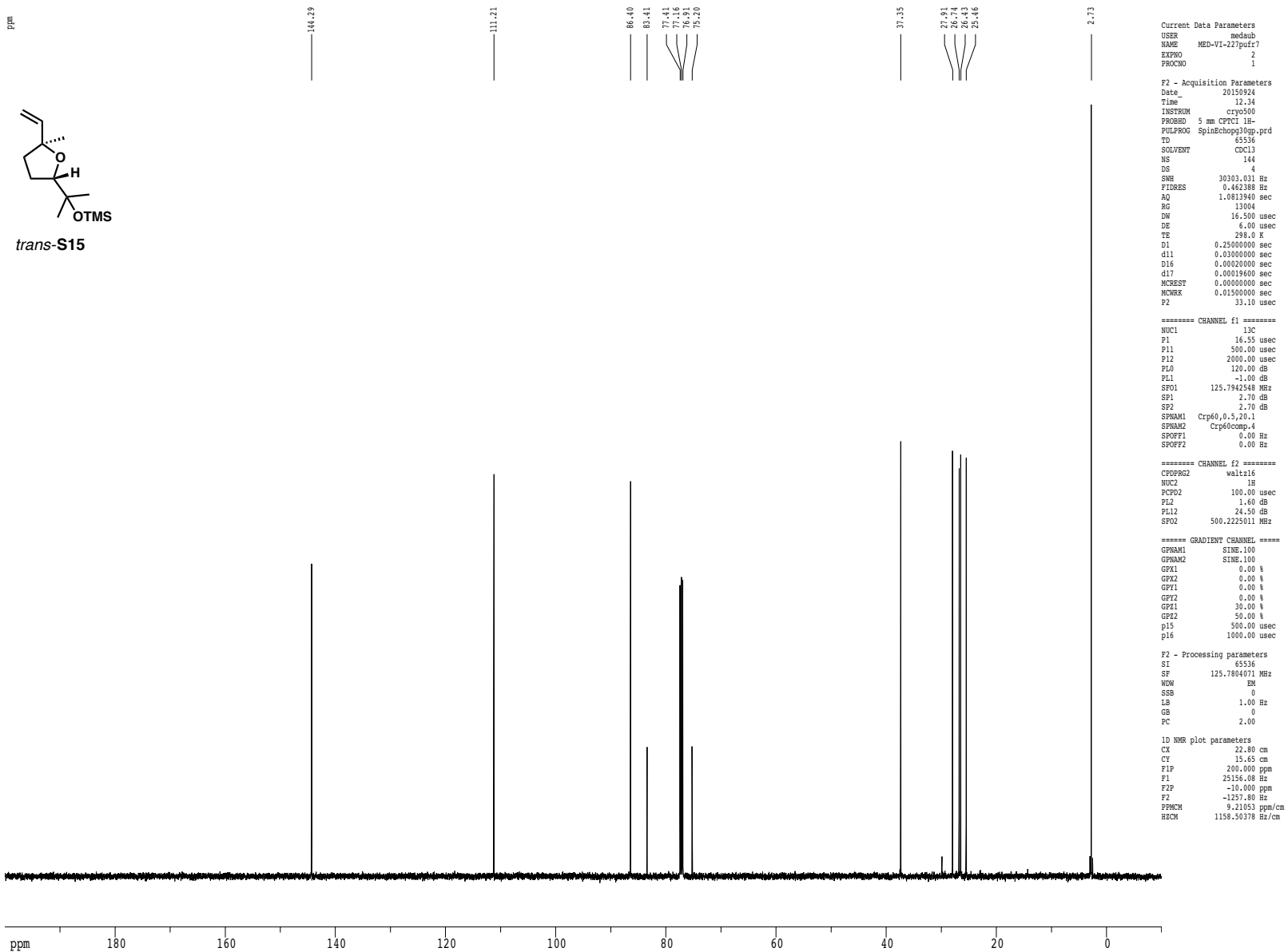
F2 - Processing parameters
SI         65536
SF         125.7804080 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

1D NMR plot parameters
CX         22.80 cm
CT         15.65 cm
PLP        200.000 ppm
F1         25156.08 Hz
F2P        -10.000 ppm
FZ         -1257.80 Hz
PPHMC      9.21053 ppm/cm
HRCM       1158.50378 Hz/cm
    
```

1H spectrum

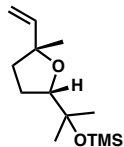


Z-restored spin-echo 13C spectrum with 1H decoupling

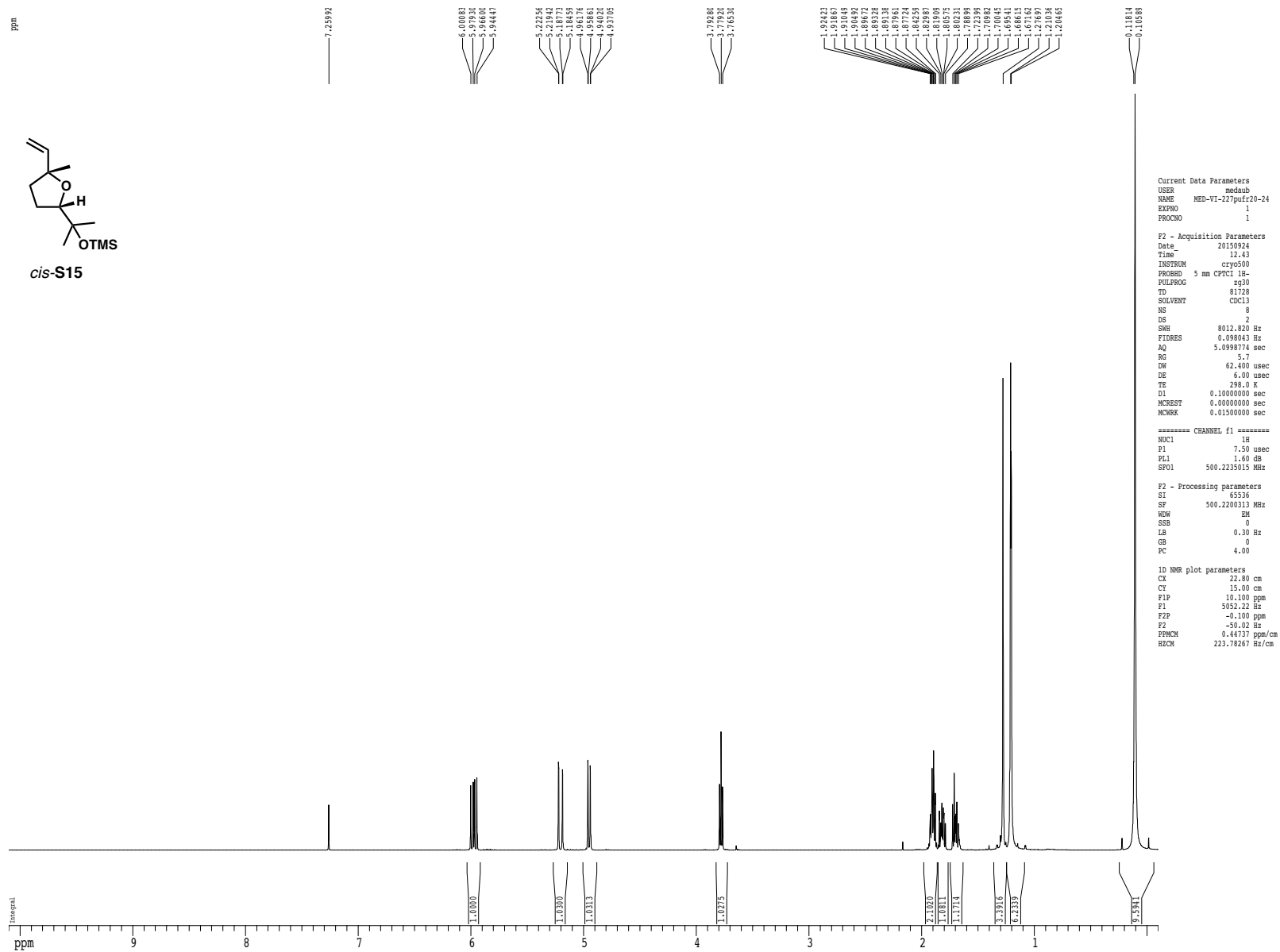


¹H spectrum

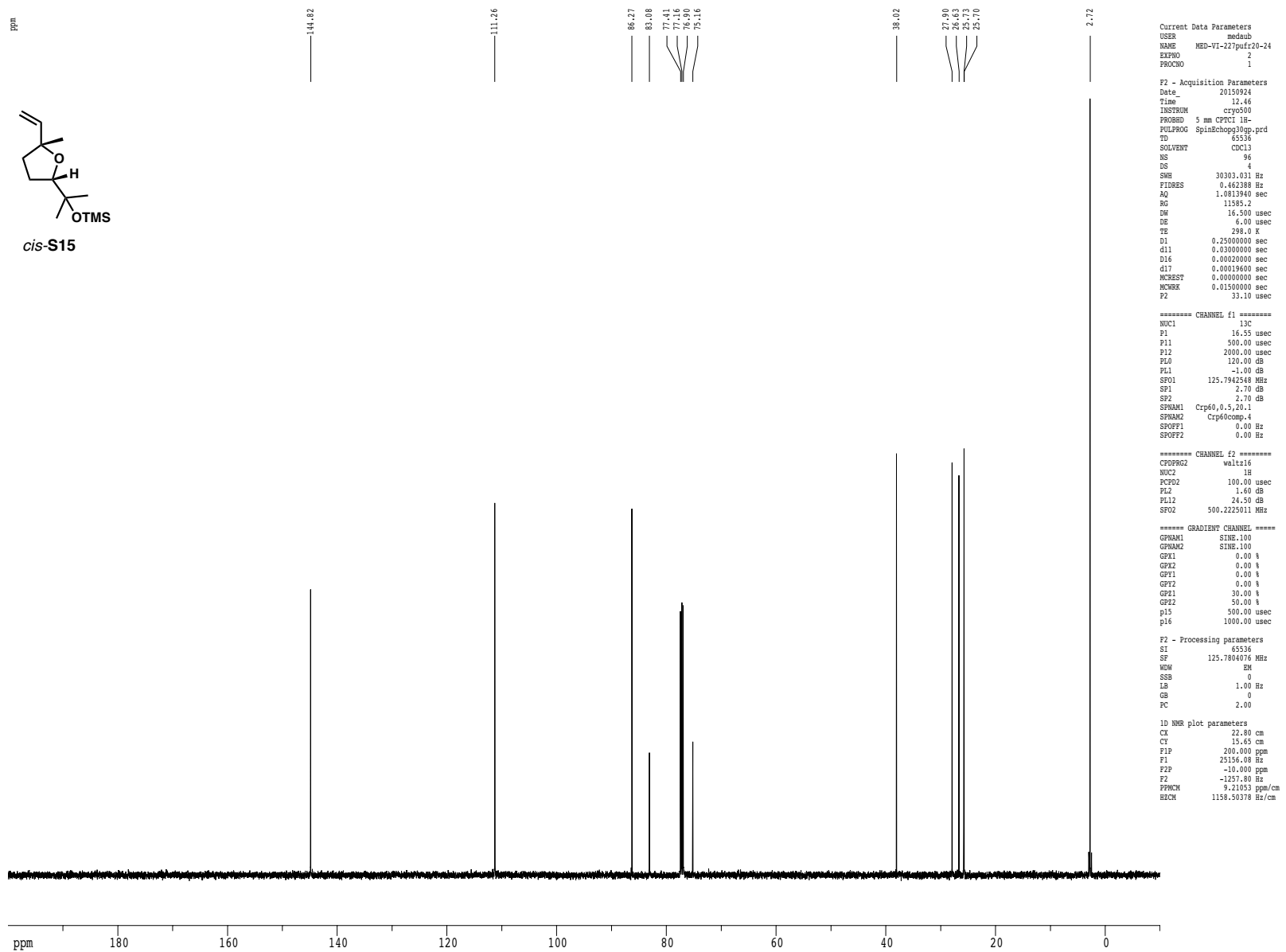
ppm



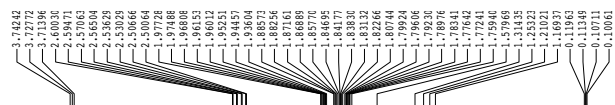
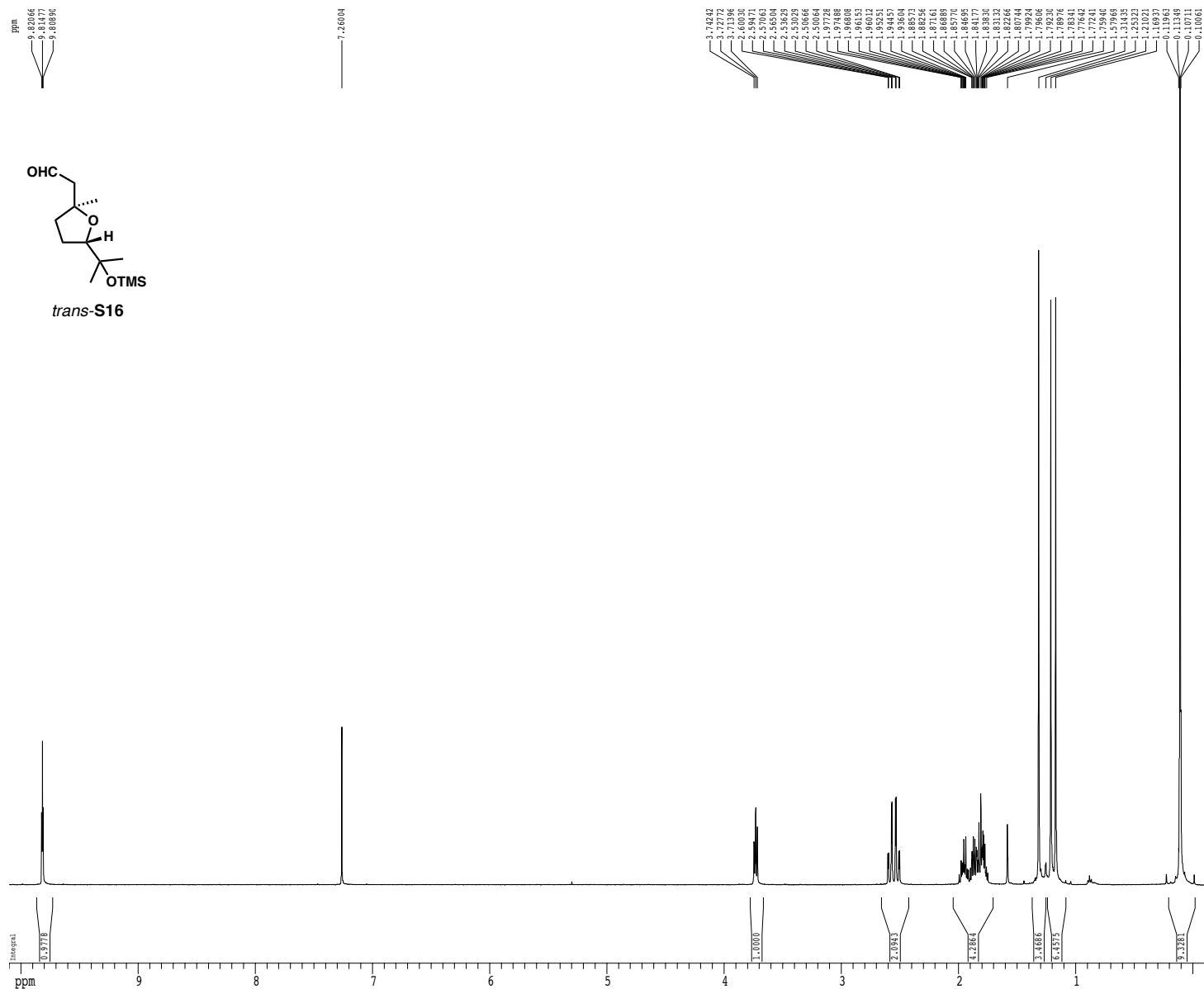
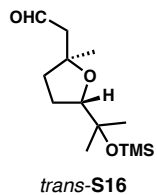
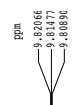
cis-S15



Z-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum



```

Current Data Parameters
USER      medaub
NAME      MED-VI-269u
EXPNO     1
PROCNO    1

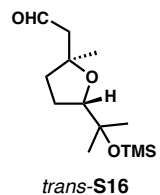
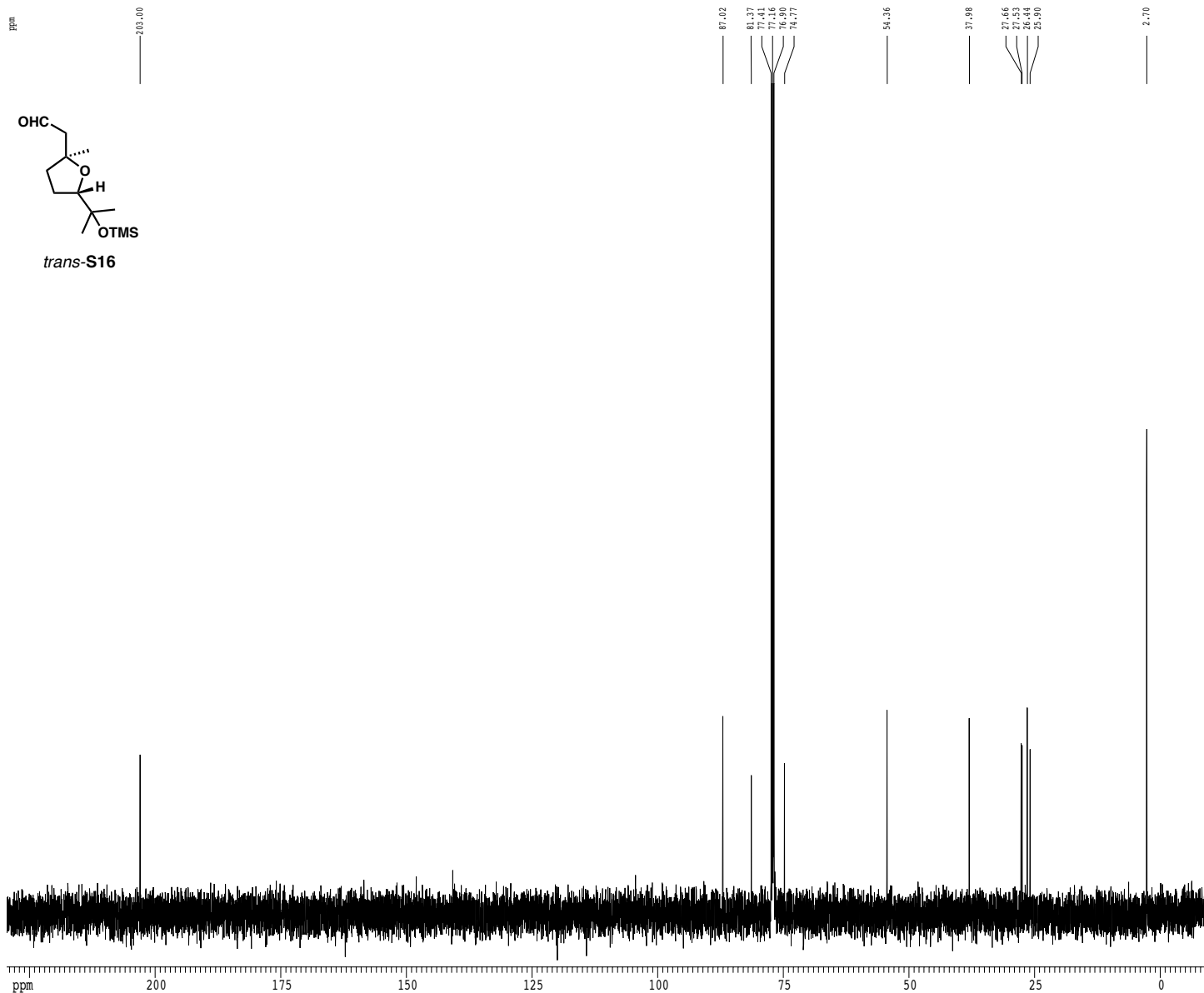
F2 - Acquisition Parameters
Date_     20151021
Time      22.03
INSTRUM   gn500
PROBHD    5 mm broadband
PULPROG   zg30
TD         81728
SOLVENT   CDCl3T
NS         8
DS         2
SNH        8012.820 Hz
FIDRES     0.098043 Hz
AQ         5.0998774 sec
RG         574.7
DW         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
MCREST     0.00000000 sec
MCWRR      0.01500000 sec

===== CHANNEL f1 =====
NUC1       1H
P1         12.00 usec
PL1        -5.80 dB
SFO1       499.1834943 MHz

F2 - Processing parameters
SI         65536
SF         499.1800270 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

1D NMR plot parameters
CX         22.80 cm
CY         50.00 cm
F1P        10.100 ppm
F1         5041.72 Hz
F2P        -0.100 ppm
F2         -49.92 Hz
PPMCM      0.44737 ppm/cm
HZCM       223.31740 Hz/cm
    
```


13C spectrum with 1H decoupling



```

Current Data Parameters
USER      medaub
NAME      MED-VI-269pu
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20151021
Time      22.06
INSTRUM   gm500
PROBHD    5 mm broadband
PULPROG   zgdc30
TD         65536
SOLVENT   CDCl3
NS         512
DS         4
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         46241
DW         16.500 usec
DE         4.50 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
MCREST     0.00000000 sec
MCMRXX     0.01500000 sec

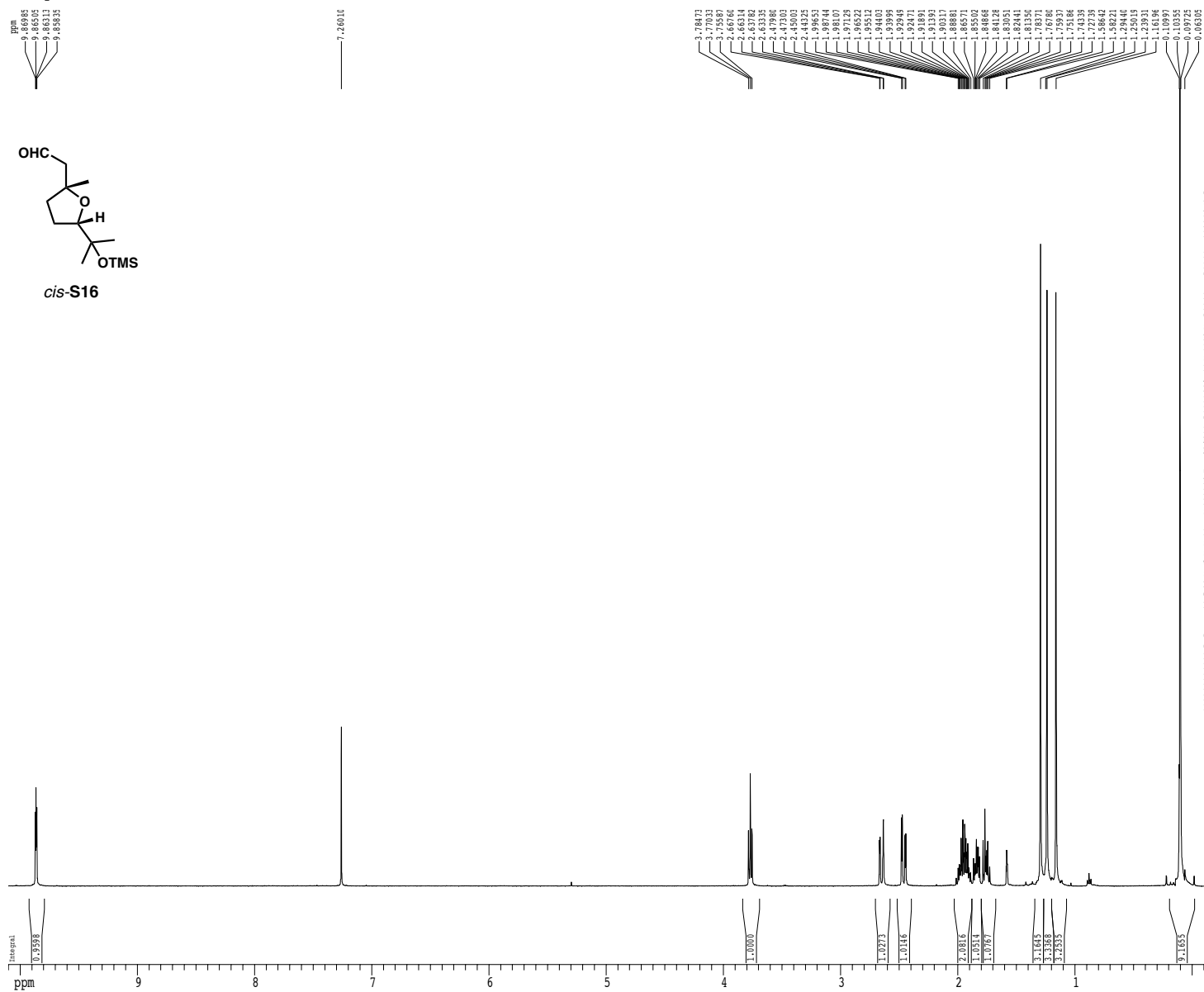
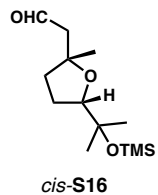
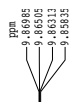
===== CHANNEL f1 =====
NUC1       13C
P1         9.00 usec
PL1        -0.60 dB
SFO1       125.5327181 MHz

===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2      80.00 usec
PL2        -3.00 dB
PL12       12.80 dB
SFO2       499.1824959 MHz

F2 - Processing parameters
SI         65536
SF         125.5188970 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

1D NMR plot parameters
CX         22.80 cm
CY         30.00 cm
FIP        229.520 ppm
F1         28809.06 Hz
F2P        -10.507 ppm
F2         -1318.78 Hz
F2MCM      10.52747 ppm/cm
H2MCM      1321.39624 Hz/cm
    
```

1H spectrum



Current Data Parameters
 USER medaub
 NAME MED-VI-270pu
 EXPNO 1
 PROCNO 1

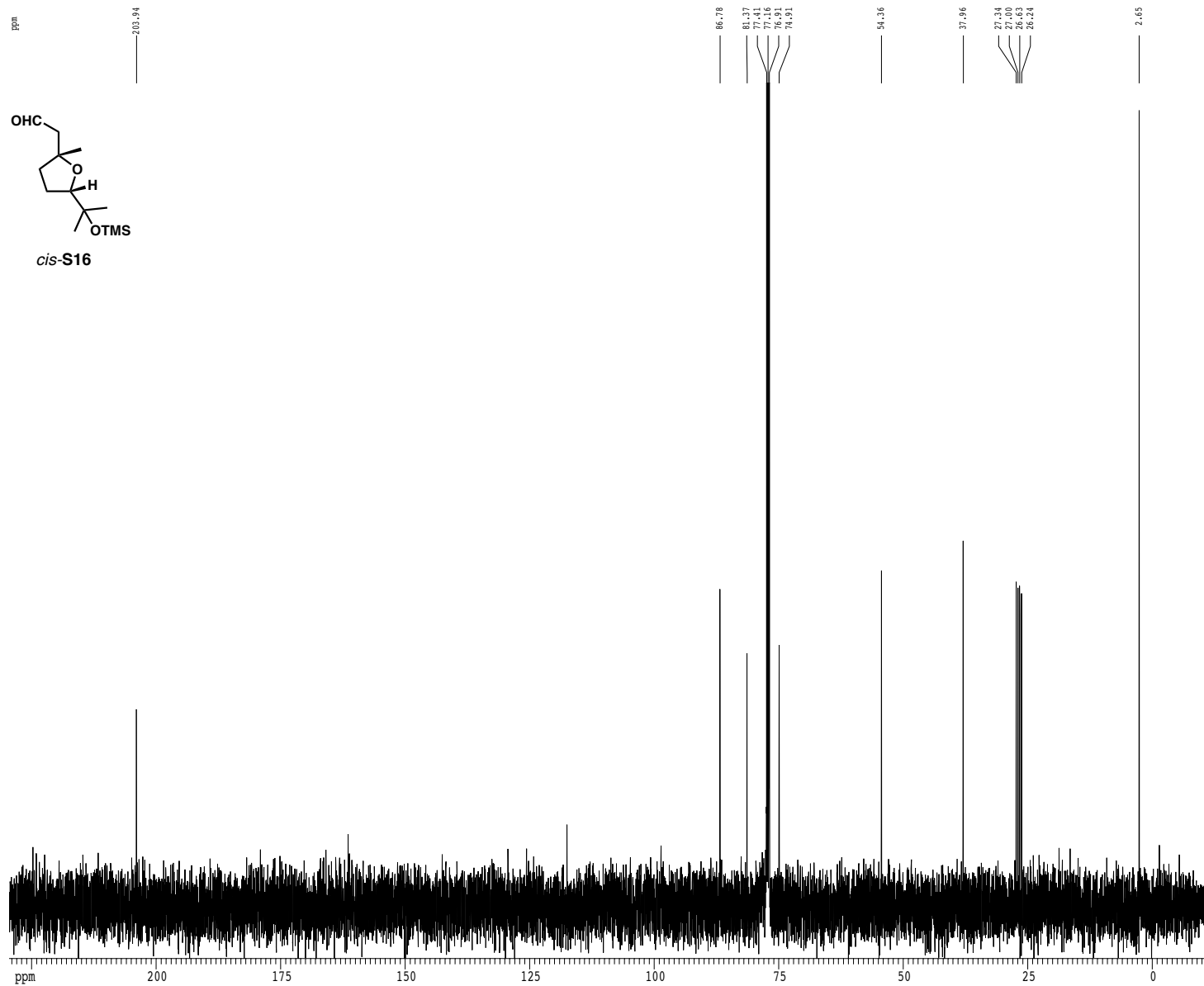
F2 - Acquisition Parameters
 Date_ 20151021
 Time 22.19
 INSTRUM gn500
 PROBHD 5 mm broadband
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3T
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 181
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 -5.80 dB
 SF01 499.1834943 MHz

F2 - Processing parameters
 SI 65536
 SF 499.1800271 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 FC 1.00

1D NMR plot parameters
 CX 22.80 cm
 CY 50.00 cm
 FIP 10.100 ppm
 F1 5041.72 Hz
 FZP -0.100 ppm
 F2 -49.92 Hz
 FWHM 0.44733 ppm/cm
 HZCN 223.31740 Hz/cm

13C spectrum with 1H decoupling



```

Current Data Parameters
USER          medaub
NAME          MED-VI-270pu
EXPNO        2
PROCNO       1

F2 - Acquisition Parameters
Date_         20151021
Time          22.23
INSTRUM      qm500
PROBHD       5 mm broadband
PULPROG      zgpg30
TD            65536
SOLVENT      CDCl3
NS            520
DS            4
SWH           30303.031 Hz
FIDRES        0.462388 Hz
AQ            1.0813940 sec
RG            46341
DM            16.500 usec
DE            4.500 usec
TE            298.0 K
D1            0.25000000 sec
d11           0.03000000 sec
MCREST       0.00000000 sec
MCWRK        0.01500000 sec

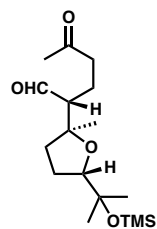
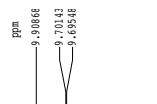
***** CHANNEL f1 *****
NUC1          13C
P1            9.00 usec
PL1           -0.60 dB
SFO1          125.5327181 MHz

***** CHANNEL f2 *****
CPDPRG2      waltz16
NUC2          1H
PCPD2        80.00 usec
PL2           -3.00 dB
PL12          12.80 dB
SFO2          499.1824959 MHz

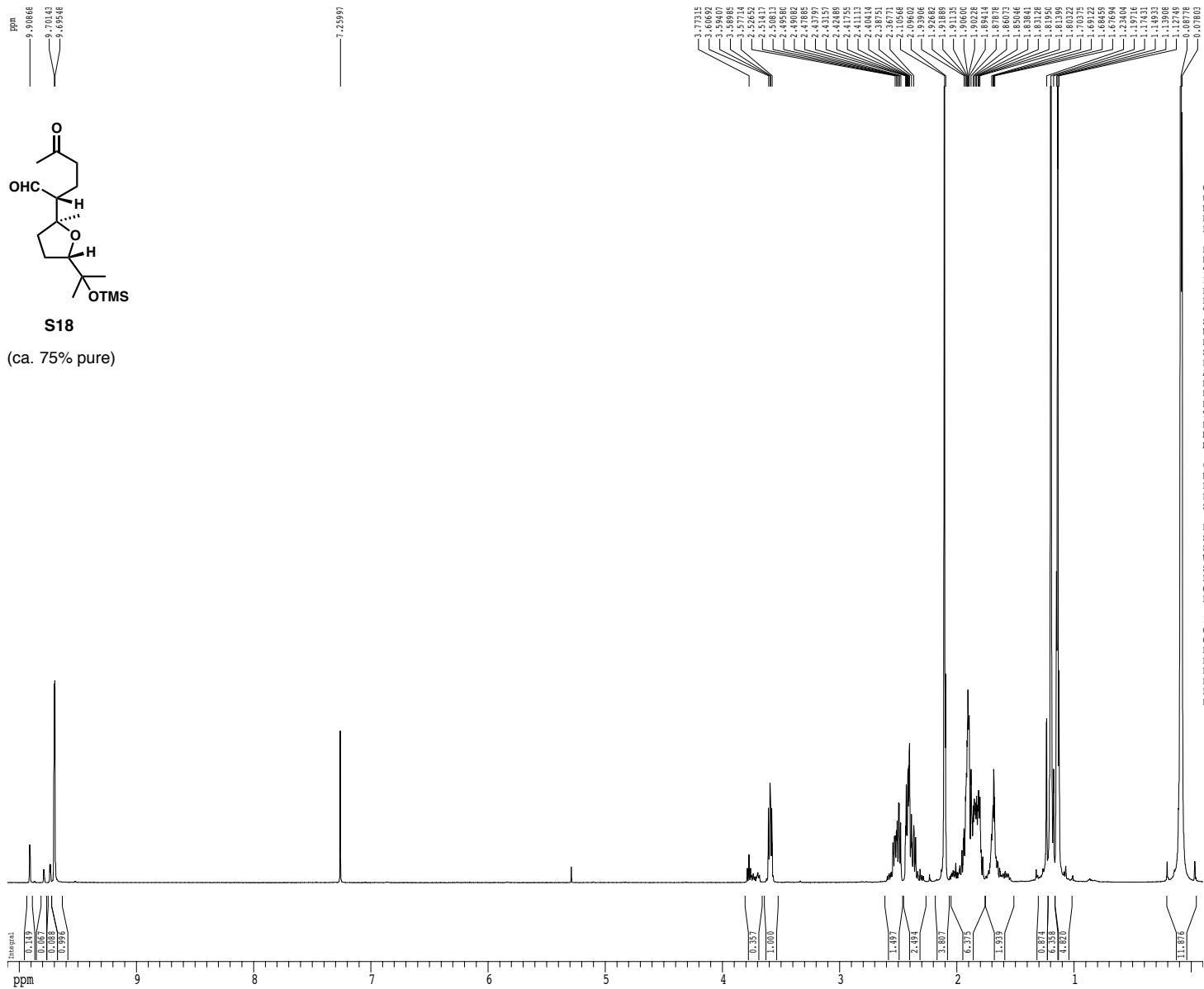
F2 - Processing parameters
SI            65536
SF            125.5188970 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            2.00

1D NMR plot parameters
CX            22.80 cm
CY            50.00 cm
P1P           229.520 ppm
F1            28809.06 Hz
F2            -10.507 ppm
F2P           -1318.78 Hz
PPMCM         10.52747 ppm/cm
HZCM          1321.39624 Hz/cm
    
```

1H spectrum



(ca. 75% pure)



```
Current Data Parameters
USER      medaub
NAME      MED-VI-233puB
EXPNO     1
PROCNO    1

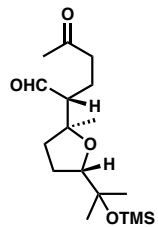
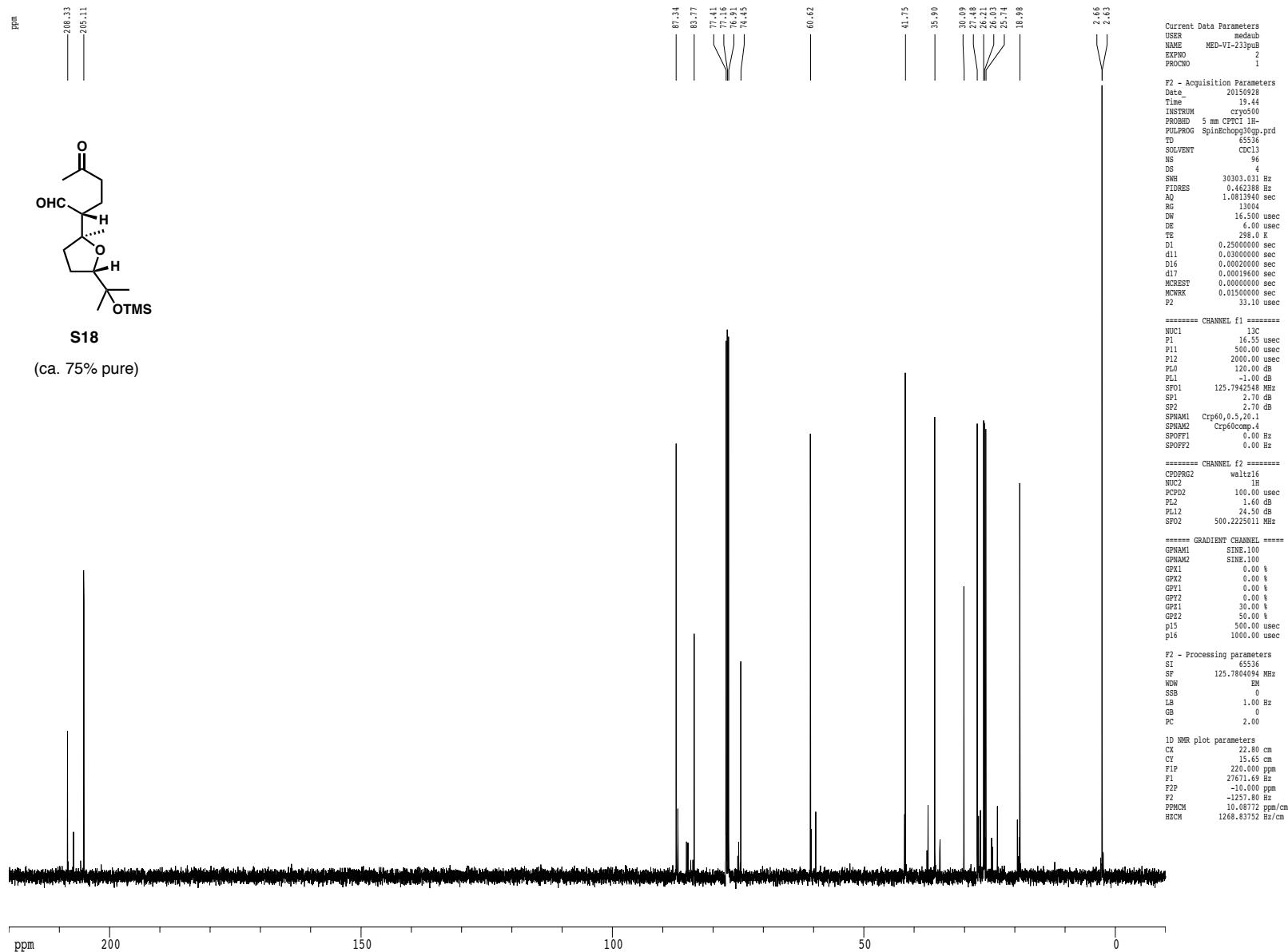
F2 - Acquisition Parameters
Date_     20150928
Time      19.41
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         81728
SOLVENT   CDCl3
NS         8
DS         2
SWH        8012.620 Hz
FIDRES     0.098043 Hz
AQ         5.0998774 sec
RG         5.7
DW         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
MCREST    0.00000000 sec
MWRK      0.01500000 sec

===== CHANNEL f1 =====
NUC1       1H
P1         7.50 usec
PI1        1.60 dB
SFO1       500.2235015 MHz

F2 - Processing parameters
SI         65536
SF         500.2200311 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00

1D NMR plot parameters
CX         22.80 cm
CY         40.00 cm
PIP        10.100 ppm
F1         5052.22 Hz
F2P        -0.100 ppm
F2         -50.02 Hz
PPHMC      0.444737 ppm/cm
HZCM       223.78267 Hz/cm
```

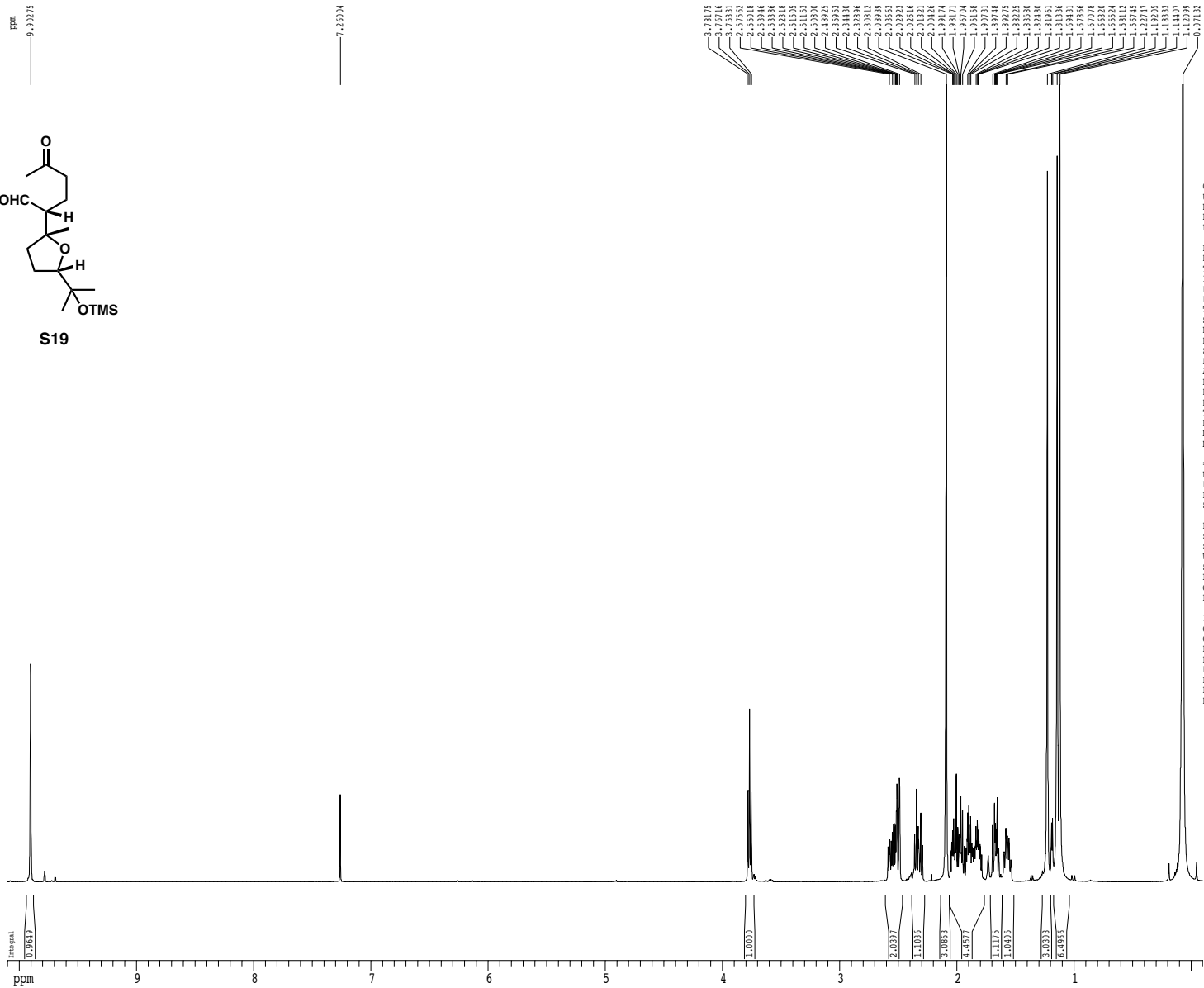
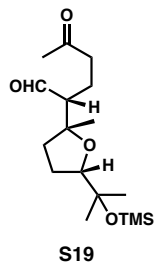
Z-restored spin-echo 13C spectrum with 1H decoupling



S18

(ca. 75% pure)

1H spectrum



```

Current Data Parameters
USER      medaub
NAME      MED-VI-233puA
EXPNO     1
PROCNO    1

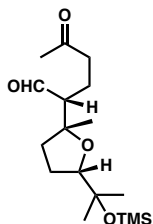
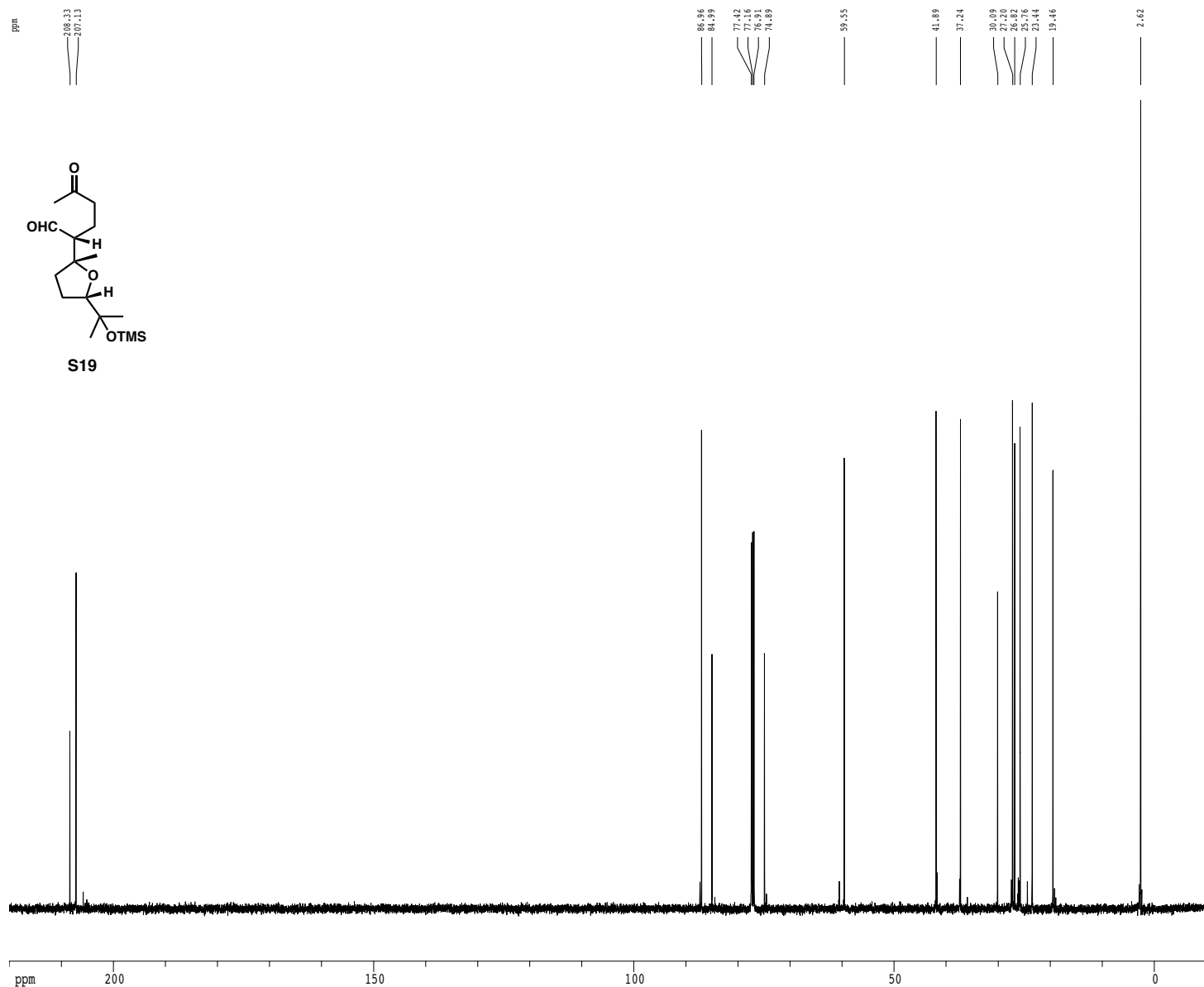
F2 - Acquisition Parameters
Date_     20150928
Time      19.22
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   zg30
TD         81728
SOLVENT   CDCl3
NS         8
DS         2
SWH        8012.620 Hz
FIDRES     0.098043 Hz
AQ         5.0998774 sec
RG         4
DW         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
MCREST    0.00000000 sec
MCWRR     0.01500000 sec

===== CHANNEL f1 =====
NUC1       1H
P1         7.50 usec
PI1        1.60 dB
SFO1       500.2235015 MHz

F2 - Processing parameters
SI         65536
SF         500.2200311 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00

1D NMR plot parameters
CX         22.80 cm
CY         30.00 cm
PI1        10.100 ppm
F1         5052.22 Hz
F2P        -0.100 ppm
F2         -50.02 Hz
PPHMC     0.44737 ppm/cm
HZCM      223.78267 Hz/cm
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



S19

```

Current Data Parameters
USER      medaub
NAME      MED-VI-233pub
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20150928
Time      19.25
INSTRUM   cryo500
PROBHD    5 mm CPCCI 1H-
PULPROG   SpinEcho30ap-gpd
TD         65536
SOLVENT   CDCl3
NS         128
DS         4
SNR        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         13004
DM         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
D16        0.00020000 sec
d17        0.00019600 sec
MCWREST    8.00000000 sec
MCWFK      0.01500000 sec
P2         33.10 usec

===== CHANNEL f1 =====
NUC1       13C
P1         16.55 usec
P11        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1       125.7942548 MHz
SF1        2.70 dB
SP2        2.70 dB
SFOAM1     Crp60,0.5,20.1
SFOAM2     Crp60comp.4
SFOFF1     0.00 Hz
SFOFF2     0.00 Hz

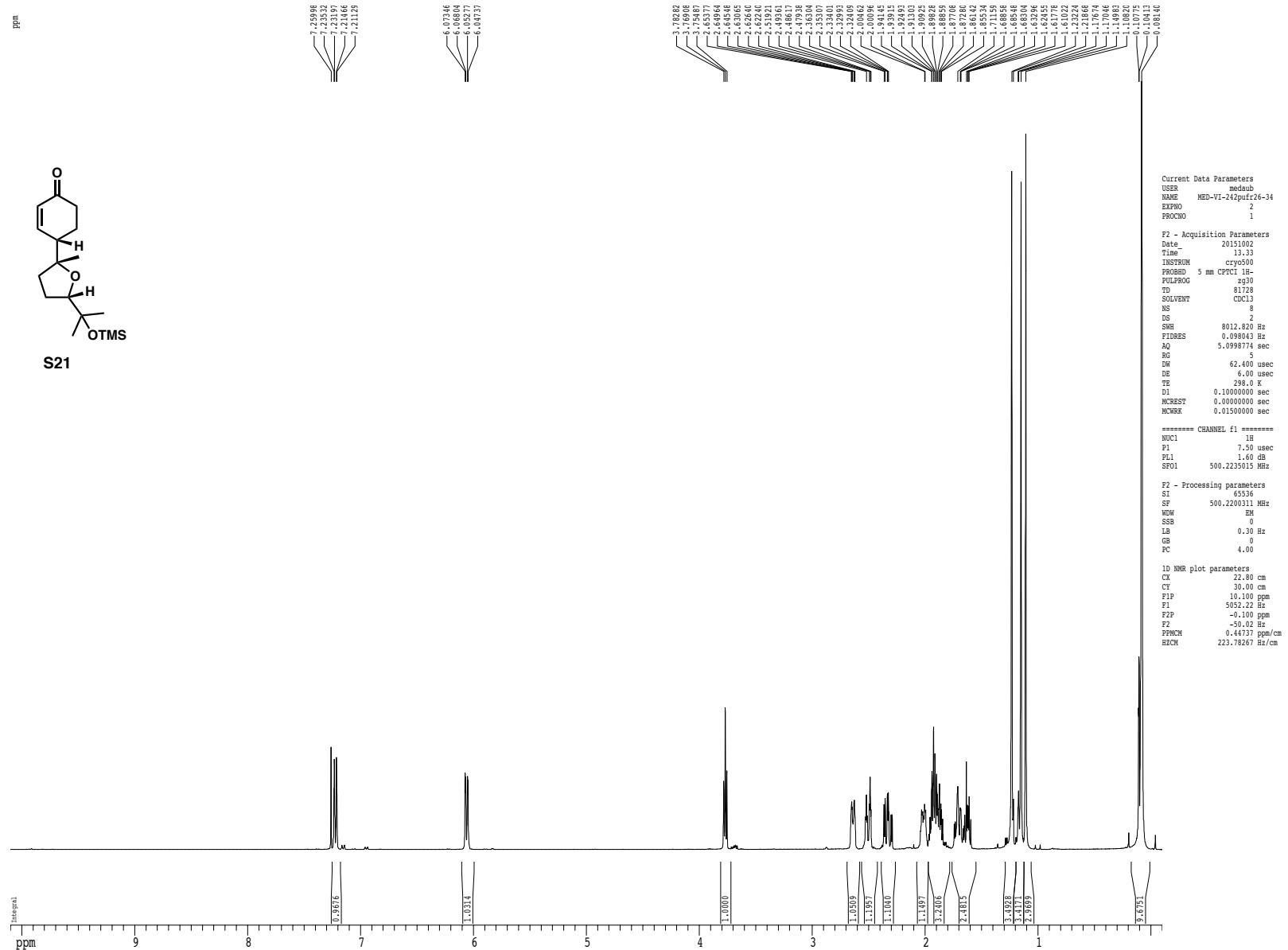
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2      100.00 usec
PL2        1.60 dB
PL12       24.50 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPMAM1     SINE.100
GPMAM2     SINE.100
GPX1       0.00 t
GPX2       0.00 t
GPY1       0.00 t
GPY2       0.00 t
GPZ1       30.00 t
GPZ2       50.00 t
p15        500.00 usec
p16        1000.00 usec

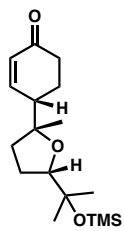
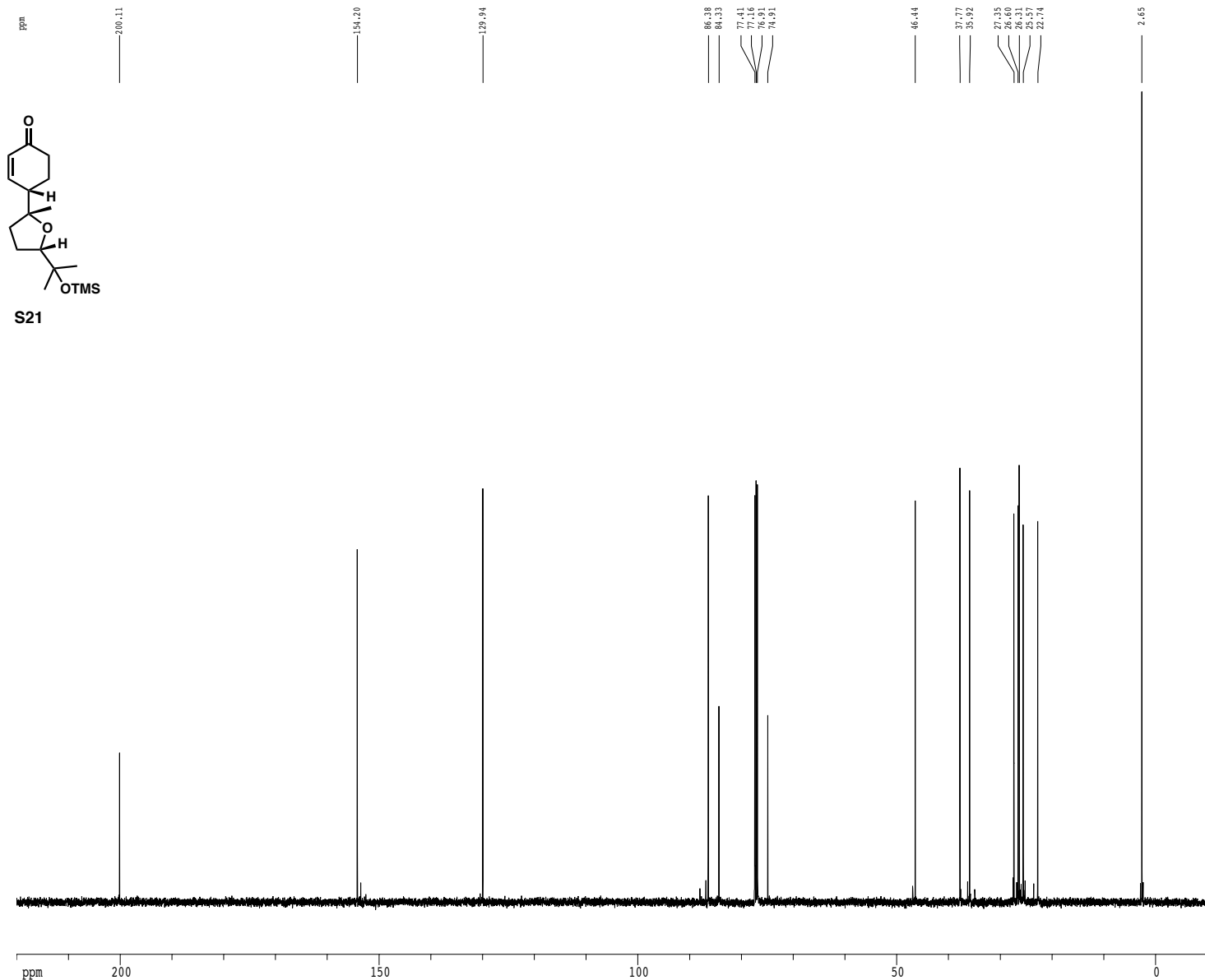
F2 - Processing parameters
SI         65536
SF         125.7804099 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

1D NMR plot parameters
CX         22.80 cm
CT         15.65 cm
PLP        220.000 ppm
F1         27671.69 Hz
F2P        -10.000 ppm
F2         -1257.80 Hz
FPMCM      10.48772 ppm/cm
HRCM       1268.83752 Hz/cm
    
```

1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling



S21

```

Current Data Parameters
USER      medaub
NAME      MED-VI-242ptr26-34
EXPNO     3
PROCNO    1

F2 - Acquisition Parameters
Date_     20151002
Time      13.37
INSTRUM   cryo500
PROBHD    5 mm CPCT 1H-
PULPROG   SpinEcho30pp.prd
TD         65536
SOLVENT   CDCl3
NS         208
DS         4
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         13004
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
D16        0.00020000 sec
d17        0.00019600 sec
MCHEST    0.00000000 sec
MCHRX     0.01500000 sec
F2         31.10 usec

===== CHANNEL f1 =====
NUC1       13C
P1         16.55 usec
P11        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SF01       125.7942548 MHz
SP1        2.70 dB
SP2        2.70 dB
SFOFF1     0.00 Hz
SFOFF2     0.00 Hz

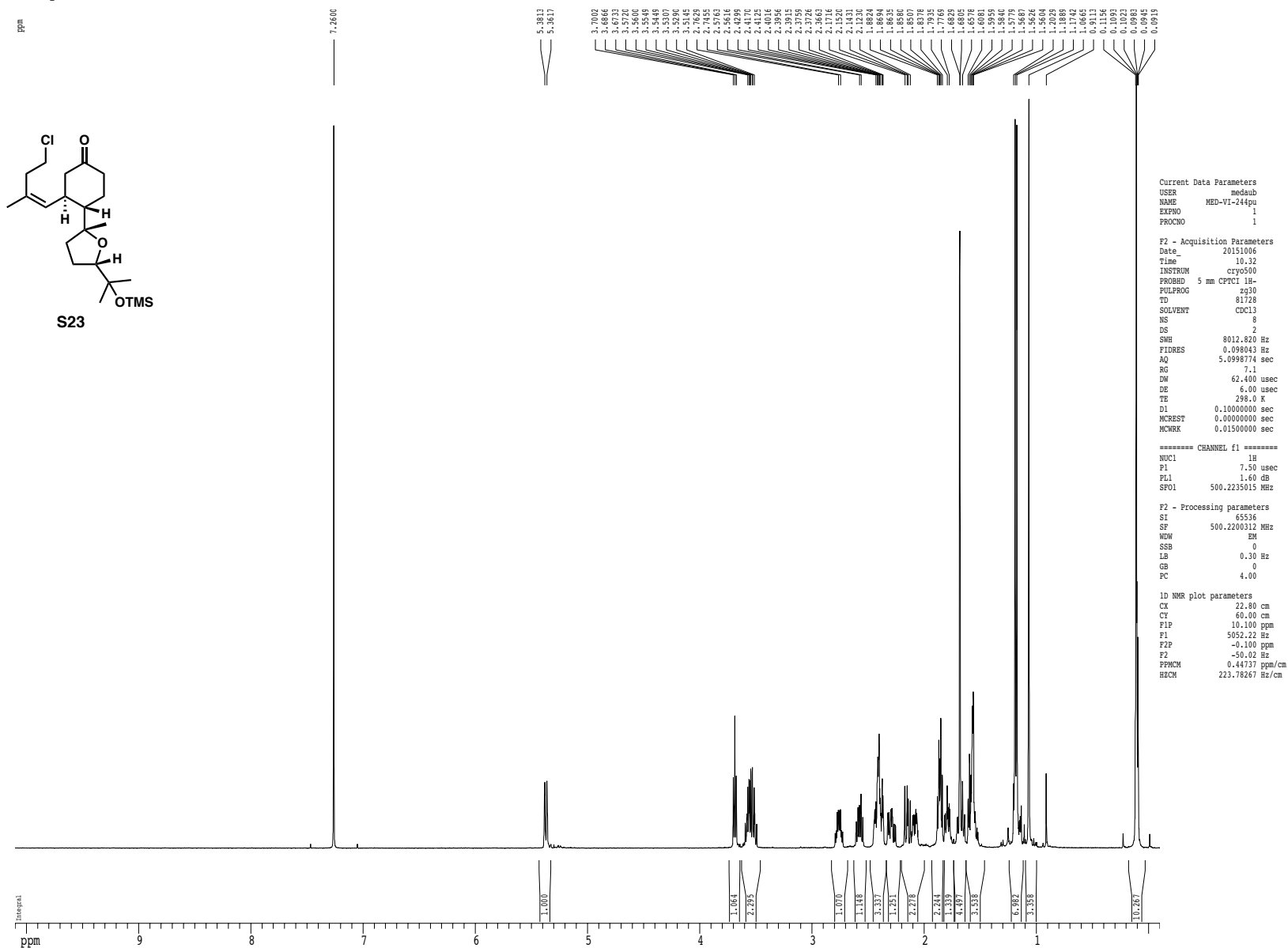
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2      100.00 usec
PL2        1.60 dB
PL12       24.50 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPMAM1     SINE.100
GPMAM2     SINE.100
GPX1       0.00 %
GPX2       0.00 %
GPY1       0.00 %
GPY2       0.00 %
GPZ1       30.00 %
GPZ2       50.00 %
p15        500.00 usec
p16        1000.00 usec

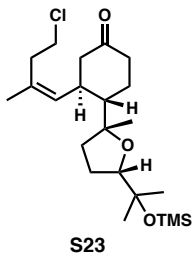
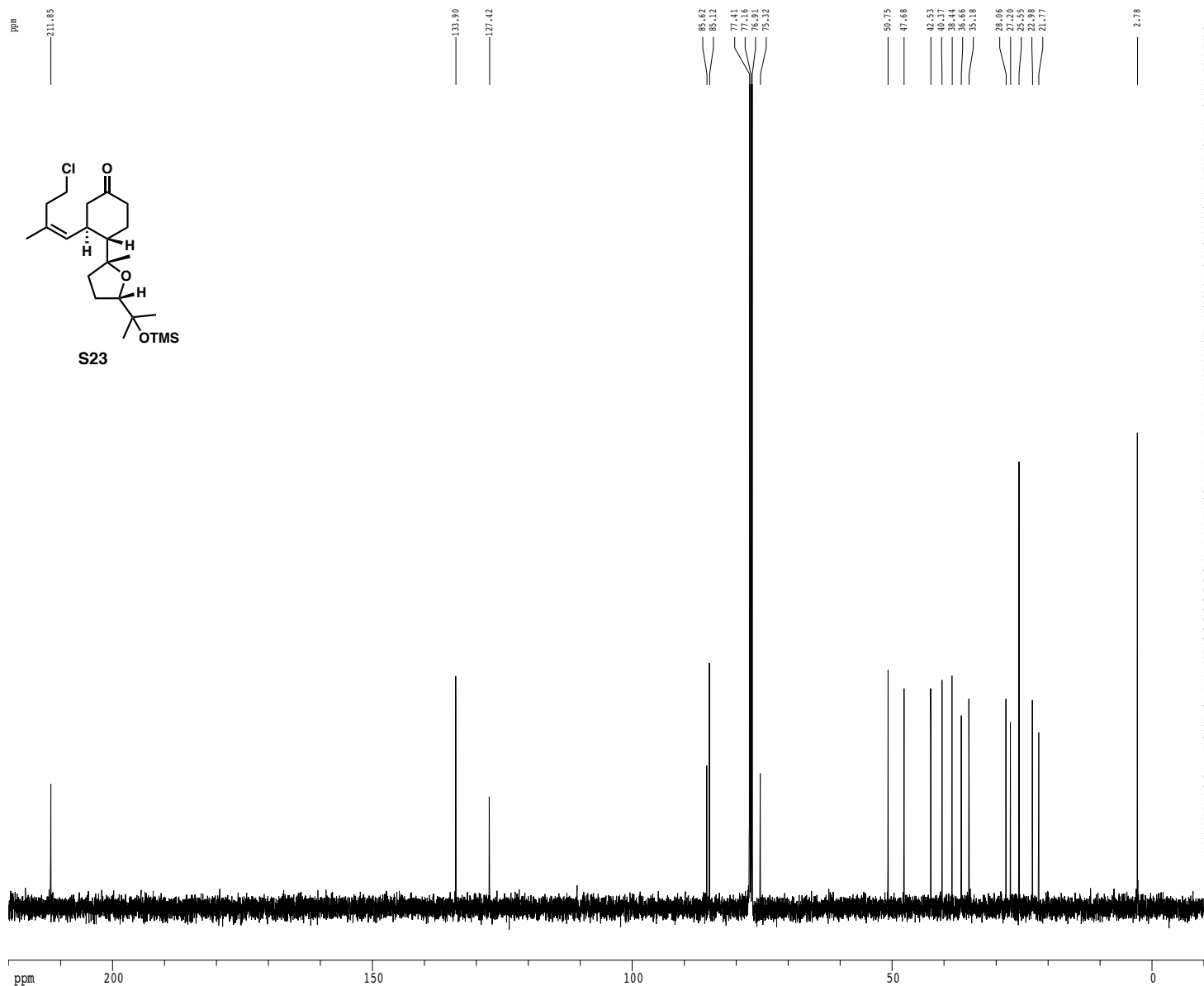
F2 - Processing parameters
SI         65536
SF         125.7804094 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

1D NMR plot parameters
CX         22.80 cm
CT         15.65 cm
F1P        220.000 ppm
F1         27671.69 Hz
F2P        -10.000 ppm
F2         -1257.80 Hz
PPMCM      10.08772 ppm/cm
HCM        1268.83752 Hz/cm
    
```

1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER      medaub
NAME      MED-VI-244pu
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20151006
Time      10.36
INSTRUM   cryo500
PROBHD    5 mm CP1C1 1H-
PULPROG   SpinEchoSgDg-prd
TD         65536
SOLVENT   CDCl3
NS         392
DS         4
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         13004
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
d16        0.00020000 sec
d17        0.00019600 sec
MCHEST    0.00000000 sec
MCMKX     0.01500000 sec
P2         33.10 usec

===== CHANNEL f1 =====
NUC1       13C
P1         16.55 usec
PL1        500.00 usec
PL2        2000.00 usec
PL0        120.00 dB
PG1        -1.00 dB
SFO1       125.7942548 MHz
SP1        2.70 dB
SP2        2.70 dB
SFOAM1     Crp60,0.5,20.1
SFOAM2     crp60comp,4
SFOFF1     0.00 Hz
SFOFF2     0.00 Hz

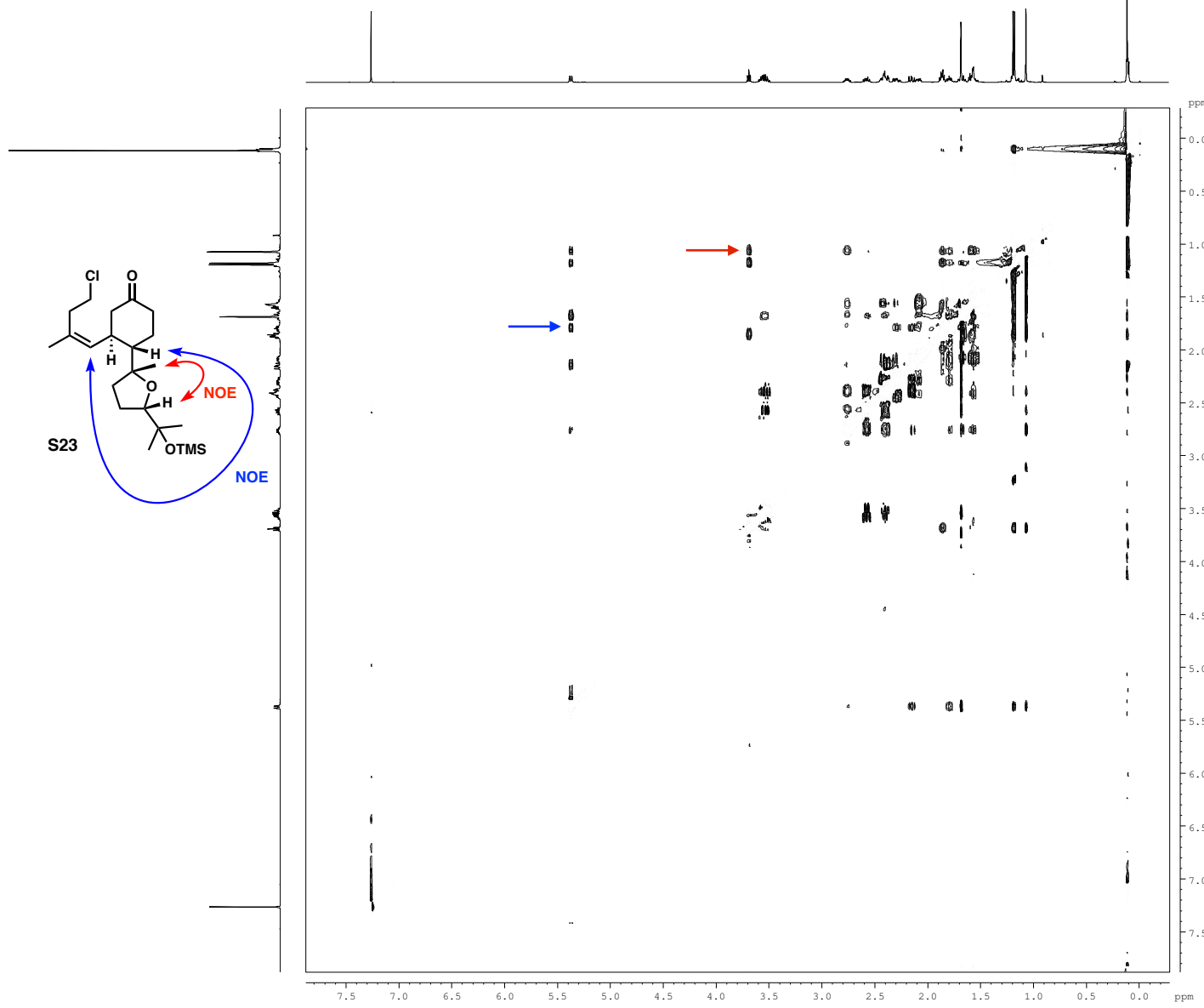
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2     100.00 usec
PL2        1.60 dB
PL12       24.50 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1     SINE.100
GPNAM2     SINE.100
GPF1       0.00 %
GPF2       0.00 %
GPF1       0.00 %
GPF2       0.00 %
GPF1       30.00 %
GPF2       50.00 %
p15        500.00 usec
p16        1000.00 usec

F2 - Processing parameters
SI         65536
SF         125.7804076 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

1D NMR plot parameters
CX         22.80 cm
CI         30.00 cm
FIP        220.000 ppm
F1         27671.69 Hz
F2P        -10.000 ppm
F2         -1257.80 Hz
F2MCH      10.48772 ppm/cm
H2CM       1268.83752 Hz/cm
    
```

gnoesy



```
Current Data Parameters
NAME      MED-VI-244pu
EXPNO     3
PROCNO    1

F2 - Acquisition Parameters
Date_     20151006
Time      10.43
INSTRUM   cryo400
PROBHD    5 mm CPTCI 1H-
PULPROG   noesygpp
TD         2048
SOLVENT   CDCl3
NS         2
DS         4
SWH        4084.967 Hz
FIDRES     1.994613 Hz
AQ         0.2506752 sec
RG         128
DW         122.400 usec
DE         6.00 usec
TE         298.0 K
D0         0.00000300 sec
D1         2.00000000 sec
D8         0.80000001 sec
D16        0.00020000 sec
d20        0.39880002 sec
IN0        0.00012240 sec

----- CHANNEL f1 -----
NUC1       1H
F1         7.50 usec
P2         15.00 usec
PL1        1.60 dB
SF01       500.2219300 MHz

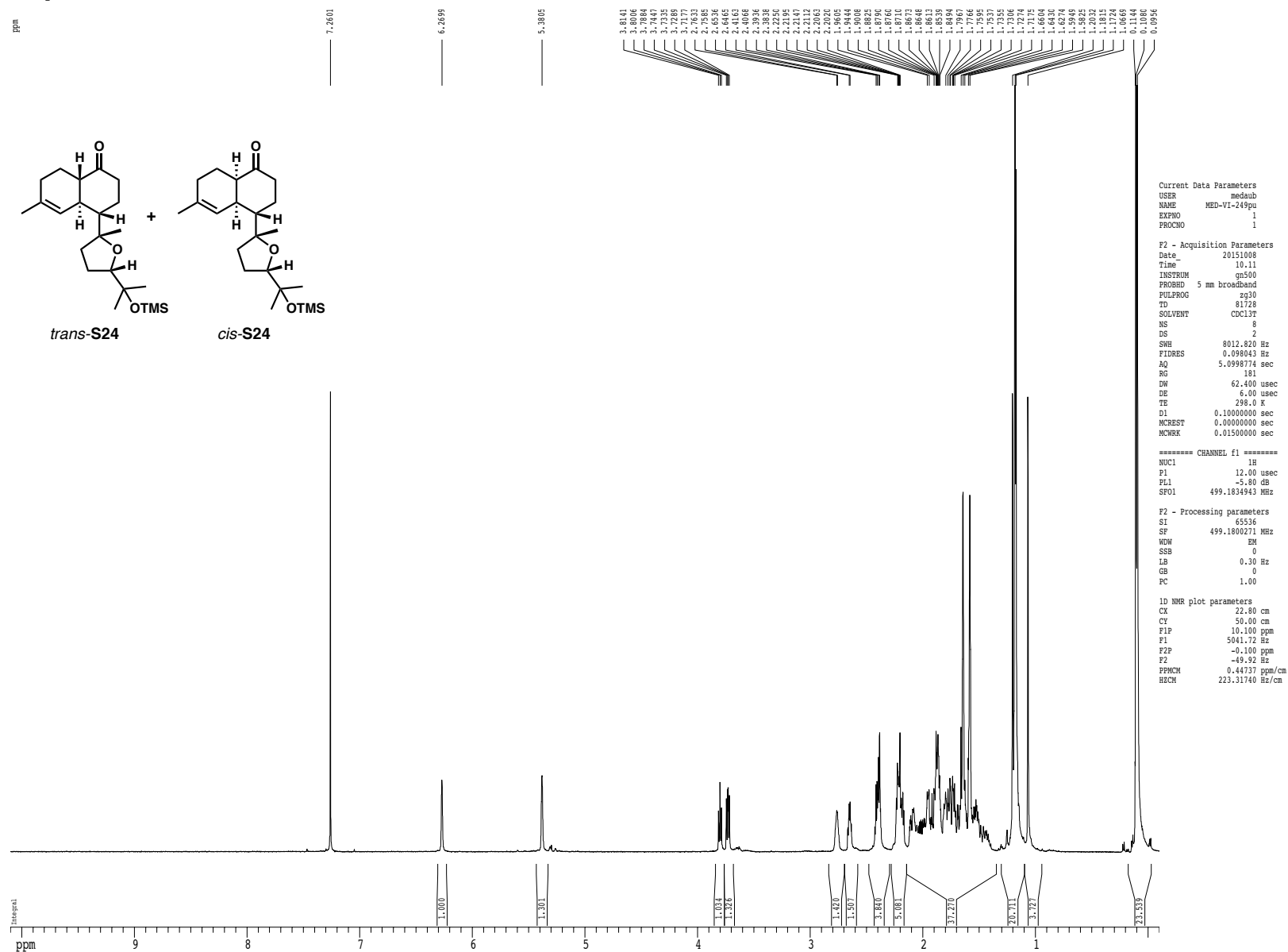
----- GRADIENT CHANNEL -----
GPNAM[1]   sine.100
GPNAM[2]   sine.100
GFX1       0 %
GFX2       0 %
GFY1       0 %
GFY2       0 %
GPZ1       40.00 %
GPZ2       -40.00 %
F16        1000.00 usec

F1 - Acquisition parameters
TD         145
SF01       500.2219 MHz
FIDRES     28.172188 Hz
SW         9.166 ppm
F0MODE     undefined

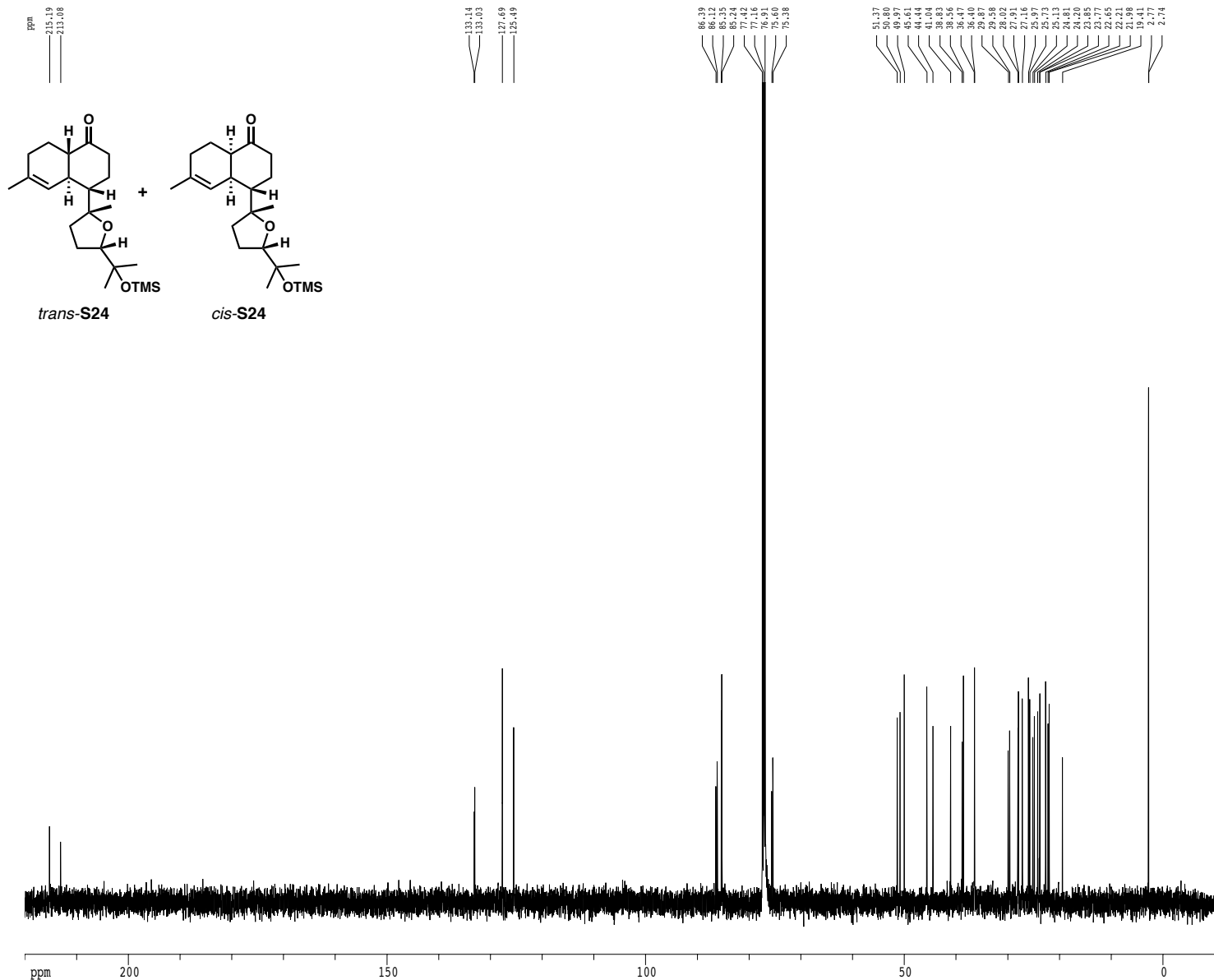
F2 - Processing parameters
SI         1024
SF         500.2200312 MHz
WDW        QSINE
SSB        2
LB         0 Hz
GB         0
PC         2.00

F1 - Processing parameters
SI         1024
MC2        TPPI
SF         500.2200312 MHz
WDW        QSINE
SSB        2
LB         0 Hz
GB         0
```

1H spectrum



13C spectrum with 1H decoupling



```

Current Data Parameters
USER      medaub
NAME      MED-VI-249pu
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20151008
Time      10.15
INSTRUM   qm500
PROBHD    5 mm broadband
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         1024
DS         4
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         2896.3
DM         16.500 usec
DE         4.500 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
MCREST     0.00000000 sec
MCWRK      0.01500000 sec

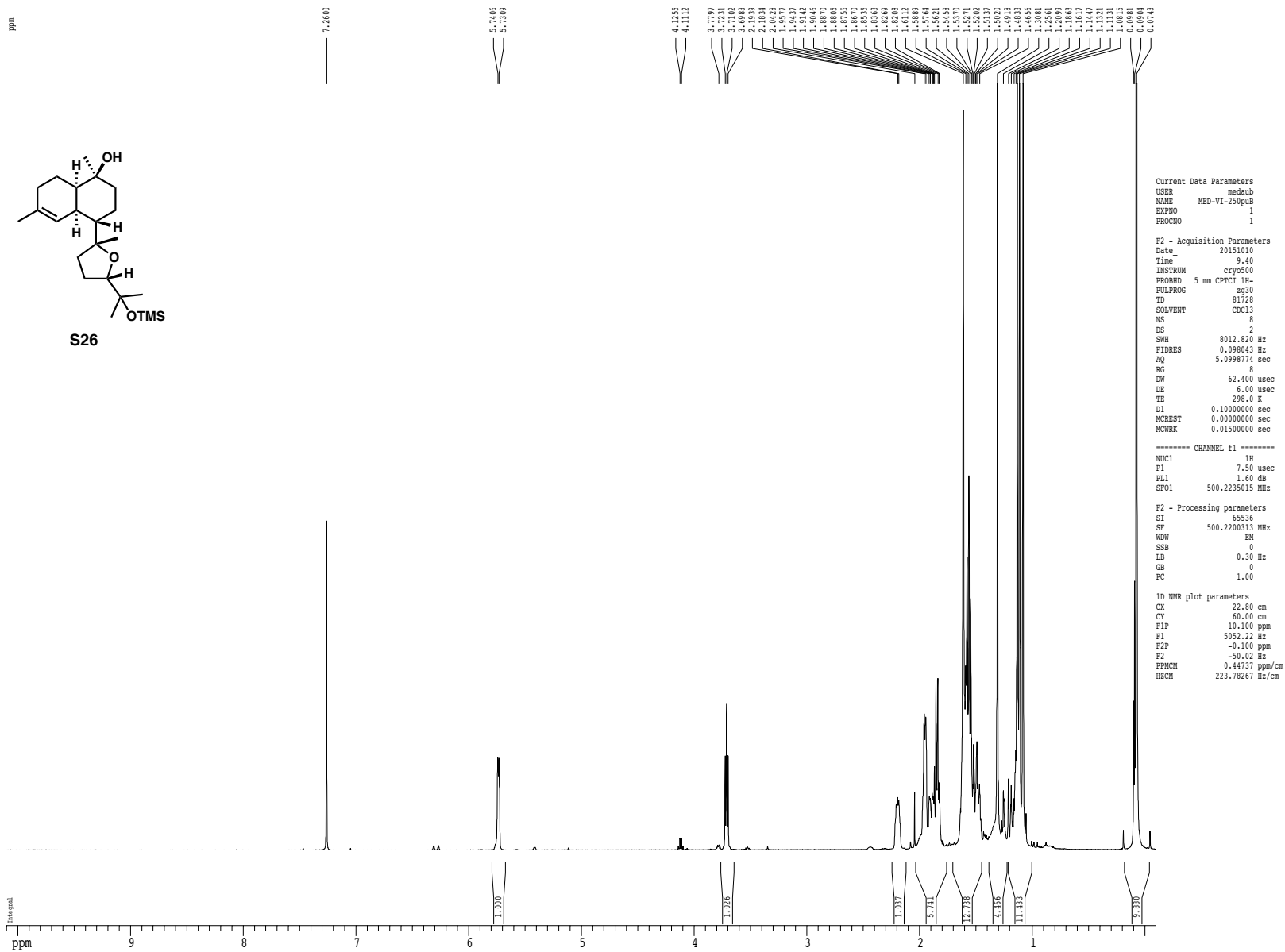
***** CHANNEL f1 *****
NUC1       13C
P1         9.00 usec
PL1        -0.60 dB
SF01       125.5327181 MHz

***** CHANNEL f2 *****
CPDPRG2    waltz16
NUC2       1H
PCPD2      80.00 usec
PL2        -3.00 dB
PL12       12.80 dB
SF02       499.1824959 MHz

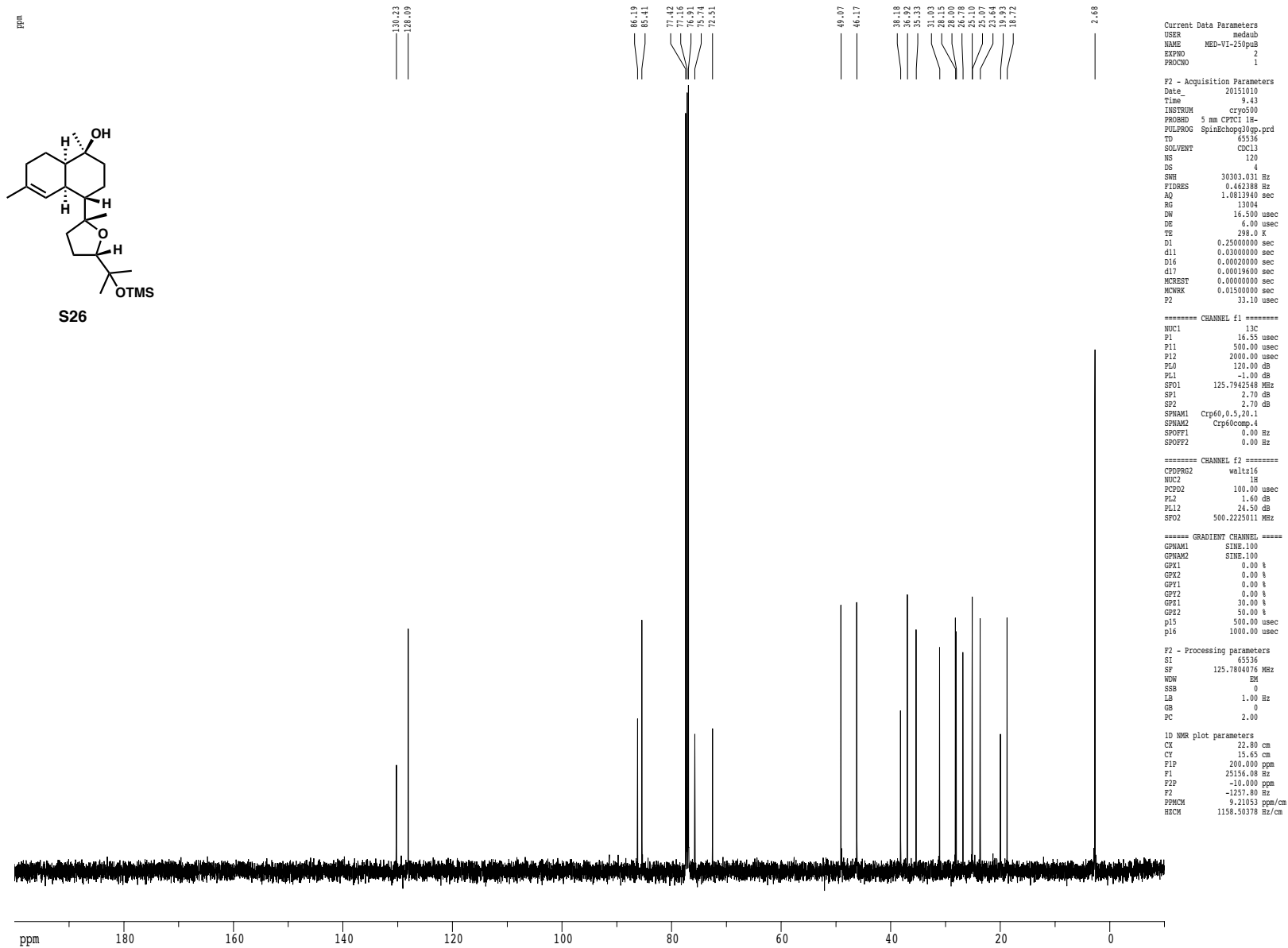
F2 - Processing parameters
SI         65536
SF         125.5188970 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

1D NMR plot parameters
CX         22.80 cm
CY         30.00 cm
F1P        220.000 ppm
F1         27614.16 Hz
F2P        -10.000 ppm
F2         -1255.19 Hz
PPMCM      10.08712 ppm/cm
HZCM       1266.19946 Hz/cm
    
```

1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER      medaub
NAME      MED-VI-250puB
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20151010
Time      9.43
INSTRUM   cryo500
PROBHD    5 mm CPCCI 1H-
PULPROG   SpinEcho30ap.prd
TD         65536
SOLVENT   CDCl3
NS         120
DS         4
SNR        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         13004
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
D16        0.00020000 sec
d17        0.00019600 sec
MCKWST     0.00000000 sec
MCKWRK     0.01500000 sec
P2         33.10 usec

===== CHANNEL f1 =====
NUC1       13C
P1         16.55 usec
P11        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1       125.7942548 MHz
SF1        2.70 dB
SP2        2.70 dB
SFOAM1     Crp60,0.5,20.1
SFOAM2     Crp60comp.4
SFOFF1     0.00 Hz
SFOFF2     0.00 Hz

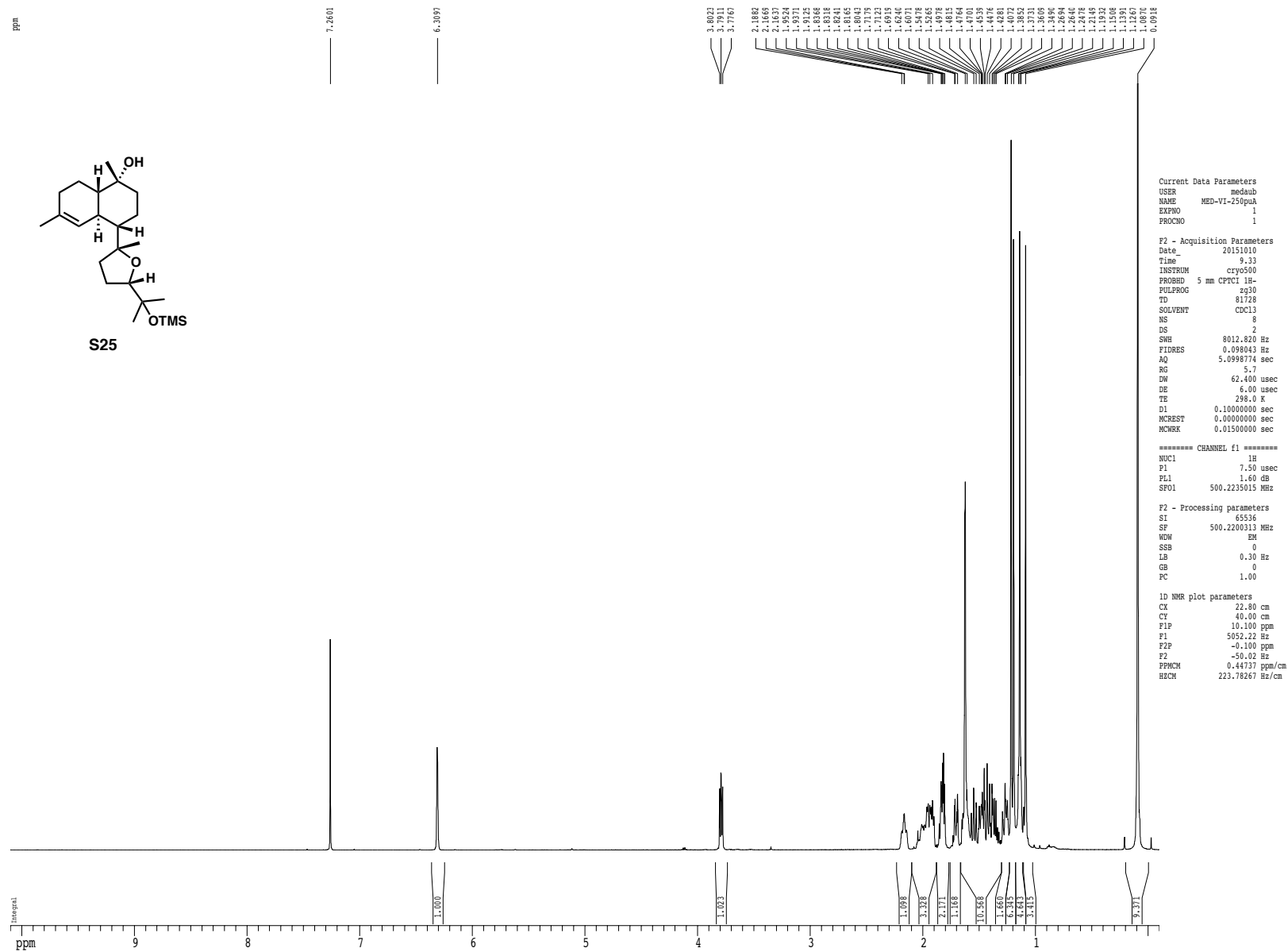
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2      100.00 usec
PL2        1.00 dB
PL12       24.50 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPMAM1     SINE.100
GPMAM2     SINE.100
GPF1       0.00 t
GPF2       0.00 t
GPF1       0.00 t
GPF2       0.00 t
GPF1       30.00 t
GPF2       30.00 t
p15        500.00 usec
p16        1000.00 usec

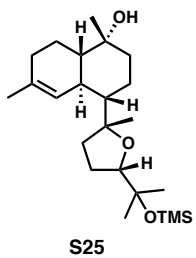
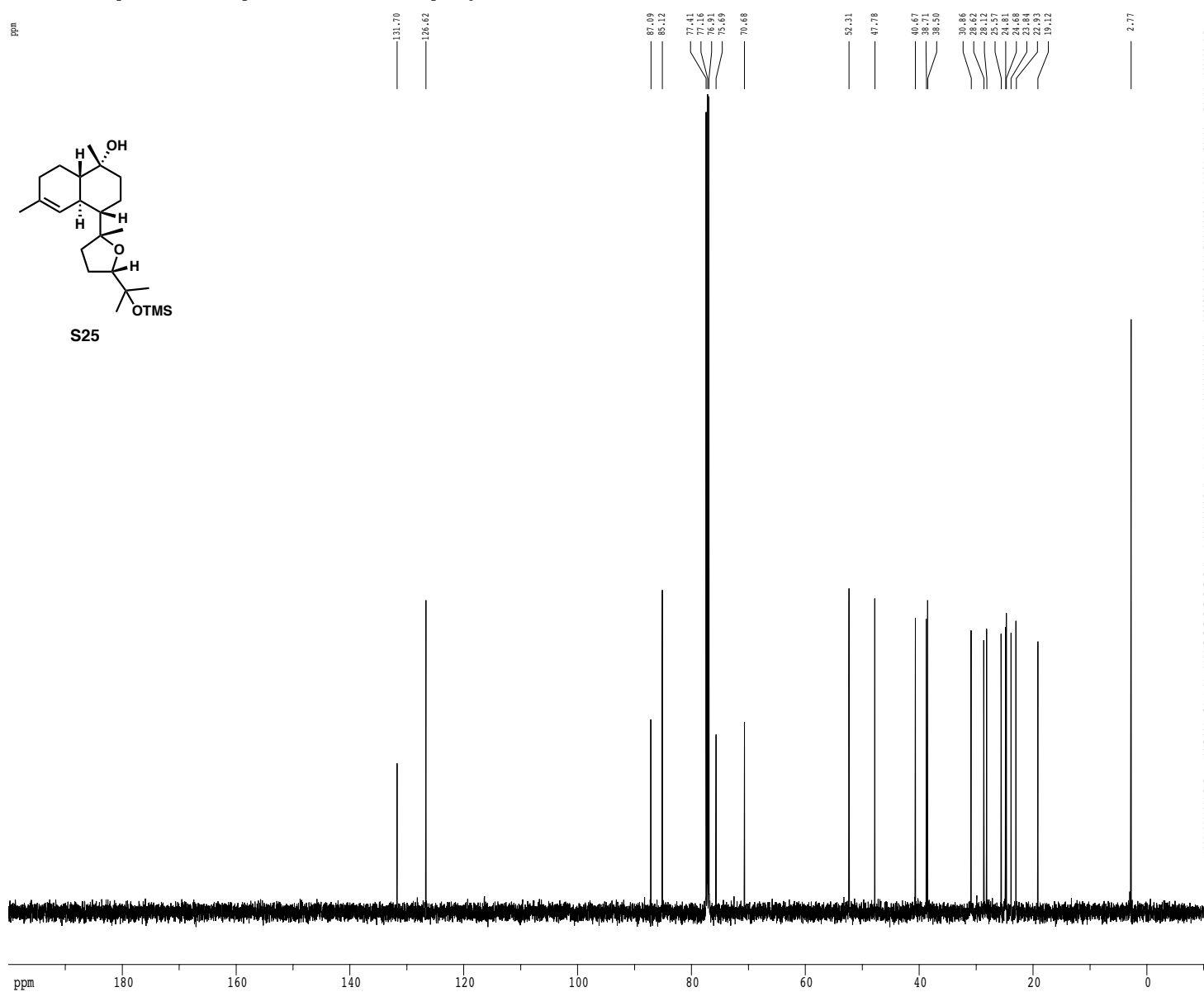
F2 - Processing parameters
SI         65536
SF         125.7804076 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

1D NMR plot parameters
CX         22.80 cm
CT         15.65 cm
PLP        200.000 ppm
P1         25156.08 Hz
F2P        -10.000 ppm
FZ         -1257.80 Hz
PPHMC      9.21053 ppm/cm
HRCM       1158.50378 Hz/cm
    
```


1H spectrum



Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER      medaub
NAME      MED-VI-250µA
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20151010
Time      9.35
INSTRUM   cryo500
PROBHD    5 mm CP1C1 1H-
PULPROG   SpinEchoSgppg-prd
TD         65536
SOLVENT   CDCl3
NS         139
DS         4
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         13004
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
d16        0.00020000 sec
d17        0.00019600 sec
MCREST     0.00000000 sec
MCMRXX     0.01500000 sec
P2         33.10 usec

===== CHANNEL f1 =====
NUC1       13C
P1         16.55 usec
PL1        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1       125.7842548 MHz
SP1        2.70 dB
SP2        2.70 dB
SFOFF1     Crp60,0.5,20.1
SFOFF2     Crp60comp,4
SFOFF1     0.00 Hz
SFOFF2     0.00 Hz

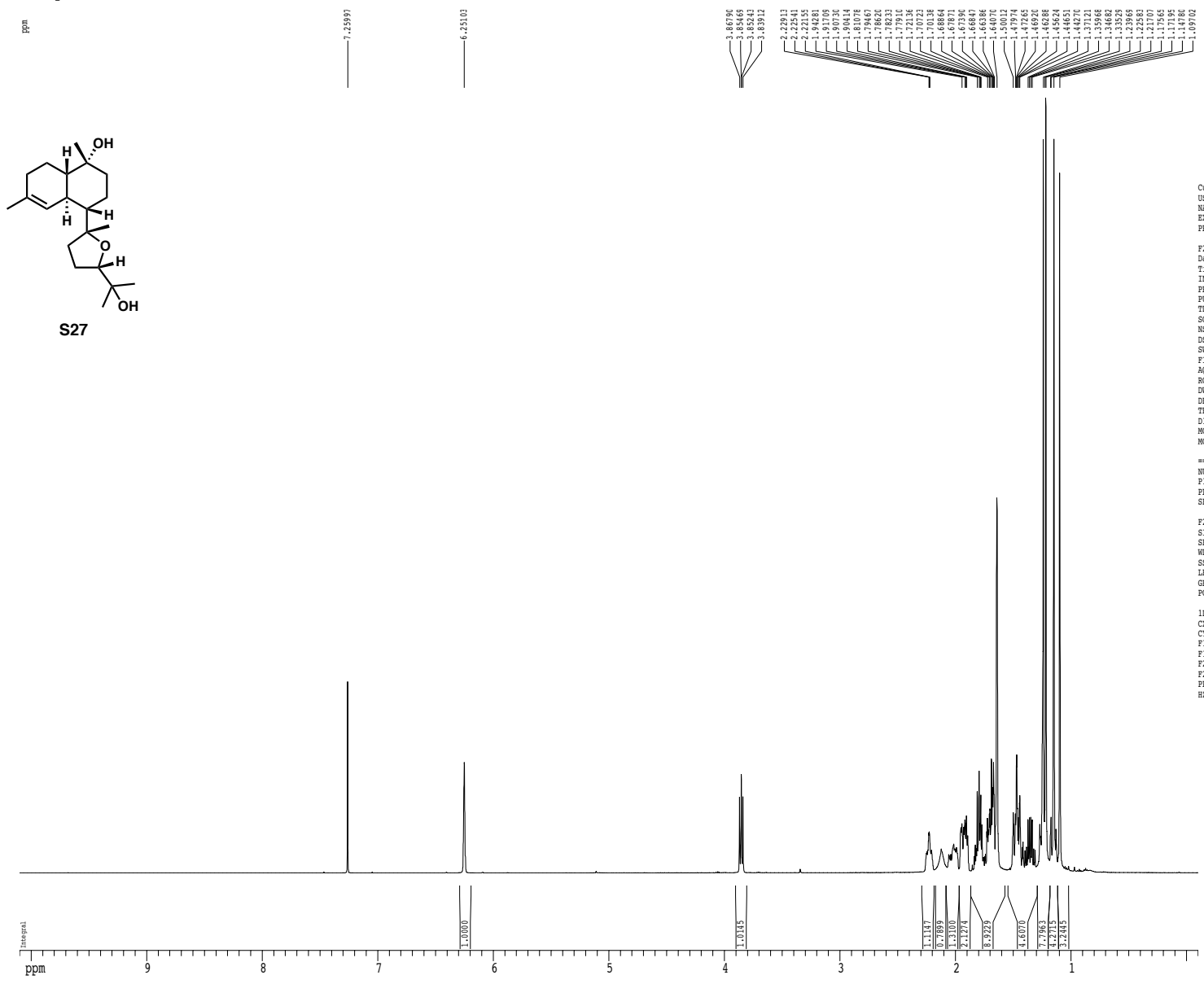
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2      100.00 usec
PL2        1.60 dB
PL12       24.50 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1     SINE.100
GPNAM2     SINE.100
GPF1       0.00 %
GPF2       0.00 %
GPF1       0.00 %
GPF2       0.00 %
GPF1       30.00 %
GPF2       50.00 %
p15        500.00 usec
p16        1000.00 usec

F2 - Processing parameters
SI         65536
SF         125.7804076 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

1D NMR plot parameters
CX         22.80 cm
CI         15.65 cm
FIP        200.000 ppm
F1         25156.08 Hz
F2P        -10.000 ppm
F2         -1257.80 Hz
F2MCH      9.21853 ppm/cm
H2CM       1158.50378 Hz/cm
    
```

1H spectrum



```

Current Data Parameters
USER      medaub
NAME      MED-VI-252pu
EXPNO     1
PROCNO    1

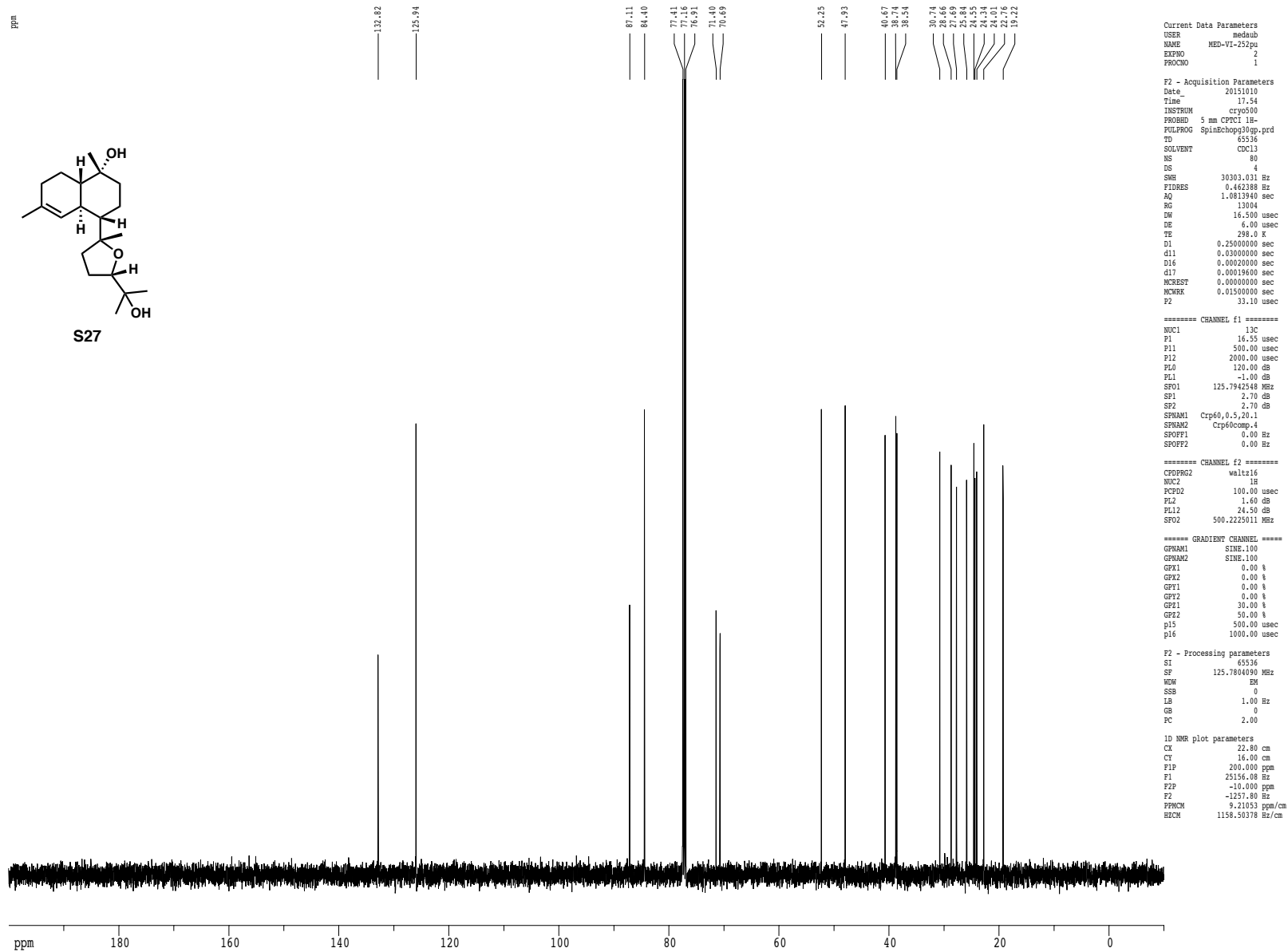
F2 - Acquisition Parameters
Date_     20151010
Time      17.52
INSTRUM   cryo500
PROBHD    5 mm CPYC1 H-
PULPROG   zg30
TD         81728
SOLVENT    CDCl3
NS         8
DS         2
SWH        8012.820 Hz
FIDRES     0.098043 Hz
AQ         5.0998774 sec
RG         5.7
DW         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
MCREST     0.00000000 sec
MCWRK     0.01500000 sec

===== CHANNEL f1 =====
NUC1       1H
P1         7.50 usec
PL1        1.60 dB
SFO1       500.2235015 MHz

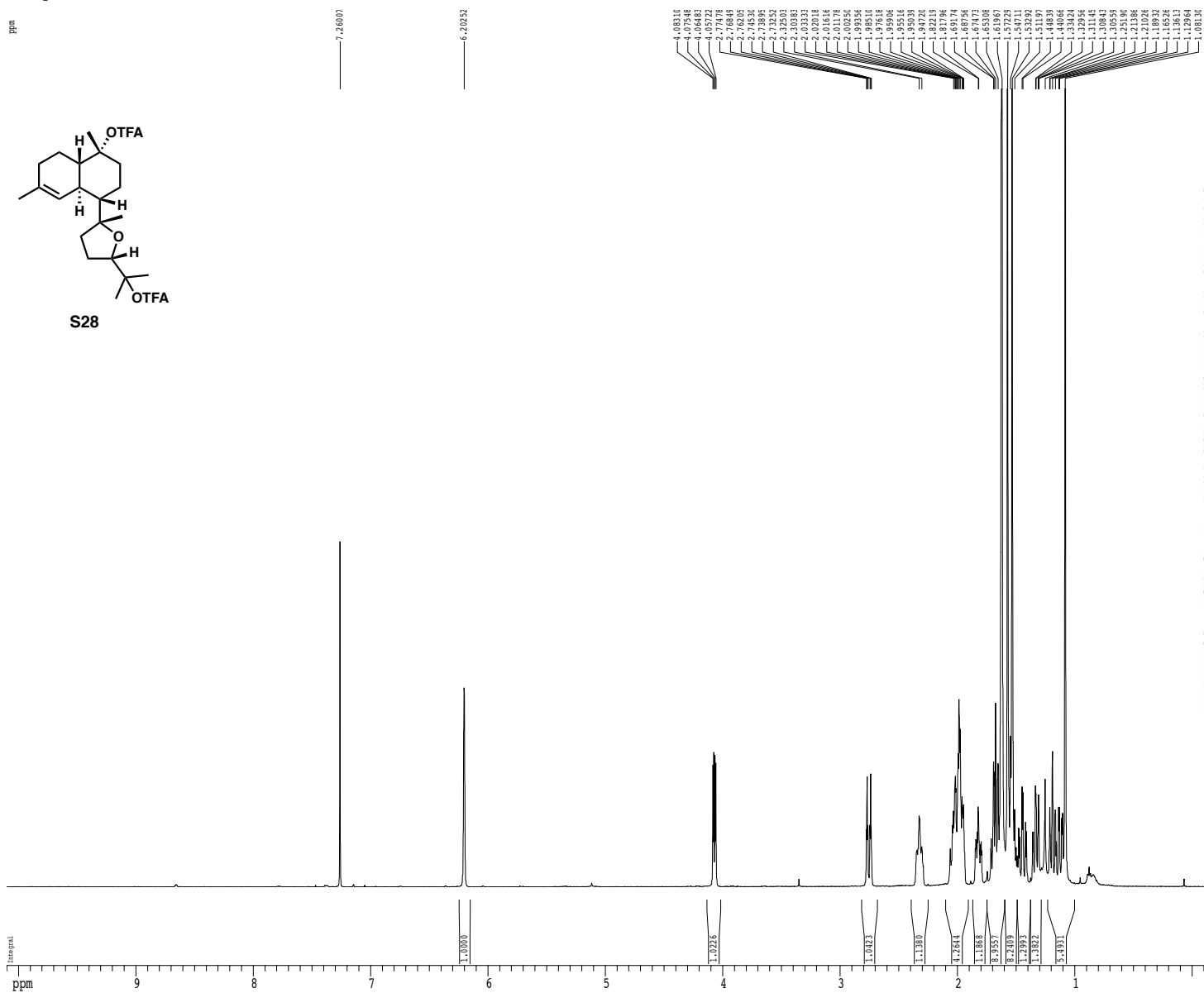
F2 - Processing parameters
SI         65536
SF         500.2200313 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00

1D NMR plot parameters
CX         22.80 cm
CY         15.00 cm
F1P        10.100 ppm
F1         5052.22 Hz
F2P        -0.100 ppm
F2         -50.02 Hz
PPMCHM    0.44737 ppm/cm
HZCM      223.78267 Hz/cm
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



1H spectrum



Current Data Parameters
 USER medaub
 NAME MED-VI-262cr
 EXPNO 1
 PROCNO 1

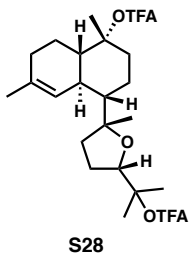
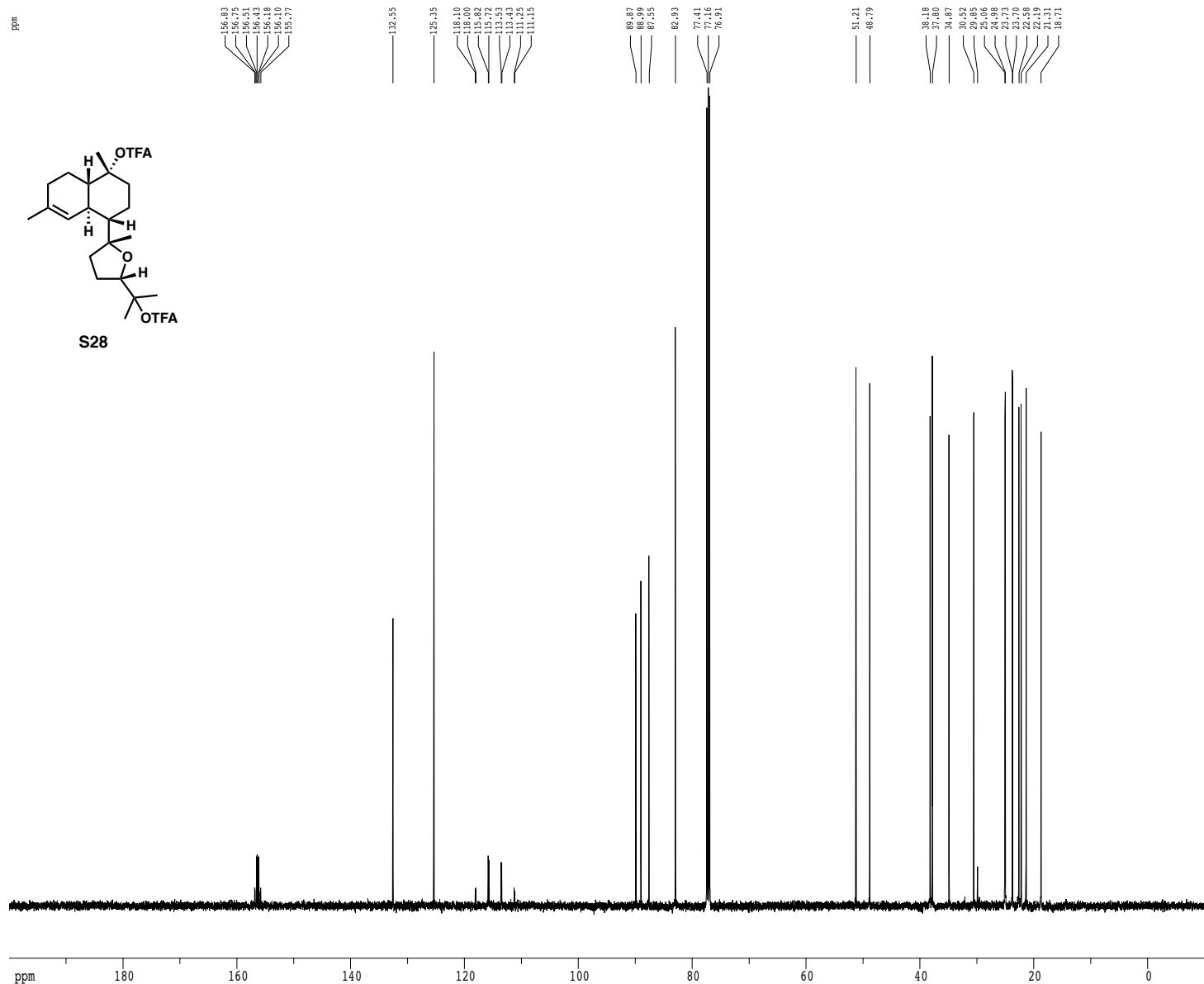
F2 - Acquisition Parameters
 Date_ 20151017
 Time 13.02
 INSTRUM cryo500
 PROBRD 5 mm CPTCI 1H-
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3
 NS 8
 DS 2
 SHF 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 5.7
 DW 62.400 usec
 DE 4.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCNRX 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.60 db
 SF01 500.2235015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2209313 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.80 cm
 CY 30.00 cm
 F1P 10.100 ppm
 F1 5052.22 Hz
 F2P -0.100 ppm
 F2 -50.00 Hz
 FPMCM 0.44737 ppm/cm
 HCM 223.78267 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER          medaub
NAME          MED-VI-262cr
EXPNO         2
PROCNO        1

F2 - Acquisition Parameters
Date_         20151017
Time          13.05
INSTRUM       cryo500
PROBHD        5 mm CPCCI 1H-
PULPROG       SpinEcho30ap.prd
TD            65536
SOLVENT       CDCl3
NS            944
DS            4
SRR           30303.031 Hz
FIDRES        0.462388 Hz
AQ            1.0813940 sec
RG            11585.2
DW            16.500 usec
DE            6.00 usec
TE            298.0 K
D1            0.25000000 sec
d11           0.03000000 sec
D16           0.00020000 sec
d17           0.00019600 sec
MCWREST       8.00000000 sec
MCWREK        0.01500000 sec
P2            33.10 usec

===== CHANNEL f1 =====
NUC1           13C
P1             16.55 usec
P11            500.00 usec
P12            2000.00 usec
PL0            120.00 dB
PL1            -1.00 dB
SFO1          125.7942548 MHz
SF1            2.70 dB
SP2            2.70 dB
SFOAM1        Crp60,0.5,20.1
SFOAM2        Crp60comp.4
SFOFF1         0.00 Hz
SFOFF2         0.00 Hz

===== CHANNEL f2 =====
CPDPRG2       waltz16
NUC2           1H
PCPD2         100.00 usec
PL2            1.00 dB
PL12           24.50 dB
SFO2          500.2225011 MHz

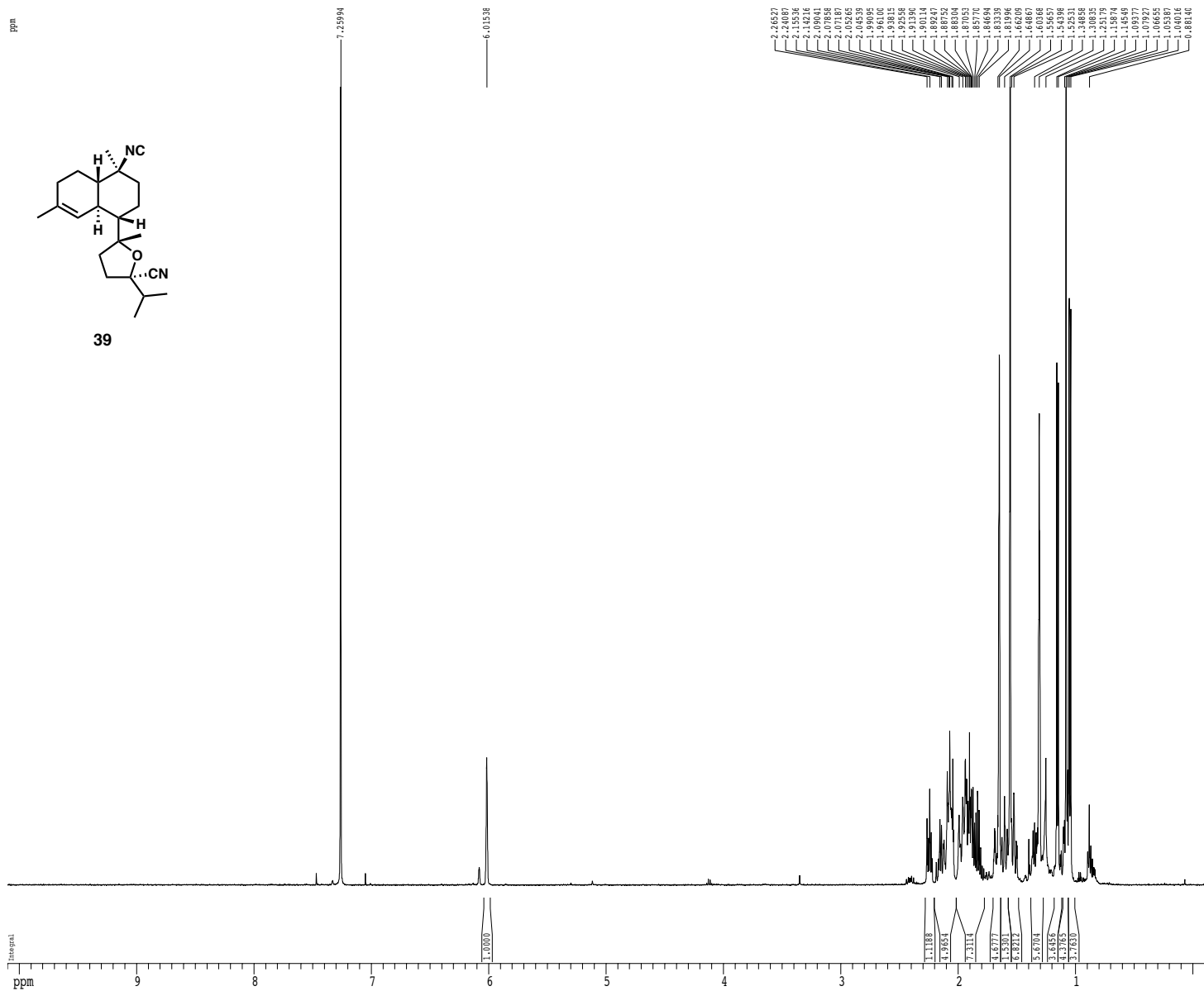
===== GRADIENT CHANNEL =====
GPMAM1        SINE.100
GPMAM2        SINE.100
GPF1           0.00 t
GPF2           0.00 t
GPF1           0.00 t
GPF2           0.00 t
GPF1           30.00 t
GPF2           50.00 t
p15            500.00 usec
p16            1000.00 usec

F2 - Processing parameters
SI            65536
SF            125.7804071 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            2.00

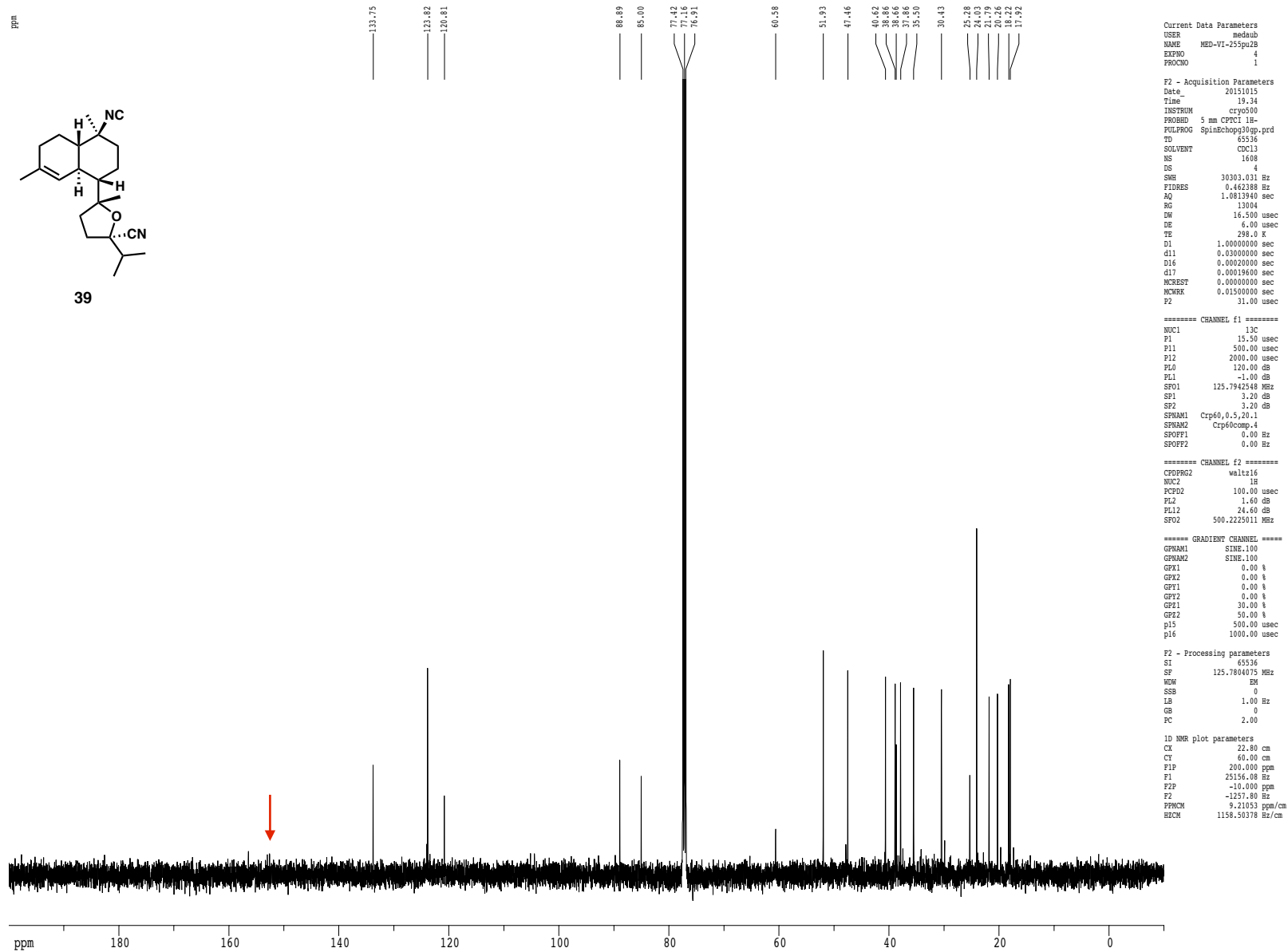
1D NMR plot parameters
CX            22.80 cm
CT            15.65 cm
PLP           200.000 ppm
P1            25156.08 Hz
F2P           -10.000 ppm
FZ            -1257.80 Hz
PPHMC         9.21053 ppm/cm
HRCM          1158.50378 Hz/cm
    
```

1H spectrum

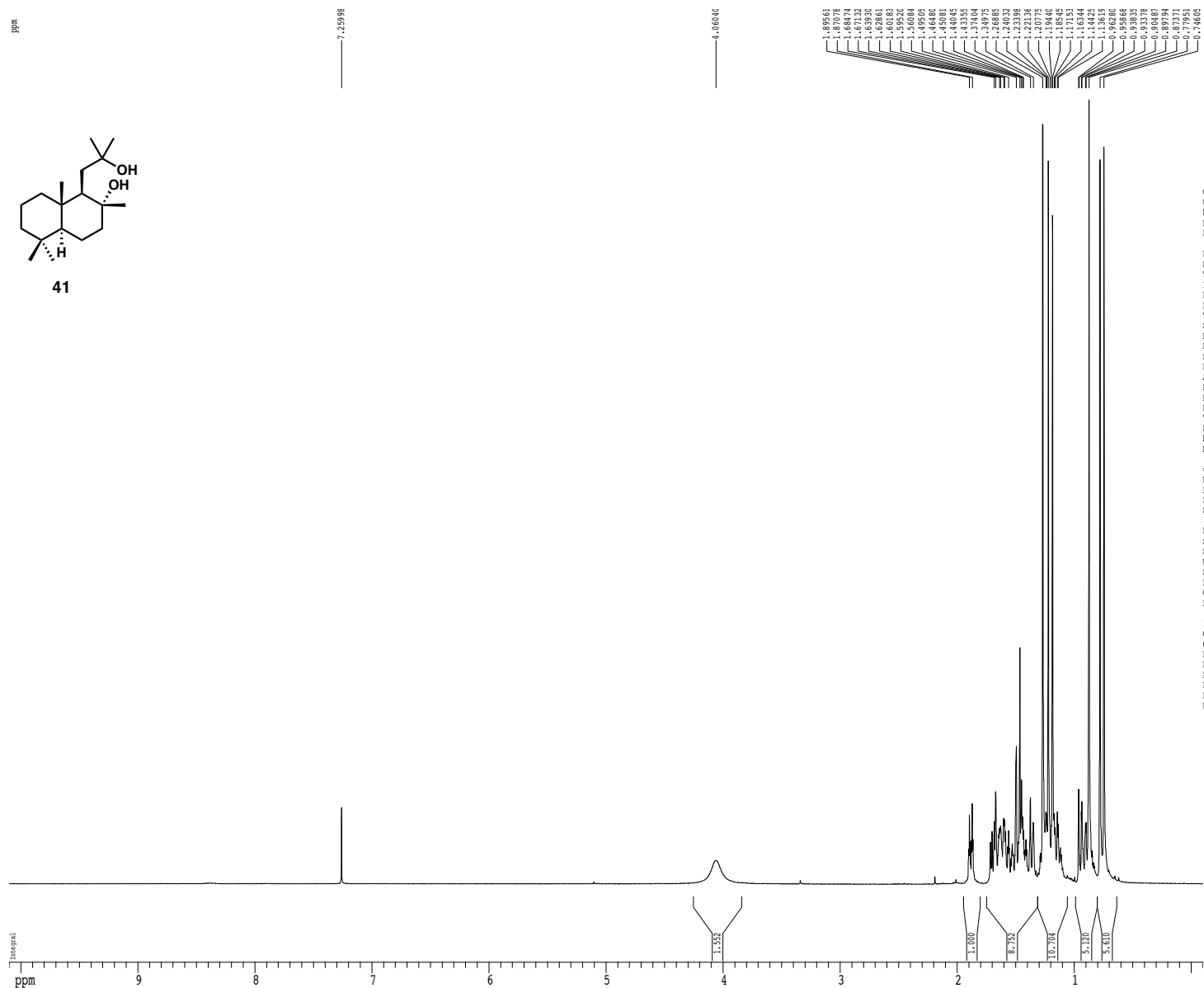
ppm



Z-restored spin-echo 13C spectrum with 1H decoupling

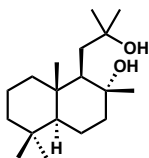


1H spectrum

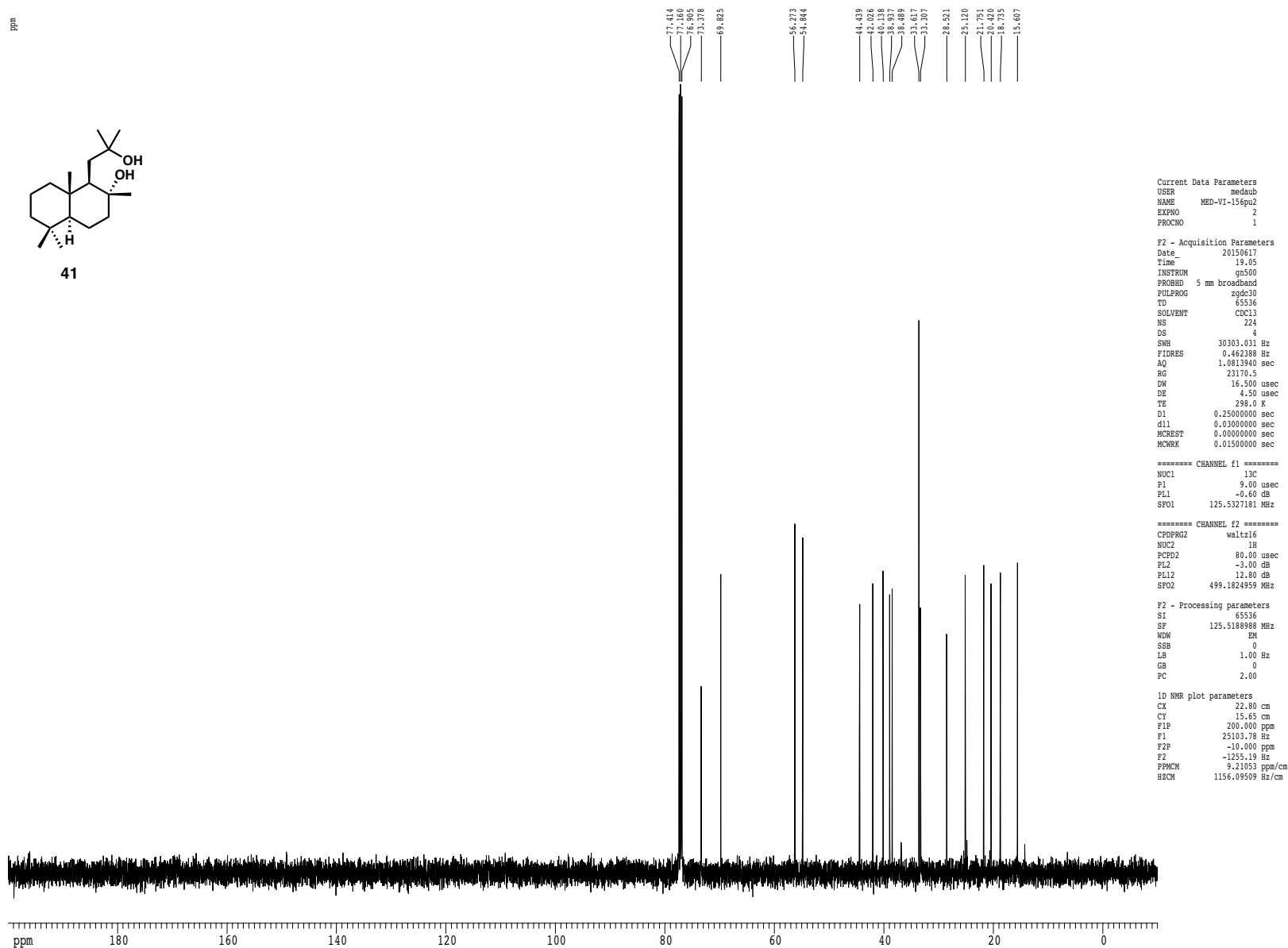


13C spectrum with 1H decoupling

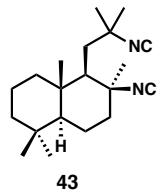
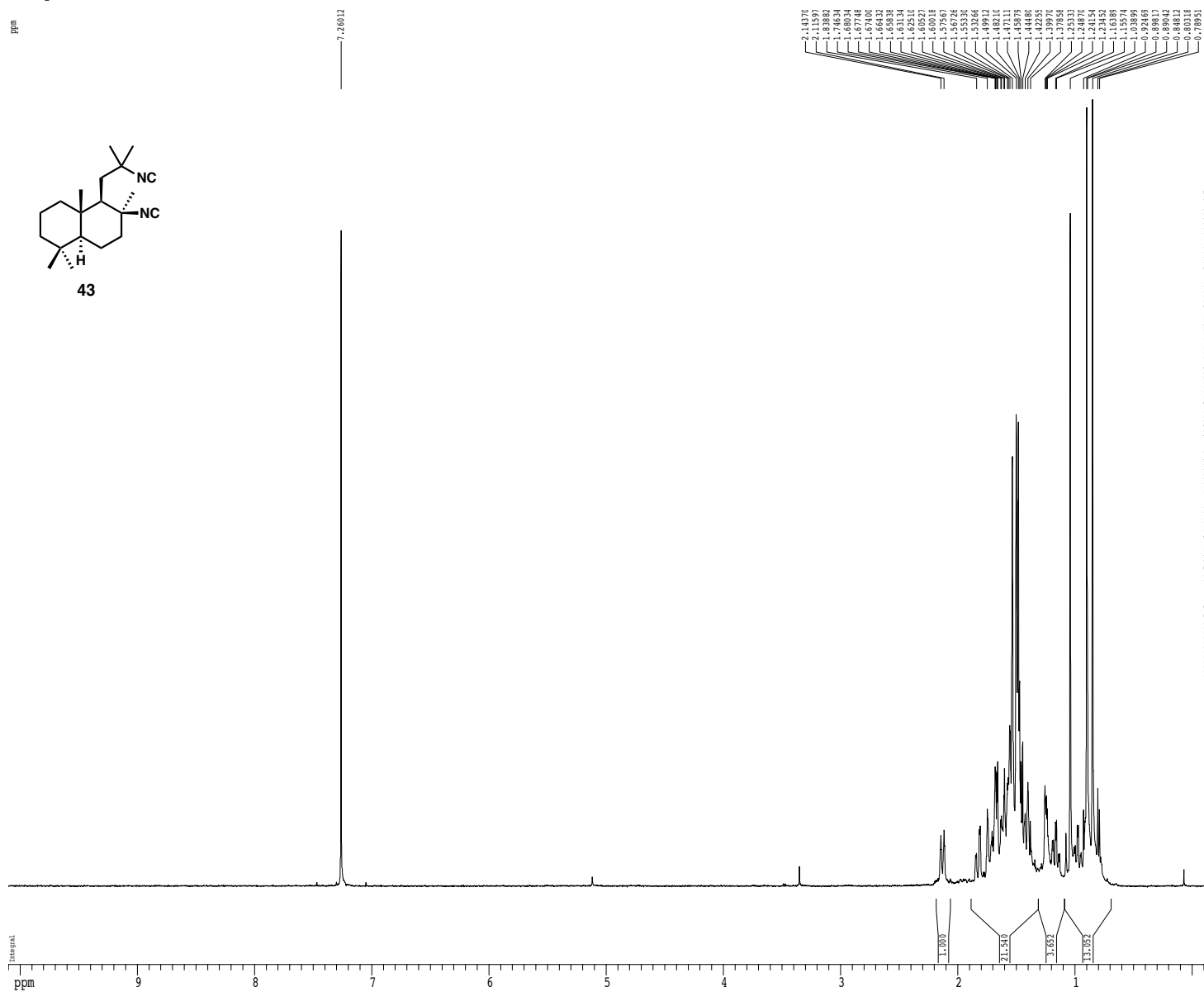
ppm



41



1H spectrum



Current Data Parameters
 USER medaub
 NAME MED-VI-159puff22-32
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150623
 Time 9.34
 INSTRUM zg30
 PROBHD 5 mm broadband
 PULPROG zg30
 TD 81328
 SOLVENT CDCl3T
 NS 8
 DS 2
 SWE 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0988774 sec
 RG 912.3
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.1000000 sec
 MCREST 0.0000000 sec
 MCWREX 0.0150000 sec

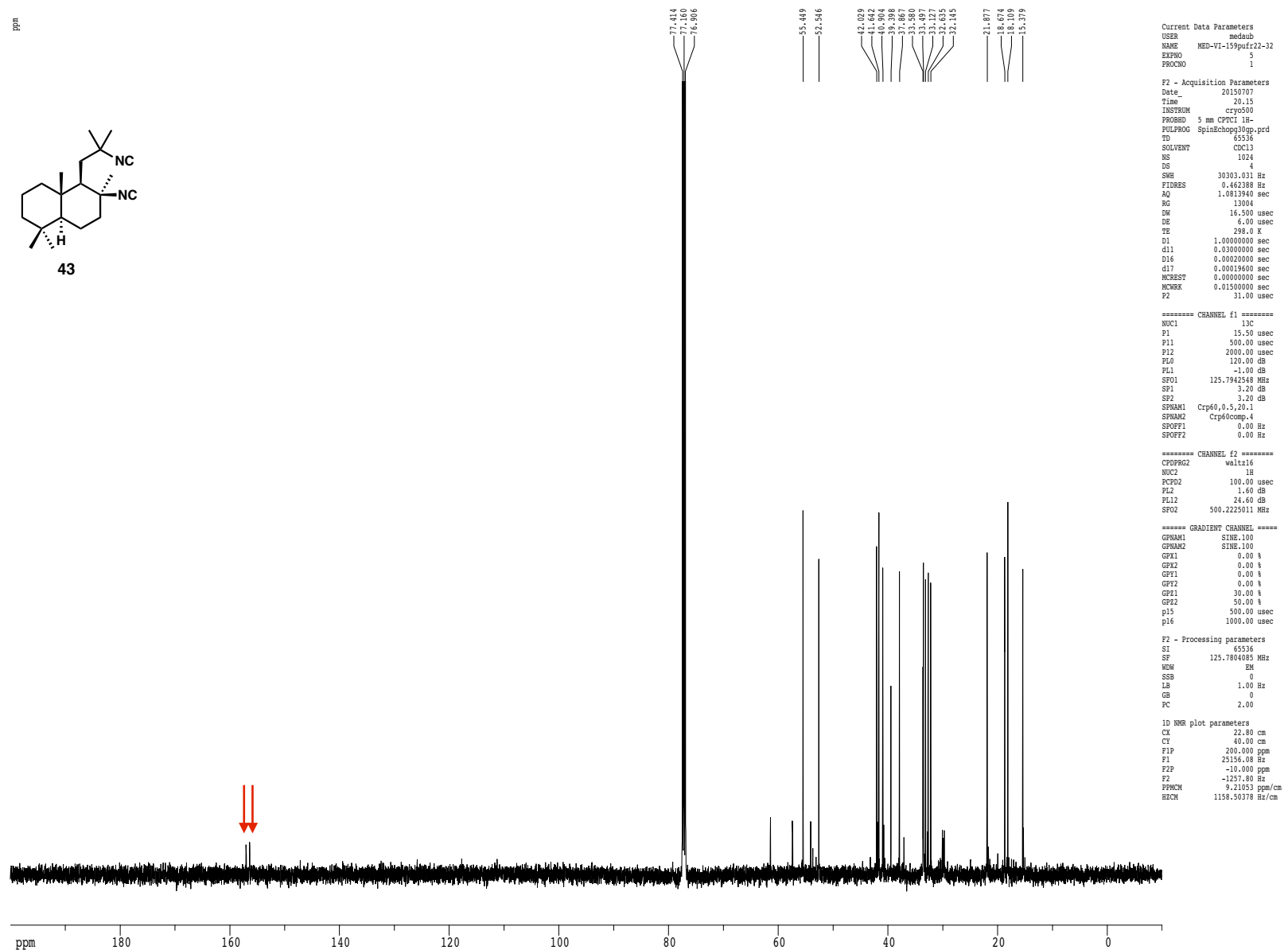
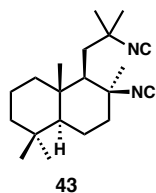
----- CHANNEL f1 -----
 NUC1 1H
 P1 12.00 usec
 PE1 -5.80 dB
 SF01 499.1834943 MHz

F2 - Processing parameters
 S1 65536
 SF 499.1830270 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 FIP 10.100 ppm
 FI 5041.72 Hz
 FZP -0.100 ppm
 F2 -49.92 Hz
 FWHM 0.44337 ppm/cm
 HSCM 223.31740 Hz/cm

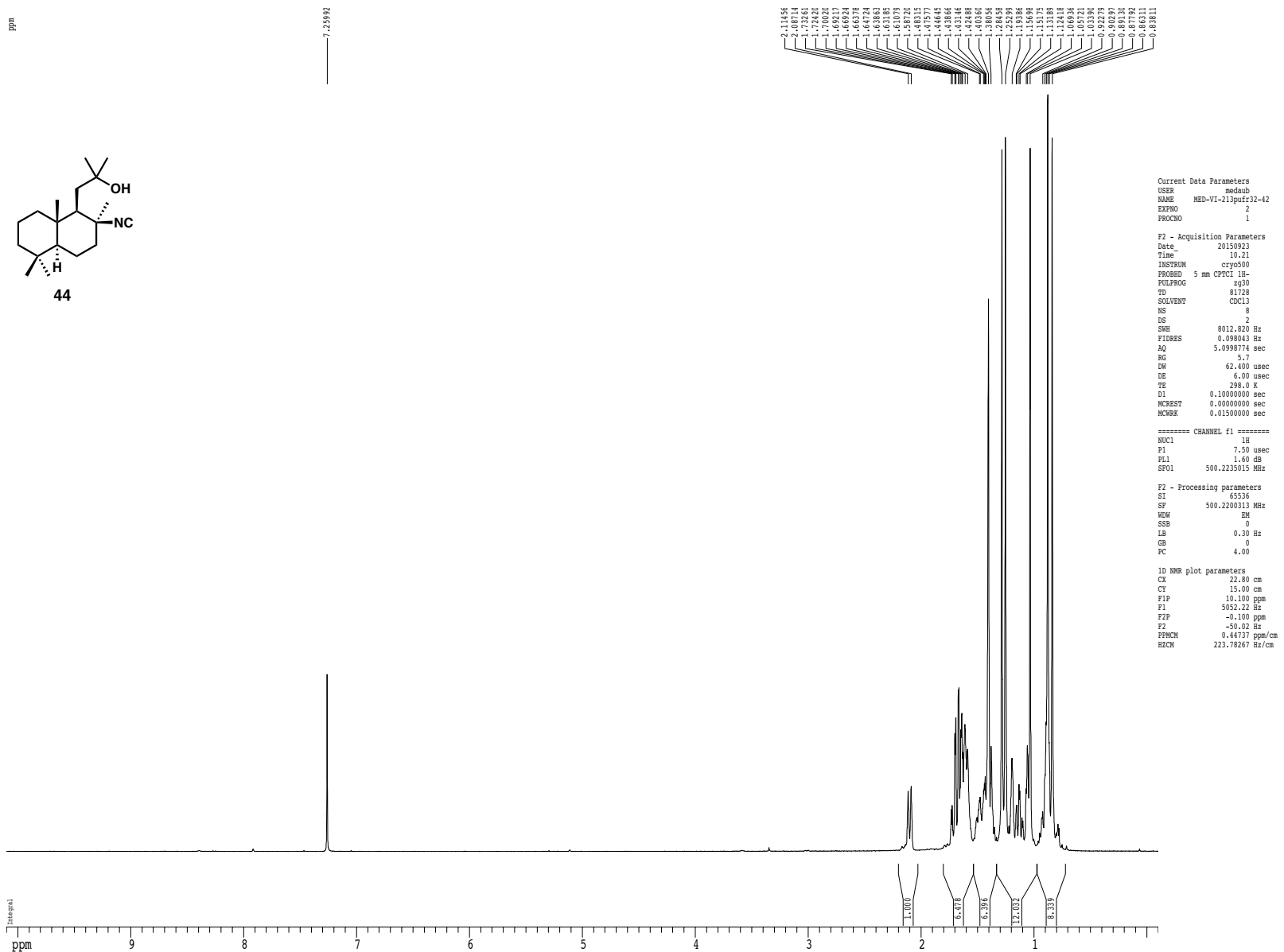
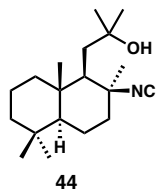
Z-restored spin-echo 13C spectrum with 1H decoupling

ppm

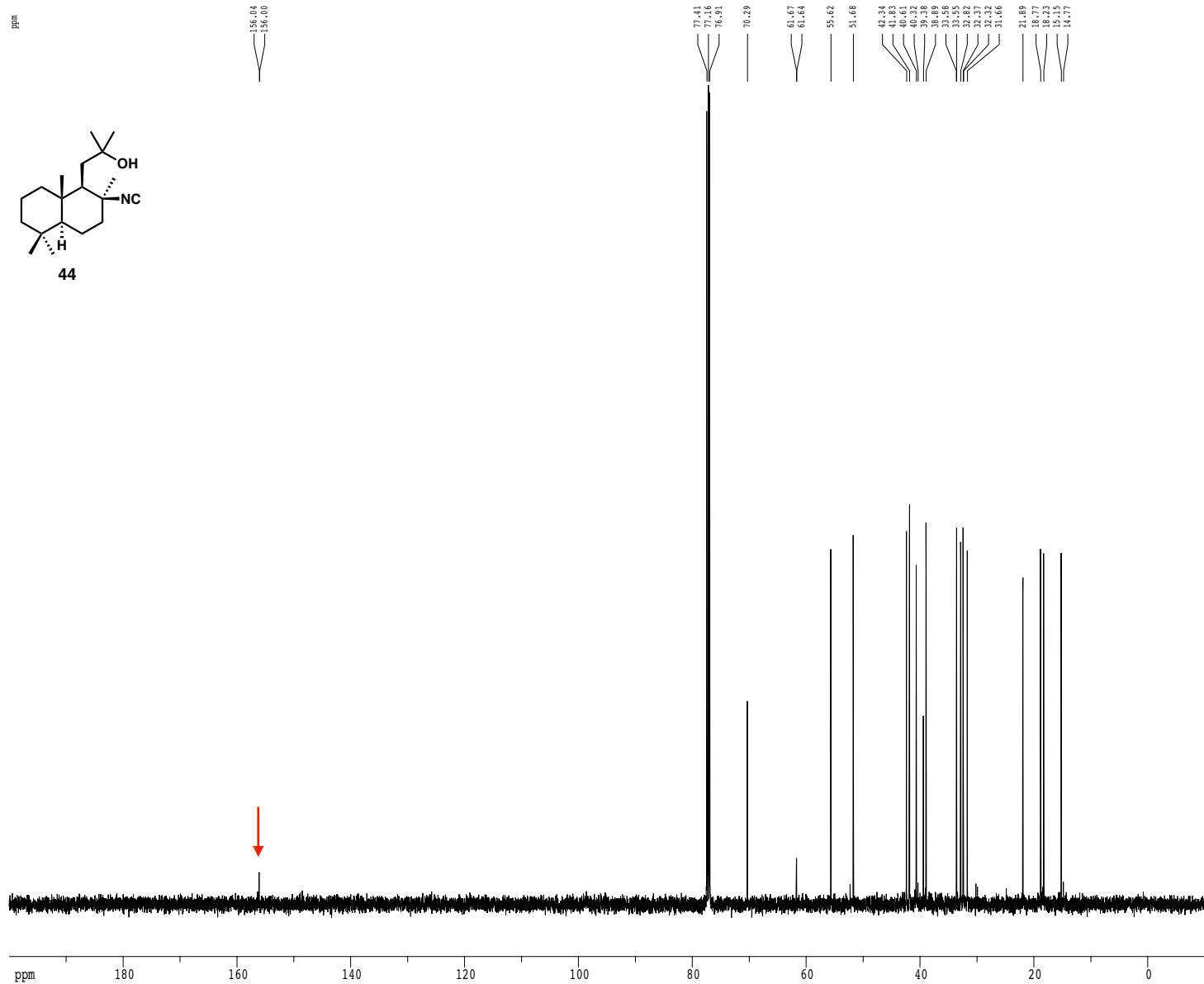


1H spectrum

ppm



Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER          medaub
NAME          MED-VI-213puf12-42
EXPNO        3
PROCNO       1

F2 - Acquisition Parameters
Date_         20150923
Time_         10.23
INSTRUM      cryo500
PROBHD       5 mm CPXI 1H-
PULPROG      SpinEchoq30op.prd
TD           65536
SOLVENT      CDCl3
NS           232
DS           4
SWH          30303.031 Hz
FIDRES       0.462388 Hz
AQ           1.0813940 sec
RG           11585.2
DW           16.500 usec
DE           6.00 usec
TE           298.0 K
D1           0.25000000 sec
d11          0.03000000 sec
D16          0.00020000 sec
d17          0.00019600 sec
MCREST       0.00000000 sec
MCRX        0.01500000 sec
F2           33.10 usec

===== CHANNEL f1 =====
NUC1          13C
P1           16.55 usec
P11          500.00 usec
P12          2000.00 usec
PL0          120.00 dB
PL1          -1.00 dB
SFO1         125.7942548 MHz
SP1          2.70 dB
SP2          2.70 dB
SFO2         125.7600000 MHz
SP1          2.70 dB
SP2          2.70 dB
SFOFF1       0.00 Hz
SFOFF2       0.00 Hz

===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2          1H
PCPD2        100.00 usec
PL2          1.60 dB
PL12         24.50 dB
SFO2         500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1       SINE.100
GPNAM2       SINE.100
GPX1         0.00 %
GPX2         0.00 %
GPY1         0.00 %
GPY2         0.00 %
GPZ1         30.00 %
GPZ2         50.00 %
p15          500.00 usec
p16          1000.00 usec

F2 - Processing parameters
SI           65536
SF           125.7804089 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           1.00

1D NMR plot parameters
CA           22.80 cm
CY           15.65 cm
F1P          200.000 ppm
F1           25156.08 Hz
F2P          -10.000 ppm
F2           -1257.80 Hz
PPMCH       9.21053 ppm/cm
H2CH        1158.50378 Hz/cm
    
```

Study details

Test concentration	1 μ M
Reference compound	Ketanserin
Test systems	human, mouse liver microsomes (from BD Gentest) with final liver microsomal protein concentration of 0.5 mg/mL
Incubation condition	0, 5, 15, 30, 45 min at 37°C
Sample size	Duplicates (n=2)
Bioanalytical method	LC-MS/MS

Calculations

$T_{1/2} = 0.693/K$ (K is the rate constant from a plot of \ln [concentration] vs. incubation time)

$Cl_{int} = (0.693/T_{1/2}) \times (1/(\text{microsomal protein concentration (0.5 mg/mL)})) \times \text{Scaling Factors (Table 1)}$

Table 1 Scaling Factors for Intrinsic Clearance Prediction in the Mouse, Rat, Dog, Monkey, and Human Microsomes

Species	Microsomal Protein	Liver Weight per Kilogram of Body Weight	Scaling Factor ^a	Hepatic Blood Flow (mL/min/kg)
	per Gram of Liver			
Mouse	45	87.5	3937.5	90
Rat	44.8	40	1792	55.2
Dog	77.9	32	2492.8	30.9
Monkey	45	32.5	1462.5	44
Human	48.8	25.7	1254.2	20.7

^aScaling Factor = (microsomal protein per gram of liver) \times (liver weight per kilogram of body weight)

Table 2a: Summary Data

Test Article	Species		Percent Remaining (%)					$T_{1/2}$ (minute)	Cl_{int} (mL/min/kg)
			0 min	5 min	15 min	30 min	45 min		
ketanserin	human	Mean	100.00	88.36	76.98	62.54	47.28	43.44	40.02
		RSD of Area Ratio	0.02	0.02	0.00	0.00	0.05		
	mouse	Mean	100.00	79.90	55.49	32.89	21.72	20.44	266.94
		RSD of Area Ratio	0.01	0.03	0.01	0.02	0.02		
MED-VII-108	human	Mean	100.00	82.25	73.80	91.55	69.01	142.48	12.20
		RSD of Area Ratio	0.17	0.25	0.15	0.17	0.14		
	mouse	Mean	100.00	100.00	86.70	77.93	71.54	87.26	62.54
		RSD of Area Ratio	0.12	0.21	0.04	0.18	0.06		

Note: Some compounds in liver microsomes showed biphasic metabolic kinetics, an initial fast disappearance followed by a slow disappearance. The data points marked in red in column "F ~ H" represented the slow disappearance rates and were excluded from half-life calculation.

Comments:

1. Historic data of ketanserin:

$$T_{1/2}(\text{Human}) = 39.69 \pm 6.36 \text{ min}$$

$$T_{1/2}(\text{Mouse}) = 18.27 \pm 3.89 \text{ min}$$

2. In this study, half lives of Ketanserin were consistent with historical results.

3. If percent remaining at 45 min was higher than 50 %, then $T_{1/2}$ was calculated using extrapolation method; If percentage remaining at 45 min was near 100% and the value $T_{1/2} < 0$ by Excel default, then $T_{1/2}$ and Cl_{int} were reported as " ∞ " and "0", respectively.

4. As summarized in table 2 (summary table), $T_{1/2}$ values marked in green suggested these compounds were stable in liver microsomes ($T_{1/2} > 120$ min);

$T_{1/2}$ values marked in orange suggested these compounds showed moderate metabolism in liver microsomes ($T_{1/2} = 30$ -120 min);

$T_{1/2}$ values marked in red suggested these compounds were susceptible to metabolism in liver microsomes ($T_{1/2} < 30$ min).

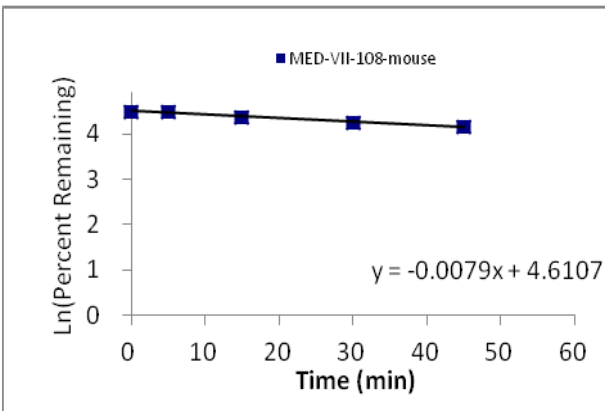
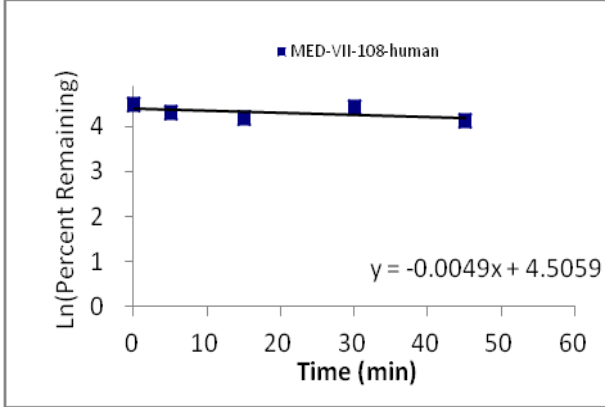
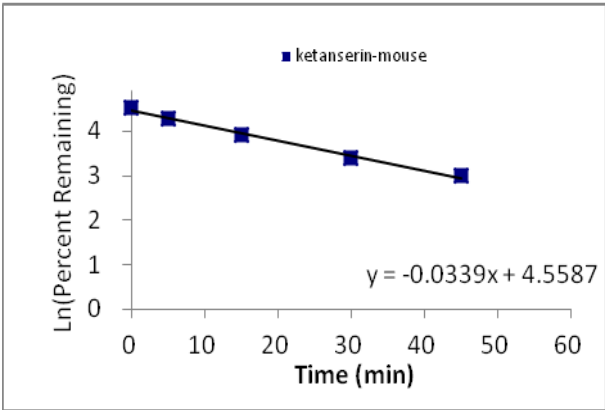
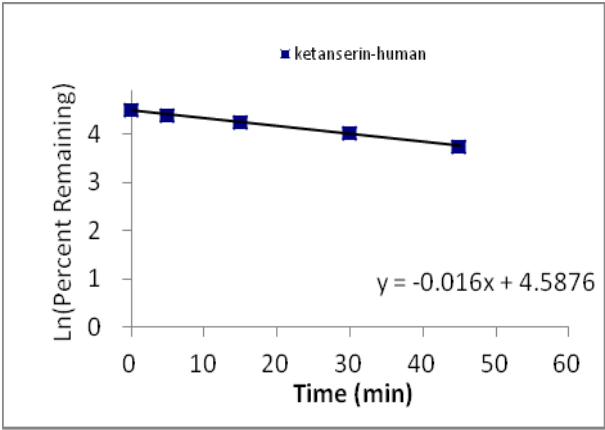


Table 3a: Raw data of reference and test compounds in human liver microsome

Cpd	Species	Time(min)	Raw Data					
			Analyte Peak Area (counts)	Analyte Peak Area (counts)	IS Peak Area (counts)	IS Peak Area (counts)	Area Ratio	Area Ratio
ketanserin	human	0	7620000	7540000	#####	#####	1.140	1.180
		5	6620000	6620000	#####	#####	1.040	1.010
		15	5640000	5650000	#####	#####	0.891	0.895
		30	4510000	4470000	#####	#####	0.723	0.728
		45	2950000	3550000	#####	#####	0.531	0.566
MED-VII-108	human	0	17600	13700	#####	#####	0.002	0.002
		5	11000	15100	#####	#####	0.001	0.002
		15	10500	12900	#####	#####	0.001	0.001
		30	12100	15300	#####	#####	0.001	0.002
		45	11400	9330	#####	#####	0.001	0.001

Table 3b: Raw data of reference and test compounds in mouse liver microsome

Cpd	Species	Time(min)	Raw Data					
			Analyte Peak Area (counts)	Analyte Peak Area (counts)	IS Peak Area (counts)	IS Peak Area (counts)	Area Ratio	Area Ratio
ketanserin	mouse	0	6460000	6490000	#####	#####	1.010	1.030
		5	5130000	5220000	#####	#####	0.795	0.835
		15	3500000	3570000	#####	#####	0.570	0.562
		30	1960000	2030000	#####	#####	0.340	0.331
		45	1140000	1290000	#####	#####	0.224	0.219
MED-VII-108	mouse	0	15500	17800	#####	#####	0.002	0.002
		5	19100	14200	#####	#####	0.002	0.002
		15	15200	14200	#####	#####	0.002	0.002
		30	10500	13300	#####	#####	0.001	0.002
		45	10700	11100	#####	#####	0.001	0.001

Detection method	UPLC-MS/MS-29(API6500+)				
Matrix	liver microsomes				
Internal standard (s)	Osalmid, imipramine				
MS conditions	Positive ion, ESI				
Mobile phase	A: 0.025%FA/1mM NH4OAc/H2O B: 0.025%FA/1mM NH4OAc/MeOH				
Column	ACQUITY-BEH-C18 (2.1*50 mm, 1.7 um)				
LC conditions	0.60 mL/min Time (min) Pump B (%) 0.2 2 0.4 98 1.4 98 1.41 2 2.5 stop				
Detection & Retention time (RT)	compound	Analyte Mass Ranges (Da)	Analyte RT (min)	IS Mass Ranges (Da)	IS RT (min)
	ketanserin	396.300/189.000 Da	0.99	281.100/193.100	1.06
	MED-VII-108	277.500/205.200 Da	0.96	230.100/121.300	0.96

Experimental procedure:

1. Buffer A: 1.0 L of 0.1 M monobasic Potassium Phosphate buffer containing 1.0 mM EDTA
Buffer B: 1.0 L of 0.1 M Dibasic Potassium Phosphate buffer containing 1.0 mM EDTA
Buffer C: 0.1 M Potassium Phosphate buffer, 1.0 mM EDTA, pH 7.4 by titrating 700 mL of buffer B with buffer A while monitoring with the pH meter.
2. Reference compounds(Ketanserin) and test compounds spiking solution:
500 μ M spiking solution: add 10 μ L of 10 mM DMSO stock solution into 190 μ L ACN.
1.5 μ M spiking solution in microsomes (0.75 mg/mL): add 1.5 μ L of 500 μ M spiking solution and 18.75 μ L of 20 mg/mL liver microsomes into 479.75 μ L of Buffer C on ice.
3. Prepare NADPH stock solution (6 mM) by dissolving NADPH into buffer C.
4. Dispense 30 μ L of 1.5 μ M spiking solution containing 0.75 mg/mL microsomes solution to the assay plates designated for different time points (0-, 5-, 15-, 30-, 45-min) on ice.
5. For 0-min, add 135 μ L of ACN containing IS to the wells of 0-min plate and then add 15 μ L of NADPH stock solution (6 mM).
6. Pre-incubate all other plates at 37 °C for 5 minutes.
7. Add 15 μ L of NADPH stock solution (6 mM) to the plates to start the reaction and timing.
8. At 5-min, 15-min, 30-min, and 45-min, add 135 μ L of ACN containing IS to the wells of corresponding plates, respectively, to stop the reaction.
9. After quenching, shake the plates at the vibrator (IKA, MTS 2/4) for 10 min (600 rpm/min) and then centrifuge at 5594 g for 15 min (Thermo Multifuge \times 3R).
10. Transfer 50 μ L of the supernatant from each well into a 96-well sample plate containing 50 μ L of ultra pure water (Millipore, ZMQS50F01) for LC/MS analysis.