

Fig. S1 – $\Delta por1$, but not $\Delta por2$, mutant exhibits increased cell death in response to different concentrations of acetic acid. Bars represent cell survival of a wild-type strain (BY4742), $\Delta por1$ and $\Delta por2$ after 200 min treatment with 100, 140 and 180 mM acetic acid. Total c.f.u. number at T0 was considered as 100 % survival. Cells lacking Por1p exhibit increased sensitivity to the different acetic acid treatments, when compared to the wild-type strain. On the other hand, absence of Por2p does not affect the resistance of yeast cells to acetic acid, with the survival of $\Delta por2$ cells being significantly different (higher) from that of $\Delta por1$ cells, and identical to that of the parental strain. Values are means \pm SD of four independent experiments. Statistical analysis was performed using a Two-way ANOVA and Bonferroni posttests (P-values: (***) P < 0.001).

Supplemental Material – Figure S2

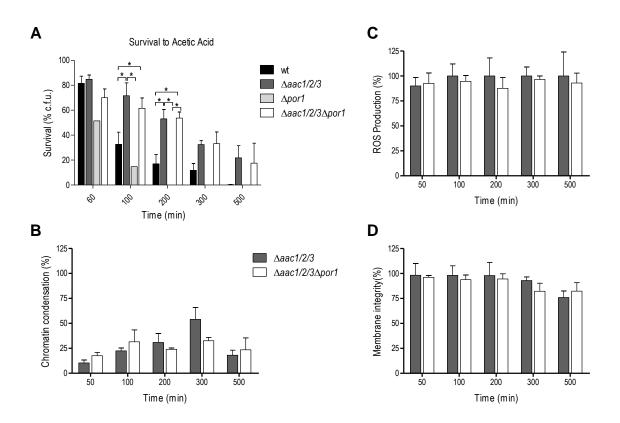


Fig. S2 – Survival of wt, $\Delta aac1/2/3$, $\Delta por1$ and $\Delta aac1/2/3\Delta por1$ cells (A), and the kinetic analyses of ROS production (B), chromatin condensation (C) and membrane integrity (D) of $\Delta aac1/2/3\Delta por1$ cells was carried out over 500 min acetic acid treatment. Absence of AAC proteins reverts the sensitivity phenotype resulting from *POR1* deletion (P-values: (*) P < 0.05). Mitochondrial ROS production, evidenced by MitoTracker Red CM-H2XRos staining, showed a high level of positive cells from the beginning of the treatment, while chromatin condensation using DAPI staining, exhibited a maximum percentage of 40% at 100 min. Finally, loss of membrane integrity using PI staining showed a low level of positive cells indicating that death was not accompanied by loss of membrane integrity. The results showed that as for $\Delta aac1/2/3$ strain, the higher resistance of $\Delta por1\Delta aac1/2/3$ strain is associated with a delayed emergence of apoptotic markers.