Figure S1

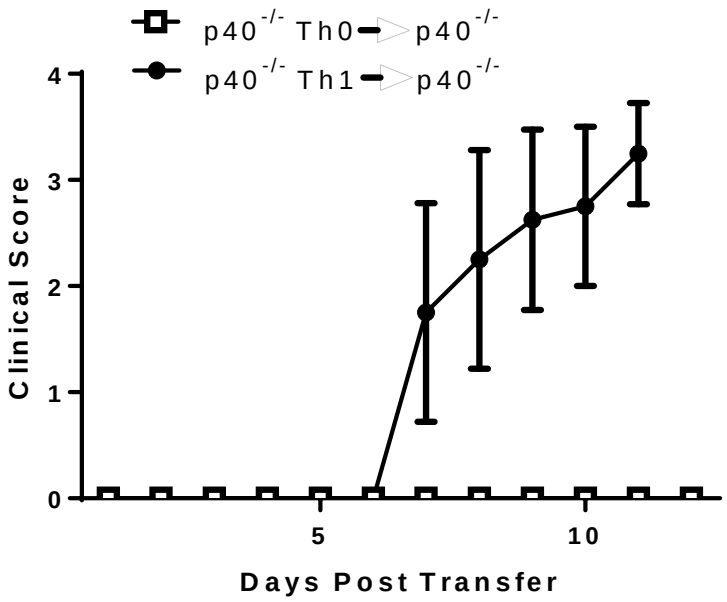

Figure S1 – MOG-primed T cells derived from IL-12p40-/- donors fail to acquire encephalitogenic properties following ex vivo under neutral conditions.
LNC from MOG/CFA-immunized IL-12p40-/- mice were cultured for 4 days with antigen under Th1 (IL-12 and IFN-γ) or neutral Th0 (anti-IL-12p40) polarizing conditions. Purified CD4+ T cells were then purified and adoptively transferred into naïve IL-12p40-/- hosts. n=3-4 mice per group. Data are shown as mean ± SEM.

Figure S2

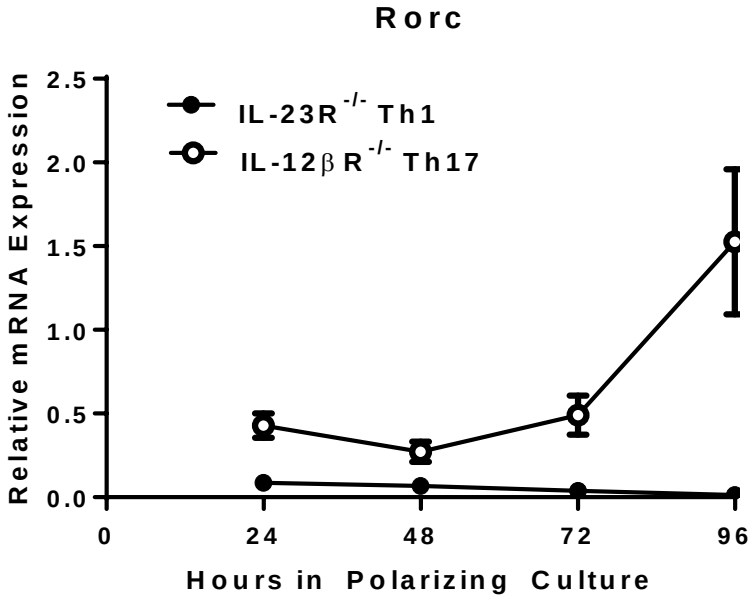

Figure S2 – *Rorc* expression during polarizing culture
LNC harvested from MOG/CFA-immunized IL-23R-/- or IL-12Rβ2-/-mice were cultured with antigen under Th1 or Th17 polarizing conditions, respectively. Cultured cells were collected at the indicated time points for RNA extraction. *Rorc* expression was measured by quantitative RT-PCR and normalized to *gapdh*. Replicates reflect separate cultures from pooled mice, and data are shown as mean ± SEM.

Figure S3

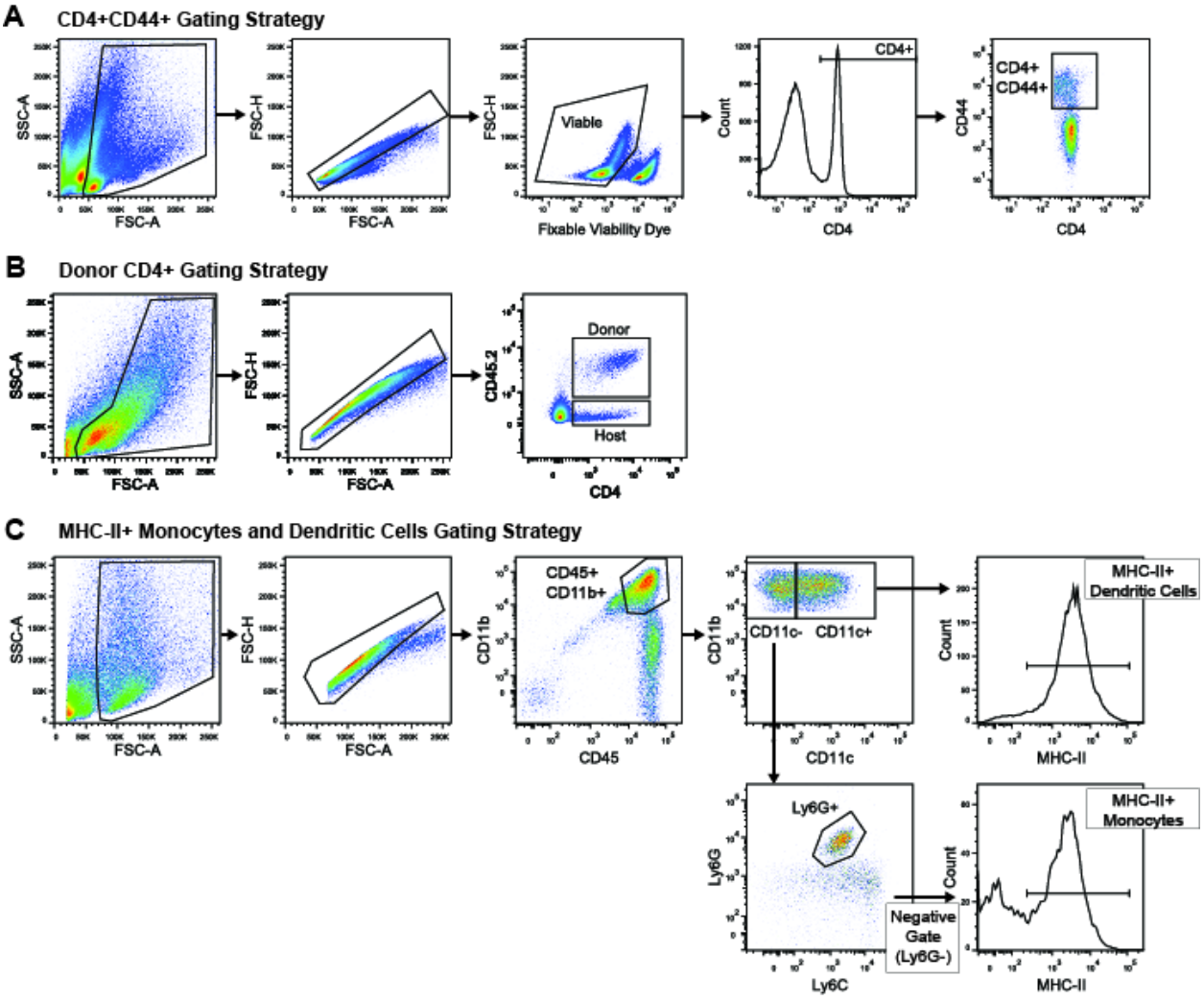

Figure S3 – Gating strategies for flow cytometry
(A) Gating strategy for $CD4^{+}CD44^{+}$ T cells analyzed post-culture. Following doublet exclusion, CD44+ cells are identified among viable CD4+ T cells. **(B)** Gating strategy for donor CD4+ T cells isolated from the CNS. Following doublet exclusion, CD4+ donor cells are distinguished from host cells by expression of CD45.2. **(C)** Gating strategy for MHC-II+ monocytes and dendritic cells. Following doublet exclusion, CD45+CD11b+ myeloid cells are assessed for expression of CD11c. Dendritic cells are identified as CD11c+. Neutrophils are excluded from the CD11c- population by gating out $Ly6G^{+}$ cells; CD11c- Ly6G- cells are classified as monocytes.