SUPPLEMENTAL INFORMATION

The supplemental data includes four supplemental figures with legends

Figure S1. Oxidant Exposure Exhibits Persistent MCU-Mediated [Ca²⁺]_m Uptake. Related to Figure 1.

(A) Mean traces for $[Ca^{2+}]_m$ (GCaMP2-mt) fluorescence measured in control, H_2O_2 and H_2O_2 + MnSOD treated HeLa cells. (B) Quantification of normalized GCaMP2-mt fluorescence at peak and 600 s. Bar represents Mean \pm SEM; * P <0.05; n = 4. (C) Mean traces of [Ca²⁺]_c (Fluo-4) fluorescence measured in control, H_2O_2 and H_2O_2 + MnSOD treated HeLa cells. (**D**) Bar graph represents peak Fluo-4 fluorescence after histamine stimulation. Data indicate Mean ± SEM; nsnot significant, n=4. (E) Representative confocal images showing HyPer-dMito fluorescence in control and LPS treated HPMVECs (left). Scatter plot of HyPer-dMito fluorescence in HPMVECs (right). Data indicate Mean ± SEM; *** P <0.001. (F) Representative confocal images showing MitoSox Red fluorescence from control and H/R subjected primary adult cardiomyocytes (left). Scatter plot of MitoSox Red fluorescence in myocytes (right). Data indicate Mean ± SEM; *** P <0.001. (G) Representative [Ca²⁺]_{out} traces in Neonatal Rat Ventricular Myocytes (NRVM) exposed to normoxia and H/R. Permeabilized cells were loaded with extramitochondrial Ca²⁺ ([Ca²⁺]_{out}) indicator Fura2FF to which a single 10 µM Ca²⁺ pulse was added to assess the $[Ca^{2+}]_{out}$ clearance rate. (H) Quantification of the rate of $[Ca^{2+}]_m$ uptake as a function of decrease in [Ca²⁺]_{out}. Data represents Mean ± SEM; **P <0.01; n = 4. (I) Representative [Ca²⁺]_{out} traces in NRVMs after addition of CCCP. (J) Quantification of total matrix $[Ca^{2+}]_m$ released following 10 μ M Ca²⁺ pulse. Data represents Mean ± SEM; **P <0.01; n = 4. (K) Oxidative stress-induced modification of MCU complex. HEK293T cells expressing Flag-tagged MCU,

MCUb, MICU1, MCUR1 and EMRE were exposed to menadione (10 μ M) for 10 min. Lysates were prepared, incubated with mPEG5 and Western blotted with FLAG antibody. (L and M) NRVMs expressing MCU-FLAG (Ad MCU) were exposed to menadione, isoproterenol (10 μ M) or H/R. Lysates were prepared, incubated with mPEG5 and Western blotted with FLAG antibody (n = 3).

Figure S2. MCU Mutants reside in the mitochondrial inner membrane and alter MCU-Mediated [Ca²⁺]_m Uptake. Related to Figure 2.

(A) Alignment of MCU sequence indicating the conserved cysteine residues among species. (B) Confocal micrographs of HeLa cells co-transfected with matrix localized GCaMP2-mt (green) and, either MCU WT-mRFP or mRFP-tagged MCU cysteine mutants. (C) Mean traces of [Ca²⁺]_c (Fluo-4) fluorescence in HeLa cells expressing Vector control, MCU^{WT}, MCU^{C97A}, and MCU^{CF}. After baseline recording, cells were treated with SERCA inhibitor, Thapsigargin (Tg; 2 μ M) and changes in fluorescence were measured. (D) Quantification of [Ca²⁺]_c fluorescence after Tg addition. Data represents Mean \pm SEM; n = 5-7. (E) Bar graph showing the mitochondrial membrane potential indicator, TMRM fluorescence in control vector, MCU, MCU^{C97A}, and MCU^{CF} HeLa cells. Data represents Mean ± SEM; n = 5. (F) Mean traces of [Ca²⁺]_m (GCaMP2-mt) fluorescence in HeLa cells expressing MCU^{WT}, MCU^{C67A}, and MCU^{C191A}. After baseline recording, cells were stimulated with histamine and changes in fluorescence were measured. (G) Quantification of normalized GCaMP2-mt peak fluorescence. Data represents Mean ± SEM; n = 5-7. (H) Mean traces of $[Ca^{2+}]_c$ (Fluo-4) fluorescence measured in HeLa cells expressing MCU, MCU^{C67A}, and MCU^{C191A}. (I) Quantification of normalized (Fluo-4) peak fluorescence. Data represents Mean \pm SEM; n = 5-7. (J) Representative [Ca²⁺]_{out} traces before and after CCCP (3 µM) addition in HeLa cells stably expressing control vector, MCU. MCU^{C97A}.

and MCU^{CF}. (**K**) Quantification of resting matrix $[Ca^{2+}]_m$ after the addition of CCCP. Data represents Mean ± SEM; **P <0.01; n = 3. (**L**) Representative $[Ca^{2+}]_{out}$ traces before and after CCCP (3 µM) addition in HPMVECs stably expressing control vector, MCU, MCU^{C97A}, and MCU^{CF}. (**M**) Quantification of resting matrix $[Ca^{2+}]_m$ after the addition of CCCP. Data represents Mean ± SEM; **P <0.01; n = 3.

Figure S3. Interaction between MCU and its regulatory components are unaffected by MCU cysteine mutation. Related to Figure 2.

(A) COS-7 cells were transfected with HA-tagged MCU and, either Flag-tagged MCU or MCU cysteine (MCU^{C97A}, and MCU^{CF}) mutants as indicated. Following immunoprecipitation with HA antibody, total cell lysates and immunoprecipitated materials were subjected to Western blot analysis. Samples were probed with anti-Flag (top) and anti-HA antibodies (bottom). (n=3). (B) COS-7 cells were transfected with V5-tagged MCUR1 and, either Flag-tagged MCU or MCU cysteine (MCU^{C97A}, and MCU^{CF}) mutants as indicated. Following immunoprecipitation with V5 antibody, total cell lysates and immunoprecipitated materials were subjected to Western blot analysis. Samples were probed with anti-Flag (top) and anti-V5 antibodies (bottom). (n=3). (C) COS-7 cells were transfected with HA-tagged EMRE and, either Flag-tagged MCU or MCU cysteine (MCU^{C97A}, and MCU^{CF}) mutants as indicated. Following immunoprecipitation with HA antibody, total cell lysates and immunoprecipitated materials were subjected to Western blot analysis. Samples were probed with anti-Flag (top) and anti-HA antibodies (bottom). (n=3). (D) COS-7 cells were transfected with HA-tagged MCUb and, either Flag-tagged MCU or MCU cysteine (MCU^{C97A}, and MCU^{CF}) mutants as indicated. Following immunoprecipitation with HA antibody, total cell lysates and immunoprecipitated materials were subjected to Western blot analysis. Samples were probed with anti-Flag (top) and anti-HA antibodies (bottom). (n=3).

(E) MCU-MICU1 interaction is intact in MCU^{C97A} mutant. COS-7 cells were transfected with HAtagged MICU1 and Flag-tagged MCU, MCU^{C97A}, and MCU^{CF} as indicated. Following immunoprecipitation with HA antibody, total cell lysates and immunoprecipitated materials were subjected to Western blot analysis. Cell lysates were probed with anti-Flag (top left) or anti-HA antibodies (bottom left) to serve as input. Immunoprecipitated samples were probed with anti-Flag (top right) and anti-HA antibodies (bottom right). (n=3).

Figure S4. Oxidation of MCU Enables MCU Complex Redistribution at the Inner Mitochondrial Membrane. Related to Figure 5.

(**A**) MCU-WT was tagged with a photo-switchable protein, mEOS3.2. HeLa cells were transfected, fixed, and imaged using super-resolution photoactivatable localization microscopy (PALM). The HeLa cells were treated with menadione for various time points (2, 5, 15, 20 minutes). The blue box marks the magnified area (right panels).

(**B**) Nanoclustering of MCU WT-mEOS3.2 was conducted as previously described (Baumgart et al., 2016). The normalized density of (ρ/ρ_0) molecules was plotted against the relative area covered by the clusters (η). The red line fitted to the graph denotes 100% randomly distributed molecules, while true clustering has a higher density (ρ/ρ_0) of molecules along a higher percentage of the area covered by the clusters (η). MCU-WT showed increased clustering with increasing time points of menadione treatment as depicted by the higher ρ/ρ_0 . (n=4-7) For each condition, we randomly quantified 5 cells for cluster analysis.



Caenorhabditis.elegans (Round worm) Danio rerio (Zebrafish) Xenopus tropicalis (Frog) Homo sapiens (Human) Pan troglodytes (Chimpanzee) Rattus norvegicus (Rat) Mus musculus (Mouse)

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