## **A Convergent Synthesis and Optical Properties of Near-infrared Emitting Bioluminescent Infra-luciferins**

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**Supplementary Information**

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General experimental

Biological materials and methods

Copies of <sup>1</sup>H, <sup>13</sup>C, <sup>31</sup>P NMR spectra: **8**, **9**, **10**, **11**, **12**, **6**, **4**, **13**, **14**, **15**, **16**, **5**

#### **General Experimental Details**

All manipulations were routinely carried out under an inert (Ar or  $N_2$ ) atmosphere. Cooling to 0 °C was effected using an ice-water bath. Cooling to temperatures below 0 °C was effected using dry iceacetone mixtures. For the purposes of thin layer chromatography (tlc), Merck silica-aluminium plates were used, with *uv* light (254 nm) and potassium permanganate or anisaldehyde for visualisation. For column chromatography Merck Geduran® Si 60 silica gel was used. Butyl lithium solutions were standardised with diphenyl acetic acid.

All reagents were used as received unless stated. Anhydrous solvents (THF, DCM, MeCN) were obtained from a solvent tower, where degassed solvent was passed through two columns of activated alumina and a 7 micron filter under 4 bar pressure. 6-( $\beta$ -Methoxyethoxymethylether)benzothiazole and ethyl *O*,*O*-diethylphosphonodithioacetate (**8**) were synthesised according to the published procedures. $1, 2$ 

Melting points are uncorrected and were recorded on a Griffin melting point machine. All NMR data was collected using a Bruker AMX 300 MHz or a Bruker AVANCE III 600 MHz as stated. All chemical shifts (δ) are reported in parts per million (ppm) relative to residual solvent peaks. Reference values for residual solvents were taken as  $\delta$  = 7.27 (CDCl<sub>3</sub>), 2.51 (DMSO –*d*6), 3.30 (MeOD- *d*4) ppm for <sup>1</sup>H NMR and δ = 77.2 (CDCl<sub>3</sub>), 39.5 (DMSO –*d*6), 49.0 (MeOD- *d*4) ppm for <sup>13</sup>C NMR. Coupling constants (*J*) are reported in Hertz and are recorded as observed in the spectrum without averaging. Infrared spectra were recorded using a Bruker Alpha ATR spectrometer. Mass spectrometry data was collected on a Micromass LCT Premier XE (ESI) or Thermo Finnigan Mat900xp (EI/CI) instrument.

#### **Biological materials and methods**

#### **Optical properties**

**Fluorescence quantum yields of analogues (Table 1).** Fluorescence spectra of 50uM analogues were measured as above and furthermore, the emission of 1uM Eosin Yellow dye (BDH Lab Supplies, Poole, UK) in basic ethanol was measured at an excitation wavelength of 490nm. Furthermore, absorption readings of 50uM analogues were also measured and used to derive fluorescence quantum yields<sup>3</sup> of analogues at  $pH$  7.8 in TEM buffer.

**pH dependence of fluorescence spectra of D-luciferin (1) and analogues 4 and 5 (Fig 3).** 10mg/ml analogues in 50% DMSO were dissolved in 500ul TEM buffer (pH 7.8) for concentrations of 500uM each. They were further diluted to 50uM in different solutions of TEM of differing pH values (pH 6.3, 6.8, 7.3, 7.8, 8.3, 8.8 & 9.3). The solutions were checked for pH after being made up with analogues (values displayed in data). Data were acquired in 2 instruments: the Varian Cary Eclipse Spectrofluorometer and also the Tecan Infinite M200 Spectrofluorometer in triplicate of triplicates on each machine.

#### **Bioluminescence specific activities of luciferin analogues with wild-type firefly luciferase (Fig 4).**

Bioluminescence assays were conducted using the PhotonIMAGER Optima (Biospace Labs, Paris, France) at 26-28°C. 50ul of 6mM ATP was dispensed onto 50ul substrates mixed with different luciferases in triplicate. Experiments were carried out several times with 0.05uM enzymes for native luciferin and 0.5uM enzymes for analogues.

**Bioluminescence spectra of wild-type luciferase enzymes with luciferin and analogues.** 1-10mM enzymes were used to determine bioluminescence spectra with luciferin and analogues at differing pH. The spectrum of Eosin Yellow in basic ethanol was compared to the absolute spectrum<sup>4</sup> to correct spectra for PMT sensitivity.

### **References**

- 1) E. Pfund, T. Lequeux, S. Masson and M. Vazeux, *Org. Lett.,* 2002, **4**, 843.
- 2) C. S. Marvel, P. de Radzitzky and J. J. Brader, *J. Am. Chem. Soc.,* 1955, **77**, 5997.
- 3) *Principles of Fluorescence Spectroscopy*, Editor: Joseph R. Lakowicz, Springer US, 2006. ISBN: 978-0-387-31278-1.
- 4) <http://omlc.org/spectra/PhotochemCAD/html/061.html> (accessed June 2016)







#### sp139-3 C13.ucl CDCI3 {C:\Bruker\TOPSPIN} jca 44





sp139-3<br>P31\_with\_proton\_couplings CDCl3 {C:\Bruker\TOPSPIN} jca 46







# sp147-3-full data<br>C13\_DayTime.ucl CDCl3 {W:\600} jca 40

















#### C13CPD.ucl CDCl3 {W:\600} tds 10









sp164-2<br>C13\_DayTime.ucl CDCl3 {W:\600} jca 8





**S14** 



sp195-frac8<br>C13\_DayTime.ucl CDCl3 {W:\600} hch3 6





**S16** 





 $\overline{\mathbf{4}}$ 





 $\overline{\mathbf{4}}$ 



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**S20** 



hg855col<br>C13CPD.ucl CDCl3 {W:\600} jca 14







hg894col<br>C13\_DayTime.ucl CDCl3 {W:\600} jca 53







hg900col<br>C13\_DayTime.ucl CDCl3 {W:\600} jca 11







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16

 $CO<sub>2</sub>Me$ 

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Solvent suppression of largest peak ( $H_2O$ ).



**S28** 



C13 DayTime.ucl D2O {W:\600} jca 44

