## **Supplementary Data**

# The Proline/Glycine-Rich Region Of The Biofilm Adhesion Protein Aap Forms An Extended Stalk That Resists Compaction

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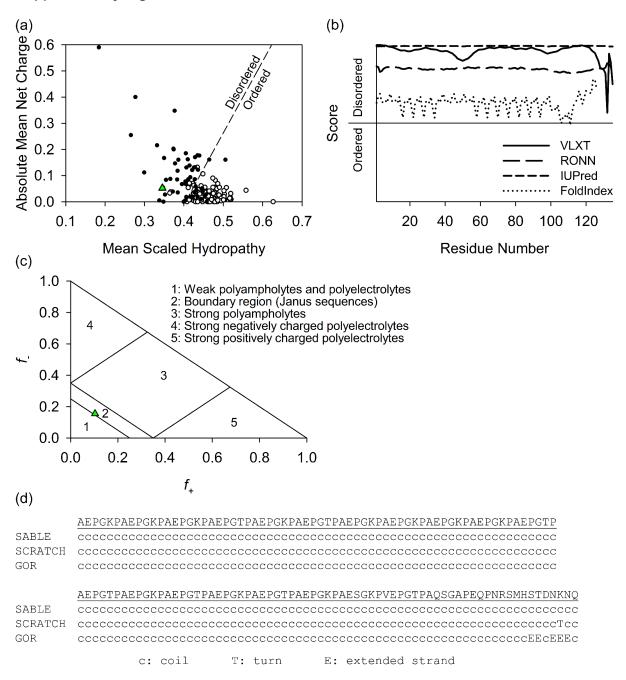
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#### Supplementary data files present in this document include:

- 1. Supplementary Figures S1—S2
- 2. Supplementary Tables S1—S5

**Supplementary Figures** 



#### Figure S1. PGR is predicted to be intrinsically disordered.

Panel (a) is the Uversky plot, showing PGR (*green triangle* – (0.3454, 0.0522)) lies on the portion of the plot where disordered proteins (*black circles*) tend to fall. Ordered proteins are shown in *white circles*. In panel (b), the results from several disorder predictions are plotted. The y-axis units should be considered arbitrary, as these algorithms have different ranges for their predictions; however, these results strongly support the hypothesis that PGR is an IDP. (c) shows PGR (*green triangle* – (0.10370, 0.15556)) in the boundary region (2) of the Das-Pappu phase plot separating weak polyampholytes/polyelectrolytes (1) and strong polyampholytes (3).

Panel (d) shows secondary structure predictions by SABLE, SCRATCH, and GOR servers, each predicting essentially complete random coil.

Q9L470_RP62A Q8CQD9_ATCC 12228 Q6UV38_5179 Q6UV37_5179-R1 A0A075IHN3_1457	EYGPTKAEPGKPAEPGKPAEPGKPAEPGTPAEPGKPAEPGTPAEPGKPAEPGKPA EYGPTKAEPGKPAEPGKPAEPGKPAEPGKPAEPGTPAEPGKPAEPGKPAEPGKPA EYGPTKAEPGKPAEPGKPAEPGKPAEPGTPAEPGTPAEPGTPAEPGKPAEPGKPA EYGPTKAEPGKPAEPGKPAEPGKPAEPGTPAEPGTPAEPGTPAEPGTPAEPGKPAEPGKPA EYGPTKAEPGKPAEPGKPAEPGKPAEPGTPAEPGTPAEPGTPAEPGTSEPGKPAEPGKPA ************************************	55 55 55 55 60
Q9L470_RP62A Q8CQD9_ATCC 12228 Q6UV38_5179 Q6UV37_5179-R1 A0A075IHN3_1457	EPGKPAEPGKPAEPGTPAEPGTPAEPGKPAEPGTPAEPGKPAEPGTPAEPGKPAESG EPGKPAEPGKPAEPGTPAEPGKPAEPGKPAEPGTPAEPGKPAEPGTPAEPGKPAEPGTPAEPGKPAESGKPV EPGKPAEPGKPAEPGTPAEPGTPAEPGKPAEPGTPAEPGKPAEPGKPAESGKPV EPGKPAEPGKPAEPGTPAEPGTPAEPGKPAEPGTPAEPGKPA	112 112 115 115 120
Q9L470_RP62A Q8CQD9_ATCC 12228 Q6UV38_5179 Q6UV37_5179-R1 A0A075IHN3_1457	<ul> <li>KPVEPGTPAQSGAPEQPNRSMHSTDNKNQLPDTG 146</li> <li>KPAEPGTPTQSGAPEQPNRSMHSTDNKNQLPDTG 146</li> <li>EPGKPVEPGTPAQSGAPEQPNRSMHSTDNKNQSPDTG 152</li> <li>EPGKPAEPGTPAQSGAPEQPNRSMHSTDNKNQLPDTG 157</li> <li>**.****:******************************</li></ul>	

**Figure S2. The sequence of PGR is highly conserved among S.** *epidermidis* strains. Identical residues are marked with an asterisk (\*), highly conserved residues with a semicolon (:), and weakly conserved residues with a period (.). The UniProt accession number and the name of the strain identify each sequence in the alignment. The NCTC 11047 strain (UniProt accession no. **E0ACJ2**) is significantly longer than the strains shown above, having an additional 7 AEPGKP repeats compared to RP62A (UniProt accession no. **Q9L470**), and thus was omitted for clarity. The Aap from strain PM221 (GenBank accession no. **CDM15051**) contains a region between the last half B-repeat and the LPXTG motif which does not resemble the PGR of any of the above strains, neither in number of residues nor in amino acid content.

#### **Supplementary Tables**

PGR Concentration	<b>S</b> <sub>20,w</sub> <sup>a</sup>	<i>f/f</i> 0 <sup>b</sup>	M <sub>calc</sub> <sup>c</sup>
25 µM	1.05	2.14	14.4 kDa
75 µM	1.05	2.11	14.0 kDa
150 µM	1.03	2.15	14.1 kDa
225 µM	1.02	2.18	14.1 kDa
300 µM	0.99	2.32	14.8 kDa

#### Table S1. Concentration dependence of sedimentation velocity AUC data

<sup>a</sup>The sedimentation coefficient standardized to 20° C and pure water.

<sup>b</sup>Frictional ratio – the experimental frictional coefficient divided by the frictional coefficient of an ideal, non-hydrated sphere

<sup>c</sup>The molecular weight calculated from the sedimentation coefficient and frictional ratio

Table S2. Temperature dependence of sedimentation velocity AUC data	Table S2. Temperature of	dependence of s	sedimentation v	elocitv AUC data
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Temperature (° C)	<b>s</b> <sub>20,w</sub> <sup>a</sup>	<i>f/f<sub>0</sub></i> <sup>b</sup>	M <sub>calc</sub> <sup>c</sup>
4° C	1.06	2.14	15.3 kDa
20° C	1.05	2.14	14.4 kDa
37° C	1.03	2.10	13.0 kDa
2			

<sup>a</sup>The sedimentation coefficient standardized to 20° C and pure water.

<sup>b</sup>Frictional ratio – the experimental frictional coefficient divided by the frictional coefficient of an ideal, non-hydrated sphere

<sup>c</sup>The molecular weight calculated from the sedimentation coefficient and frictional ratio

Temperature (° C)	NaCI Concentration	<b>S</b> <sub>20,w</sub> <sup>a</sup>	<i>f/f<sub>0</sub></i> <sup>b</sup>	M <sub>calc</sub> <sup>c</sup>
4° C	30 mM	1.04	2.05	13.8 kDa
	100 mM	1.06	2.02	13.9 kDa
	300 mM	1.06	2.00	13.8 kDa
	1 M	1.03	2.00	13.2 kDa
20° C	30 mM	1.04	2.09	13.7 kDa
	100 mM	1.05	2.08	13.8 kDa
	300 mM	1.04	2.08	13.7 kDa
	1 M	1.00	2.19	13.8 kDa
37° C	30 mM	1.04	2.04	12.6 kDa
	100 mM	1.05	2.05	12.9 kDa
	300 mM	1.03	2.09	13.0 kDa
2	1 M	0.98	2.12	12.2 kDa

#### Table S3. Salt dependence of sedimentation velocity AUC data

<sup>a</sup>The sedimentation coefficient standardized to 20° C and pure water.

<sup>b</sup>Frictional ratio – the experimental frictional coefficient divided by the frictional coefficient of an ideal, nonhydrated sphere

<sup>c</sup>The molecular weight calculated from the sedimentation coefficient and frictional ratio

Sequence	N <sup>a</sup>	Net Charge	$R_h$ predicted <sup>b</sup>	f <sub>PPII</sub> c	$R_h  \text{Obs}^d$	Reference
Aap-PGR	135	7	38.50	0.5350	37.06 <sup>e</sup>	This work
p53(1-93)	93	15	29.51	0.4890	32.4	[1]
p53(1-93) ALA-	93	15	28.66	0.4581	30.4	[1]
p53 TAD	73	14	24.79	0.4500	23.8	[2]
Securin	202	1	42.57	0.4130	39.7	[3]
PDE-γ	87	4	26.51	0.4122	24.8	[4]
Cad136	136	9	33.77	0.4025	28.1	[5]
HIF1-α-403	202	29	42.13	0.4024	44.3	[6]
Tau-K45	198	19	41.52	0.3988	45.0	[7]
HIF1-α-530	170	10	37.81	0.3899	38.3	[6]
Fos-AD	168	16	37.17	0.3783	35.0	[8]
ShB-C	146	4	34.32	0.3764	32.9	[9]
α-synuclein	140	9	33.47	0.3744	28.2	[10]
Mlph(147-403)	260	28	47.00	0.3703	49.0	[11]
CFTR-R-region	189	5	39.18	0.3644	32.0	[12]
p57-ID	73	6	23.14	0.3636	24.0	[13]
prothymosin-α	110	43	29.02	0.3633	33.7	[14]
LJIDP1	94	4	26.46	0.3565	24.5	[15]
Mlph(147-240)	97	15	26.85	0.3528	28.0	[11]
SNAP25	206	14	40.60	0.3513	39.7	[16]
Hdm2-ABD	97	29	26.47	0.3345	25.7	[17]
Vmw65	89	19	25.13	0.3278	28.0	[18]
p53(1-93) PRO-	93	15	24.93	0.2832	27.4	[1]

Table S4. Comparison of hydrodynamic properties for PGR to a dataset of studied IDPs

<sup>a</sup>The number of amino acids in the sequence

<sup>b</sup>The predicted  $R_h$  from sequence and according to equation 6 in the main text

<sup>c</sup>The fractional number of PPII residues from sequence and according to intrinsic PPII propensities [19] <sup>d</sup>The  $R_h$  of the IDP as measured experimentally in the reference listed in the final column

<sup>e</sup>As measured in this study, listed is the average of SEC and DLS measurements.

IDP dataset adapted from Tomasso et al. [19] and sorted by f<sub>PPII</sub>

Sequence	N <sup>a</sup>	$R_h^{b}$	Reference
staphylococcal nuclease	151 <sup>‡</sup>	22.5	[20]
human recombinant lysozyme	132 <sup>‡</sup>	21.8	[20]
bovine erythrocyte carbonic anhydrase	267 <sup>‡</sup>	26.8	[20]
bovine pancreatic trypsin inhibitor	58	15.8	[21]
SH3 domain of PI3 kinase	90	18.6	[21]
horse heart cytochrome c	104	17.8	[21]
hen lysozyme	129	20.5	[21]
horse myoglobin	153	21.2	[21]
bovine alpha-lactalbumin	123	18.8	[21]
bovine pancreatic ribonuclease A	124	19.0	[21]
sperm whale apomyoglobin	153	20.9	[21]
ubiquitin	76	16.5	[21]
(apo)cytochrome C	104	18.5	[21]
α-lactalbumin	123	18.5	[21]
tumor supressor, p16	156	20.0	[21]
(apo)myoglobin	154	20.9	[21]
β-lactoglobulin	162	22.0	[21]
sarcoplasmic calcium binding	174	21.5	[21]
adenylate kinase	194	21.9	[21]
tryptophan synthase	268	24.2	[21]
β-lactamase	257	23.7	[21]
carbonic anhydrase B	260	23.3	[21]
RTEM β-lactamase	263	24.5	[21]

 Table S5. Folded proteins and hydrodynamic measurements from literature

<sup>a</sup>The number of amino acids in the sequence, taken from [21] unless otherwise noted <sup>b</sup>Hydrodynamic radius, in Å, reported by the reference listed in the final column <sup>‡</sup>The number of residues was estimated from the *MW* using the average of 111.6 Da/residue Folded protein data set adapted from [21], see [22-24] for additional details and individual references

### **Supplemental References**

 R.B. Perez, A. Tischer, M. Auton, S.T. Whitten, Alanine and proline content modulate global sensitivity to discrete perturbations in disordered proteins, Proteins. 82 (2014) 3373-3384.
 D.F. Lowry, A. Stancik, R.M. Shrestha, G.W. Daughdrill, Modeling the accessible conformations of the intrinsically unstructured transactivation domain of p53, Proteins. 71 (2008) 587-598.
 N. Sanchez-Puig, D.B. Veprintsev, A.R. Fersht, Human full-length Securin is a natively unfolded protein, Protein science : a publication of the Protein Society. 14 (2005) 1410-1418.
 V.N. Uversky, S.E. Permyakov, V.E. Zagranichny, I.L. Rodionov, A.L. Fink, A.M. Cherskaya, et al., Effect of zinc and temperature on the conformation of the gamma subunit of retinal phosphodiesterase: a natively unfolded protein, Journal of proteome research. 1 (2002) 149-159. [5] S.E. Permyakov, I.S. Millett, S. Doniach, E.A. Permyakov, V.N. Uversky, Natively unfolded C-terminal domain of caldesmon remains substantially unstructured after the effective binding to calmodulin, Proteins. 53 (2003) 855-862.

[6] N. Sanchez-Puig, D.B. Veprintsev, A.R. Fersht, Binding of natively unfolded HIF-1alpha ODD domain to p53, Molecular cell. 17 (2005) 11-21.

[7] A. Soragni, B. Zambelli, M.D. Mukrasch, J. Biernat, S. Jeganathan, C. Griesinger, et al., Structural characterization of binding of Cu(II) to tau protein, Biochemistry. 47 (2008) 10841-10851.

[8] K.M. Campbell, A.R. Terrell, P.J. Laybourn, K.J. Lumb, Intrinsic structural disorder of the C-terminal activation domain from the bZIP transcription factor Fos, Biochemistry. 39 (2000) 2708-2713.

[9] E. Magidovich, I. Orr, D. Fass, U. Abdu, O. Yifrach, Intrinsic disorder in the C-terminal domain of the Shaker voltage-activated K+ channel modulates its interaction with scaffold proteins, Proceedings of the National Academy of Sciences. 104 (2007) 13022-13027.

[10] K.E. Paleologou, A.W. Schmid, C.C. Rospigliosi, H.Y. Kim, G.R. Lamberto, R.A. Fredenburg, et al., Phosphorylation at Ser-129 but not the phosphomimics S129E/D inhibits the fibrillation of alpha-synuclein, The Journal of biological chemistry. 283 (2008) 16895-16905.

[11] N.C. Geething, J.A. Spudich, Identification of a minimal myosin Va binding site within an intrinsically unstructured domain of melanophilin, The Journal of biological chemistry. 282 (2007) 21518-21528.
[12] J.M.R. Baker, Structural characterization and interactions of the CFTR regulatory region. (2009).
[13] J.N. Adkins, K.J. Lumb, Intrinsic structural disorder and sequence features of the cell cycle inhibitor

p57Kip2, Proteins. 46 (2002) 1-7.

[14] S. Yi, B.L. Boys, A. Brickenden, L. Konermann, W.Y. Choy, Effects of zinc binding on the structure and dynamics of the intrinsically disordered protein prothymosin alpha: evidence for metalation as an entropic switch, Biochemistry. 46 (2007) 13120-13130.

[15] S. Haaning, S. Radutoiu, S.V. Hoffmann, J. Dittmer, L. Giehm, D.E. Otzen, et al., An unusual intrinsically disordered protein from the model legume Lotus japonicus stabilizes proteins in vitro, The Journal of biological chemistry. 283 (2008) 31142-31152.

[16] U.B. Choi, J.J. McCann, K.R. Weninger, M.E. Bowen, Beyond the random coil: stochastic conformational switching in intrinsically disordered proteins, Structure (London, England : 1993). 19 (2011) 566-576.

[17] S.G. Sivakolundu, A. Nourse, S. Moshiach, B. Bothner, C. Ashley, J. Satumba, et al., Intrinsically unstructured domains of Arf and Hdm2 form bimolecular oligomeric structures in vitro and in vivo, Journal of molecular biology. 384 (2008) 240-254.

[18] L. Donaldson, J.P. Capone, Purification and characterization of the carboxyl-terminal transactivation domain of Vmw65 from herpes simplex virus type 1, The Journal of biological chemistry. 267 (1992) 1411-1414.

 [19] M.E. Tomasso, M.J. Tarver, D. Devarajan, S.T. Whitten, Hydrodynamic Radii of Intrinsically Disordered Proteins Determined from Experimental Polyproline II Propensities, PLoS Comput Biol. 12 (2016) e1004686.

[20] T.D. Langridge, M.J. Tarver, S.T. Whitten, Temperature effects on the hydrodynamic radius of the intrinsically disordered N-terminal region of the p53 protein, Proteins. 82 (2014) 668-678.

[21] J.A. Marsh, J.D. Forman-Kay, Sequence determinants of compaction in intrinsically disordered proteins, Biophys. J. 98 (2010) 2383-2390.

[22] D.K. Wilkins, S.B. Grimshaw, V. Receveur, C.M. Dobson, J.A. Jones, L.J. Smith, Hydrodynamic Radii of Native and Denatured Proteins Measured by Pulse Field Gradient NMR Techniques, Biochemistry. 38 (1999) 16424-16431.

[23] O. Tcherkasskaya, E.A. Davidson, V.N. Uversky, Biophysical Constraints for Protein Structure Prediction, J. Proteome Res. 2 (2003) 37-42.

[24] O. Tcherkasskaya, V.N. Uversky, Denatured collapsed states in protein folding: Example of apomyoglobin, Proteins: Structure, Function, and Bioinformatics. 44 (2001) 244-254.