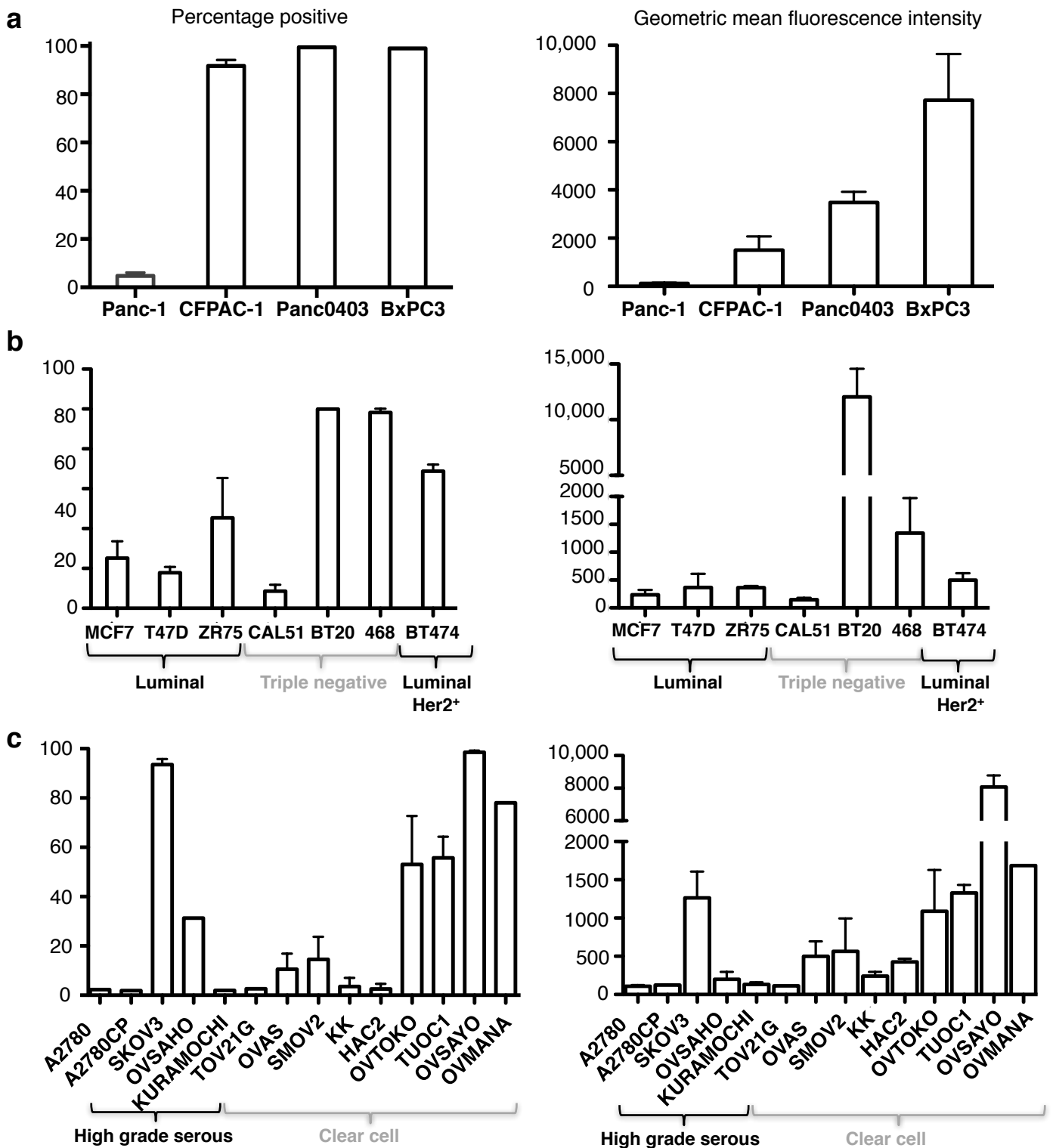


Supplemental Information

Targeting of Aberrant $\alpha v \beta 6$ Integrin Expression in Solid Tumors Using Chimeric Antigen Receptor-Engineered T Cells

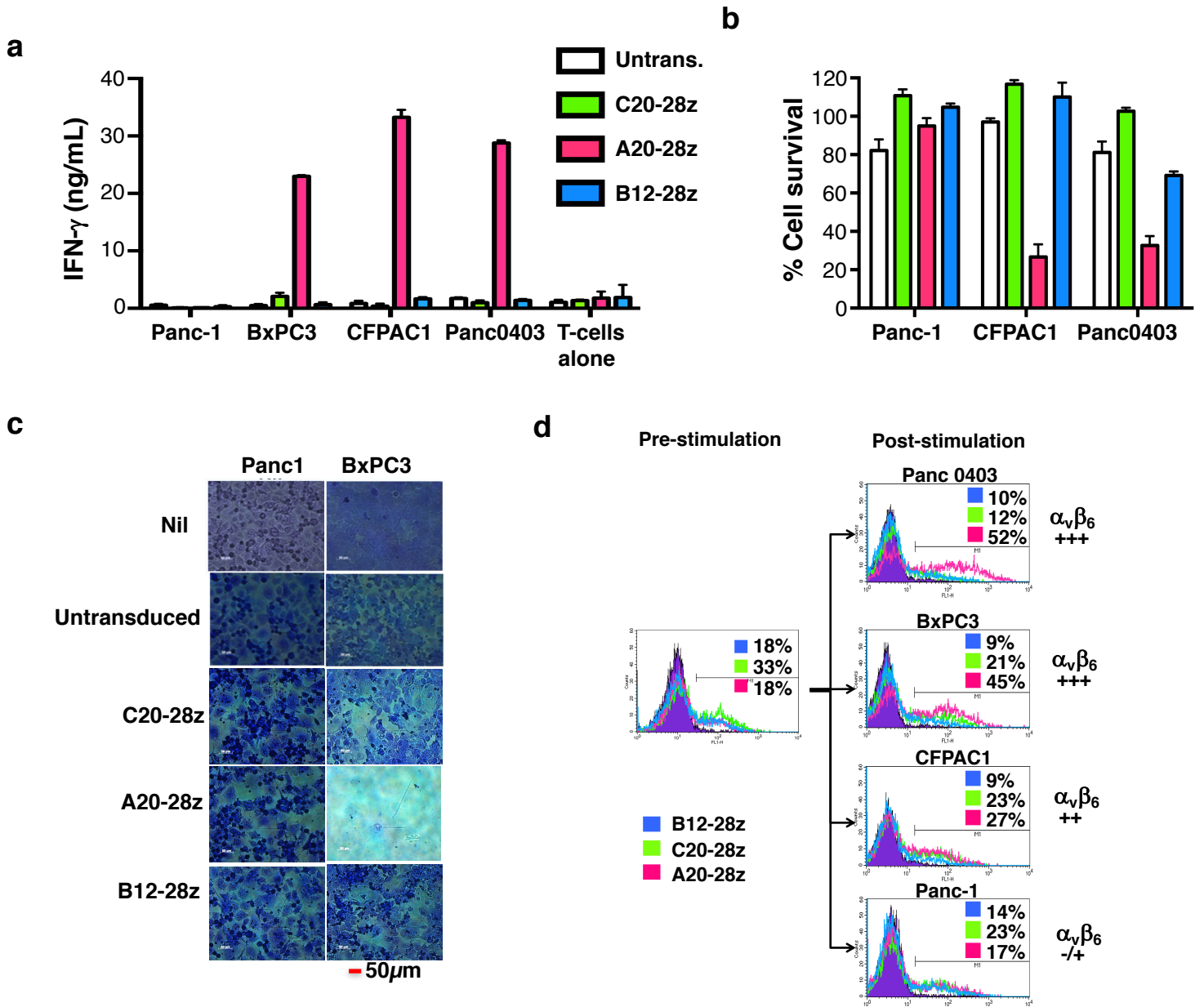
Lynsey M. Whilding, Ana C. Parente-Pereira, Tomasz Zabinski, David M. Davies, Roseanna M.G. Petrovic, Y. Vincent Kao, Shobhit A. Saxena, Alex Romain, Jose A. Costa-Guerra, Shelia Violette, Hiroaki Itamochi, Sadaf Ghaem-Maghani, Sabari Vallath, John F. Marshall, and John Maher

Supplementary Figure S1



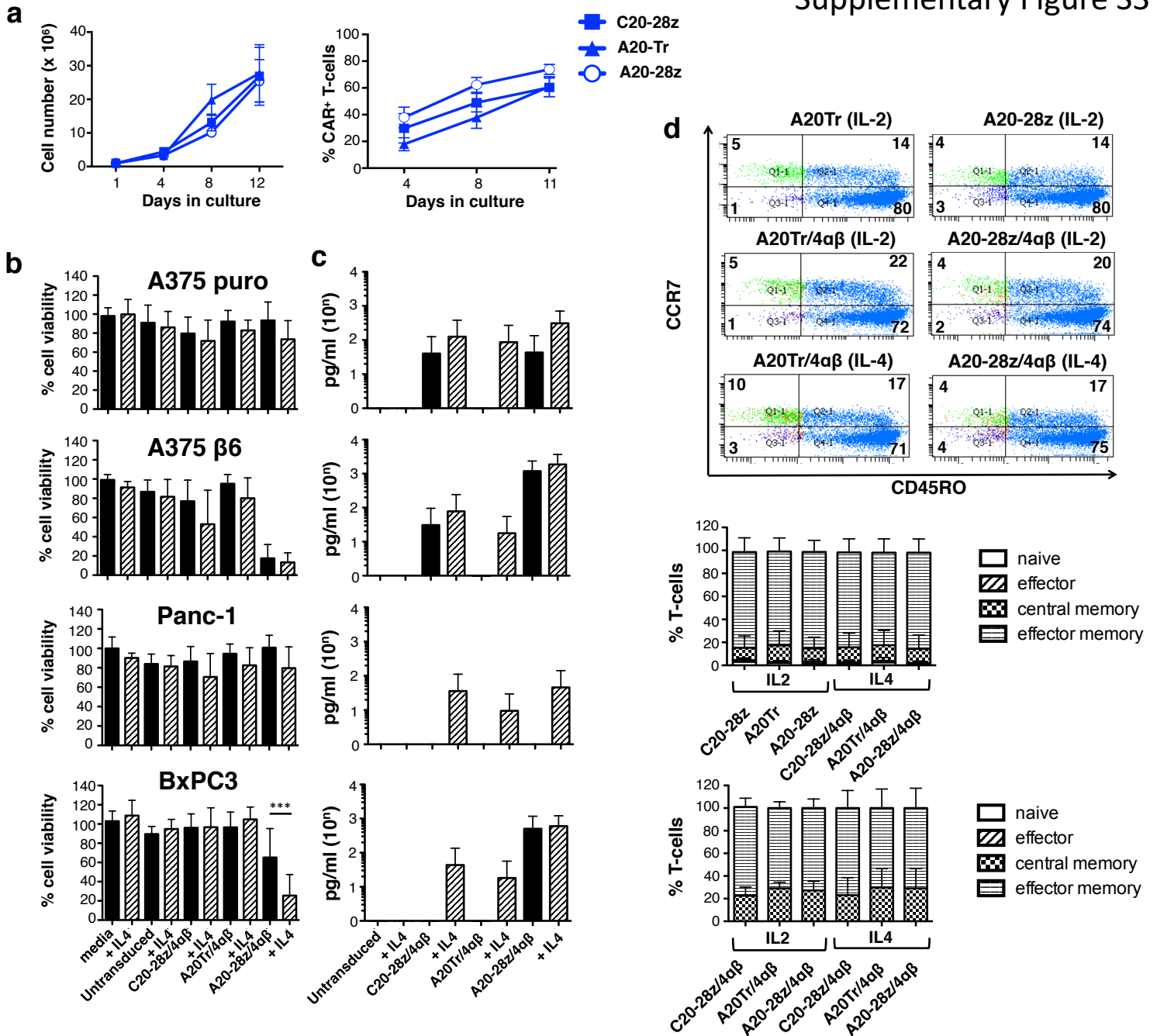
Supplementary Figure S1: Expression of $\alpha\beta6$ integrin by tumor cell lines. Immortalized pancreatic (a), breast (b) and ovarian cancer cell lines (c) cells were analyzed for expression of the $\beta6$ integrin subunit by flow cytometry after incubation with the 6.3G9 antibody followed by goat anti-mouse IgG-PE. Cells stained with secondary antibody alone served as negative control. Data show the mean \pm SD from 2-8 independent experiments.

Supplementary Figure S2

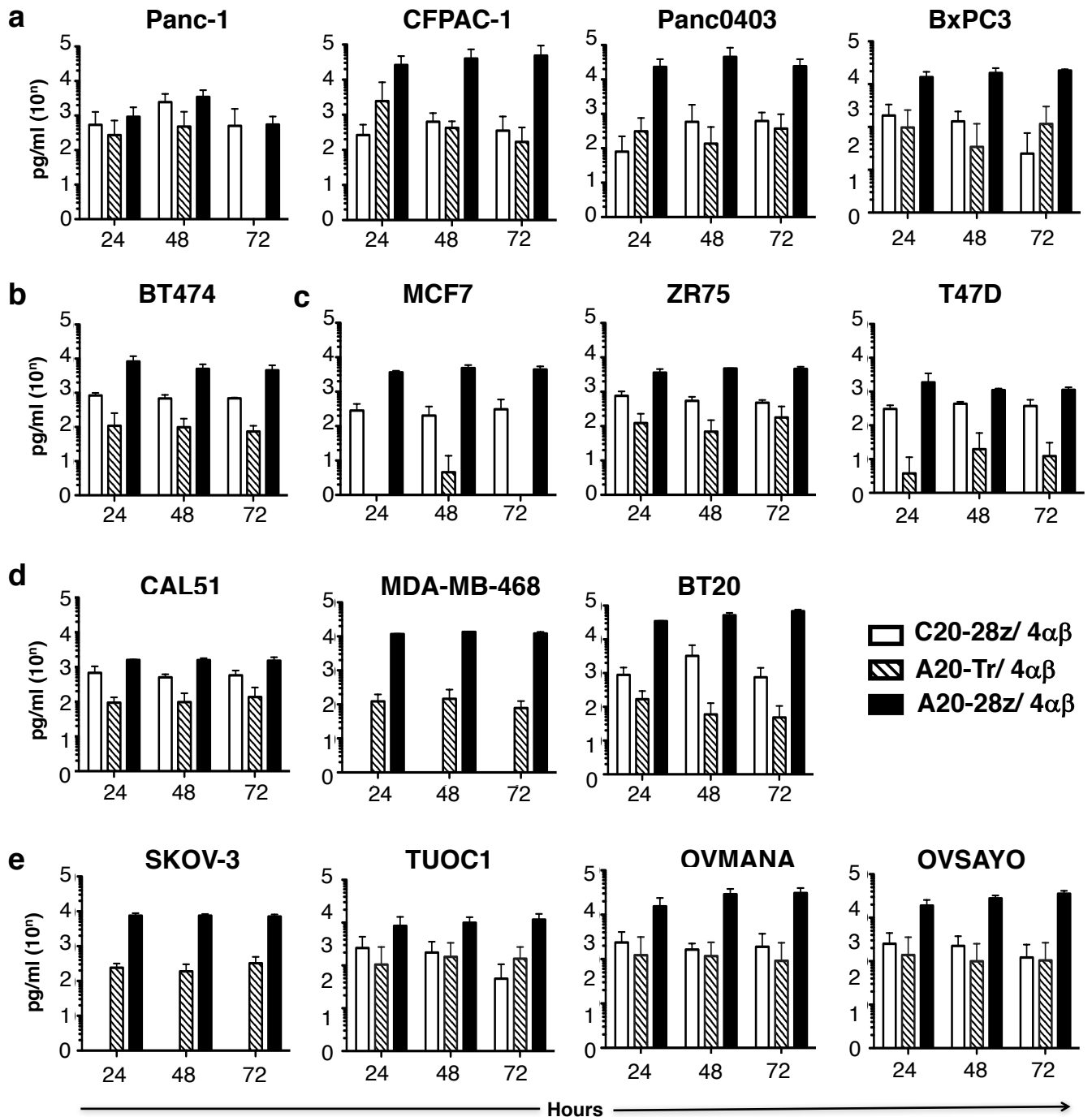


Supplementary Figure S2: *In vitro* comparison of anti-tumor activity of candidate $\alpha_v\beta_6$ -specific CARs. (a) T-cells that expressed indicated CARs were co-cultivated at a 10:1 ratio with the specified pancreatic cell lines. After 24h, IFN- γ was measured in harvested supernatants while residual tumor cell viability was assessed by MTT assay (b). Both datasets show mean \pm SD of 6 replicates. (c) 1×10^6 T-cells that expressed indicated CARs were co-cultivated with a confluent monolayer (24 well plate) of the indicated pancreatic tumor cell line, making comparison with untransduced T-cells or no addition (nil). After 24h, cultures were supplemented with IL-2 (100U/mL), which was added thereafter three times per week. Residual tumor monolayers were stained by crystal violet on day 8. (d) T-cells described in c were analyzed by flow cytometry for CAR expression before and after culture with the indicated monolayer for 8 days. Crystal violet and CAR T-cell enrichment data are representative of three independent cultures with $\alpha_v\beta_6$ -expressing pancreatic tumor cell lines.

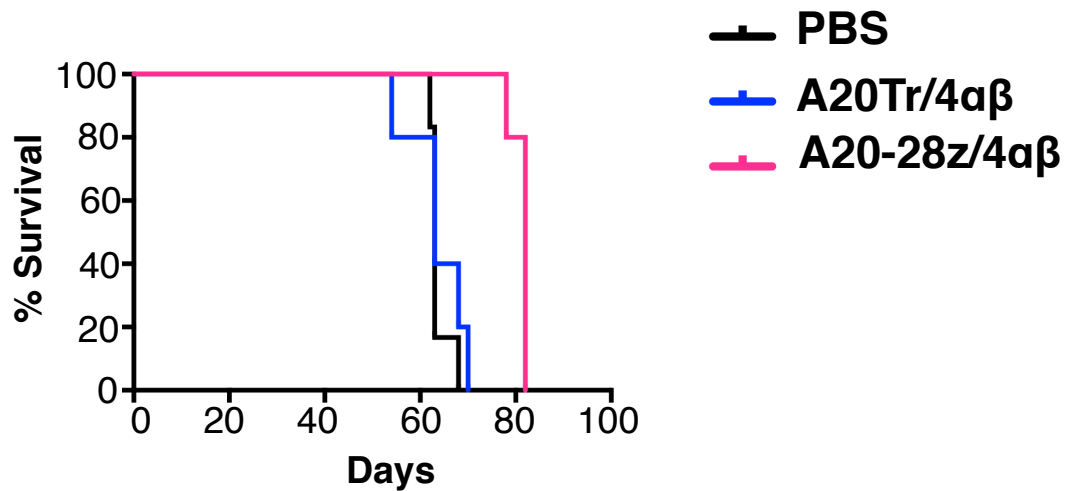
Supplementary Figure S3



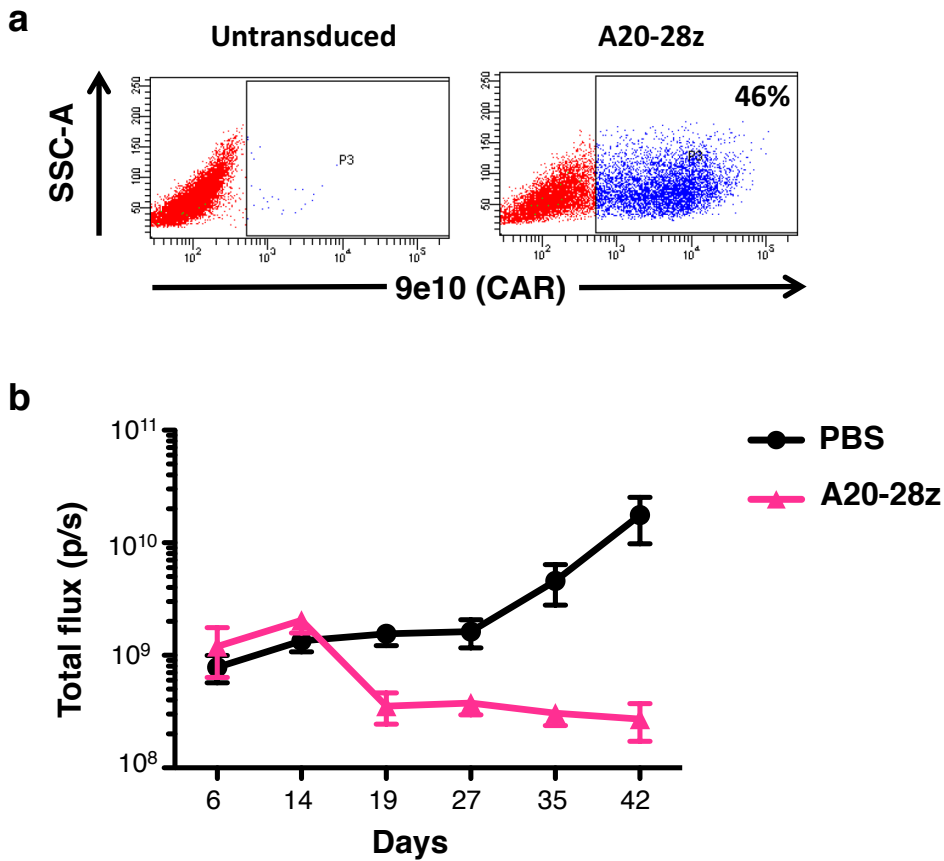
Supplementary Figure S3: Expansion and enrichment of CAR T-cells in IL-4. (a) Culture of 4 $\alpha\beta$ -expressing CAR T-cells in IL-4, demonstrating expansion and enrichment of the indicated CAR T-cell populations (mean \pm SD, $n=5-6$ independent replicates). Tumour cells were co-cultivated at a 1:1 ratio with the indicated CAR/4 $\alpha\beta$ -engineered T-cells in the absence or presence of exogenous cytokine (30ng/ml), following *ex vivo* expansion and enrichment of CAR T-cells using IL-4. (b) Cytotoxicity and (c) IFN- γ release was analysed at 48hrs. Data show the mean mean \pm SD of 5 independent experiments, each performed in triplicate. (d) The phenotype of T-cells cultured in either IL-2 or IL-4 for 12 days was assessed by flow cytometry. Data show representative plots and pooled data from T-cells derived from 5 independent donors.



Supplementary Figure S4: Production of interferon (IFN)- γ by $\alpha v\beta 6$ re-targeted CAR T-cells. Firefly luciferase-expressing pancreatic (a), HER2⁺ breast (b), luminal breast (c), triple negative breast (d), or ovarian tumor cells (e) were co-cultivated at a 1:1 ratio with the indicated CAR/ $4\alpha\beta$ -engineered T-cells in the absence of exogenous cytokine, following *ex vivo* expansion and enrichment of CAR T-cells using IL-4. Supernatant was harvested at each time-point and analyzed for IFN- γ . Data shows the mean \pm SD of 3-6 independent replicates.

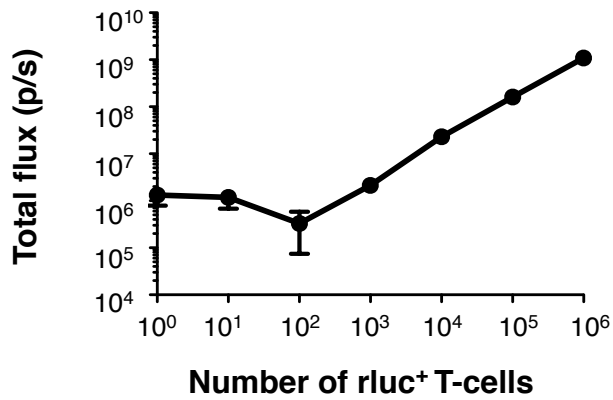


Supplementary Figure S5: Survival curve. Mice were injected i.p. with 1×10^6 SKOV-3-ffluc cells and tumors were allowed to establish for 21 days before i.p. treatment with 10×10^6 of the indicated gene-modified T-cells. Control mice received PBS. Animals were culled when humane endpoints were reached. $p = 0.0014$ comparing A20-28z/4αβ treated mice with both other groups using the Log-rank (Mantel-Cox) test.

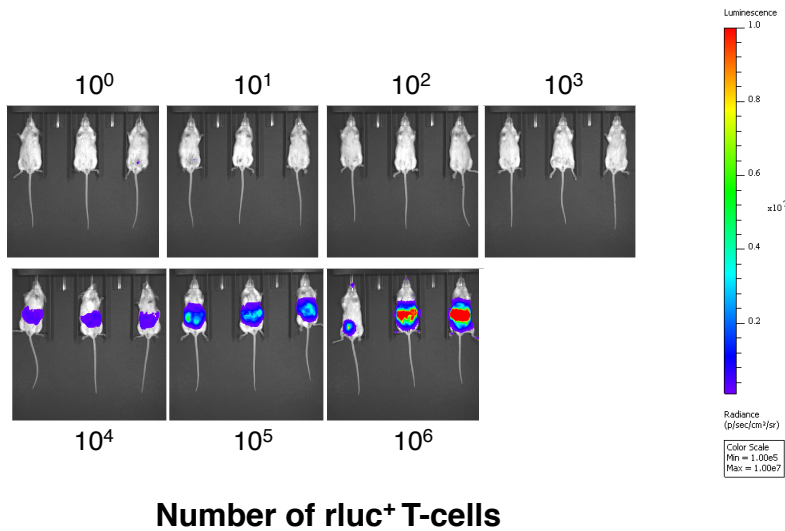


Supplementary Figure S6: *In vivo* anti-tumor activity of $\alpha\beta 6$ re-targeted CAR T-cells against Panc0403 PDAC xenografts. (a) T-cells were transduced with a retroviral vector encoding for A20-28z (e.g. without $4\alpha\beta$). After culture for 6 days in IL-2, cells were analyzed by flow cytometry. 9e10 detects a myc epitope tag in the CAR ectodomain. SSC – side scatter. Gates were set using untransduced T-cells cultured in IL-2. (b) Mice were injected i.p. with 2×10^6 Panc0403-ffluc cells and tumors were allowed to establish for 14 days before i.p. treatment with 20×10^6 of A20-28z T-cells or PBS as control. Bioluminescence imaging using d-luciferin (substrate for ffluc) was used to monitor tumor status. Data show the mean \pm SD of tumor-derived total flux ($n=5$ mice per group). The arrow indicates the day of treatment with CAR T-cells.

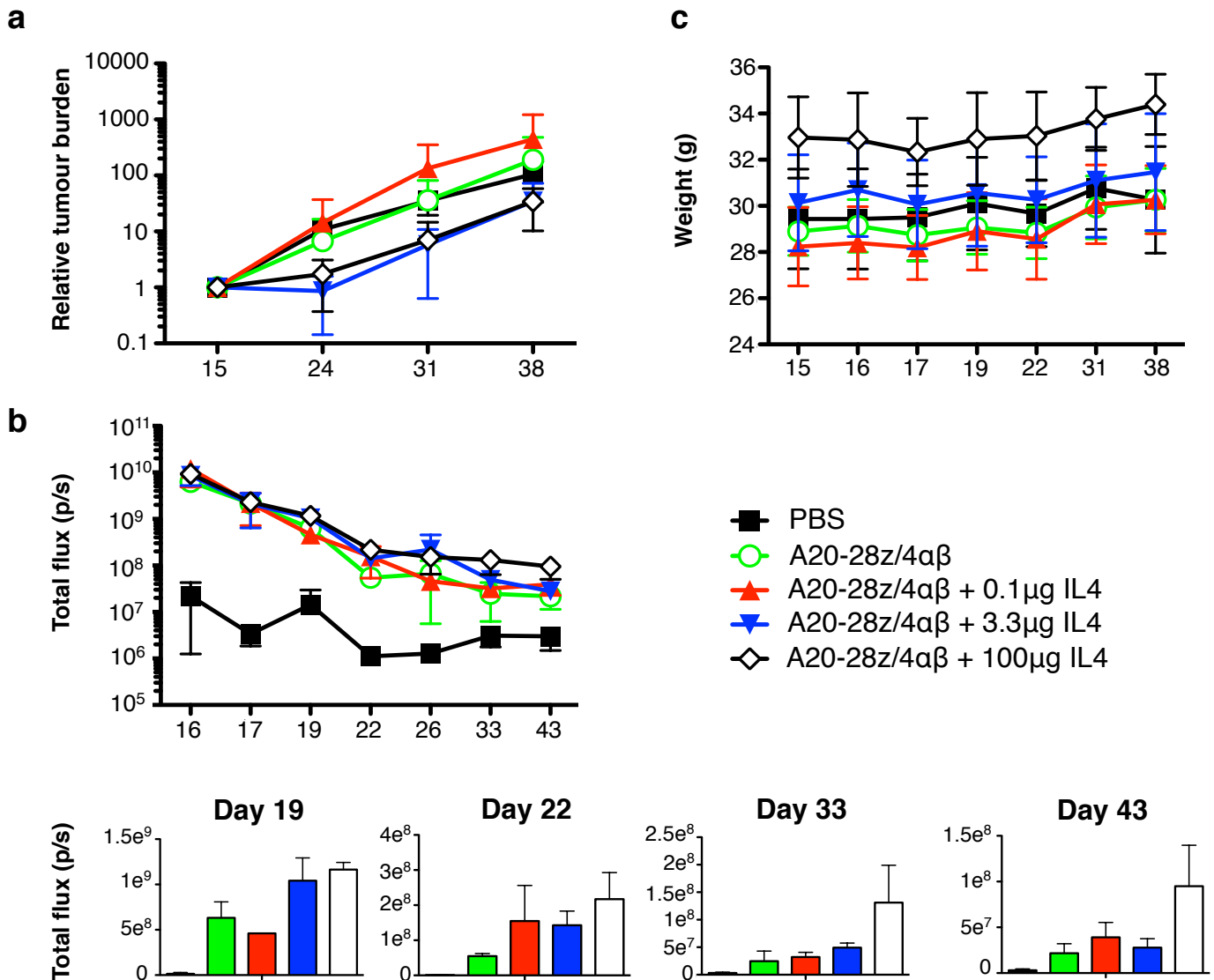
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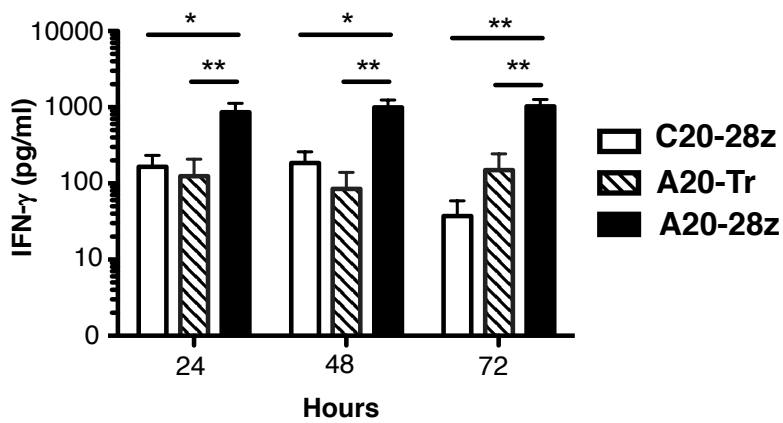
b



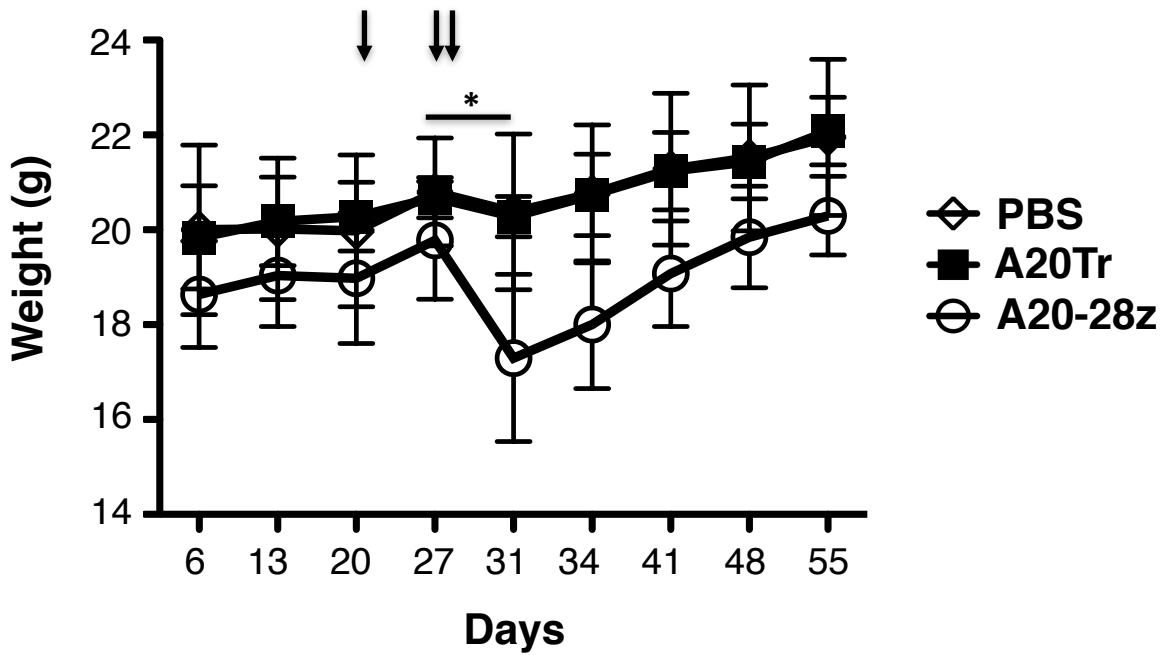
Supplementary Figure S7: *In vivo* imaging of adoptively transferred T-cells that express *Renilla luciferase*. (a) T-cells were transduced with a retroviral vector encoding rluc/GFP. The indicated number of rluc⁺ T-cells were injected i.p. into SCID-Beige mice, followed by i.p. injection of substrate (coelenterazine) and BLI imaging after 30 minutes. The graph shows the mean \pm SD of 3 mice/group. Images of individual mice are shown in (b).



Supplementary Figure S8: Tumor stress test to evaluate effect of exogenous IL-4 treatment on T-cell activity *in vivo*. 1×10^5 BxPC3-ffluc cells were injected i.p. in NSG mice. Tumors were allowed to establish for 16 days prior to injection of 2.5×10^6 CAR T-cells i.p. Either PBS or the indicated dose of human IL-4 was administered 3 times per week thereafter using the i.p. route. T-cells were transduced with A20-28z/4 $\alpha\beta$ CAR and a retroviral vector encoding rLuc/GFP to enable imaging of CAR T-cells. **(a)** Tumour burden was assessed weekly by BLI. Tumour burden is expressed relative to pre-treatment levels. **(b)** T-cells were imaged using coelenterazine administered i.p.. All data show the mean \pm SD of 3 mice/group. **(c)** Weight of mice during the study.



Supplementary Figure S9: Production of IFN- γ by $\alpha\beta 6$ re-targeted human CAR T-cells when stimulated by mouse tumor cells that naturally express $\alpha\beta 6$ integrin. 4T1 cells were co-cultivated at a 1:1 ratio with the indicated CAR/4 $\alpha\beta$ -engineered T-cells in the absence of exogenous cytokine, following *ex vivo* expansion and enrichment of CAR T-cells using IL-4. Supernatant was harvested at each time-point and analyzed for IFN- γ . Data shows the mean \pm SD of 3 independent experiments, each performed in duplicate. * $p < 0.05$; ** $p < 0.01$.



Supplementary Figure S10: *In vivo* safety testing of human $\alpha\beta 6$ -retargeted CAR T-cells. SCID Beige mice received the indicated CAR T-cell populations that had been enriched to homogeneity following *ex vivo* culture in IL-4. Bolus doses of 20 million CAR T-cells were administered by iv (tail vein) injection at timepoints indicated by the overhead arrows. Animals were weighed at the indicated intervals (mean \pm SD, n=5 mice per group). * $p < 0.05$ for comparison of weight of A20-28z-treated mice on day 31 versus day 27.