

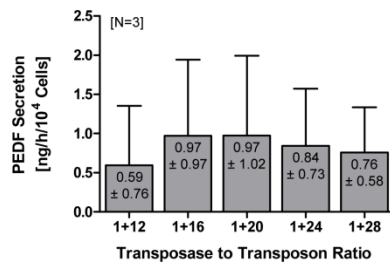
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## Supplemental Information

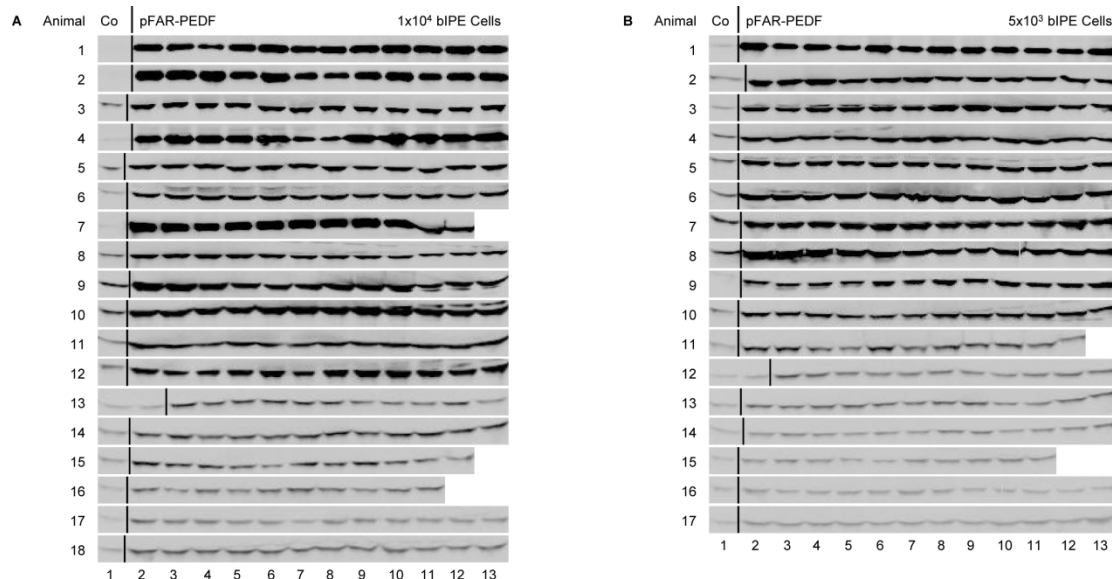
### Engineering of PEDF-Expressing Primary Pigment Epithelial Cells by the *SB* Transposon System Delivered by pFAR4 Plasmids

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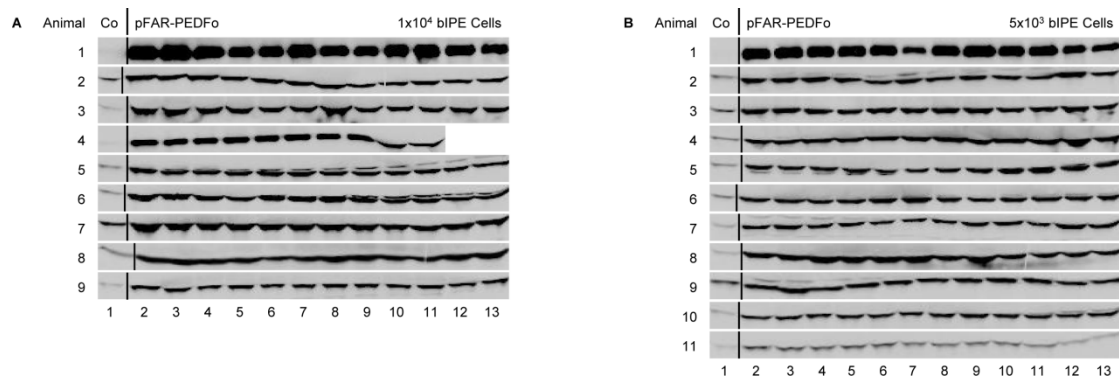
## Supplemental Figures and Legends



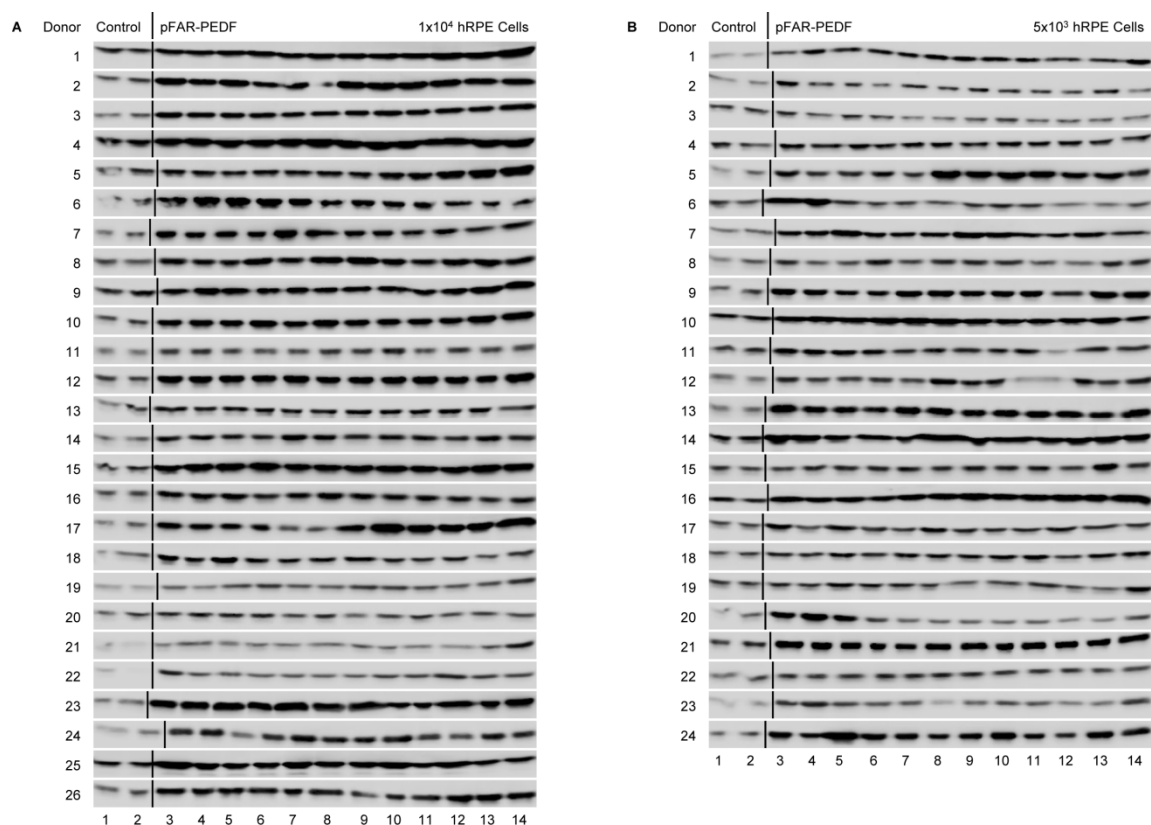
**Figure S1.** ELISA quantification of total PEDF secreted by primary human RPE cells transfected with varying ratios of SB100X transposase plasmid to PEDF transposon plasmid. For each donor sample, 2 transfections of  $1 \times 10^5$  cells with 0.038  $\mu\text{g}$  to 0.017  $\mu\text{g}$  pFAR4-CMV SB100X SV40 transposase and 0.462  $\mu\text{g}$  to 0.483  $\mu\text{g}$  pFAR4-ITRs CMV PEDF BGH plasmid DNA were carried out. PEDF secretion was analyzed in 3 human donor eyes (age:  $61.7 \pm 13.3$  years; gender: 2 males and 1 female; time postmortem:  $20.1 \pm 1.4$  hours; cultivation time before transfection:  $45.7 \pm 17.0$  days). Data are presented as mean  $\pm$  SD. Statistical analysis by one-way ANOVA with Bonferroni's Multiple Comparison Test showed no significant differences.



**Figure S2.** Western blot analysis of PEDF secretion by primary bovine IPE cells transfected with the pFAR4-ITRs CMV PEDF BGH transposon miniplasmid. Each analysis included 1 or 2 control transfections without the addition of miniplasmid DNA (column 1-2) and 10-12 transfections using 0.03  $\mu\text{g}$  pFAR4-CMV SB100X SV40 transposase and 0.47  $\mu\text{g}$  pFAR4-ITRs CMV PEDF BGH transposon miniplasmid DNA (columns 2/(3)-13). Culture supernatants were analyzed for total PEDF secretion using anti-PEDF antibodies 21 days after transfection. (A) Western blots of PEDF secreted by  $1 \times 10^4$  transfected IPE cells isolated from 18 bovine eyes (cultivation time before transfection:  $26.9 \pm 9.76$  days). (B) Western blots of PEDF secreted by  $5 \times 10^3$  transfected IPE cells isolated from 17 bovine eyes (cultivation time before transfection:  $27.9 \pm 13.6$  days). Note that the same exposure time was used for all chemiluminescence reactions.

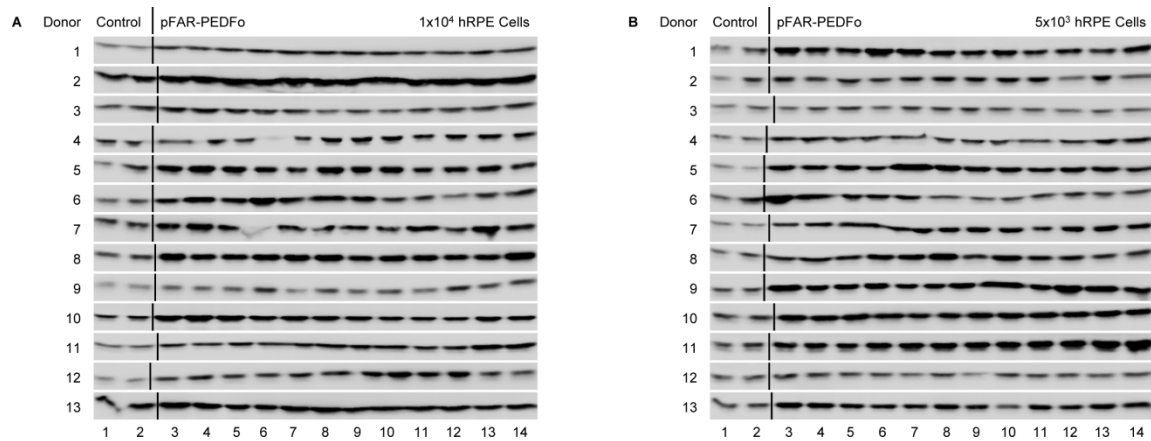


**Figure S3. Western blot analysis of PEDF secretion by primary bovine IPE cells transfected with the pFAR4-ITRs CMV PEDFOptimized BGH transposon miniplasmid.** Each analysis included 1 control transfection without the addition of miniplasmid DNA (column 1) and 10-12 transfections using 0.03  $\mu\text{g}$  pFAR4-CMV SB100X SV40 transposase and 0.47  $\mu\text{g}$  pFAR4-ITRs CMV PEDFOptimized BGH transposon miniplasmid DNA (columns 2-13). Culture supernatants were analyzed for total PEDF secretion using anti-PEDF antibodies 21 days after transfection. **(A)** Western blots of PEDF secreted by  $1 \times 10^4$  transfected IPE cells isolated from 9 bovine eyes (cultivation time before transfection:  $22.6 \pm 7.15$  days). **(B)** Western blots of PEDF secreted by  $5 \times 10^3$  transfected IPE cells isolated from 11 bovine eyes (cultivation time before transfection:  $29.2 \pm 15.2$  days). Note that the same exposure time was used for all chemiluminescence reactions.



**Figure S4. Western blot analysis of PEDF secretion by primary human RPE cells transfected with the pFAR4-ITRs CMV PEDF BGH transposon miniplasmid.** Each analysis included 2 control transfections without the addition of miniplasmid DNA (columns 1-2) and 12 transfections using 0.03  $\mu\text{g}$  pFAR4-CMV SB100X SV40 transposase and 0.47  $\mu\text{g}$  pFAR4-ITRs CMV PEDF BGH transposon miniplasmid DNA (columns 3-14). Culture supernatants were analyzed for total PEDF secretion using anti-PEDF antibodies 21 days after transfection. **(A)** Western blots of PEDF secreted by  $1 \times 10^4$  transfected RPE cells isolated from 26 human donor eyes (age:  $67.5 \pm 12.0$  years; gender: 13 males and 13 females; time postmortem:  $31.7 \pm 14.4$  hours; cultivation time before transfection:  $42.0 \pm 14.4$  days). **(B)** Western blots of PEDF secreted by  $5 \times 10^3$  transfected RPE cells isolated from 24 human donor eyes (age:  $66.2 \pm 14.6$  years; gender: 14 males and 10 females; time postmortem:

32.9 ± 14.5 hours; cultivation time before transfection: 45.2 ± 15.8 days). Note that the same exposure time was used for all chemiluminescence reactions.



**Figure S5. Western blot analysis of PEDF secretion by primary human RPE cells transfected with the pFAR4-ITRs CMV PEDFOptimized BGH transposon miniplasmid.** Each analysis included 2 control transfections without the addition of miniplasmid DNA (columns 1-2) and 12 transfections using 0.03 µg pFAR4-CMV SB100X SV40 transposase and 0.47 µg pFAR4-ITRs CMV PEDFOptimized BGH transposon miniplasmid DNA (columns 3-14). Culture supernatants were analyzed for total PEDF secretion using anti-PEDF antibodies 21 days after transfection. **(A)** Western blots of PEDF secreted by 1x10<sup>4</sup> transfected RPE cells isolated from 13 human donor eyes (age: 61.1 ± 16.3 years; gender: 7 males and 6 females; time postmortem: 33.5 ± 16.2 hours; cultivation time before transfection: 39.8 ± 17.1 days). **(B)** Western blots of PEDF secreted by 5x10<sup>3</sup> transfected RPE cells isolated from 13 human donor eyes (age: 61.1 ± 16.3 years; gender: 8 males and 5 females; time postmortem: 32.5 ± 16.1 hours; cultivation time before transfection: 42.6 ± 18.6 days). Note that the same exposure time was used for all chemiluminescence reactions.

## Supplemental Tables

**Table S1. Western blot-based quantification of long-term PEDF secretion by primary bovine IPE cells after SB100X-mediated transfection.** Each analysis included 2 control transfections without the addition of miniplasmid DNA and 2 transfections using 0.03 µg pFAR4-CMV SB100X SV40 transposase and 0.47 µg pFAR4-ITRs CMV PEDF BGH or pFAR4-ITRs CMV PEDFoptimized BGH miniplasmid DNA. Culture supernatants were analyzed for total PEDF secretion at times from 5 to 20 weeks. Western blot signal intensities of PEDF-transfected and PEDFo-transfected cells were normalized to the signal intensities of the control cells. Using an unpaired two-tailed t test, a statistical difference was observed between control and transfected cells, but not between cells transfected with pFAR-PEDF and pFAR-PEDFo at any time period.

		Time [Days]	N [Samples]	Signal Intensities			pFAR-PEDF versus pFAR-PEDFo
				Mean ± SD	Range		
<b>1x10<sup>4</sup> bIPE Cells</b>	Control	49 ± 1	18	1.00 ± 0.25	0.54-1.46		
	pFAR-PEDF			19.9 ± 22.8	1.95-74.9	**P=0.0013	
	pFAR-PEDFo			17.1 ± 16.1	2.29-58.1	***P=0.0002	Not significant
	Control	102 ± 9	18	1.00 ± 0.41	0.24-1.76		
	pFAR-PEDF			17.4 ± 13.2	1.34-42.6	****P<0.0001	
	pFAR-PEDFo			22.6 ± 18.8	0.89-58.5	****P<0.0001	Not significant
<b>1x10<sup>4</sup> bIPE Cells</b>	Control	136 ± 10	18	1.00 ± 0.30	0.44-1.56		
	pFAR-PEDF			15.2 ± 14.5	2.93-65.6	***P=0.0002	
	pFAR-PEDFo			13.5 ± 8.74	2.98-29.7	****P<0.0001	Not significant
	Control	221 ± 10	16	1.00 ± 0.36	0.30-1.70		
	pFAR-PEDF			5.97 ± 4.20	1.16-18.4	****P<0.0001	
	pFAR-PEDFo			6.00 ± 4.54	1.53-16.9	***P=0.0001	Not significant
<b>5x10<sup>3</sup> bIPE Cells</b>	Control	365 ± 3	16	1.00 ± 0.24	0.49-1.51		
	pFAR-PEDF			13.5 ± 23.7	0.50-73.8	*P=0.0499	
	pFAR-PEDFo			12.2 ± 22.1	0.19-76.3	Not significant	Not significant
	Control	49 ± 1	11	1.00 ± 0.24	0.54-1.46		
	pFAR-PEDF			15.2 ± 14.4	1.75-55.6	****P<0.0001	
	pFAR-PEDFo			12.4 ± 12.5	1.28-57.0	****P<0.0001	Not significant
<b>5x10<sup>3</sup> bIPE Cells</b>	Control	103 ± 9	11	1.00 ± 0.40	0.24-1.76		
	pFAR-PEDF			13.9 ± 12.3	2.56-43.9	****P<0.0001	
	pFAR-PEDFo			19.1 ± 17.3	1.00-50.6	****P<0.0001	Not significant
	Control	136 ± 9	11	1.00 ± 0.29	0.44-1.56		
	pFAR-PEDF			10.2 ± 6.87	2.08-25.1	****P<0.0001	
	pFAR-PEDFo			8.60 ± 7.55	1.84-27.8	****P<0.0001	Not significant
<b>5x10<sup>3</sup> bIPE Cells</b>	Control	221 ± 9	10	1.00 ± 0.33	0.30-1.70		
	pFAR-PEDF			7.92 ± 7.18	1.09-30.2	***P=0.0001	
	pFAR-PEDFo			7.88 ± 8.28	0.52-26.1	***P=0.0007	Not significant
	Control	365 ± 3	10	1.00 ± 0.39	0.00-2.00		
	pFAR-PEDF			8.64 ± 11.1	0.44-37.8	**P=0.0040	
	pFAR-PEDFo			8.79 ± 19.2	0.20-86.8	Not significant	Not significant

**Table S2. Western blot-based quantification of long-term PEDF secretion in primary human RPE cells after SB100X-mediated transfection.** Each analysis included 2 control transfections without the addition of miniplasmid DNA and 2 transfections using 0.03 µg pFAR4-CMV SB100X SV40 transposase and 0.47 µg pFAR4-ITRs CMV PEDF BGH or pFAR4-ITRs CMV PEDFoptimized BGH miniplasmid DNA. Culture supernatants were analyzed for total PEDF secretion at the times indicated. Western blot signal intensities of PEDF-transfected and PEDFo-transfected cells were normalized to the signal intensities of the control cells. Statistical analysis by unpaired two-tailed t test showed a significant difference between transfected and non-transfected cells. No statistical significant difference was observed between cells transfected with pFAR-PEDF and pFAR-PEDFo for the first 110 days after transfection. However, at 160 days, a statistical significant decrease in PEDF secretion by pFAR-PEDF transfected cells compared to pFAR-PEDFo transfected cells was observed.

		Time [Days]	N [Samples]	Signal Intensities			pFAR-PEDF versus pFAR-PEDFo
				Mean ± SD	Range		
1x10 <sup>4</sup> hRPE Cells	Control	31 ± 5	10	1.00 ± 0.15	0.74-1.26		
	pFAR-PEDF			6.36 ± 3.27	2.08-11.3	****P<0.0001	
	Control	27 ± 1	4	1.00 ± 0.22	0.73-1.27		
	pFAR-PEDFo			10.7 ± 4.86	5.07-16.7	**P=0.0071	Not significant
	Control	61 ± 10	30	1.00 ± 0.25	0.56-1.44		
	pFAR-PEDF			6.03 ± 4.57	0.93-19.3	****P<0.0001	
	Control	64 ± 8	14	1.00 ± 0.27	0.61-1.39		
	pFAR-PEDFo			8.07 ± 4.14	1.71-15.5	****P<0.0001	Not significant
	Control	107 ± 8	30	1.00 ± 0.24	0.43-1.57		
	pFAR-PEDF			6.84 ± 7.03	0.41-27.1	****P<0.0001	
	Control	108 ± 7	14	1.00 ± 0.23	0.51-1.49		
	pFAR-PEDFo			8.13 ± 4.98	1.13-15.5	****P<0.0001	Not significant
	Control	162 ± 14	20	1.00 ± 0.15	0.75-1.25		
	pFAR-PEDF			2.51 ± 1.72	0.34-6.03	***P=0.0004	
Control	154 ± 4	8	1.00 ± 0.17	0.76-1.24			
pFAR-PEDFo			4.83 ± 2.49	1.66-8.12	***P=0.0007	**P=0.0098	
Control	215 ± 20	10	1.00 ± 0.10	0.85-1.15			
pFAR-PEDF			2.61 ± 2.12	0.78-7.33	*P=0.0278		
Control	193 ± 0	2	1.00 ± 0.08	0.94-1.06			
pFAR-PEDFo			8.10 ± 3.92	5.32-10.9	Not significant	*P=0.0135	
5x10 <sup>3</sup> hRPE Cells	Control	33 ± 5	14	1.00 ± 0.24	0.65-1.35		
	pFAR-PEDF			7.51 ± 4.84	2.13-19.4	****P<0.0001	
	Control	31 ± 6	8	1.00 ± 0.28	0.65-1.35		
	pFAR-PEDFo			6.88 ± 4.86	3.30-16.7	**P=0.0042	Not significant
	Control	62 ± 12	30	1.00 ± 0.23	0.56-1.44		
	pFAR-PEDF			7.29 ± 6.92	1.70-33.2	****P<0.0001	
	Control	64 ± 13	20	1.00 ± 0.19	0.76-1.24		
	pFAR-PEDFo			4.79 ± 2.75	0.81-10.7	****P<0.0001	Not significant
	Control	109 ± 10	26	1.00 ± 0.23	0.55-1.45		
	pFAR-PEDF			8.66 ± 11.4	1.32-55.8	**P=0.0012	
	Control	111 ± 11	16	1.00 ± 0.17	0.75-1.25		
	pFAR-PEDFo			8.47 ± 8.03	0.98-26.7	***P=0.0008	Not significant
	Control	162 ± 18	20	1.00 ± 0.25	0.56-1.44		
	pFAR-PEDF			3.20 ± 2.01	0.73-7.79	****P<0.0001	
Control	153 ± 12	10	1.00 ± 0.29	0.56-1.44			
pFAR-PEDFo			7.33 ± 7.90	1.35-23.8	*P=0.0208	*P=0.0379	
Control	216 ± 20	12	1.00 ± 0.40	0.43-1.57			
pFAR-PEDF			2.87 ± 2.02	0.63-6.61	**P=0.0048		
Control	201 ± 11	4	1.00 ± 0.47	0.43-1.57			
pFAR-PEDFo			7.53 ± 1.32	6.33-9.40	****P<0.0001	***P=0.0010	

**Table S3. Primer sequences for integration site profiling of the pFAR4-ITRs CMV PEDF BGH transposon in human RPE cells.**

<b>SB_20_hmr</b>	AACTTAAGTGTATGTAAACTTCCGACT
<b>4mer primers</b>	
<b>atcc_asPE</b>	GTGACTGGAGTTCAGACGTGTGCTCTTCCGATCTHNNNNNatcc
<b>ggat_asPE</b>	GTGACTGGAGTTCAGACGTGTGCTCTTCCGATCTBBNNNnggat
<b>gaca_asPE</b>	GGACTGGAGTTCAGACGTGTGCTCTTCCGATCTVVVNNNgaca
<b>gcaa_asPE</b>	GTGACTGGAGTTCAGACGTGTGCTCTTCCGATCTVNVNVNgcaa
<b>tgte_asPE</b>	GTGACTGGAGTTCAGACGTGTGCTCTTCCGATCTHHNNNtgte
<b>ttgc_asPE</b>	GTGACTGGAGTTCAGACGTGTGCTCTTCCGATCTHHNNNttgc
<b>SB-7</b>	cttAAGTGTATGTAAACTTCCGACTTCA
<b>PE_first</b>	Gtgactggagttcagacgtg
<b>SB_PE_noTA_BC_1</b>	ACACTCTTCCCTACACGACGCTCTTCCGATCTATCACGatGTAAACTTCCGACTTCAACTG
<b>SB_PE_noTA_BC_2</b>	ACACTCTTCCCTACACGACGCTCTTCCGATCTCGATGTatGTAAACTTCCGACTTCAACTG
<b>SB_PE_noTA_BC_3</b>	ACACTCTTCCCTACACGACGCTCTTCCGATCTTTAGGCatGTAAACTTCCGACTTCAACTG
<b>SB_PE_noTA_BC_4</b>	ACACTCTTCCCTACACGACGCTCTTCCGATCTTGACCAatGTAAACTTCCGACTTCAACTG
<b>PE nest</b>	CAAGCAGAAGACGGCATAACGAGAT_reverse_complement_of_Illumina_Truseq_barcode_GTGACTGG AGTTCAGACGTGTGCTCTTCCGATCT
<b>Illumina 1</b>	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTTCCGATCT Oligonucleotide sequences© 2006-2010 Illumina, Inc All rights reserved.