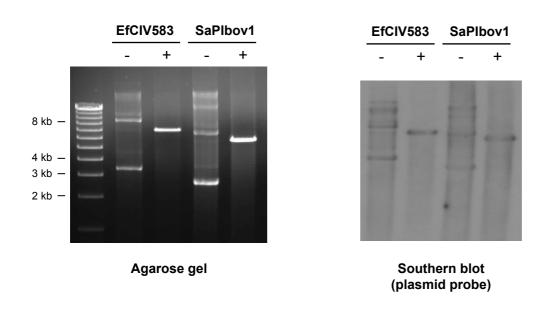
## A. PICI replication origins

PICI	Iteron sequence	Iteron orga	anization
Gram-positive		AT-rich region	
SaPlbov1	gtaccc	<del>+++</del> <del>++</del>	<del></del>
EfCIV583	gggag	<del>+++</del>	<del>+++</del>
SpnCIST556	tgtgac	<del></del>	<del>***</del>
SpnCITCH8431	tgtgac	<del></del>	<del>+++++</del>

## B. PICI autonomous replication



**Figure S1. Characterization of the PICI replication origins.** (A) Comparative map of the replication origins of several PICIs. The iterons are represented by arrows, and their sequences are shown at left. Note that there are always two sets of iterons flanking an AT-rich region, which could be the melting site. (B) Plasmid DNA carrying the Pri-Rep-*ori* segment of EfCIV583 or SaPlbov1 was isolated from overnight cultures of *S. aureus* RN4220, in presence of erythromycin. Plasmids were analyzed in agarose gels (left) or transferred for Southern blot studies using a probe specific for the plasmid (right). (-): non-digested plasmid; (+) plasmids digested with *Bam*HI, which cuts one in the plasmids.

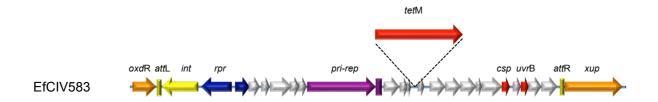
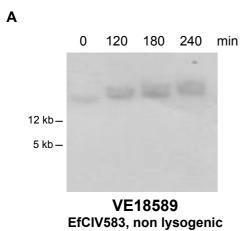
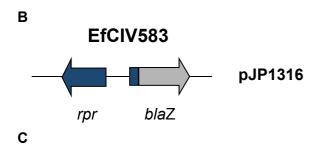
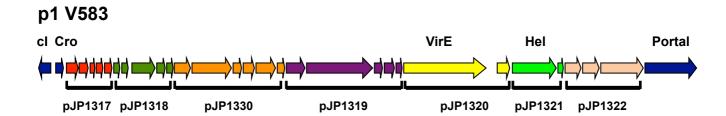
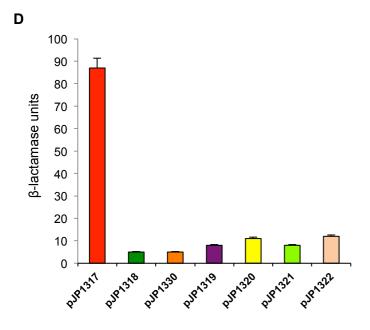


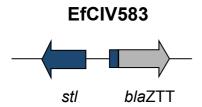
Figure S2. Location of the *tet*M marker in the EfCIV583 genome.













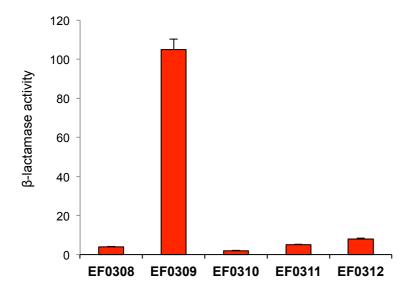
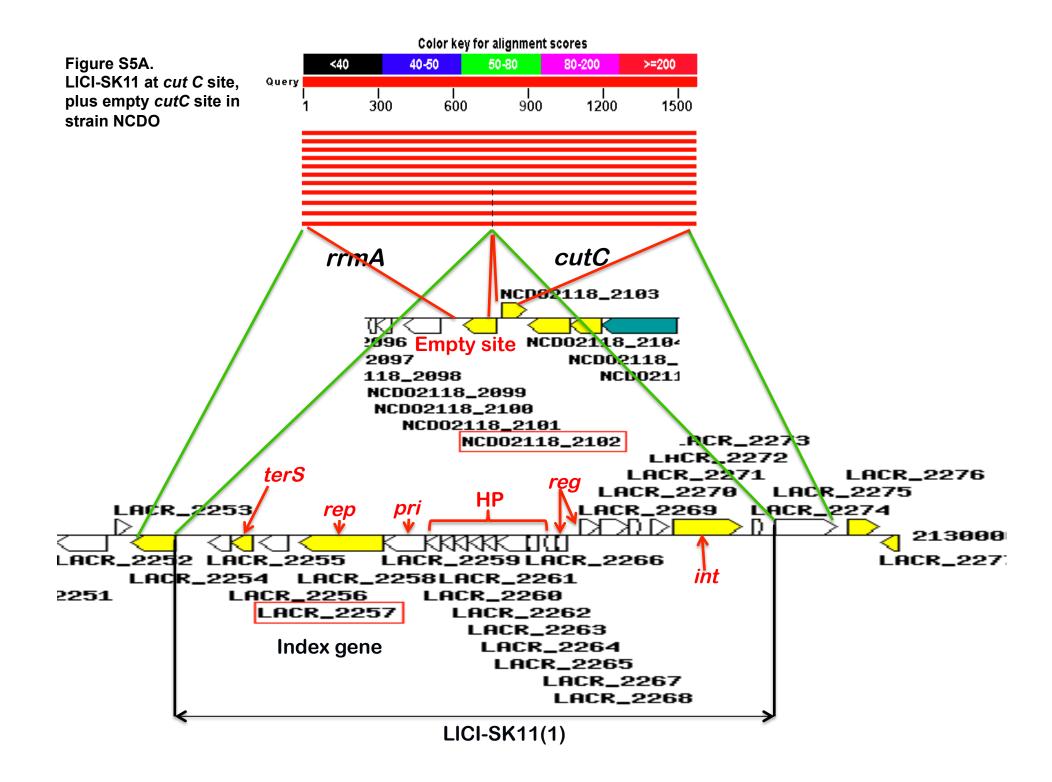
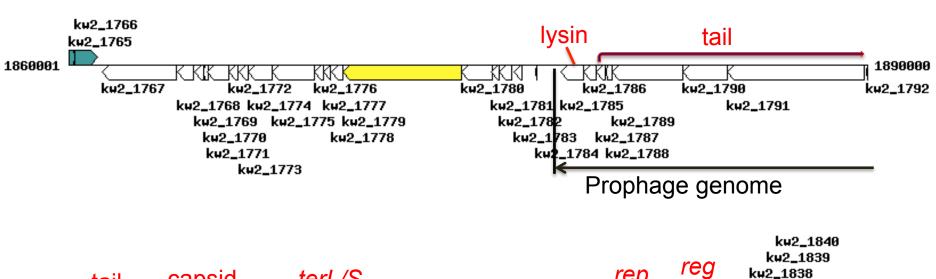


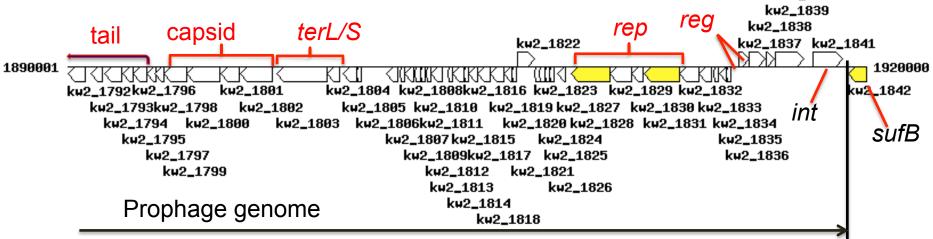
Figure S3. Identification of p1 region containing the derepressor of EfCIV583. (A) Failure of MC to induce excision/replication of EfCIV583. A culture of VE18589 (EfCIV538 positive) was induced with MC and samples taken during the subsequent incubation were used to prepare minilysates that were separated on agarose and Southern blotted with a EfCIV538 probe. (B) Construct used to test for de-repression. (C) Map of p1 showing regions that were cloned and tested for de-repression. *S. aureus* RN4220 derivative strains containing pJP1316 and pCN51 derivative plasmids (Pcad promoter) containing the different regions of p1 were assayed for β-lactamase activity after induction with 5 μM CdCl<sub>2</sub>. Samples were normalized for total cell mass. (D) β-lactamase activity generated by the above reporter construct in the presence of each of the cloned p1 regions, following induction with 5 μm CdCl<sub>2</sub>. (E) Demonstration of the de-repression activity of a subclone of pJP1317. Tests were performed with the construct shown in B. Values presented are the averages (±SD) of three independent assays.

## Enterococcus faecalis oxdR attL int attR xup csp uvrB EfCIV583 attL pri-rep attR hipB EfCILA3B-2 oxdR attL int attR xup EfCI918 oxdR attL int pri-rep EfCIB16457 oxdR attL int pri-rep csp uvrB EfCISF105 **→**|||+ Integration-excision (*int/xis*) Replication(pri/rep) Accesory Attachment sites (attL/attR) Replication origin (ori) Hypothetical Regulatory Flanking genes

Figure S4. Enterococcus faecalis PICI genomes.







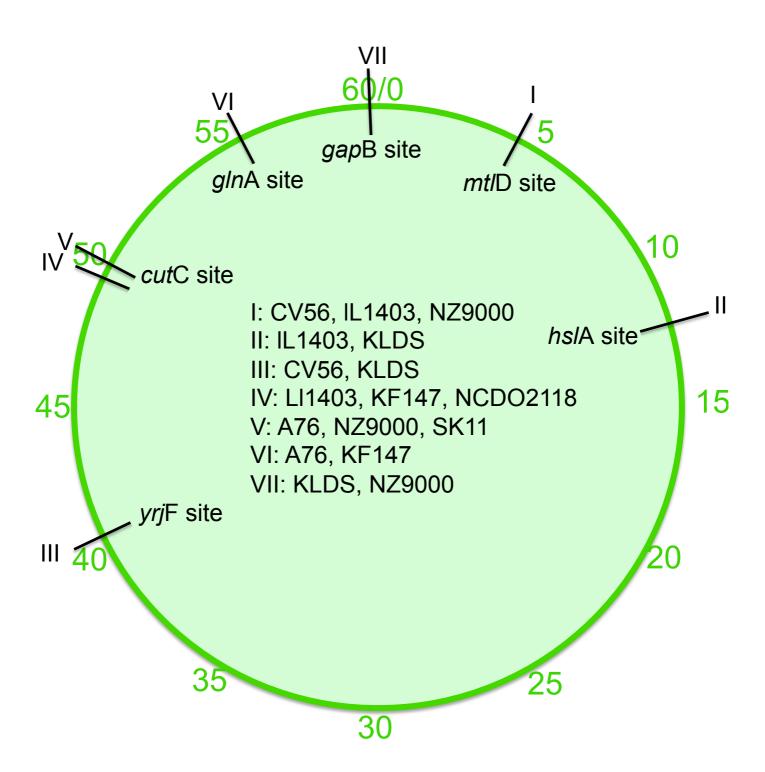


Figure S6. Locations of PICI att sites in lactococci.

Phage bIL286 LlCI-bIL310	TTTTAATAACCCTCCCCCGTATCTTTTCACAGGGAAACCACACACA
Phage bIL286 LlCI-bIL310	GCGTGAAAACCCATTTTTCAAAATTTTTATATAGGGGGGGTCAAAACACTAAAA GCGTGAAAGTCCATTTTTGAAAATTTTTATATAGG-GGGGTCAAAAATCTAAAA ******** ******* *****************
Phage bIL285 L1CINZ9000-2	AAAGAACCGAGTGAGTTTAGCTTTTTCCAAGTGTGAGGAAATTTGAAAATATTTTTTTAC AAAGAACCGAGTGAGTTTAGCTTTTTCCAAGTGTACAAAGTCCTGAATCTATTTTTTTAC ************************
Phage bIL309 LlCI-A76-2	CCCCCGTCATCGCTTTTAGGAATACCGTATACCAATGGTGGCTTCCTGAGTAAAAAAGT CCCCCGTCATTGCTTTTAGGAATACCGTATACCAATAGTGGCTCCCTGAGTAAAAAAGT ******** ****************************
Phage bIL309 LlCI-A76-2	GATTTTTTAAAATTTTTGCATAGGGGGGT GGTTTTTTAAAATTTTTGCATAGGGGGGGT *.*****************************

**Figure S7. Cos alignment.** The predicted latococcal PICI and phage *cos* sites and their flanking sequences are aligned using ClustalW2.