Photoredox Radical Conjugate Addition of Dithiane-2-carboxylate Promoted by an Iridium(III) Phenyltetrazole Complex: a Formal Radical Methylation of Michael Acceptors

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# **Electronic Supplementary Information**

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General methods and materials.  $^{1}$ H NMR spectra were recorded on Varian Gemini 200, Varian Mercury 400. Chemical shifts are reported in ppm from TMS with the solvent resonance as the internal standard (deuterochloroform:  $\delta = 7.24$  ppm). Data are reported as follows: chemical shift, multiplicity (s = singlet, d = duplet, t = triplet, q = quartet, dd = double duplet, dt = double triplet, bs = broad signal, m = multiplet), coupling constants (Hz).  $^{13}$ C NMR spectra were recorded on Varian Gemini 200, Varian MR400 spectrometers. Chemical shifts are reported in ppm from TMS with the solvent as the internal standard (deuterochloroform:  $\delta = 77.0$  ppm). GC-MS spectra were taken by EI ionization at 70 eV on a Hewlett-Packard 5971 with GC injection. LC-electrospray ionization mass spectra (ESI-MS) were obtained with Agilent Technologies MSD1100 single-quadrupole mass spectrometer. They are reported as: m/z (rel. intense). IR spectra were recorded using Bruker Alpha FTIR spectrometer equipped with single reflection diamond ATR accessory. Chromatographic purification was done with 240-400 mesh silica gel. Purification on preparative thin layer chromatography was done on Merck TLC silica gel 60 F<sub>254</sub>.

**Materials.** If not otherwise stated, all reactions were carried out in flame dried glassware under nitrogen atmosphere. Anhydrous solvents were supplied by Aldrich in Sureseal® bottles and were used as received avoiding further purification.

Reagents were purchased from Aldrich and AlfaAesar, and used without further purification unless otherwise stated. Compounds 3e, 3f, 3h, 3i, 3l, 3m, 3o, 3p, 3q and 25 were commercially available. 1,3-dithianecarboxylic acid 4a was prepared according reported procedure. Iridium complexes 1,2 2,3 19-20,4 21,3 22,2 and 23-244 were prepared according to described procedures.

# Optimization of radical conjugate addition of dithiane-2-carboxylate

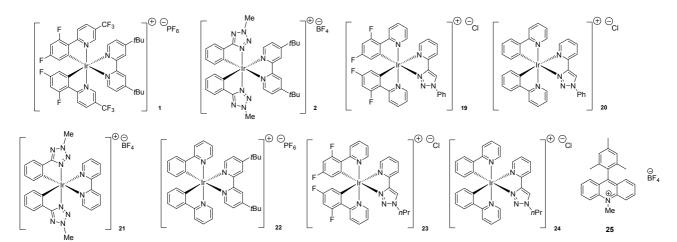


Table S1. Iridium catalyst screening.<sup>a</sup>

Entry	Catalyst	Conversion (%)b
1	1	93
2	2	97
3	19	18
4	20	17
5	21	72
6	22	47
7	23	13
8	24	13
9	25	0

 $<sup>^</sup>a$  The reactions were performed at r.t. with 0.1 mmol of 4a, 2 equiv of malonate (3a) and 1 equiv of  $K_2HPO_4$ , in the presence of 1 mol% of photocatalyst and in DMSO (0.2 M), and stopped after 16 h.  $^b$  Determined by  $^1H$  NMR using internal standard.

Table S2. Solvent screening.a

Entry	Solvent	Conversion (%)b
1	DMSO	97
2	DMF	53
3	DCE	11
4	EtOH	34
5	EtOH:H <sub>2</sub> O (8:2)	45

 $<sup>^</sup>a$  The reactions were performed at r.t. with 0.1 mmol of  ${\bf 4a}, 2$  equiv of malonate (3a) and 1 equiv of  $K_2HPO_4$ , in the presence of 1 mol% of iridium complex 2 and in reported solvent (0.2 M), and stopped after 16 h.  $^b$  Determined by  $^1H$  NMR using internal standard.

Table S3. Base screening.a

Entry	Base (equiv.)	Solvent	Conversion (%)b
1	Lutidine (1)	DMSO	65
2	Lutidine (0.5)	DMSO	35
3	-	DMSO	2
4	K <sub>2</sub> HPO <sub>4</sub> (1.5)	DMSO	75
5	K <sub>2</sub> HPO <sub>4</sub> (1)	DMSO	97
6	CsCO <sub>3</sub> (1)	DMSO	0
7	DBU (1)	DMSO	45
8	CsCO <sub>3</sub> (1)	DMF	0
9	DBU (1)	DMF	30

<sup>&</sup>lt;sup>a</sup> The reactions were performed at r.t. with 0.1 mmol of **4a**, 2 equiv of malonate (**3a**) and base, in the presence of 1 mol% of iridium complex **2** and in reported solvent (0.2 M), and stopped after 16 h. <sup>b</sup> Determined by <sup>1</sup>H NMR using internal standard.

Table S4

Entry	Conversion (%)b	Variation from standard conditions
1	97	-
2	0	No Light
3	0	No iridium complex
4	37	<b>4a:3a</b> 1.5:1
5	42	<b>4a:3a</b> 1:1
6	60	<b>4a:3a</b> 1:1.5
7	94	Temperature controlled at 25±3°C

 $<sup>^{\</sup>rm a}$  Standard conditions: 0.1 mmol of **4a**, 2 equiv of malonate (**3a**) and 1 equiv of  $K_2HPO_4$ , in the presence of 1 mol% of iridium complex **2** and in DMSO (0.2 M), and stopped after 16 h.  $^{\rm b}$  Determined by  $^{\rm l}H$  NMR using internal standard.

#### Preparation of starting Michael acceptors

Compounds **3a-d** were prepared following reported procedure.<sup>5</sup> To a solution of proline (1 mmol, 0.115 g) in DMSO (3.3 mL) aldehyde (10 mmol) was added. After 10 min. malonate (10 mmol) was slowly added and the reaction stirred overnight. The reaction mixture was diluted with Et<sub>2</sub>O (30 mL) and washed with water (20 mL), NaHCO<sub>3</sub> sat. sln. (20 mL) and brine (20 mL). The organic phase was concentrated under reduced pressure and the residue was purified by column chromatography on silica (cyclohexane:ethyl acetate from 95:5 to 90:10) to give pure **3a-d** as white oils (65-84% yields). Spectroscopic data were according to the literature.<sup>6</sup>

To a solution of itaconic acid (3.9 mmol, 0.500 g) in MeOH (10 mL) some drops of concentrated sulfuric acid were added. The reaction was refluxed for 8 hours, cooled at room temperature and the solvent was evaporated under reduced pressure. The residue was dissolved in  $Et_2O$  (30 mL) and washed with NaHCO<sub>3</sub> sat. sln. (2 x 10 mL) and brine (20 mL). The organic phase was concentrated under reduced pressure to give 3g (94%, 3.7 mmol, 0.576 g) as colorless oil. The product was used in the next step without purification.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 6.27 (s, 1H), 5.67 (s, 1H), 3.71 (s, 3H), 3.64 (s, 3H), 3.29 (s, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  =171.0, 166.5, 133.6, 128.4, 52.0, 51.9, 37.4.

OMe paraformaldehyde 
$$K_2CO_3$$
  $OMe$   $M_2CO_3$   $OMe$   $M_3$ ,  $M_4$   $M_5$   $M_5$   $M_6$   $M_6$ 

Compounds **3j,k** were prepared following reported procedure.<sup>7</sup> A mixture of ester (7.0 mmol, 1.044 g, 1 mL), paraformaldehyde (10.5 mmol, 0.313 g) and K<sub>2</sub>CO<sub>3</sub> (7.0 mmol, 0.960 g) in DMF (5 mL) was heated at 100°C for 6h. The mixture was cooled at room temperature, water (20 mL) and Et<sub>2</sub>O (30 mL) were added and the organic layer was separated. The aqueous layer was extracted with Et<sub>2</sub>O (2 x 20 mL). The collected organic layers were washed with brine (10 mL), dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure to give the crude products. Column chromatography on silica (cyclohexane:Et<sub>2</sub>O from 100:0 to 90:5) afforded pure **3j,k** as colorless oils.

3j: colorless oil; 73% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 7.45 – 7.28 (m, 5H), 6.36 (d, J=1.2, 1H), 5.88 (d, J=1.2, 1H), 3.81 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 167.2, 141.3, 136.6, 128.2 (2C), 128.1, 128.0 (2C), 126.8, 52.1.

3k: colorless oil, 85% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 7.33 – 7.28 (m, 2H), 7.19 – 7.14 (m, 2H), 6.31 (d, J=1.3, 1H), 5.86 (d, J=1.3, 1H), 3.81 (s, 3H), 2.36 Ar = 4-Me-C<sub>6</sub>H<sub>4</sub> (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 167.4, 141.1, 138.0, 133.8, 128.8 (2C), 128.1 (2C), 126.0, 52.1, 21.1.

Compounds **3n** were prepared following reported procedure.<sup>8</sup> A mixture of deoxibenzoin (5.1 mmol, 1.0 g), formaldehyde (38% in water, 1.8 mL), piperidine (0.66 mmol, 65  $\mu$ L) and acetic acid (1.1 mmol, 63  $\mu$ L) in MeOH (13 mL) was refluxed for 6h. The mixture was cooled at room temperature and the volatiles were evaporated under reduced pressure. The residue was dissolved in ethyl acetate (30 mL) and washed with 1 M HCl (10 mL), NaHCO<sub>3</sub> sat. sln. (10 mL) and brine (20 mL). The organic layer was dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure to give the crude product. Column chromatography on silica (cyclohexane:ethyl acetate from 95:5 to 80:20) afforded pure **3n** as colorless oil (94%, 4.8 mmol, 1.0 g).

(3n): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 7.95 – 7.87 (m, 2H), 7.58 – 7.47 (m, 1H), 7.47 – 7.41 (m, 4H), 7.41 – 7.24 (m, 3H), 6.06 (s, 1H), 5.63 (s, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  =

241.8, 216.7, 167.4, 156.2, 156.2, 152.2, 149.1 (2C), 147.8 (2C), 147.6 (2C), 146.2 (2C), 140.0.

#### Preparation of 2-substituted 1,3-dithiane-2-carboxylate 4b,c

To a solution of 1,3-dithiane-2carboxylate (0.95 mmol, 0.183 g, 154  $\mu$ L) in THF (3 mL), NaH (60% suspension in mineral oil, 1.9 mmol, 0.076 g) was slowly added at 0°C. The reaction mixture was gently heated at 70°C for 20 min. and cooled at room temperature. Alkylating agent (1.4 mmol) was added and the reaction mixture was stirred overnight. After that, the reaction was cooled at 0°C, water (10 mL) and ethyl acetate (20 mL) were added and the organic layer was separated. The aqueous layer was extracted with ethyl acetate (2 x 20 mL). The collected organic layers were washed with brine (10 mL), dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure to give the crude ester product.

The residue was dissolved in tBuOH (4 mL) and 1M KOH aq. (4 mL) was added. After 24 hours volatiles were evaporated under reduced pressure, water (10 mL) and Et<sub>2</sub>O (20 mL) were added and the organic layer was separated. The aqueous layer was extracted with Et<sub>2</sub>O (2 x 10 mL). 5 M HCl aq. was slowly added to aqueous layer until pH = 1, and they were extracted with DCM (4 x 20 mL). The collected organic layers were washed with brine (10 mL), dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure to give pure compounds as white solids, that were used in the photocatalytic reaction without further purification.

**4b**: White solid, 87% yield (two steps); m.p. = 79°C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C): 
$$\delta = 7.39 - 7.32$$
 (m, 2H),  $7.31 - 7.25$  (m, 3H),  $3.42 - 3.31$  (s, 2H),  $3.26$  (ddd,  $J=14.8$ , 12.8, 2.6, 2H),  $2.76 - 2.63$  (m, 2H),  $2.18 - 2.08$  (m, 1H),  $1.91 - 1.76$  (m, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta = 176.7$ , 134.0, 130.8 (2C), 128.0 (2C), 127.5, 53.2, 44.3, 29.6, 28.0, 24.1; HRMS calcd for  $C_{12}H_{14}O_{2}S_{2}$ : 254.0435; found: 254.0430.

4c: White solid, 83% yield (two steps); m.p. =  $56^{\circ}$ C; H NMR (400 MHz, CDCl<sub>3</sub>,  $25^{\circ}$ C):  $\delta = 11.57$  (bs, 1H), 5.95 - 5.78 (m, 1H), 5.26 - 5.10 (m, 2H), 3.33 (ddd, J=14.1, 12.9, 2.6, 2H), 2.73 (d, J=7.1, 2H), 2.63 (ddd, J=14.2, 4.2, 3.1, 2H), 2.26 - 2.00 (m, 1H), 1.91 - 1.70 (m, 1H);  $^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>,  $25^{\circ}$ C):  $\delta = 177.2$ , 131.1, 119.9, 50.2, 42.7,

#### Photoredox radical conjugate addition of carboxylates

General procedure on 0.2 mmol scale: In a Schlenk tube with rotaflo stopcock under argon atmosphere at r.t., catalyst 2 (1.7 mg, 0.002 mmol), 1,3-dithiane-2-carboxylic acid 4a (0.2 mmol, 0.032g) and K<sub>2</sub>HPO<sub>4</sub> (0.2 mmol, 0.034 g), were dissolved in 400 μL DMSO. After 2 min, Michael acceptor (0.4 mmol) was added. The reaction mixture was carefully degassed via freeze-pump thaw (three times), and the vessel refilled with argon. The Schlenk tube was stirred and irradiated with a blu LED positioned approximately at 10 cm distance from the reaction vessel. After 16 h of irradiation, NaHCO<sub>3</sub> sat. sln. (2 mL) was added and the mixture was extracted with ethyl acetate (4 x 5 mL). The collected organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure to give crude products. Column chromatography on silica (cyclohexane:ethyl acetate or cyclohexane:Et<sub>2</sub>O) afforded pure compounds.

**General procedure on 2.4 mmol scale:** The reaction was conducted applying the same protocol used for 0.2 mmol scale but using: catalyst **2** (10.4 mg, 0.012 mmol), 1,3-dithiane-2-carboxylic acid **4a** (2.4 mmol, 0.393 g) and K<sub>2</sub>HPO<sub>4</sub> (2.4 mmol, 0.417 g) and Michael acceptor **5a** (4.8 mmol, 0.960 g), were dissolved in 4.8 mL DMSO. Product **5a** was obtained in 84% yield.

**5a**: colorless oil, 86% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C): 
$$\delta$$
 = 4.31 (d,  $J$ =4.4, 1H), 3.77 (d,  $J$ =7.0, 1H), 3.73 (s, 3H), 3.71 (s, 3H), 2.94 – 2.71 (m, 4H), 2.71 – 2.55 (m, 1H), 2.14 – 1.98 (m, 1H), 1.93 – 1.75 (m, 1H), 1.68 (ddd,  $J$ =14.0, 7.3, 5.7, 1H), 1.59 (dt,  $J$ =13.4, 6.6, 1H), 1.36 – 1.26 (m, 1H), 0.90 (d,  $J$ =6.5, 3H), 0.87 (d,  $J$ =6.5, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 169.0, 168.9, 53.6, 52.6, 52.4, 52.2, 41.1, 38.5, 31.0, 30.5, 26.5, 25.9, 22.5, 22.4.; HRMS calcd for C<sub>14</sub>H<sub>24</sub>O<sub>4</sub>S<sub>2</sub>: 320.1116; found: 320.1108.

**5b**: colorless oil, 87% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C): 
$$\delta$$
 = 4.33 (d,  $J$ =4.3, 1H), 4.25 – 4.11 (m, 4H), 3.72 (d,  $J$ =7.0, 1H), 2.93 – 2.72 (m, 4H), 2.68 – 2.57 (m, 1H), 2.11 – 2.00 (m, 1H), 1.89 – 1.73 (m, 1H), 1.73 – 1.55 (m, 3H), 1.26 (t,  $J$ =7.2, 6H), 0.88 (dd,  $J$ =10.0, 6.4, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$ = 168.7, 168.5, 61.5, 61.4, 54.1, 52.4, 40.9, 38.6, 31.1, 30.6, 26.5, 26.0, 22.6, 22.5, 14.0 (2C); EI-MS:  $m/z$  = 347 (2), 303 (10, M-OEt), 257 (26), 188 (76), 145 (100), 119 (65); HRMS calcd for C<sub>16</sub>H<sub>28</sub>O<sub>4</sub>S<sub>2</sub>: 348.1429; found: 348.1422.

**5c**: colorless oil, 85% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 7.30 – 7.22 (m, 2H), 7.21 – 7.08 (m, 3H), 4.31 (d, J=5.3, 1H), 3.86 (d, J=6.8, 1H), 3.73 (s, 3H), 3.72 (s, 3H), 2.87 – 2.77 (m, 4H), 2.76 – 2.63 (m, 3H), 2.17 – 1.99 (m, 2H), 1.94 – 1.74 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 169.0, 168.8,

141.7, 128.4 (2C), 128.3 (2C), 125.9, 53.4, 52.7, 52.5, 51.5, 42.7, 34.9, 31.8, 30.7, 30.3, 25.9; EI-MS: m/z = 236 (56), 145 (100), 119 (65), 91 (45); HRMS calcd for  $C_{18}H_{24}O_4S_2$ : 368.1116; found: 368.119.

5d: white solid; m.p. = 77 °C, 84% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 7.60 – 6.97 (m, 5H), 4.57 (d, J=6.1, 1H), 4.21 (q, J=6.9, 2H), 4.17 (d, J=10.7, 1H), 3.90 (dd, J=10.7, 6.1, 1H), 3.84 (q, J=7.2, 2H), 2.91 – 2.67 (m, 4H), 1.98 (m, 1H), 1.76 (m, 1H), 1.25 (t, J=6.9, 3H), 0.89 (t, J=7.1, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 167.9, 167.4, 136.7, 129.4 (2C), 127.9 (2C), 127.8, 61.7, 61.3, 54.9, 51.3, 49.4, 30.5, 30.4, 25.6, 14.0, 13.6; HRMS calcd for C<sub>18</sub>H<sub>24</sub>O<sub>4</sub>S<sub>2</sub>: 368.1116; found: 368.1109.

**5e**: colorless oil, 91% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 4.26 – 4.15 (m, 3H), 4.15 – 4.08 (m, 2H), 3.37 (dddd, J=9.5, 7.1, 4.8, 0.9, 1H), 3.01 – 2.70 (m, 6H), 2.13 – 1.99 (m, 1H), 1.88 (m, 1H), 1.25 (m, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 171.6, 171.5, 61.3, 60.8, 47.6, 46.1, 34.1, 29.3, 29.1, 25.3, 14.1, 14.0; ESI-MS: m/z = 292 (53), 263 (2, M-Et), 218 (43), 158 (34), 145 (36), 119 (100); HRMS calcd for C<sub>12</sub>H<sub>20</sub>O<sub>4</sub>S<sub>2</sub>: 292.0803; found: 292.0796.

5f: yellowish oil, 49% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 4.29 (d, J=5.3, 1H), 4.15 – 4.08 (m, 4H), 2.89 – 2.79 (m, 6H), 2.74 (dd, J=16.5, 5.7, 1H), 2.55 (dd, J=12.4, 6.4, 1H), 2.48 (dd, J=16.5, 7.3, 1H), 2.11 – 2.02 (m, 1H), 1.89 – 1.77 (m, 1H), 1.33 – 1.16 (m, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 172.0 (2C), 60.6 (2C), 51.1 (2C), 39.8, 36.0 (2C), 29.9, 25.9, 14.2 (2C); EI-MS: m/z = 306 (36), 261 (43) 218 (47), 145 874), 119 (100); HRMS calcd for C<sub>13</sub>H<sub>22</sub>O<sub>4</sub>S<sub>2</sub>: 306.0960; found: 306.0955.

5g: colorless oil, 90% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 3.99 (dd, J=8.0, 6.9, 1H), 3.68 (s, 3H), 3.64 (s, 3H), 3.18 – 3.08 (m, 1H), 2.86 – 2.75 (m, 4H), 2.68 (dd, J=16.5, 8.2, 1H), 2.54 (dd, J=16.5, 5.7, 1H), 2.19 (ddd, J=14.4, 7.7, 6.9, 1H), 2.05 (m, 1H), 1.89 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 174.2, 171.6, 52.1, 51.8, 44.4, 38.4, 36.7, 35.6, 29.7, 29.6, 25.7; HRMS calcd for C<sub>11</sub>H<sub>18</sub>O<sub>4</sub>S<sub>2</sub>: 278.0647; found:

278.0639.

Sh: yellowish oil, 79% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 7.29 – 7.11 (m, 5H), 4.12 (d, J=8.7, 1H), 3.56 (s, 3H), 3.24 (dd, J=13.4, 4.2, 1H), 3.12 (m, 1H), 2.97 – 2.87 (m, 3H), 2.86 – 2.73 (m, 2H), 2.17 – 2.01 (m, 1H), 2.00 – 1.86 (m, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 172.6, 138.6, 128.8 (2C), 128.4 (2C), 126.5, 52.6, 51.7, 47.6, 36.4, 29.7, 28.9, 25.6; EI-MS: m/z = 282 (42), 191 (35), 159 (29), 119 (100), 91 (87); HRMS calcd for C<sub>14</sub>H<sub>18</sub>O<sub>2</sub>S<sub>2</sub>: 282.0748; found: 282.0741.

**5i**: colorless oil, 39% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 4.03 (t, J=7.0, 1H), 2.88 – 2.73 (m, 4H), 2.43 (dd, J=8.0, 7.1, 2H), 2.11 – 1.97 (m, 3H), 1.92 – 1.81 (m, 1H), 1.43 (s, 9H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 172.8, 77.2, 46.4, 32.5, 30.5, 29.9 (2C), 28.1 (3C), 25.8; HRMS calcd for C<sub>11</sub>H<sub>20</sub>O<sub>2</sub>S<sub>2</sub>: 248.0905; found: 248.0901.

5j: yellowish oil; 90% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 7.36 – 7.21 (m, 5H), 3.96 (t, J=7.6, 1H), 3.75 (dd, J=8.7, 6.3, 1H), 3.64 (s, 3H), 2.87 – 2.73 (m, 4H), 2.55 (ddd, J=14.2, 8.7, 7.6, 1H), 2.21 (ddd, J=14.2, 8.0, 7.6, 1H), 2.05 (m, 1H), 1.86 (m, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 173.6, 137.9, 128.8 (2C), 128.0 (2C), 127.5, 52.1, 48.1, 44.3, 38.4, 29.6, 29.4, 25.8; ESI-MS: m/z = 251.0 [M-OMe]<sup>+</sup>, 283.0 [M+H]<sup>+</sup>, 305.0 [M+Na]<sup>+</sup>; HRMS calcd for C<sub>14</sub>H<sub>18</sub>O<sub>2</sub>S<sub>2</sub>: 282.0748; found: 282.0741.

**5k**: yellowish oil, 88% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$ =7.21 – 7.15 (m, 2H), 7.15 – 7.09 (m, 2H), 3.92 (t, J=7.6, 1H), 3.75 (dd, J=8.7, 6.2, 1H), 3.64 (s, 3H), 2.90 – 2.65 (m, 4H), 2.52 (ddd, J=14.1, 8.7, 7.1, 1H), 2.19 (ddd, J=14.1, 8.1, 6.2, Ar = 4-Me-C<sub>6</sub>H<sub>4</sub> 1H), 2.31 (s, 3H), 2.12 – 1.97 (m, 1H), 1.94 – 1.78 (m, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 173.7, 137.2, 134.9, 129.5 (2C), 127.8 (2C), 52.1, 47.7, 44.3, 38.4, 29.6, 29.3, 25.8, 21.0; HRMS calcd for C<sub>15</sub>H<sub>20</sub>O<sub>2</sub>S<sub>2</sub>: 296.0905; found: 296.0899.

Ph NH<sub>2</sub> **5l**: sticky solid, 45% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 7.31 – 7.15 (m, 5H), 5.32 (bs, 1H), 5.24 (bs, 1H), 4.27 (d, J=8.4, 1H), 3.24 (dd, J=13.5, 4.8, 1H), 2.99 – 2.82 (m, 5H), 2.68 (ddd, J=13.5, 8.5, 4.7, 1H), 2.18 – 2.07 (m, 1H), 1.97 – 1.82 (m, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 173.2, 138.8, 129.0 (2C), 128.6 (2C), 126.6, 54.0, 48.3, 36.3, 30.3, 29.7, 25.8; HRMS calcd for C<sub>13</sub>H<sub>17</sub>NOS<sub>2</sub>: 267.0752; found: 267.0745.

**5m**: sticky solid, 75% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 7.94 – 7.86 (m, 2H), 7.71 – 7.61 (m, 1H), 7.61 – 7.52 (m, 2H), 4.00 (t, J=7.0, 1H), 3.37 – 3.21 (m, 2H), 2.90 – 2.72 (m, 4H), 2.28 – 2.12 (m, 2H), 2.14 – 1.97 (m, 1H), 1.91 – 1.74 (m, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 138.8, 133.8, 129.4 (2C), 128.0 (2C), 53.3, 44.9, 29.5 (2C), 28.4, 25.5; EI-MS: m/z = 288 (64), 146 (100), 119 (76), 77 (87); HRMS calcd for C<sub>12</sub>H<sub>16</sub>O<sub>2</sub>S<sub>3</sub>: 288.0312; found: 288.0306.

5n: colorless oil, 85% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 8.18 – 7.88 (m, 2H), 7.63 – 6.99 (m, 8H), 4.98 (t, J=7.2, 1H), 3.83 (dd, J=8.2, 6.9, 1H), 2.97 – 2.57 (m, 5H), 2.24 (dt, J=14.1, 7.2, 1H), 2.13 – 1.94 (m, 1H), 1.94 – 1.74 (m, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 198.6, 138.4, 136.4, 132.9, 129.0 (2C), 128.8 (2C), 128.5 (2C), 128.3 (2C), 127.3, 50.0, 44.7, 38.8, 29.6, 29.5, 25.9; ESI-MS: m/z = 329.2 [M+H]<sup>+</sup>, 351.0 [M+Na]<sup>+</sup>; HRMS calcd for C<sub>19</sub>H<sub>20</sub>OS<sub>2</sub>: 328.0956; found: 328.0947.

**50**: colorless oil, 76% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 9.72 (s, 1H), 4.13 (d, J=4.4, 1H), 2.88 – 2.66 (m, 4H), 2.40 (dd, J=17.6, 7.0, 1H), 2.35 – 2.21 (m, 1H), 2.12 – 1.93 (m, 1H), 1.86 – 1.70 (m, 1H), 1.69 – 1.52 (m, 1H), 1.52 – 1.44 (m, 1H), 1.43 – 1.29 (m, 1H), 0.86 (t, J=7.5, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 201.1, 52.0, 45.6, 39.5, 30.5, 30.4, 26.0, 24.7, 11.6; HRMS calcd for C<sub>9</sub>H<sub>16</sub>OS<sub>2</sub>: 204.0643; found: 204.0638.

**5p**: colorless oil, 93% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 9.75 (t, J=1.5, 1H), 4.08 (d, J=4.9, 1H), 2.87 – 2.82 (m, 4H), 2.80 (d, J=4.9, 1H), 2.63 – 2.50 (m, 1H), 2.44 (ddd, J=17.2, 8.0, 1.9, 1H), 2.17 – 2.00 (m, 1H), 1.93 – 1.71 (m, 1H), 1.13 (d, J=6.8, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 200.9, 53.9, 48.0, 33.1, 30.5, 30.3, 26.0, 17.8; HRMS calcd for C<sub>8</sub>H<sub>14</sub>OS<sub>2</sub>: 190.0486; found: 150.0481.

**5q**: colorless oil, 85% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 4.07 (d, J=5.0, 1H), 2.97 – 2.75 (m, 4H), 2.54 (ddt, J=14.2, 4.2, 2.0, 1H), 2.48 – 2.32 (m, 2H), 2.30 – 2.03 (m, 5H), 1.92 – 1.76 (m, 1H), 1.74 – 1.51 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 210.4, 53.5, 45.2, 43.5, 41.1, 30.7, 30.7, 28.4, 26.1, 24.8; HRMS calcd for C<sub>10</sub>H<sub>16</sub>OS<sub>2</sub>: 216.0643; found: 216.0638.

5r: The product was obtained using general procedure with 4b instead of 4a; yellowish oil, 89% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta = 7.49 - 6.93$  (m,

10H), 3.98 (dd, J=9.1, 2.4, 1H), 3.63 (s, 3H), 3.24 (dd, J=14.5, 8.7, 1H), 3.19 (d, J=12.8, 1H), 3.14 (d, J=14.2, 1H), 2.99 (ddd, J=14.5, 10.3, 2.9, 1H), 2.87 (ddd, J=14.2, 10.2, 3.0, 1H), 2.77 – 2.63 (m, 2H), 2.06 (dd, J=15.0, 2.4, 1H), 1.98 (m, 1H), 1.83 (m, 1H);  $^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 174.5, 139.8, 135.1, 131.0 (2C), 128.7 (2C), 127.9 (2C), 127.8 (2C), 127.2, 127.0, 53.2, 52.3, 48.2, 45.9, 41.6, 26.9, 26.2, 24.4; HRMS calcd for  $C_{21}H_{24}O_{2}S_{2}$ : 372.1218; found: 372.1210.

5s: The product was obtained using general procedure with 4b instead of 4a; colorless oil, 87% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 8.03 – 7.91 (m, 2H), 7.57 – 7.43 (m, 1H), 7.41 – 7.34 (dd, J=8.3, 6.8, 2H), 7.31 – 7.20 (m, 9H), 7.17 – 7.10 (m, 1H), 5.03 (dd, J=9.1, 1.7, 1H), 3.61 (dd, J=14.9, 9.1, 1H), 3.32 (d, J=14.0, 1H), 3.29 (d, J=14.0, 1H), 3.02 – 2.82 (m, 2H), 2.87 (ddd, J=14.4, 6.9, 3.1, 1H), 2.63 (ddd, J=14.4, 6.8, 3.2, 1H), 2.10 (dd, J=14.9, 1.7, 1H), 2.02 – 1.87 (m, 1H), 1.9 – 1.78 (m, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 198.7, 139.7, 137.1, 135.4, 132.5, 131.1 (2C), 128.9 (2C), 128.8 (2C), 128.4 (2C), 128.3 (2C), 127.9 (2C), 127.0, 126.9, 53.6, 49.8, 46.8, 42.2, 26.9, 26.3, 24.5; HRMS calcd for C<sub>26</sub>H<sub>26</sub>OS<sub>2</sub>: 418.1425; found: 418.1416.

5t: The product was obtained using general procedure with 4c instead of 4a; yellowish oil, 86% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 8.03 – 7.93 (m, 2H), 7.51 – 7.43 (m, 1H), 7.42 – 7.34 (m, 2H), 7.34 – 7.29 (m, 2H), 7.28 – 7.20 (m, 2H), 7.20 – 7.11 (m, 1H), 5.97 – 5.79 (m, 1H), 5.18 – 5.12 (m, 1H), 5.12 – 5.07 (m, 1H), 4.99 (dd, J=8.9, 1.9, 1H), 3.54 (dd, J=14.9, 8.9, 1H), 2.87-2.81 (m, 2H), 2.78 – 2.67 (m, 1H), 2.67 – 2.55 (m, 3H), 2.10 (dd, J=14.9, 1.9, 1H), 1.96-1.93 (m, 1H), 1.91 – 1.78 (m, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 198.6, 140.0, 137.1, 132.6, 132.4, 129.0 (2C), 128.7 (2C), 128.4 (2C), 128.2 (2C), 127.0, 118.8, 52.6, 49.7, 44.3, 42.2, 26.5, 26.0, 24.7; HRMS calcd for C<sub>22</sub>H<sub>24</sub>OS<sub>2</sub>: 368.1269; found: 368.1260.

14: The product was obtained using general procedure with cyclohexylcarboxylic acid instead of **4a** and catalyst **1** replaced **2**; colorless oil, 46% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 3.69 (d, J=5.9, 6H), 3.45 (d, J=7.5, 1H), 2.14 – 2.07 (m, 1H), 1.76 – 1.56 (m, 5H), 1.38 – 1.25 (m, 3H), 1.21 – 1.01 (m, 6H), 0.85 (d, J=4.0, 3H), 0.83 (d, J=4.0, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  =170.0, 169.9, 54.4, 52.3, 52.1, 41.7, 40.7, 38.2, 30.5, 29.20, 26.9, 26.8, 26.6, 26.5, 22.8, 22.6.

16: The product was obtained using general procedure with *rac*-Cbz-proline instead of 4a; sticky solid, 87% yield; The molecules present different diastereoisomer and different conformers, NMR signal appears very broad and some <sup>13</sup>C NMR signals were missed. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 7.56 – 6.76 (m, 10H), 5.11 (bs, 2H), 4.22 – 3.91 (m, 1H), 3.75 – 3.50 (m, 9H), 3.19 (m, 1H), 3.09 (dq, *J*=8.5, 4.3, 1H), 2.80 – 2.37 (m, 3H), 2.07 – 1.90 (m, 1H), 1.90 – 1.59 (m, 3H); <sup>13</sup>CNMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 169.2, 155.6, 141.8, 136.6, 128.4 (2C), 128.3 (2C) 128.2 (2C), 128.1, (2C), 127.9 (2C), 125.8, 66.8, 60.3, 58.7, 52.3, 48.2, 46.9, 40.6, 31.9, 29.7, 27.0; ESI-MS: m/z = 454.2 [M+H]<sup>+</sup>, 476.2 [M+Na]<sup>+</sup>.

#### Reductive removal of the dithiane moiety

General Procedure: To a solution of 5a (0.1 mmol, 0.032 g) in methanol (2 mL), Ni-Raney (0.450 g, slurry in water) was added and the reaction was keep under H<sub>2</sub> atmosphere (1 atm). After 3h the reaction mixture was filtered through a Celite® pad washed with DCM and the organic solvent was removed under reduce pressure. The residue was diluted with ethyl acetate (10 mL), the organic layer was separated, and the aqueous layer was extracted with ethyl acetate (2 x 5 mL). The collected organic layers were washed with brine (5 mL), dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduce pressure.

Flash chromatography (cyclohexane:diethyl ether, 95:5) of the residue give **6** in 91% yield (0.091 mmol, 0.02 g).

OME 6: yellowish oil, 90% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 3.71 (s, 6H), 3.22 (d, J=7.8, 1H), 2.35 – 2.24 (m, 1H), 1.61 (m, 1H), 1.15 – 1.05 (m, 2H), 0.94 (d, J=6.7, 3H), 0.87 (d, J=6.5, 3H), 0.85 (d, J=6.5, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 169.3 (2C), 57.8, 52.2, 52.1, 31.4, 29.7, 25.2, 23.7, 21.3, 16.9; EI-MS: m/z = 201 (6, M-Me), 185 (10, M-OMe), 132 (100), 100 (56), 83 (45), 69 (50); HRMS calcd for C<sub>11</sub>H<sub>20</sub>O<sub>4</sub>: 216.1362; found: 216.1351.

7: yellowish oil, 84% yield; Spectral proprieties were in according with literature. <sup>9</sup>

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta = 4.24 - 4.04$  (m, 4H), 2.95 – 2.81 (m, 1H), 2.70 (dd, J=16.4, 8.1, 1H), 2.37 (dd, J=16.4, 6.1, 1H), 1.26 – 1.21 (m, 6H), 1.20 (d, J=7.1, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta = 175.2$ , 171.8, 60.5, 60.4, 37.7, 35.8, 16.9, 14.1, 14.0. 8: yellowish oil, 88% yield; Spectral proprieties were in according with literature. 8: 9

9: yellowish oil, 90% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 7.38 – 7.23 (m, 4H), 7.27 – 7.09 (m, 6H), 7.13 – 7.02 (m, 3H), 7.05 – 6.91 (m, 2H), 4.68 (d, J=8.4, 1H), 2.97 – 2.80 (m, 1H), 2.47 (ddd, J=14.5, 8.6, 6.4, 1H), 2.34 (ddd, J=14.5, 8.8, 6.9, 1H), 1.67 – 1.53 (m, 1H), 1.49 – 1.24 (m, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 142.7, 142.2, 141.1, 128.8 (2C), 128.6 (2C), 128.3 (2C), 128.2 (2C), 128.1 (2C), 127.8, 127.0, 126.9 (2C), 126.4, 125.5, 78.8, 54.2, 35.6, 29.1; ESI-MS: m/z = 317.1 [M+H]<sup>+</sup>, 339.2 [M+Na]<sup>+</sup>; HRMS calcd for C<sub>23</sub>H<sub>24</sub>O: 316.1827; found: 316.1821.

#### Hydrolysis of the dithiane moiety

**PhI(OAc)**<sub>2</sub> **method**:<sup>11</sup> Bis(trifluoroacetoxy)iodobenzene (52 mg, 0.16 mmol) was added at 0°C to a stirred solution of **5a** or **5t** (0.1 mmol), water (1 mL) and CH<sub>3</sub>CN (9 mL). After it was stirred at room temperature for 30 min, the reaction was quenched with NaHCO<sub>3</sub> sat. sln. (10 ml) and the volatiles were evaporated. The residue was dissolved in ethyl acetate (20 mL) and the organic layer was separated. The aqueous layer was extracted with ethyl acetate (3 x 10 mL). The collected organic layers were washed with brine (10 mL), dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure to give the crude product. Column chromatography on silica (cyclohexane:ethyl acetate from 95:5 to 80:20) afforded pure products.

OME 10: yellowish oil, 76% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 9.77 (d, J=1.2, 1H), 3.75 (s, 3H), 3.73 (d, J=7.8, 1H), 3.72 (s, 3H), 3.17 – 3.01 (m, 1H), 1.62 – 1.64 (m, 1H), 1.37 – 1.20 (m, 2H), 0.94 (d, J=6.3, 3H), 0.91 (d, J=6.2, 3H); <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 198.0, 168.5 (2C), 52.8, 52.1, 48.3, 36.5, 29.7, 25.9, 22.9, 22.1; HRMS calcd for C<sub>11</sub>H<sub>18</sub>O<sub>5</sub>: 230.1154; found: 230.1149.

12: yellowish oil, 91% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 8.08 – 7.76 (m, 2H), 7.47 – 7.42 (m, 1H), 7.39 – 7.31 (m, 2H), 7.30 – 7.21 (m, 4H), 7.21 – 7.15 (m, 1H), 5.89 (m, 1H), 5.16 (m, 2H), 5.13 – 5.06 (m, 1H), 3.59 (dd, J=18.0, 10.0, 1H), 3.27 – 3.13 (m, 2H), 2.76 (dd, J=18.0, 4.0, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 206.7, 198.8, 138.5, 136.3, 132.9, 130.2, 129.1 (2C), 128.9 (2C), 128.4 (2C), 128.1 (2C), 127.3, 119.0, 48.7, 47.7, 46.8; ESI-MS: m/z = 279.0 [M+H]<sup>+</sup>, 301.0 [M+Na]<sup>+</sup>; HRMS calcd for C<sub>19</sub>H<sub>18</sub>O<sub>2</sub>: 278.1307; found: 278.1302.

**HgO/HBF**<sub>4</sub> **method:** To a suspension of HgO (0.18 mmol, 0.039 g) in HBF<sub>4</sub> (48% in water, 0.2 mL) and THF (1 mL) a solution of **5r** (0.09 mmol, 0.034 g) in THF (1 mL) was added. After 2 hours the reaction was quenched adding 10 mL of NaHCO<sub>3</sub> sat. sln. until basic pH. The mixture was filtered through a Celite® pad washing with ethyl acetate. Water (10 mL) was added and the organic layer was separated. The aqueous layer was extracted with ethyl acetate (3 x 10 mL). The collected organic layers were washed with brine (10 mL), dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure to give the crude product. Column chromatography on silica (cyclohexane:ethyl acetate from 95:5 to 90:10) afforded pure product **11** (90%, 0.08 mmol, 0.021 g) as yellowish oil.

11: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta = 7.49 - 6.86$  (m, 10H), 4.07 (dd, J=10.2, 4.5, 1H), 3.70 (s, 2H), 3.62 (s, 3H), 3.36 (dd, J=18.0, 10.2, 1H), 2.71 (dd, J=18.0, 4.5, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta = 205.9$ , 173.6, 138.0, 133.7, 129.4 (2C), 128.8 (2C), 128.7 (2C), 127.7 (2C), 127.5, 127.1, 52.3, 50.1, 46.2, 45.5; HRMS calcd for  $C_{18}H_{18}O_3$ : 282.1256; found: 282.1251.

#### **Reaction in presence of TEMPO**

Evidence of a possible radical mechanism was determinate by performing the reaction in the presence of radical scavengers like TEMPO (2,2,6,6-tetramethylpiperidine 1-oxyl).

Two different experiments were conducted in the presence of a catalytic (20 mol%) and stoichiometric amount of TEMPO. In both cases no formation of product or adduct between TEMPO and reaction intermediates were observed.

#### Preparation of ammonium salts 17, 18, 19

$$\mathsf{R} \xrightarrow{\mathsf{COOH}} \underbrace{ \overset{\mathsf{nBu_4}\mathsf{N}^{\oplus}\,\ominus\,\mathsf{OH}}{\mathsf{H_2O}}}_{\mathsf{H_2O}} \mathsf{R} \xrightarrow{\mathsf{COO}} \overset{\ominus}{\mathsf{nBu_4}\mathsf{N}^{\oplus}}$$

**General procedure:** <sup>12</sup> To a solution of **4a** (0.3 mmol, 0.05g) in water (5 mL) *n*Bu<sub>4</sub>NOH\*30H<sub>2</sub>O (0.3 mmol, 0.240 g) was added and the reaction mixture was heated at 60°C for 2 hours. Water was evaporated under reduced pressure and the residue was dried under high vacuum for 2 hours to give pure **17** (96%, 0,29 mmol, 0.117 g) as white solid.

17: sticky solid, quantitative; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 4.70 (s, 1H), 3.45 – 3.23 (m, 8H), 2.99 (ddd, J=14.0, 6.3, 3.0, 2H), 2.75 (ddd, J=13.7, 10.5, 2.7, 2H), 2.08 – 1.94 (m, 1H), 1.90 – 1.76 (m, 1H), 1.68 – 1.55 (m, 8H), 1.53 – 1.33 (m, 8H), 0.99 (t, J=7.3, 12H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 170.6, 58.5 (4C), 52.9, 30.2 (2C), 26.2, 23.9 (4C), 19.6 (4C), 13.6 (4C).

18: sticky solid, quantitative; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 3.46 – 3.17 (m, 8H), 2.90 (bs, 1H), 2.11 – 2.00 (m, 1H), 1.95 – 1.85 (m, 2H), 1.77 – 1.54 (m, 10H), 1.51 – 1.30 (m, 10H), 1.31 – 1.10 (m, 3H), 0.97 (t, J=7.3, 12H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 58.9 (4C), 48.0, 31.0 (2C), 26.6 (2C), 24.2 (4C), 19.9, 19.8 (4C), 13.7 (4C), carboxylic carbon was not detected also using long acquisition time with 4s delay.

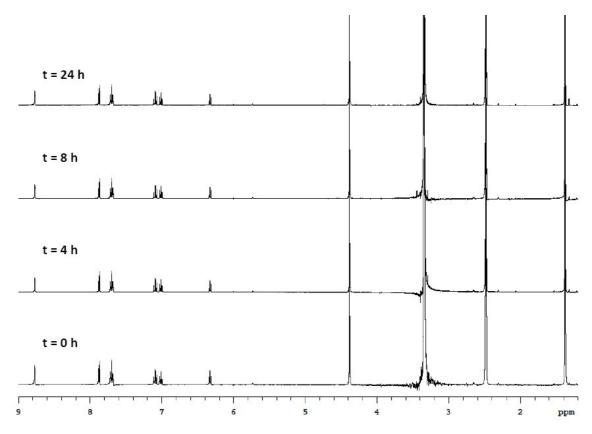
19: sticky solid, quantitative; The compound is present as two different compound conformers (A and B). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  = 7.42 – 7.01 (m, 10H, A+B), 5.20 (d, J=13.2, 1H), 5.11 (d, J=12.7, 1H, B), 4.99 (d, J=13.0, 1H, B), 4.27 (t, J=5.9, 1H, A), 4.15 (dd, J=7.8, 3.1, 2H, A), 3.74 – 3.58 (m, 2H, B), 3.42 (td, J=17.5, 8.1, 2H, A), 3.35 – 3.16 (m, 8H, A+B), 2.26 – 2.00 (m, 2H, A+B), 1.81 – 1.67 (m, 2H, A+B), 1.68 – 1.54 (m, 8H, A+B), 1.47 – 1.27 (m, 8H, A+B), 0.96 (t, J=7.3, 12H, A+B); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25°C):  $\delta$  =171.2 (A), 171.1 (B), 154.7 (A+B), 128.4 (B, 2C), 128.3 (A, 2C), 127.9 (A, 2C), 127.6 (B, 2C), 127.4 (A), 127.2 (B), 66.9 (A), 66.5 (B), 58.8 (A+B, 4C), 46.8 (A), 46.4 (B), 30.6 (B), 29.6 (A), 24.4 (A), 24.1 (B), 24.0 (A+B, 4C), 23.3 (A), 23.2 (B), 23.1 (C), 19.6 (A+B, 4C), 13.6 (A+B, 4C), carboxylic carbon was not detected also using long acquisition time with 4s delay.

### Photoredox radical conjugate addition of salt 17

To determinate if tetrabutylammonium salt 17 is a suitable substrate for the photoredox radical conjugate addition we tested it in the reaction using general procedure without the presence of  $K_2HPO_4$  as base. Product 5a was obtained in 72% of yield, demonstrating the possibility to use tetrabutylammonium carboxylate as substrate.

#### Photostability of Ir(III) complex 2

In order to test its photostability during the reaction, complex **2** was dissolved in degassed deuterated DMSO and was irradiated using blue light LED for 24 hours in order to replicate the reaction conditions. Any possible photolysis was monitored by <sup>1</sup>H NMR spectroscopy. As reported in Figure S1, less than 5% of complex **2** undergoes to photochemical decomposition after 24 hours of irradiation, revealing its good photostability under reaction conditions, differently from complex **1.**<sup>13</sup>



**Figure S1.** <sup>1</sup>H NMR spectra for complex **2** in deuterated DMSO solution irradiated with blue light LED for increasing times. The solution was degassed by nitrogen gas for 15 minutes.

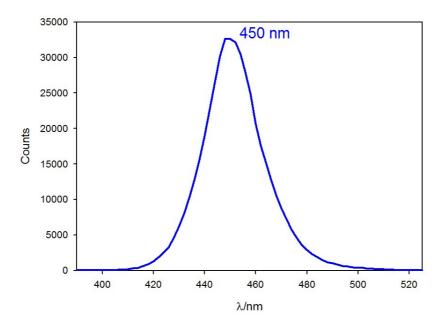
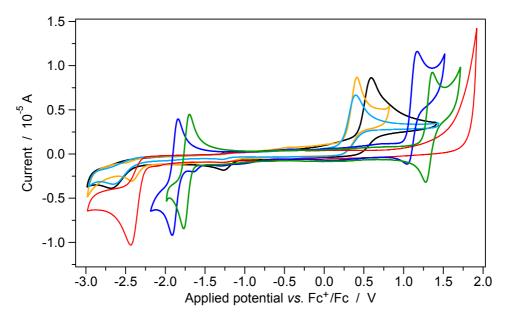
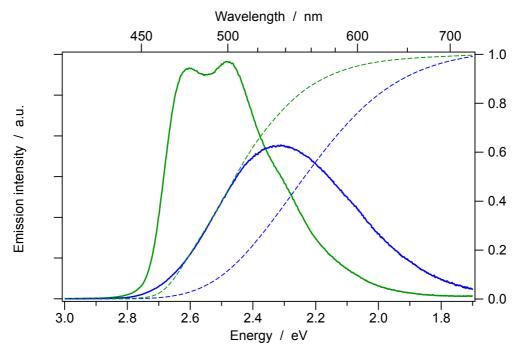


Figure S2. Emission profile of the 24W Blue LED strip used to irradiate the solutions.

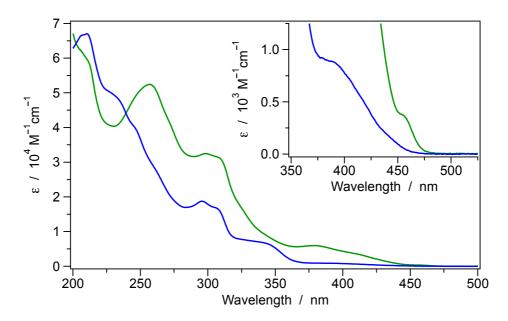
#### Photophysical and electrochemical data



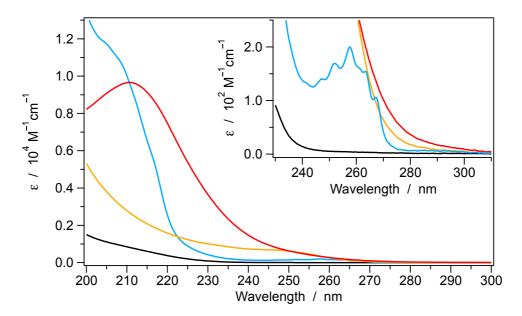
**Figure S3.** Cyclic voltammograms of the **17** (orange), **18** (black) and **19** (light blue) substrates, together with the one of the Michael acceptor **3a** (red) and of the iridium(III) photocatalysts **1** (green) and **2** (dark blue). The voltammograms are recorded at a scan rate of 200 mV s<sup>-1</sup> in room-temperature acetonitrile in the presence of TBAPF<sub>6</sub> 0.1 M as supporting electrolyte.



**Figure S4.** Full lines: corrected and normalized emission spectra of photocatalyst **1** (green) and **2** (dark blue) in room-temperature acetonitrile solution, reported in relative quanta per energy interval. Dashed lines: integrals of the spectra; the mean photon energy is calculated as the value at which the integral is 50%.



**Figure S5.** Absorption spectra of photocatalyst **1** (green) and **2** (dark blue) in room-temperature acetonitrile solution. The lowest-energy electronic transition, attributed to the direct population of the emitting state, is magnified in the inset for each of the two iridium(III) photocatalysts.



**Figure S6.** Absorption spectra of the **17** (orange), **18** (black) and **19** (light blue) substrates, together with the one of the Michael acceptor **3a** (red), in room-temperature acetonitrile solution. The lowest-energy transitions are magnified in the inset for the sake of clarity.

#### **Determination of quantum yield**

The reaction was carried out in spectrofluorimetric grade DMSO and placed in a Suprasil® quartz cuvette with a 2.00 mm path length. The cuvette was charged with the ammonium carboxylate 17 (81 mg, 0.20 mmol, 1 equiv), the Michael acceptor 3a (80 mg, 0.40 mmol, 2 equiv), the iridium photocatalyst 2 (1.7 mg, 0.0020 mmol), and 500 µl of DMSO. Before irradiating the sample, dissolved oxygen was removed from the cuvette by argon bubbling for 15 minutes. The reaction mixture was excited at 334 nm for exactly 7 hours, using a 100 W Hg lamp equipped with an appropriate dichroic filter. During this process, the solution was stirred and kept at constant temperature.

The photon flux coming from the lamp to the sample in the cuvette was estimated using the ferrioxalate actinometer (photon flux: 1.97 nmol/s). This experimental value was also confirmed by using a calibrated silicon diode (light power: 0.6 mW, corresponding to a photon flux of 2 nmol/s at 334 nm). In our experimental conditions, all the incident light is quantitatively absorbed by the iridium photocatalyst (A > 3 at 334 nm).

The product formation was determined by <sup>1</sup>H-NMR based on the relative conversion of the carboxylate derivative.

After irradiation, the estimated conversion was 7%, corresponding to 14 µmol of product formed.

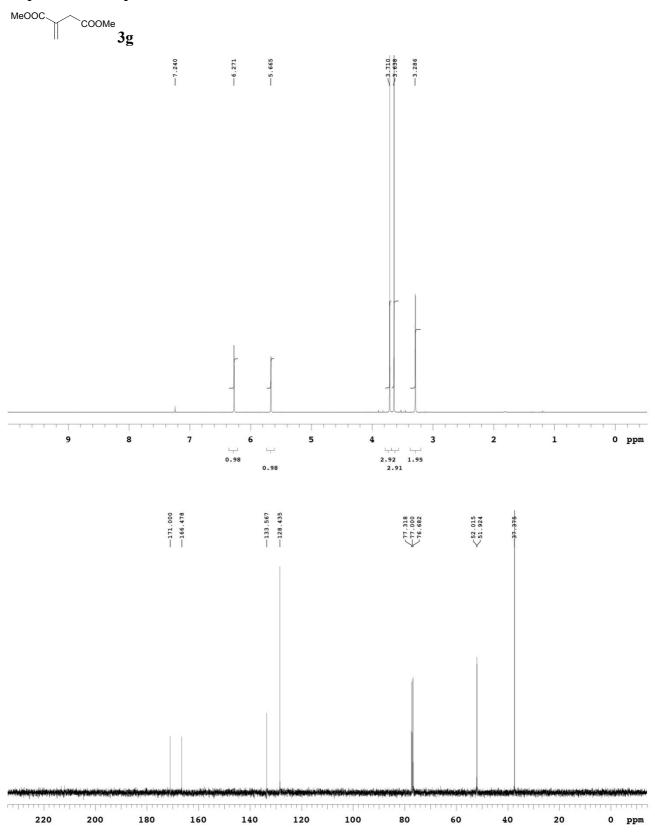
$$\Phi = \frac{moles\ of\ product\ formed}{moles\ of\ photons\ absorbed} = \frac{14\cdot 10^{-6}mol}{1.97\cdot 10^{-9}\frac{mol}{s}\cdot 2.5\cdot 10^{4}\ s} = 0.28\pm 0.05$$

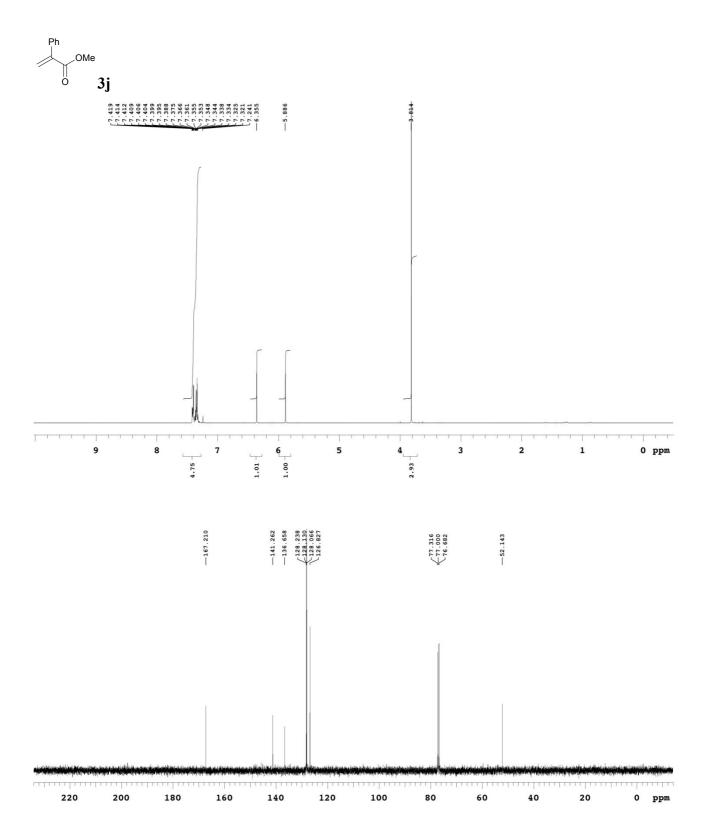
#### References

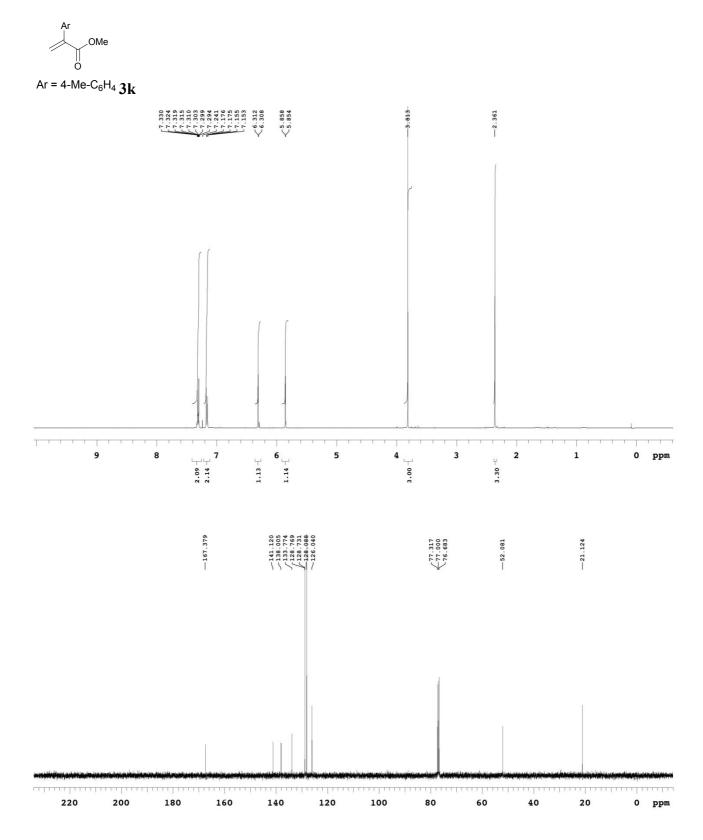
- E. Massolo, M. Benaglia, A. Genoni, R. Annunziata, G. Celentano and N. Gaggero, *Org. Biomol. Chem.*, 2015, **13**, 5591-5596.
- <sup>2</sup> M. S. Lowry, J. I. Goldsmith, J. D. Slinker, R. Rohl, R. A. Pascal Jr., G. G. Malliaras and S. Bernhard, *Chem. Mater.*, 2005, **17**, 5712-5719.

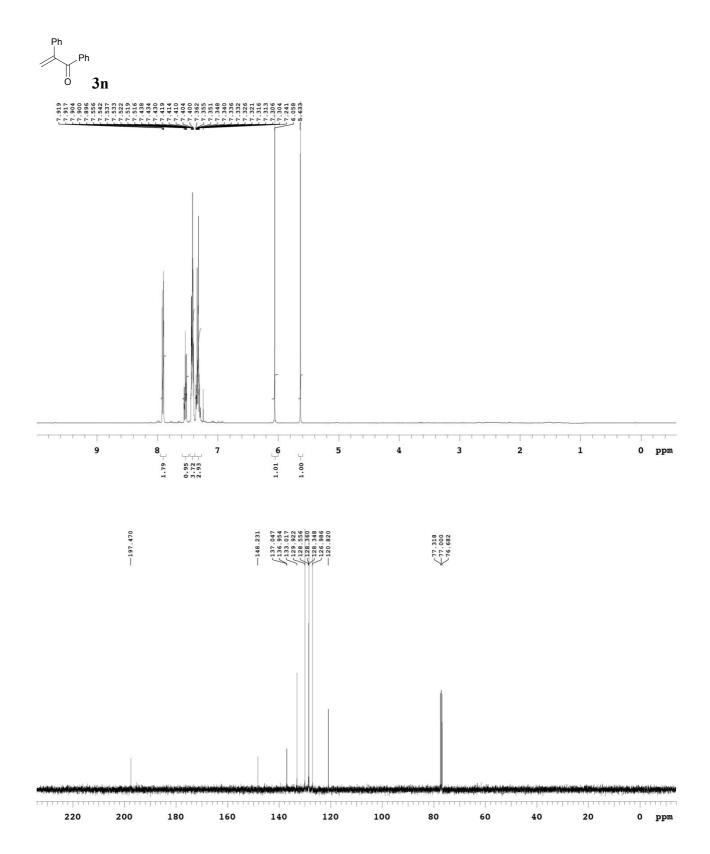
- F. Monti, A. Baschieri, I. Gualandi, J. J. Serrano-Pérez, J. M. Junquera-Hernández, D. Tonelli, A. Mazzanti, S. Muzzioli, S. Stagni, C. Roldan-Carmona, A. Pertegás, H. J. Bolink, E. Ortí, L. Sambri and N. Armaroli, *Inorg. Chem.*, 2014, **53**, 7709-7721.
- M. Felici, P. Contreras-Carballada, Y. Vida, J. M. M. Smits, R. J. M. Nolte, L. De Cola, R.
   M. Williams, M. C. Feiters, *Chem. Eur. J.*, 2009, 15, 13124–13134.
- G. Cardillo, S. Fabbroni, L. Gentilucci, M. Gianotti and A. Tolomelli, *Synth. Comm.* 2003, **33**, 1587-1594.
- a) G. Cardillo, L. Gentilucci, M. Gianotti, R. Perciaccante and A. Tolomelli, *J. Org. Chem.* 2001, 66, 8657-8660; b) P. Ramesh, B. Shalini and N. W. Fadnavis, *RSC Adv.*, 2014, 4, 7368-7373
- a) M. J. Lavecchia, R. Puig de la Bellacasa, J. I. Borrell and C. N. Cavasotto, *Biorg. Med. Chem.*, 2016, **24**, 768-778; b) J. Pietruszka and M. Schölzel, *Adv. Synth. Catal.*, 2012, **354**, 751-756.
- <sup>8</sup> V. Gembus, J.-J. Bonnet, F. Janin, P. Bohn, V. Levacher and J.-F. Briere, *Org. Biomol. Chem.*, 2010, **8**, 3287-3293.
- <sup>9</sup> Y. Chen, E. Kervio and J. Retey, *Helv. Chim. Acta*, 2002, **85**, 552-558.
- K. Kamata, K. Yonehara, Y. Nakagawa, K. Uehara and N. Mizuno, *Nature Chemistry* 2010,2, 478–483.
- <sup>11</sup> Z. Meng and S. Danishefsky, *Angew. Chem. Int. Ed.*, 2005, **44**, 1511-1513.
- C. R. Allen, P. L. Richard, A. J. Ward, L. G. A. van de Water, A. F. Masters and T. Maschmeyer, *Tetrahedron Letters*, 2006, 47, 7367-7370
- C. Yang, F. Mehmood, T. L. Lam, S. L.-F. Chan, Y. Wu, C.-S. Yeung, X. Guan, K. Li, C. Y.-S. Chung, C.-Y. Zhou, T. Zou, and C.-M. Che, *Chem. Sci.* 2016, 7, 3123-3136.
- M. Montalti, A. Credi, L. Prodi , and M. T. Gandolfi, Handbook of Photochemistry, Third Edition, CRC Press, 2006, chapter 12.

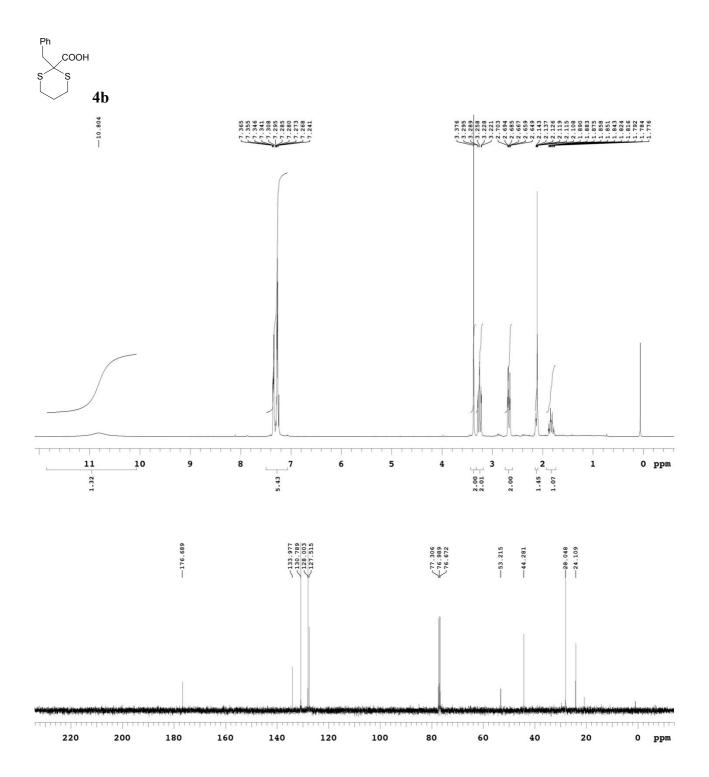
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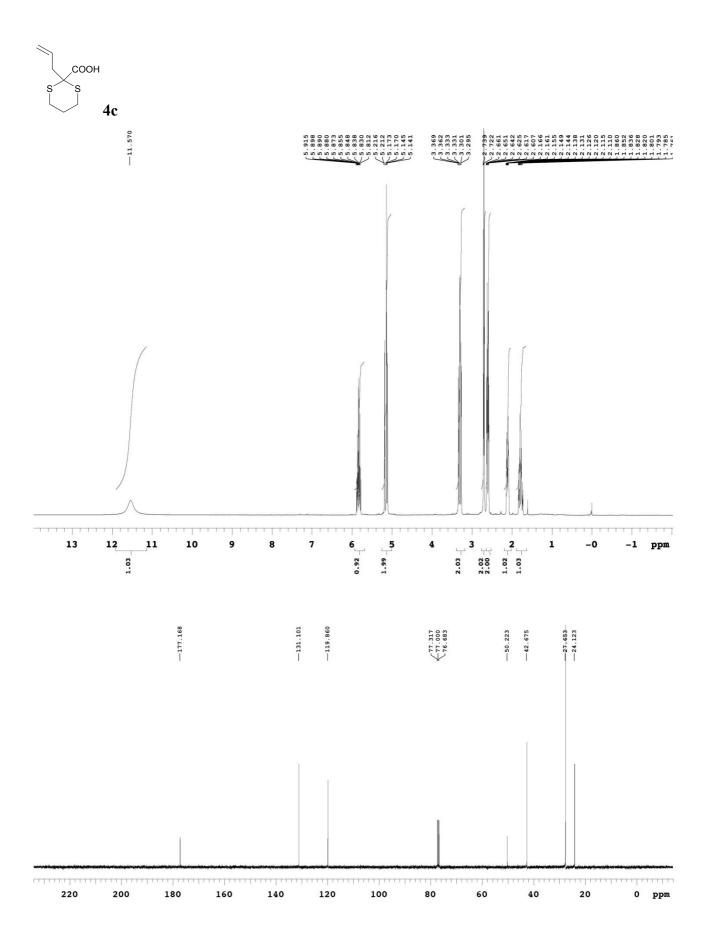


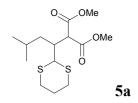


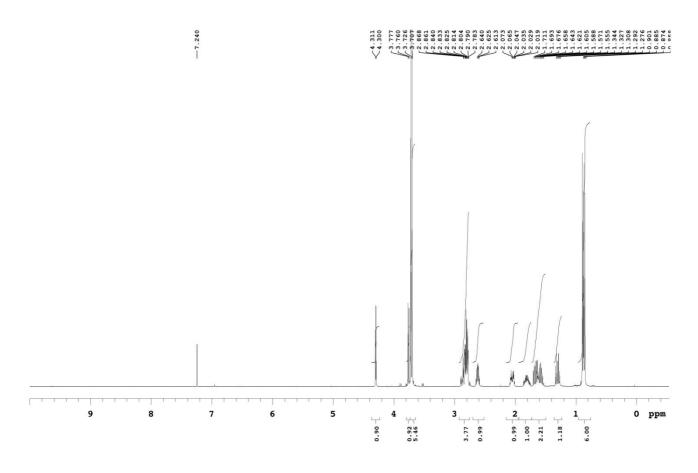


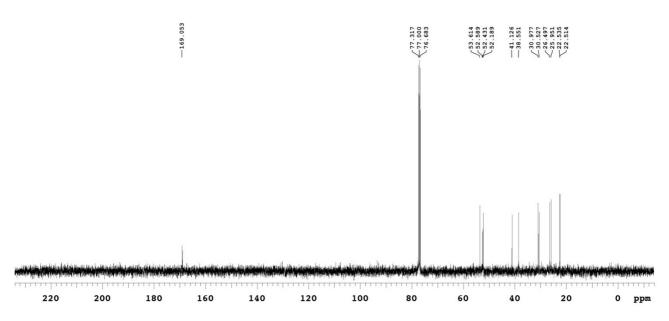


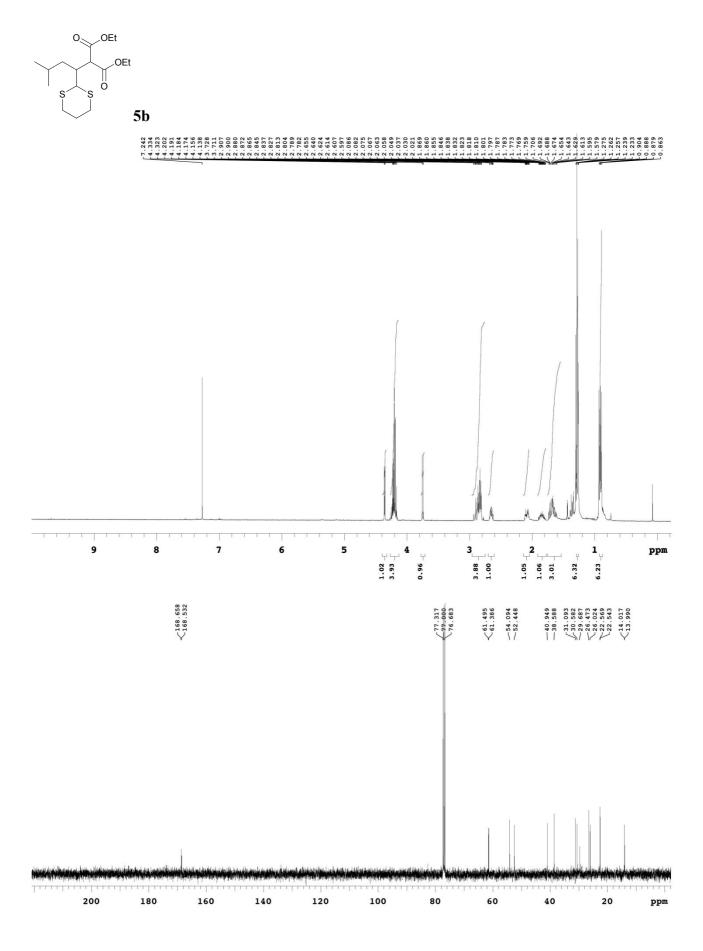


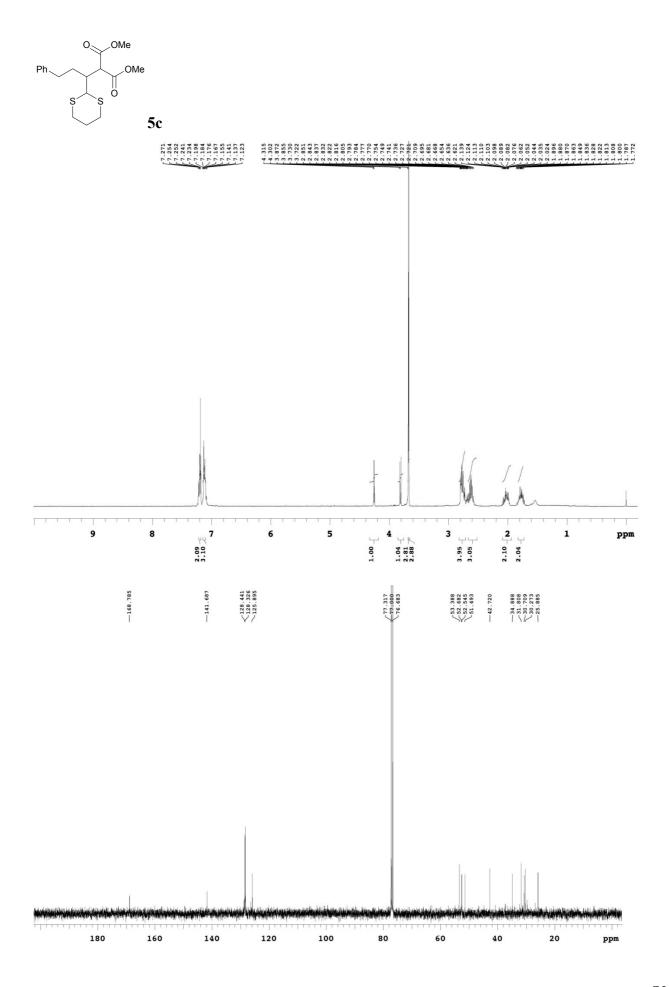




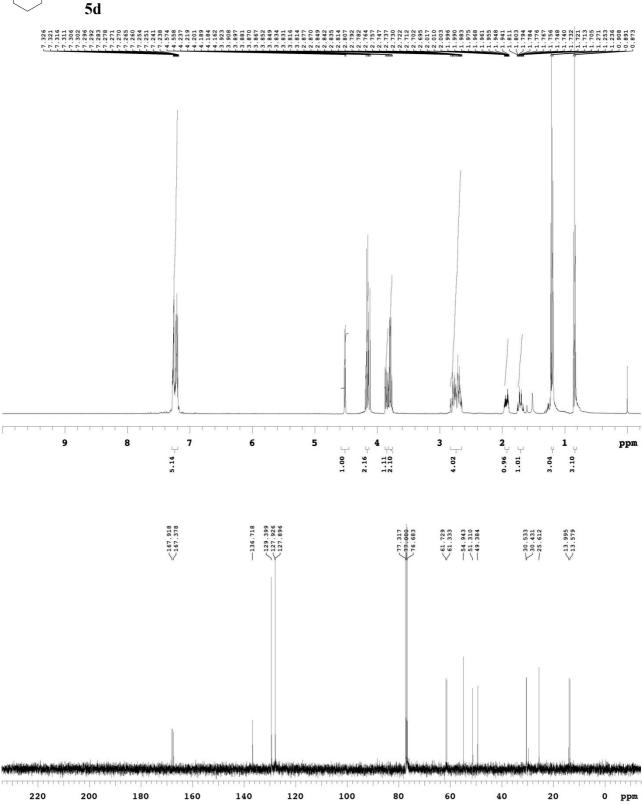


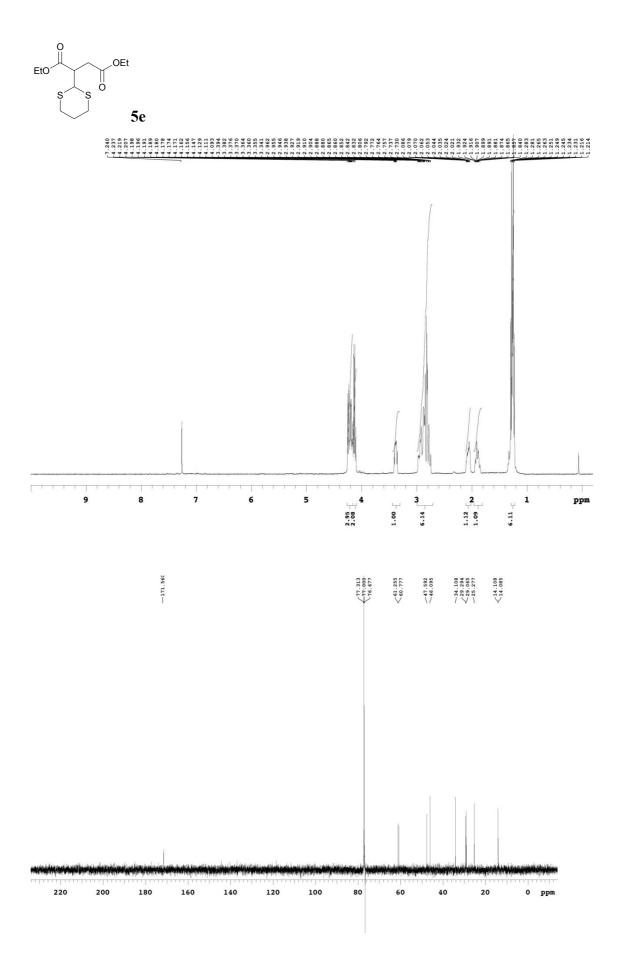


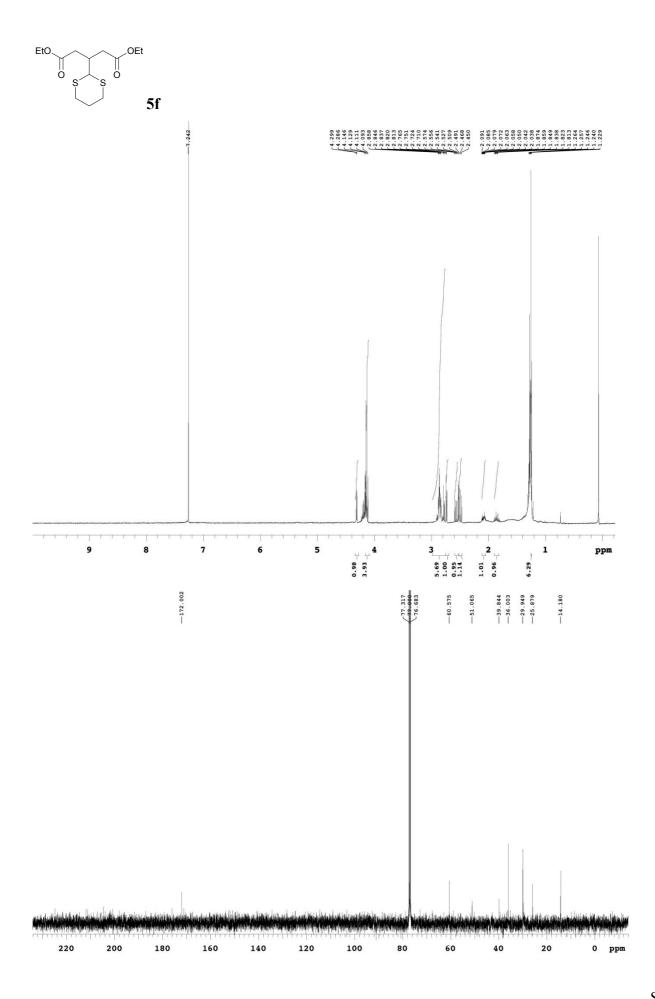


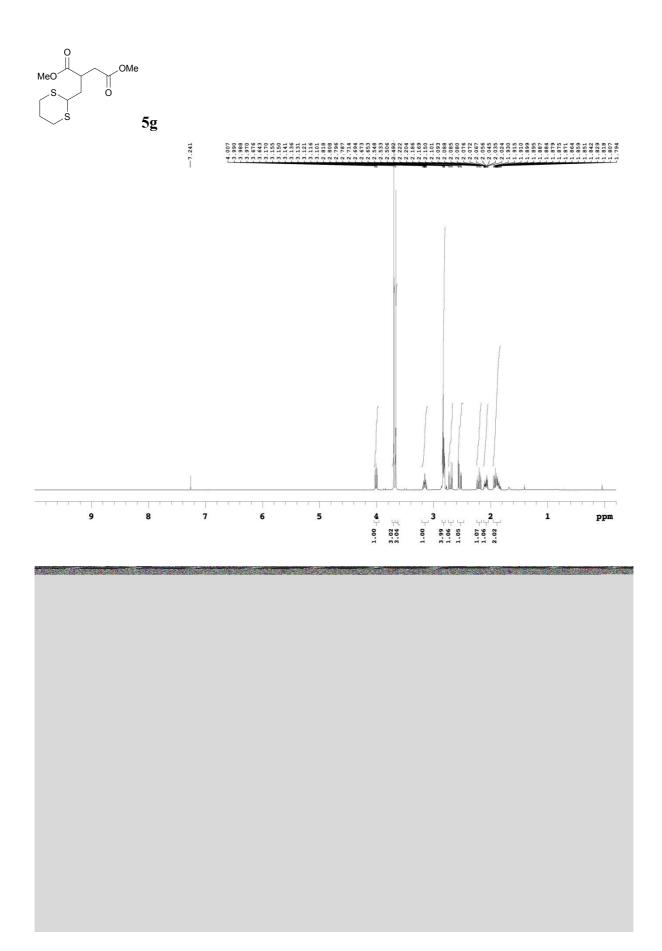


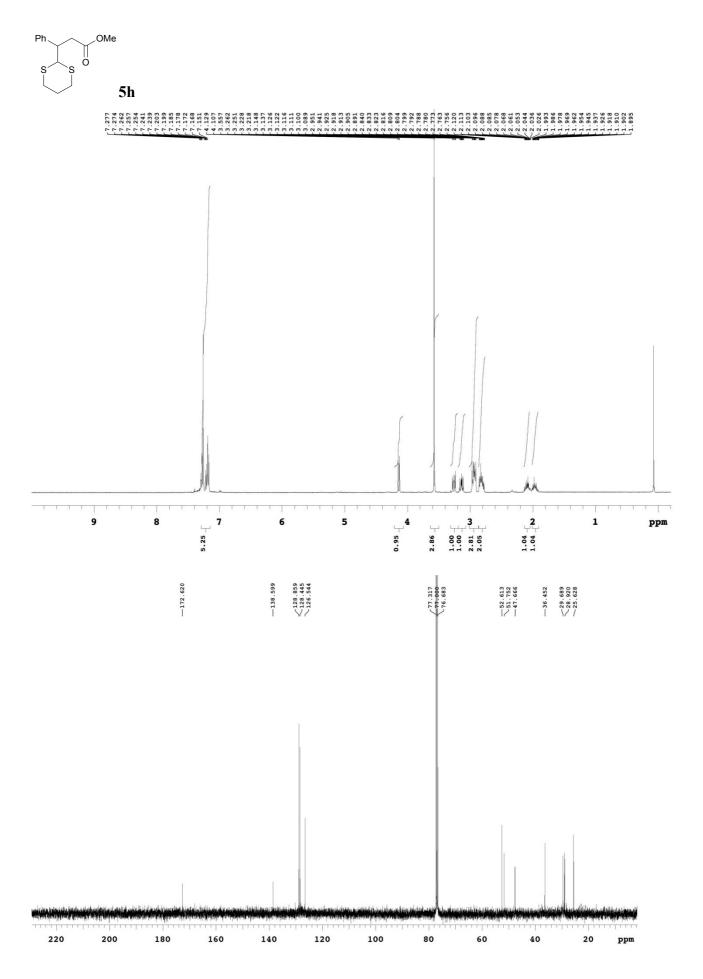


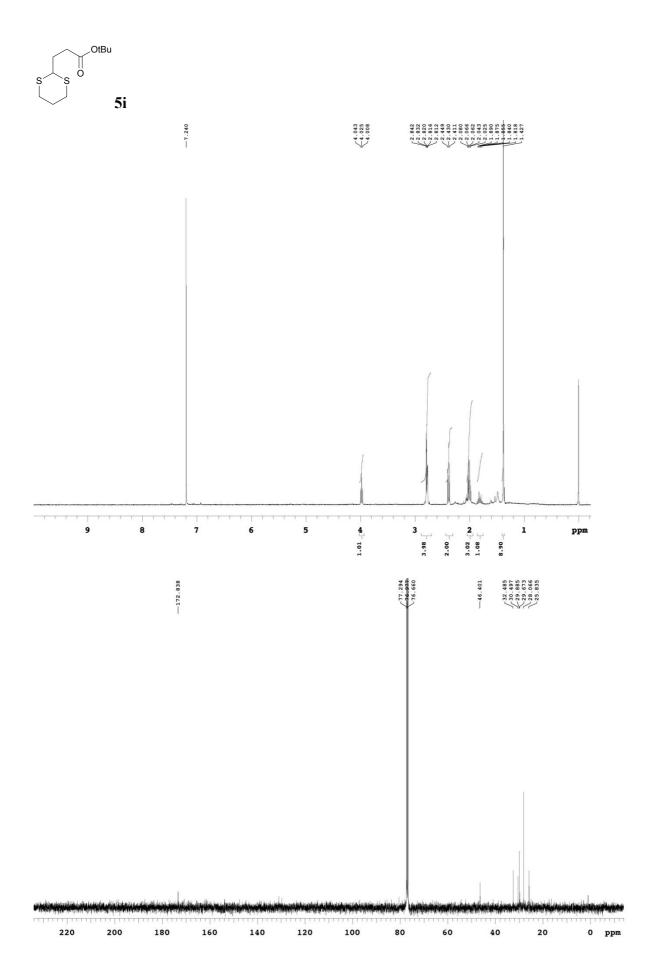


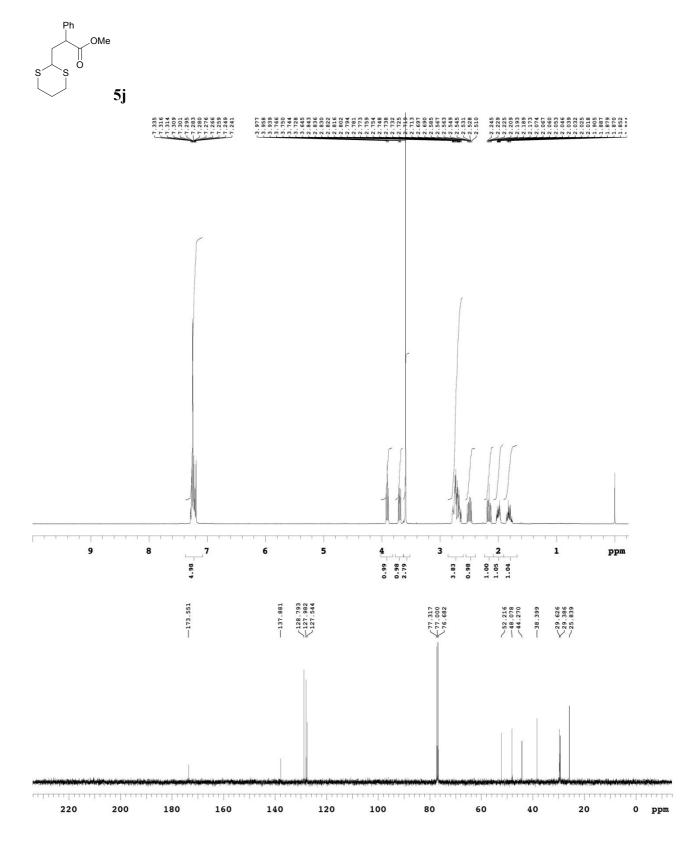


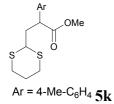


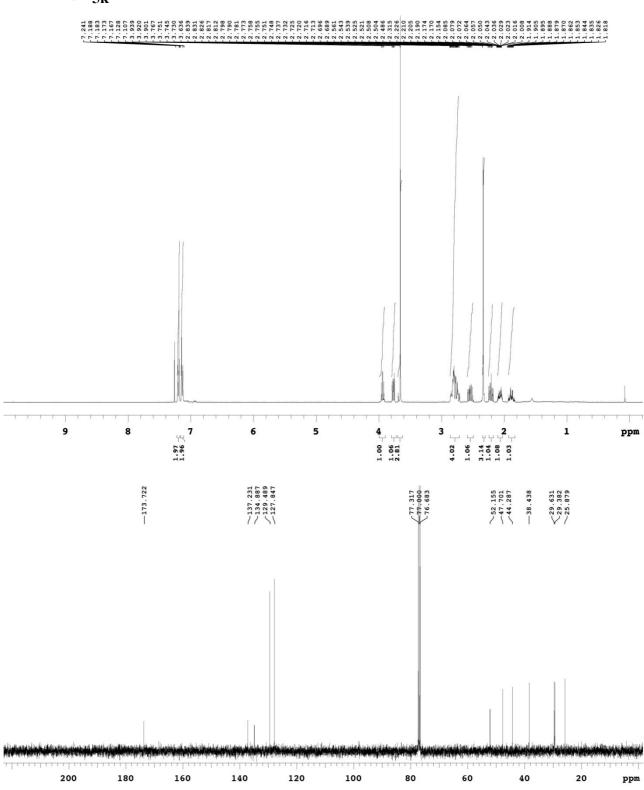


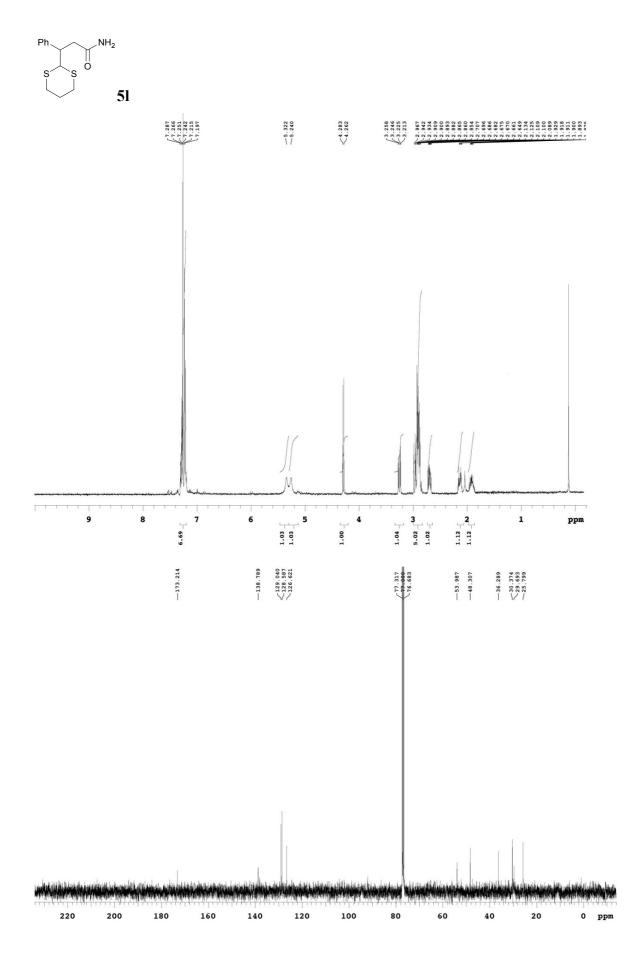


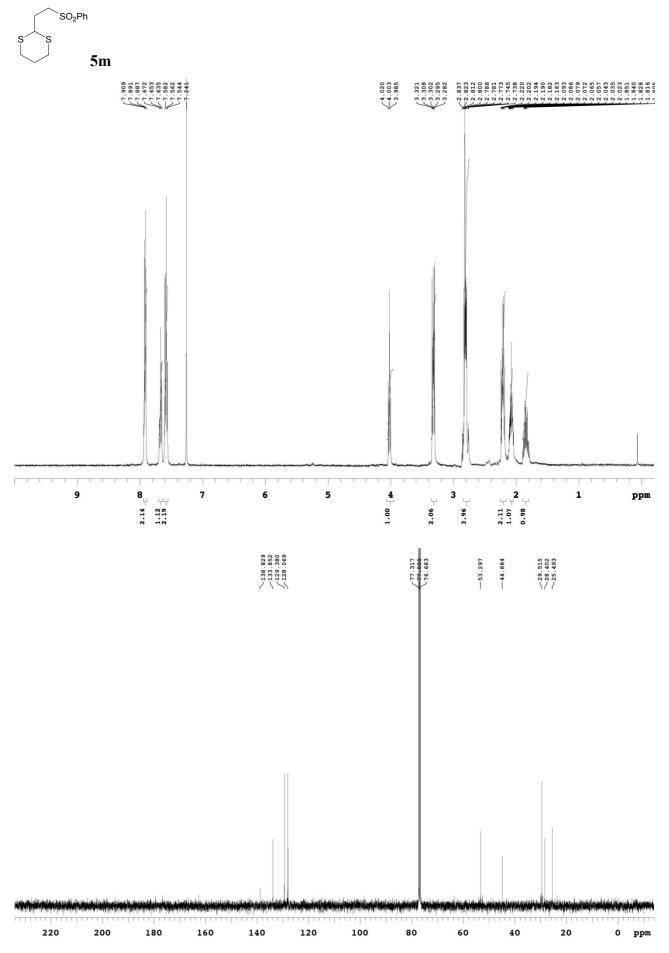


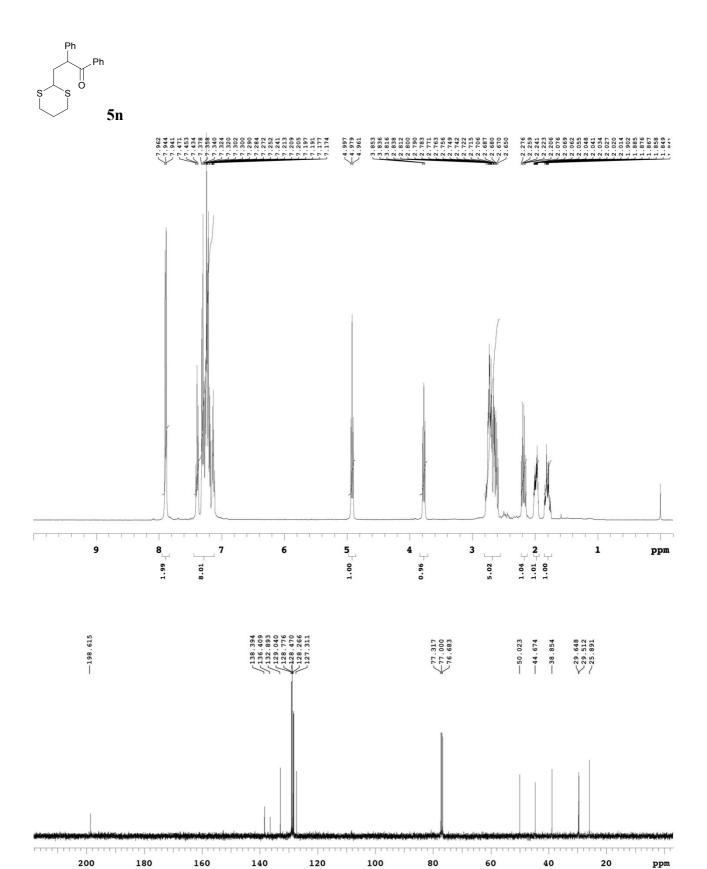


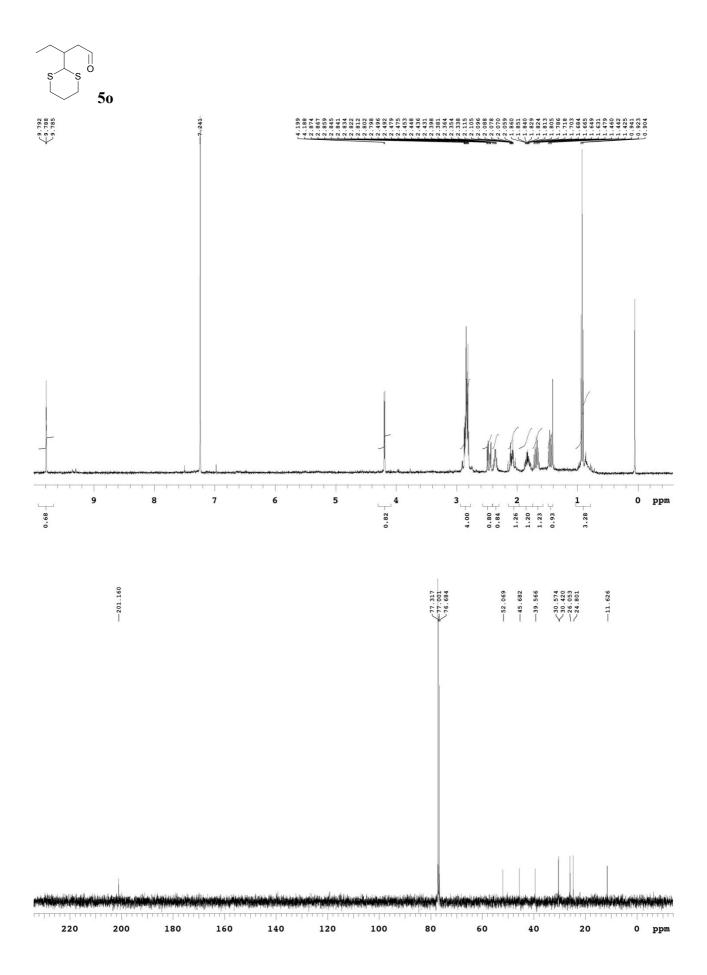


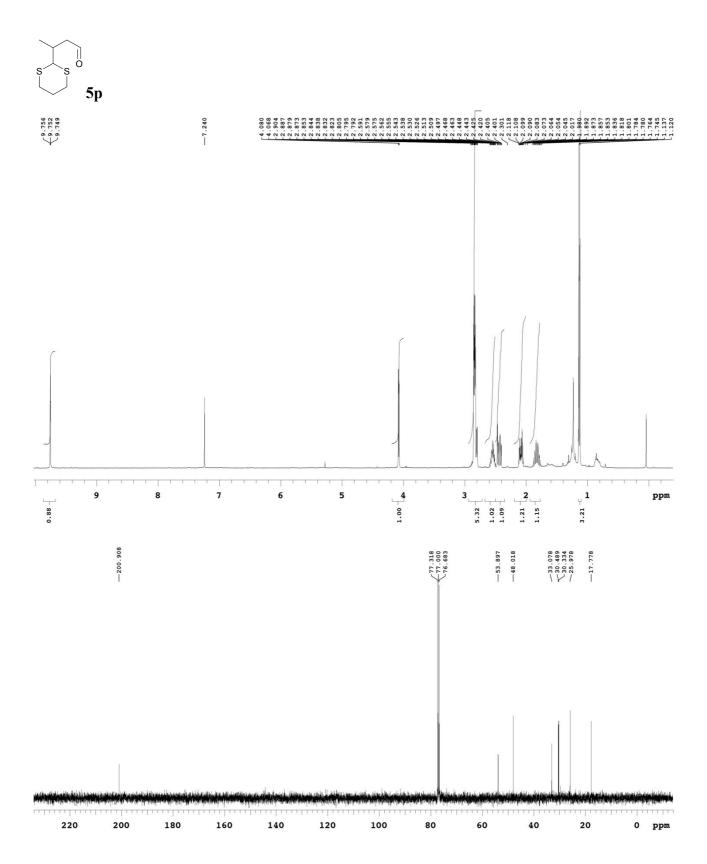


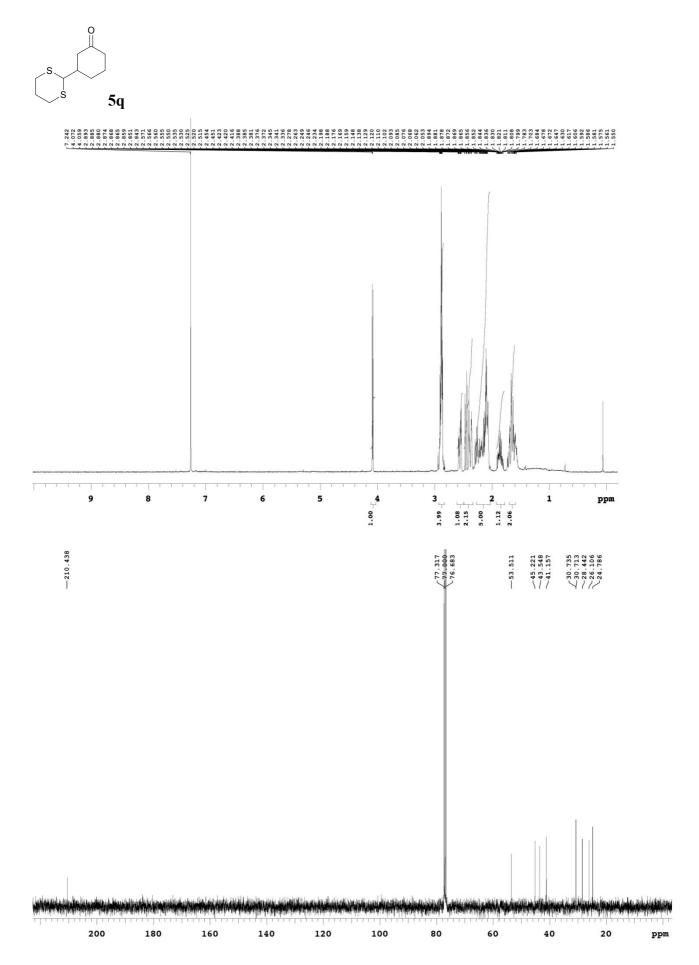




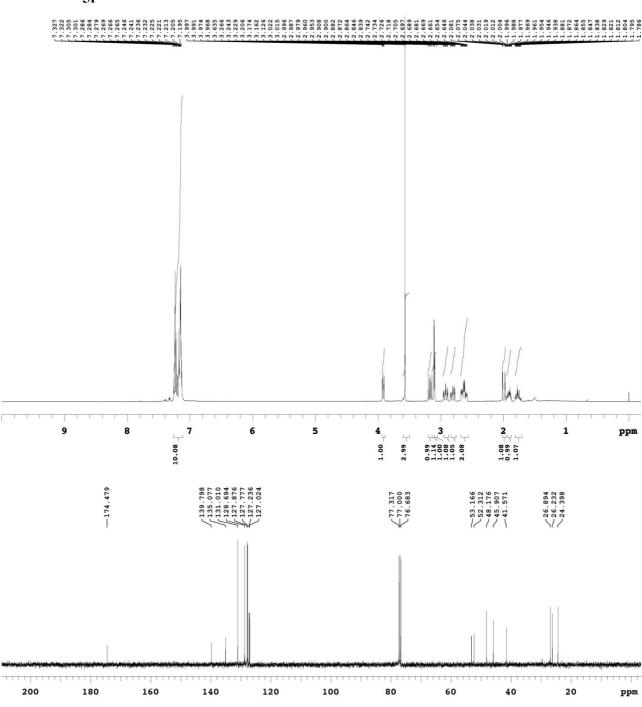




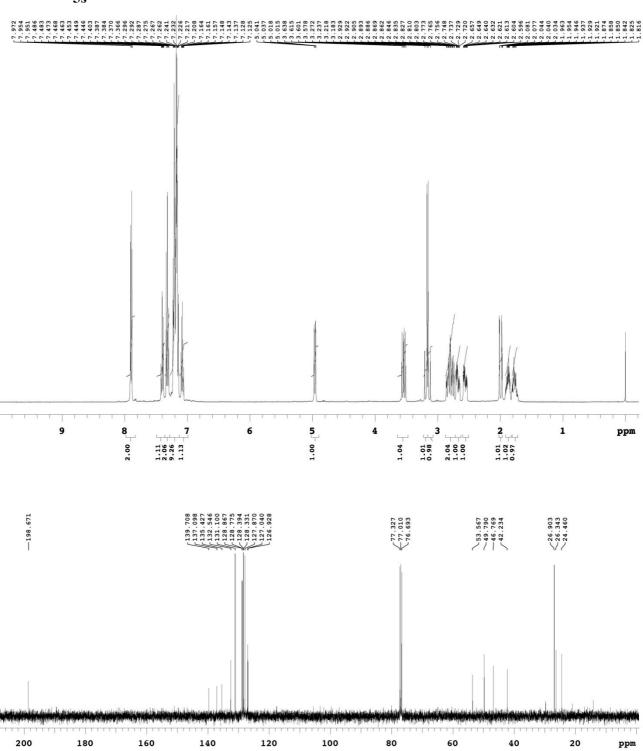


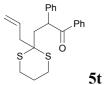


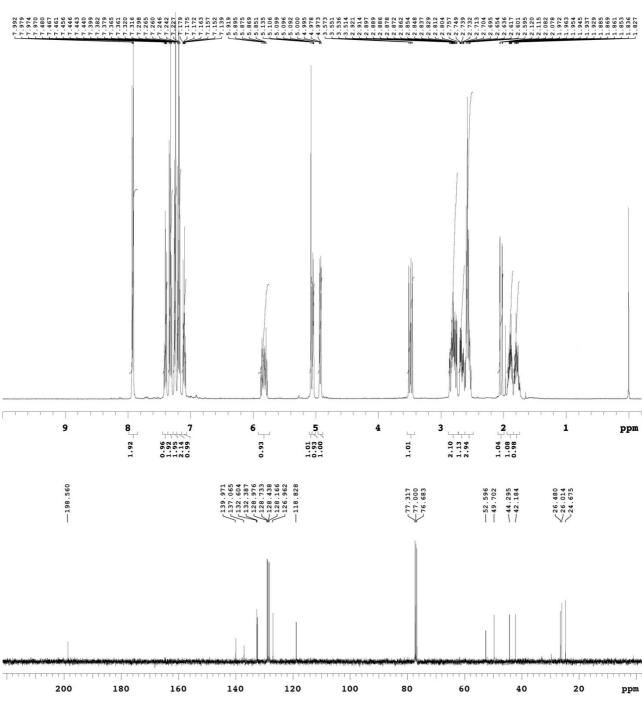


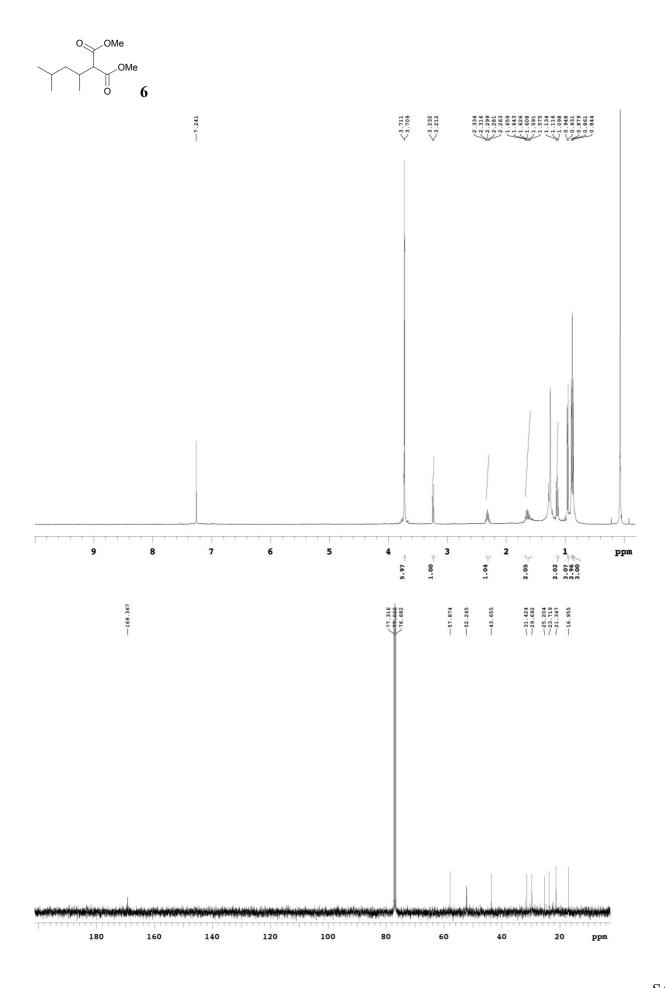


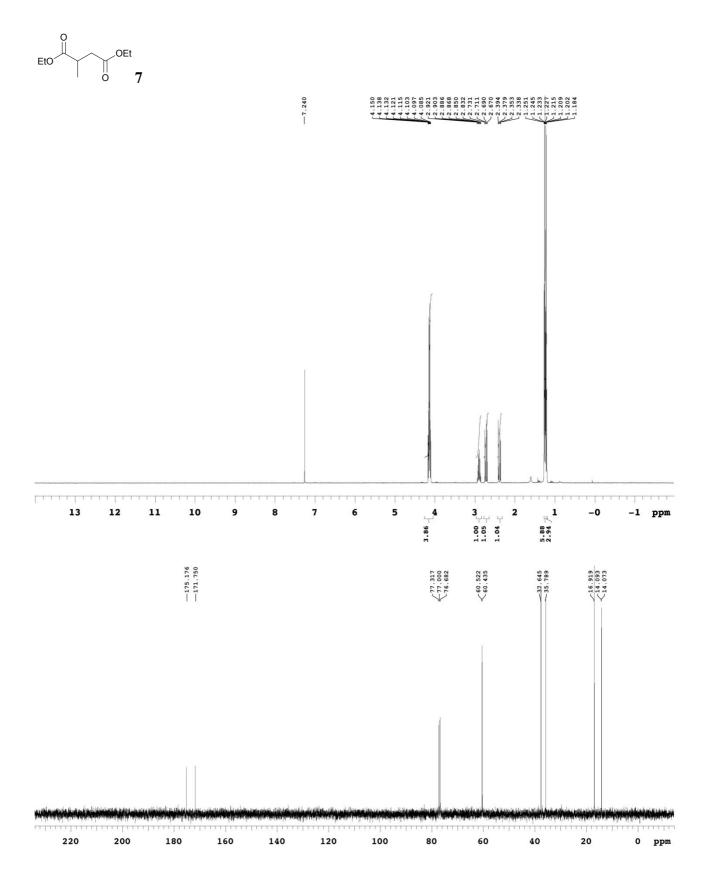




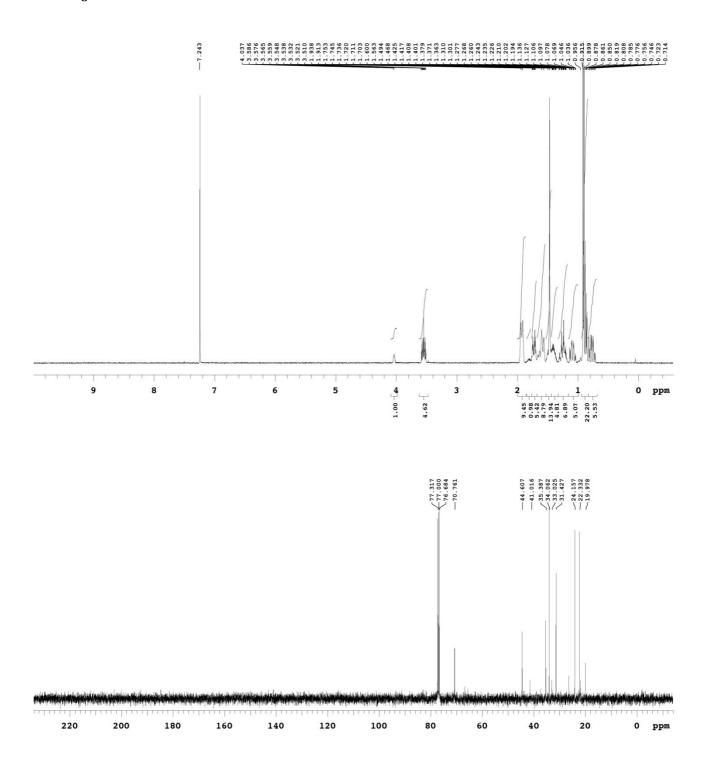


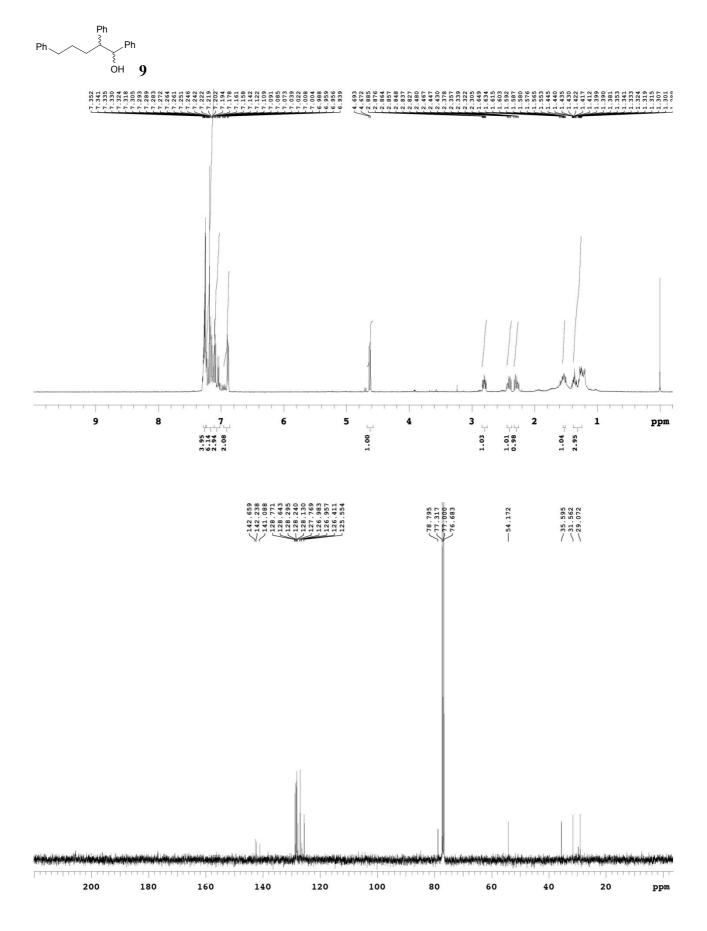


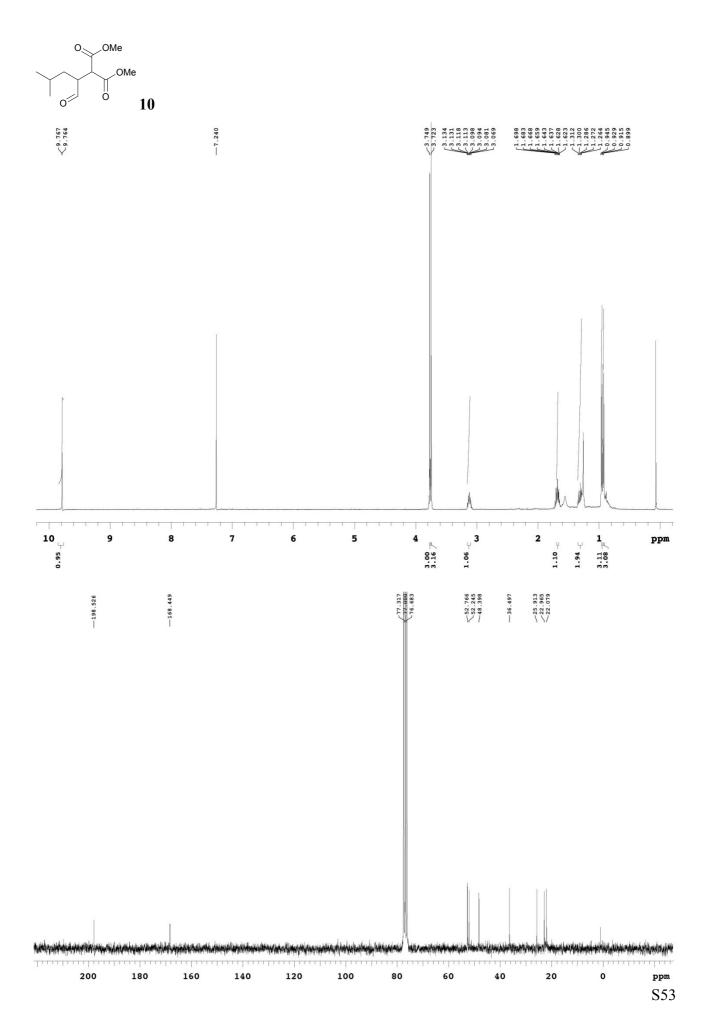


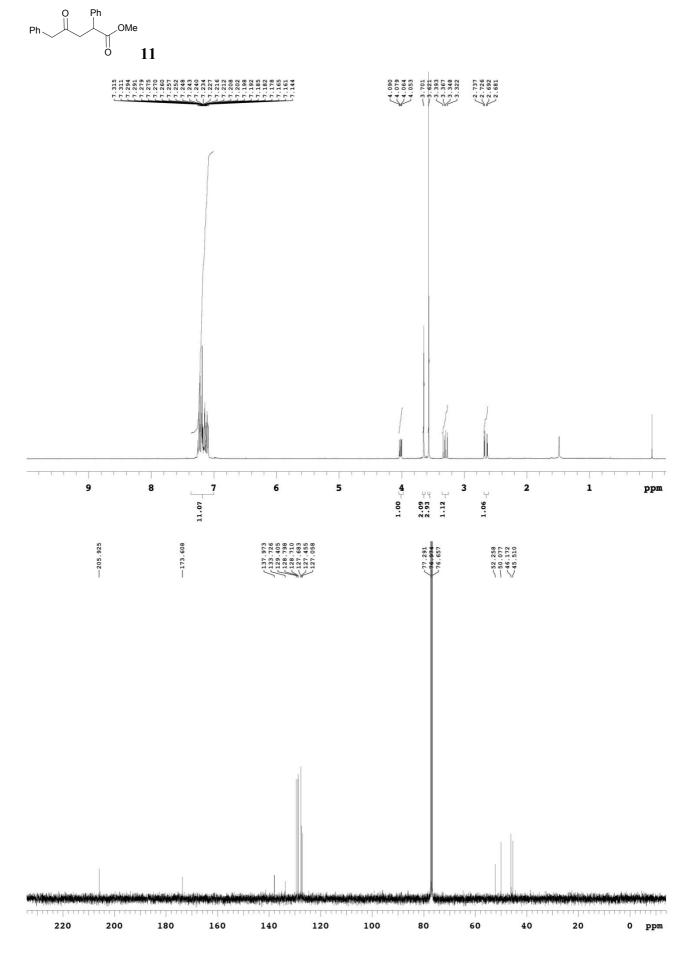


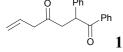


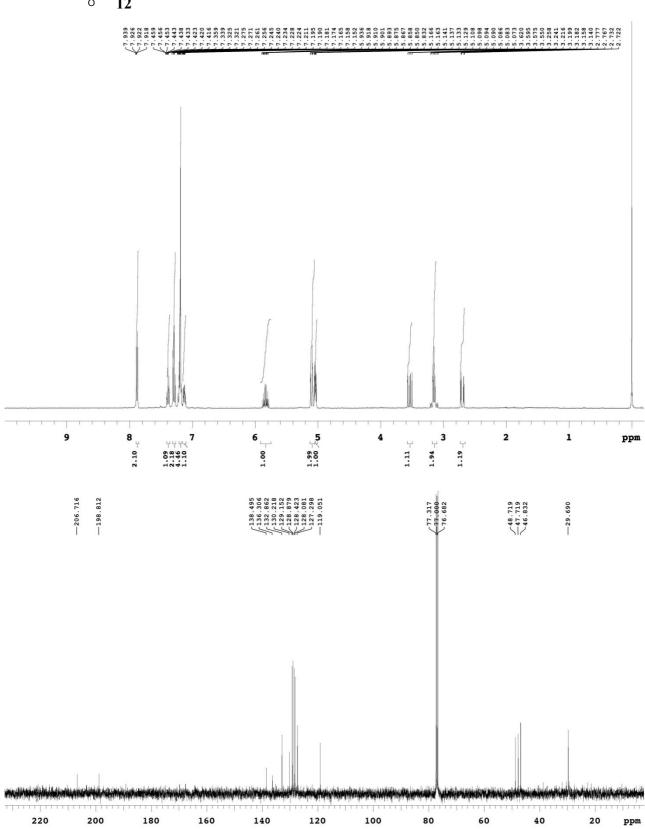


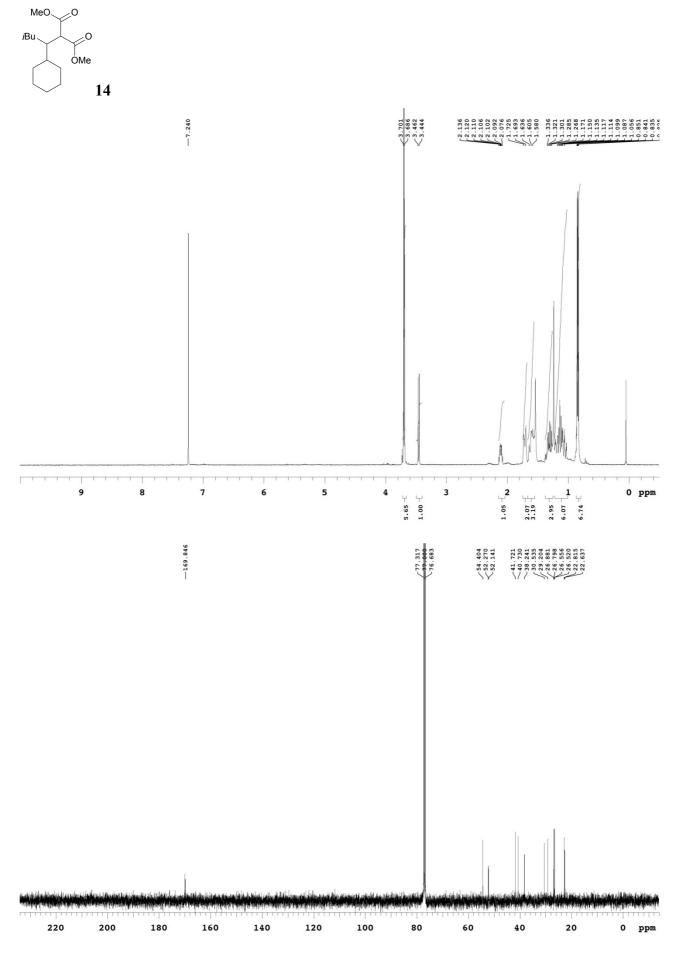


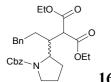


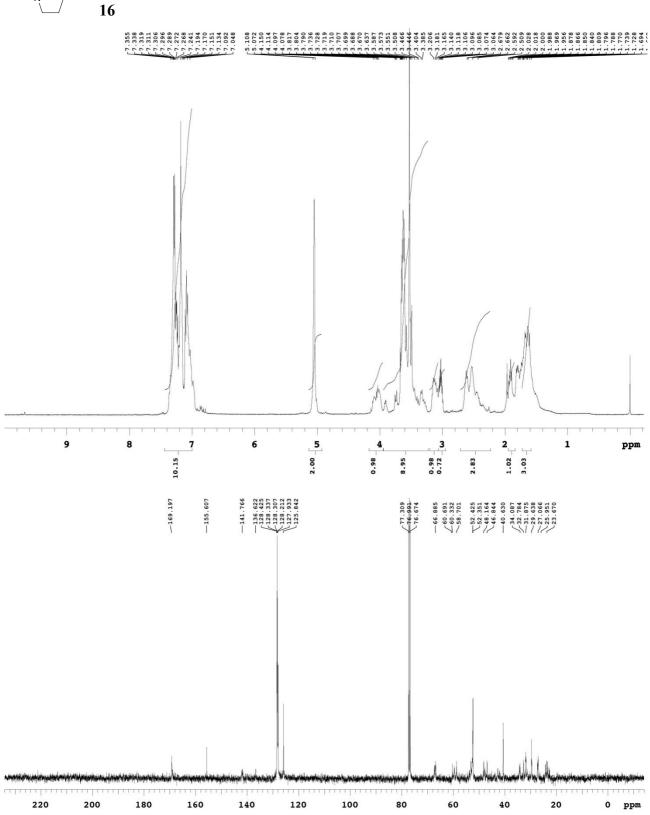


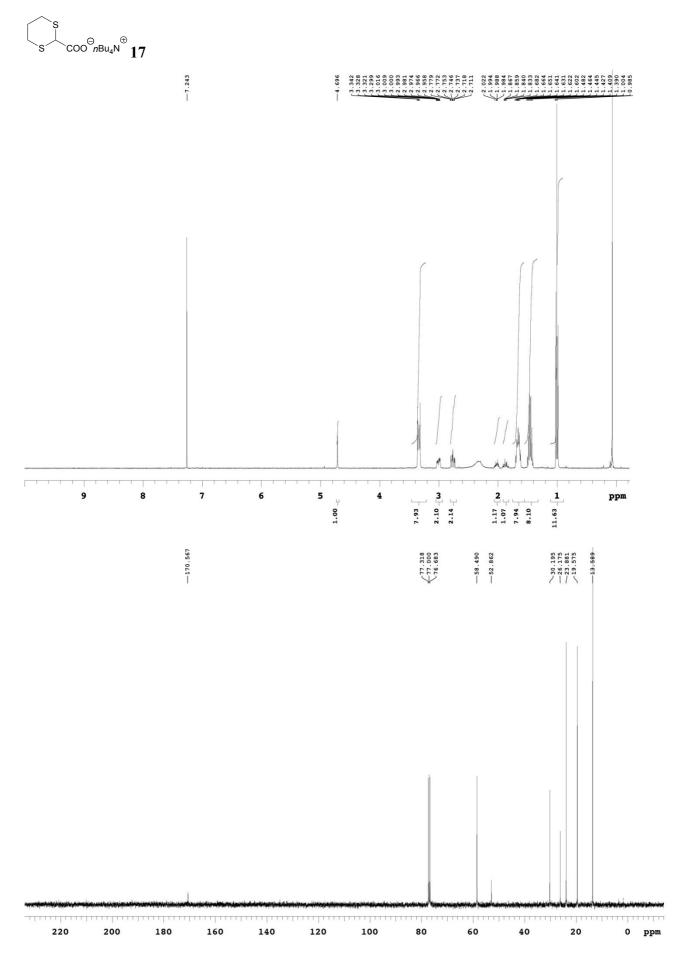


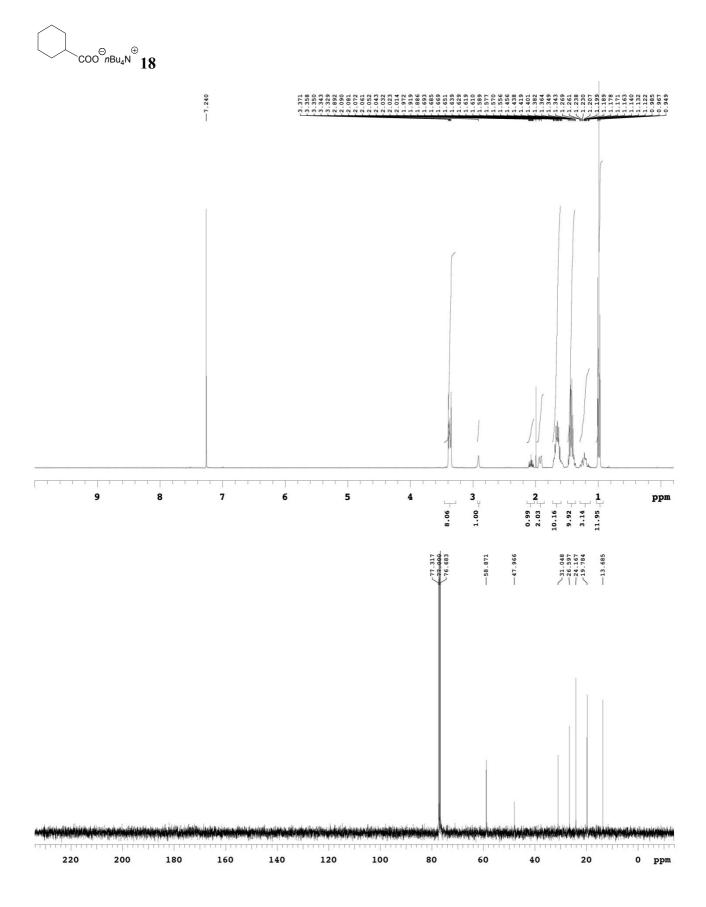


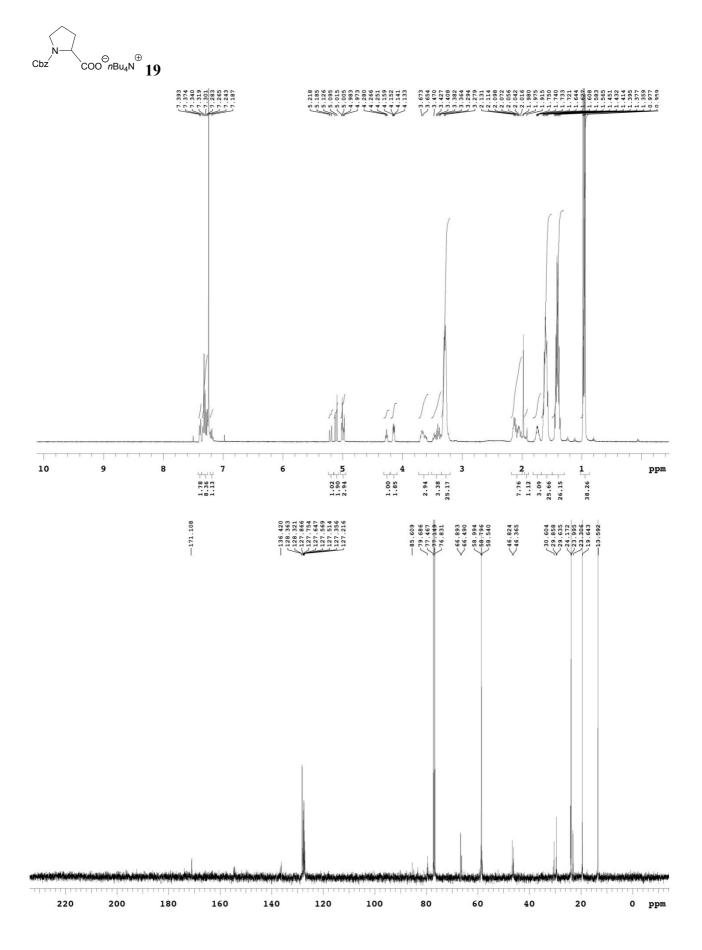






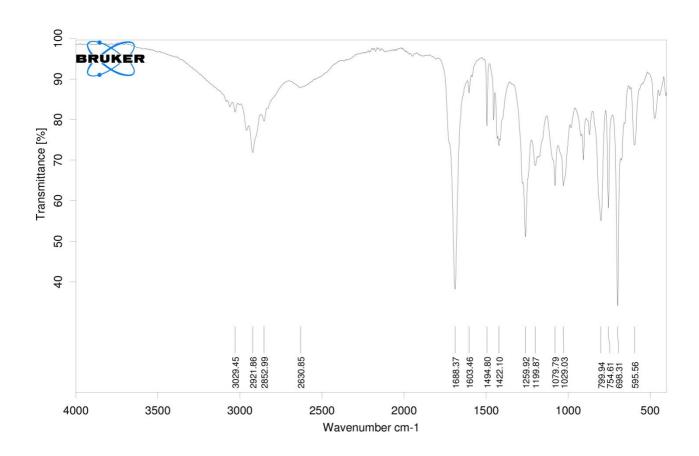




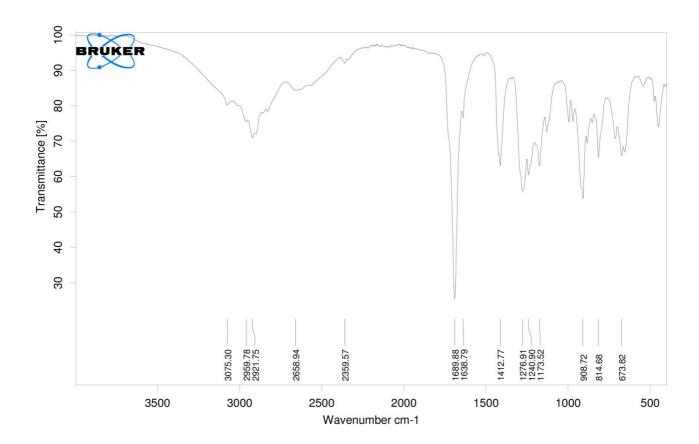


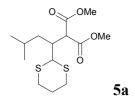
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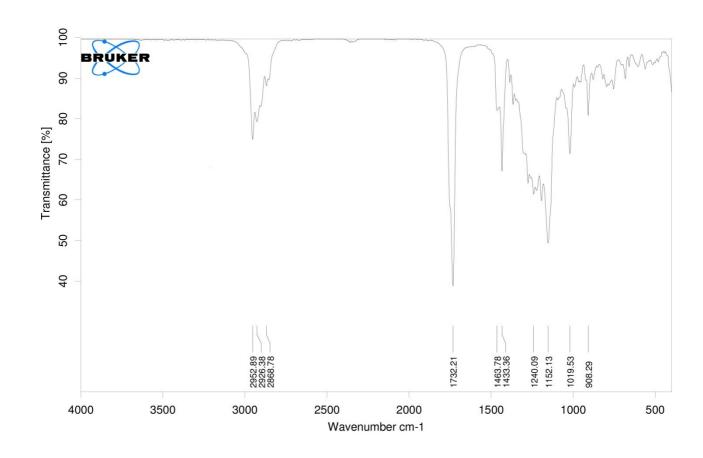


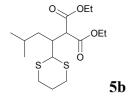


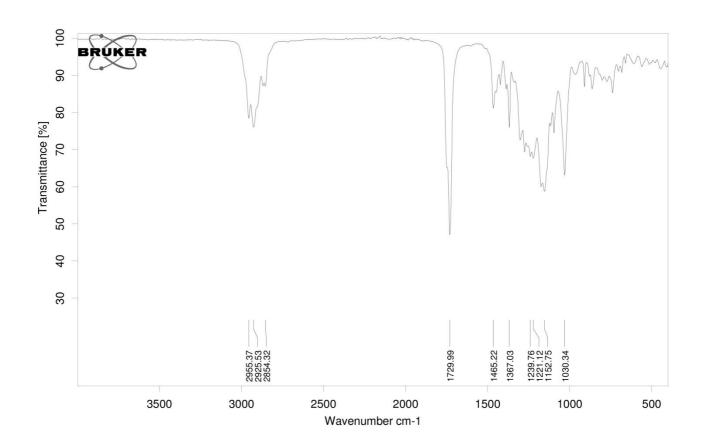


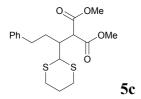


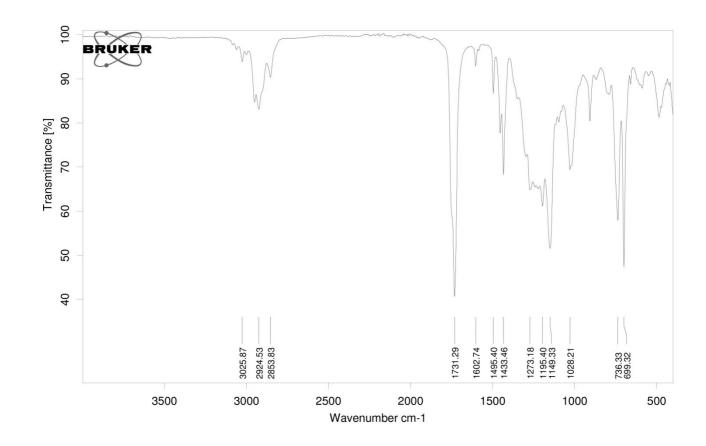


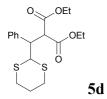


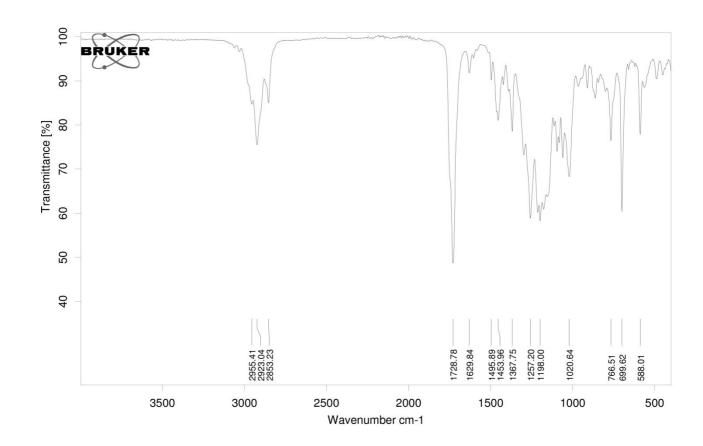


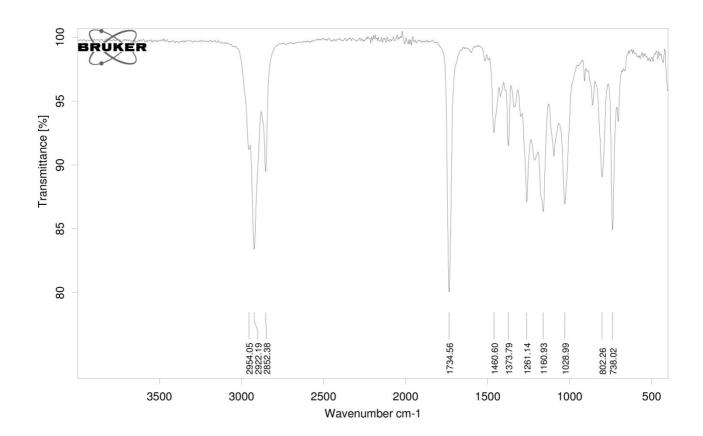


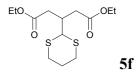


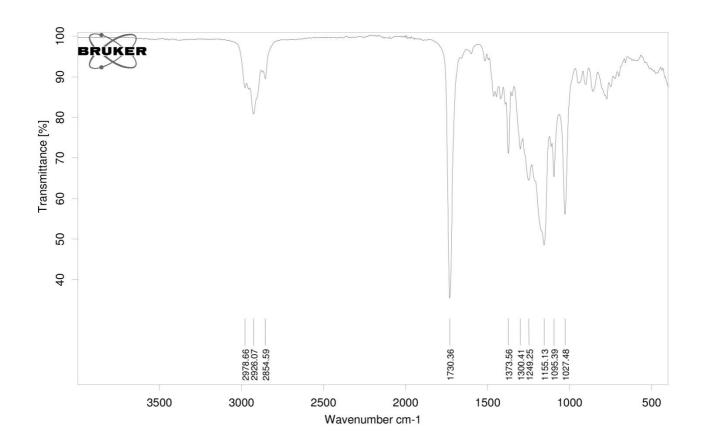


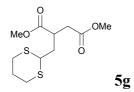


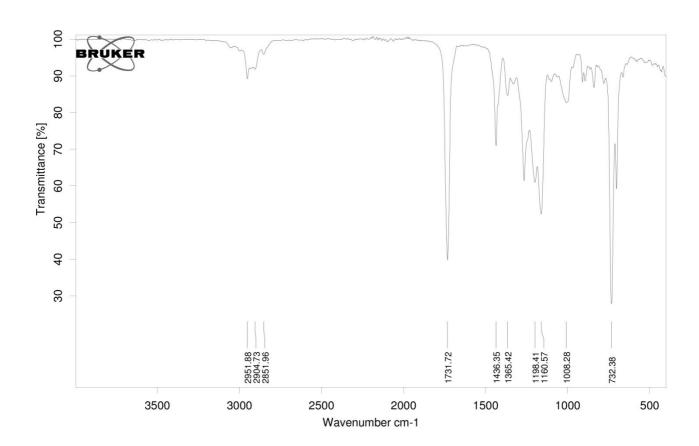


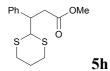


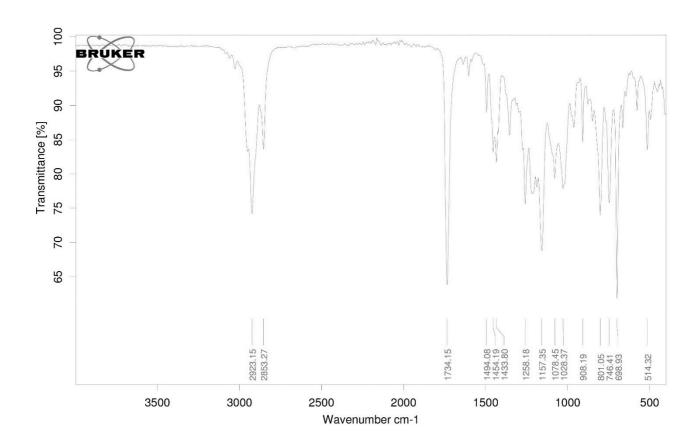


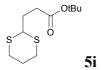


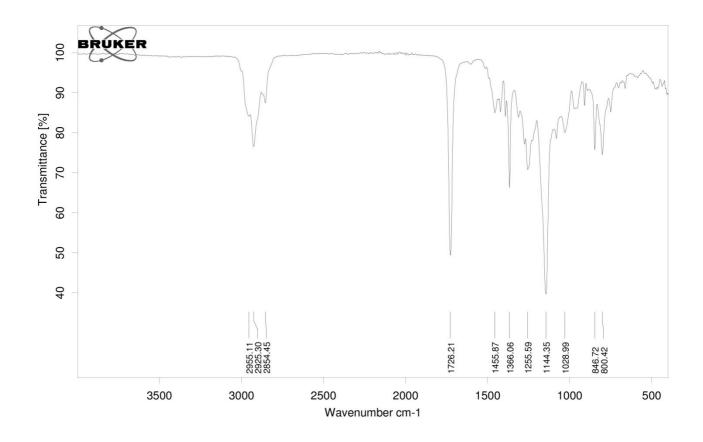


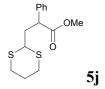


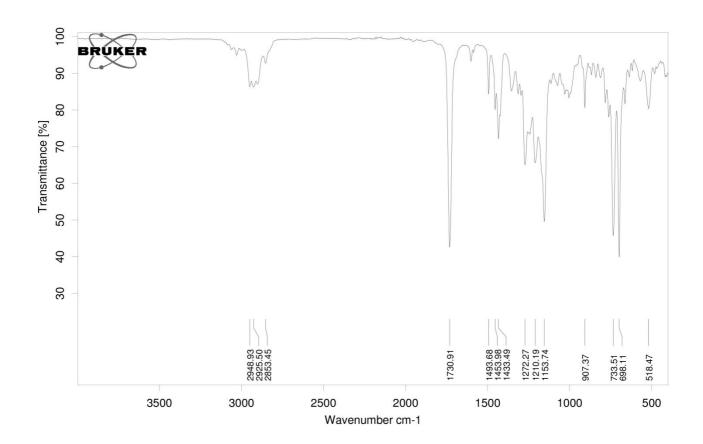












Ar = 
$$4$$
-Me- $C_6H_4$   $5k$ 

