

## Supplemental Document

### Mathematical modeling complete methods:

**Model development.** To quantitatively characterize the inoculum effect with and without functional quorum sensing, previously developed pharmacodynamic models for colistin (1) were used as the basis to model the two different strains. Three pre-existing subpopulations with differencing susceptibilities to polymyxin B or colistin were considered as previously described (1) with the modification of the growth model. A life cycle growth model was used to describe the bacterial growth (2, 3). We assumed that bacteria existed in two states: the vegetative state (S1) (preparing for replication) and the replicating state (S2) (state immediately before replication). The transition from S2 to S1 ( $k_{21}$ ) is assumed to be faster than transition from S1 to S2 ( $k_{12}$ ) making  $k_{12}$  the rate-limiting factor. Hence, it is represented on the growth rate constant of bacteria. The growth model is described by equations 3 and 4. The replication factor (REP) (2) is a mathematical function to limit the total population ( $CFU_{ALL}$ ) to the maximum population size or carrying capacity of *in vitro* system ( $CFU_{MAX}$ ) (eq. 5).

$$\frac{d(S1)}{dt} = Rep \cdot k_{21} \cdot S2 - k_{12} \cdot S1, \quad IC: CFU_0 \quad \text{eq. (S1)}$$

$$\frac{d(S2)}{dt} = -k_{21} \cdot S2 + k_{12} \cdot S1 \quad IC: 0 \quad \text{eq. (S2)}$$

$$REP = 2 \cdot \left( 1 - \frac{CFU_{ALL}}{CFU_{MAX} + CFU_{ALL}} \right) \quad \text{eq. (S3)}$$

Previously developed target site binding model was used to describe the competitive interaction of polymyxin with  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$  at the outer membrane of *P. aeruginosa* —initial site of polymyxin action (1). Free polymyxin concentration ( $f_p$ ) is described by eq. 6 where  $C_{\text{MGCA}}$  is the concentration of cation in the media,  $\text{KD}_{\text{MC}}$  is the binding affinity of polymyxin to charged phosphate groups of lipid A,  $C_P$  is polymyxin concentration,  $\text{KD}_P$  is the polymyxin binding affinity. Binding parameters were fixed to literature values (1), and we assumed that the binding affinity of colistin and polymyxin B are the same. The effective polymyxin concentration ( $C_{p\text{eff}}$ ) was computed as a function of  $f_p$  (fraction of receptors unoccupied by cation),  $\text{EC}_{50}$  (concentration required to achieve 50% of effective colistin concentration relative to the concentration in broth), H (Hill coefficient), and  $C_P$  (polymyxin concentration) (eq. 7) (1). The polymyxin killing activity (KKS) was modeled as a second-order killing process as represented by eq. 8, where  $k_2$  is a second-order rate constant, assumed to be different for each bacterial subpopulation.

$$f_p = 1 - \frac{C_{\text{MGCA}}}{\text{KD}_{\text{MC}} + C_{\text{MGCA}} + \frac{C_P}{\text{MW}_P} + \frac{\text{KD}_{\text{MC}}}{\text{KD}_P}} \quad \text{eq. (S4)}$$

$$C_{p\text{eff}} = \frac{f_p^H}{\text{EC}_{50}^H + f_p^H} \cdot C_P \quad \text{eq. (S5)}$$

$$\text{KKS} = k_2 \cdot C_{p\text{eff}} \quad \text{eq. (S6)}$$

Inoculum effect is incorporated by assuming a hypothetical signaling compartment that inhibits the growth rate ( $\text{INH}_{k12}$ ) and killing activity ( $\text{INH}_{\text{kil}}$ ) of polymyxin, as described in eq. 9 and eq. 10, respectively.

$$\text{INH}_{k12} = 1 - \frac{\text{Imax}_{k12}}{\text{IC}_{50} + C_{\text{sig}}} \cdot C_{\text{sig}} \quad \text{eq. (S7)}$$

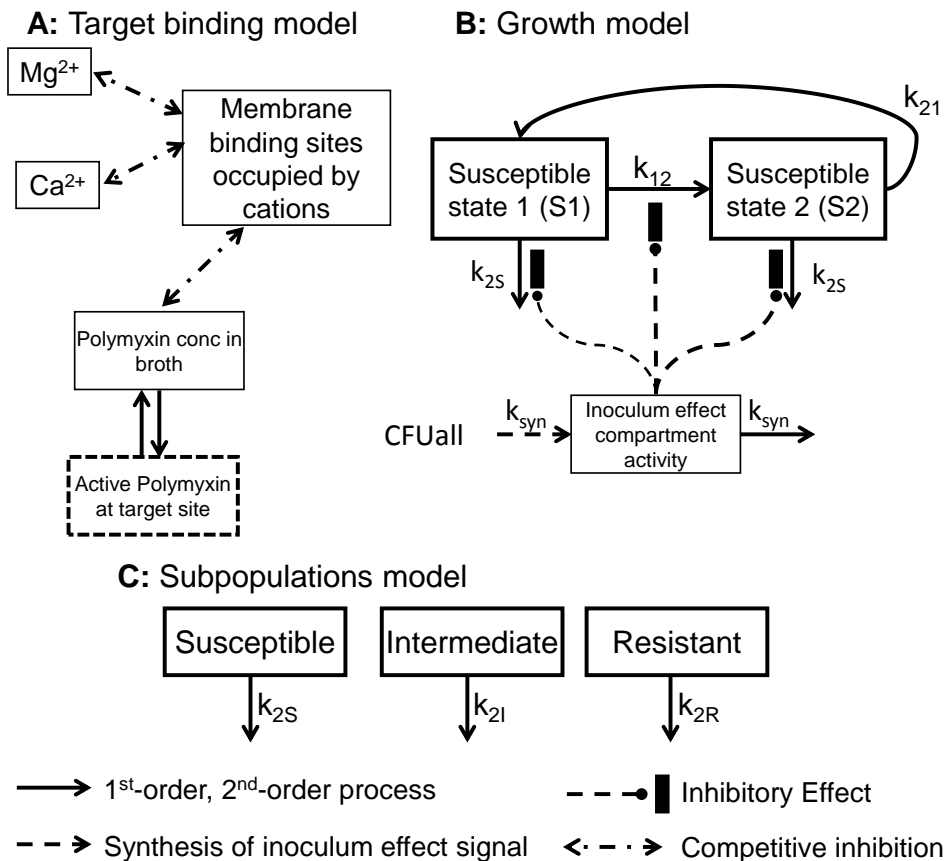
$$\text{INH}_{\text{kill}} = 1 - \frac{\text{Imax}_{\text{kill}}}{\text{IC}_{50} + C_{\text{sig}}} \cdot C_{\text{sig}} \quad \text{eq. (S8)}$$

**Estimation:** Data were modeled using population approach in S-ADAPT software (version 1.57) with SADAPT-TRAN—pre- and post-processing tool to fit all data simultaneously. Importance sampling Monte Carlo parametric expectation maximization method (pmethod=4) algorithm (4, 5) in SADAPT was used. The additive residual error variance model on log<sub>10</sub>-scale and the Poisson error model (eq. 9)) were used for the bacterial count greater or equal to 100 CFU/mL and below 100 CFU/mL, respectively (1, 3).

$$\text{Var} = \left( \text{CFU}_{\text{Plate}} \cdot \text{SD}_{\text{CFU}} \cdot \text{Ln}(10) + \sqrt{\text{CFU}_{\text{Plate}}} \cdot \text{SD}_{\text{Pois}} + \text{SD}_{\text{Add}} \right)^2 \quad \text{eq. (11.)}$$

where, Var is residual variance, CFU<sub>plate</sub> is number of colonies on the plate, SD<sub>CFU</sub> is the standard deviation used to describe the additive error on log<sub>10</sub> scale, and SD<sub>Pois</sub> is standard deviation of Poisson error with a value of 1, and SD<sub>Add</sub> is standard deviation of additive error with a value of 0.25.

**Figure S1:** The pharmacodynamic model for polymyxin antibiotic against *P. aeruginosa*. The model consisted of target binding site model of polymyxin (A), growth, kill and inoculum effect models (B), and a subpopulation model (C). Target site model consisted of a competitive interaction between polymyxin,  $Mg^{2+}$  and  $Ca^{2+}$  at the LPS binding site of outer membrane. The growth model assumed that bacteria existed in two states: vegetative state (state 1) and prior replicate state (state 2). The transition between state 2 to state 1 was assumed to be rapid while state 1 to state 2 is rate limiting step. The attenuation of bacteria killing was assumed due to the hypothetical inoculum effect compartment where the compartment inhibits the activity of polymyxin killing and bacterial growth. The bacterial population was assumed to consist of three pre-existing subpopulations: susceptible, intermediate, and least-susceptible ('resistant').



**Table S1:** Parameter estimates from the pharmacodynamic model

Parameter	Symbol	Polymyxin B		Colistin	
		WT (BCV)	lasRrhIR KO (BCV)	WT (BCV)	lasRrhIR KO (BCV)
Fraction of receptors occupied by cation, resulting in an effective colistin concn of 50% relative to the colistin concn in broth	EC <sub>506</sub>	0.154 (0.503)	0.0290(1.12)	0.375 (0.407)	0.399 (0.0336)
	EC <sub>508</sub>	0.320 (0.112)	0.510 (0.248)	0.473 (0.036)	0.334 (0.0437)
	EC <sub>509</sub>	0.999 (0.777)	0.998 (0.583)	0.999 (0.735)	0.997 (0.348)
Second-order killing rate constant Susceptible population	k <sub>2S</sub> (L/(mg·h))	4.23 (0.263)	5.22 (0.0587)	8.02 (0.0686)	5.00 (0.219)
Intermediate population	k <sub>2I</sub> (L/(mg·h))	0.373 (0.0694)	0.336 (0.0962)	1.28 (0.0214)	0.911 (0.103)
Resistant population	k <sub>2R</sub> (L/(mg·h))	0.00344 (0.0767)	0.0159 (1.21)	0.00145 (0.279)	0.0288 (0.0316)
Maximal inhibition of bacterial killing by signal molecule	I <sub>maxKill</sub>	0.970 (0.134)	0.961 (0.359)	0.999 (0.510)	0.995 (0.606)
Maximal inhibition of bacterial growth by signal molecule	I <sub>maxRep</sub>	0.835 (1.19)	0.938 (0.078)	0.993 (0.993)	0.942 (0.0866)
Signal molecule concn resulting in 50% of Max. effect	Log <sub>10</sub> IC <sub>50</sub>	7.29 (0.158)	7.62 (0.439)	7.16 (0.02)	7.37 (0.122)
Degradation half-life of signal molecule	T <sub>1/2</sub> (K <sub>deg</sub> ) (min)	5.59 (0.259)	10.62 (0.241)	81.6 (1.19)	31.6 (1.51)
Maximum population size	Log <sub>10</sub> POP <sub>max</sub>	9.19 (0.201)	9.5 (0.145)	11.2 (1.11)	9.6 (0.237)
Initial inoculum Size: 10 <sup>6</sup> inoculum	Log <sub>10</sub> CFU <sub>06</sub>	5.75(0.00649)	5.81 (0.0221)	5.87 (0.0401)	6.16 (0.131)
10 <sup>8</sup> inoculum	Log <sub>10</sub> CFU <sub>08</sub>	7.97 (0.0420)	8.04 (0.0338)	8.08 (0.0197)	8.19 (0.0124)
10 <sup>9</sup> inoculum	Log <sub>10</sub> CFU <sub>09</sub>	9.25(0.0300)	9.13 (0.0379)	9.14 (0.0126)	9.32 (0.0709)
Intermediate population as fraction of initial inoculum	Log <sub>10</sub> Fr <sub>I</sub>	-3.69 (0.316)	-4.94 (0.316)	-3.75 (0.316)	-4.49 (0.316)
Resistant population as fraction of initial inoculum	Log <sub>10</sub> Fr <sub>R</sub>	-6.90 (0.807)	-7.32 (0.316)	-6.09 (0.316)	-6.87 (0.316)
Bacterial growth: Mean generation time susceptible population	MTT <sub>12S</sub> (min)	17.6 (0.288)	20.7 (0.15)	24.3 (0.0176)	24.4 (0.0205)
Mean generation time intermediate population	MTT <sub>12I</sub> (min)	18.4 (0.331)	21.5 (0.12)	6.075 (0.475)	9.45 (0.0866)
Mean generation time resistant population	MTT <sub>12R</sub> (min)	112 (0.170)	135 (0.0683)	101 (0.577)	60.0 (1.08)

**Table S2:** Parameter estimates for the hill-type functions corresponding to manuscript Figure 2 which shows the comparative pharmacodynamic responses between PAO1 wild-type and the isogenic *lasR/rhIR* QS double knockout at each inoculum to either polymyxin B or colistin. E<sub>max</sub> is maximal effect, E<sub>o</sub> represents baseline activity, IC<sub>50</sub> is antibiotic concentration (mg/L) which achieves half of the maximum inhibition, and Hill is the Hill coefficient. The parameter estimates of the hill-type models are outlined (C) to mirror A1-B3.

Inoculum			10 <sup>6</sup> CFU/mL		10 <sup>8</sup> CFU/mL		10 <sup>9</sup> CFU/mL	
			Wild Type	$\Delta lasRrhIR$	Wild Type	$\Delta lasRrhIR$	Wild Type	$\Delta lasRrhIR$
Polymyxin B	E <sub>max</sub>	-	4.198 (12.8 %)	5.28 (3.59 %)	2.67 (11.2 %)	3.36 (3.22 %)	2.69 (11.7 %)	1.74 (3.77 %)
	E <sub>o</sub>	-	-0.135 (33.8 %)	-0.18 (12.0 %)	-0.169 (10.6 %)	-0.057 (7.01 %)	-0.014 (6.96 %)	-0.103 (4.67 %)
	IC <sub>50</sub>	mg/L	4.24 (14.0 %)	5.58 (6.12 %)	12.5 (16.0 %)	5.61 (5.71 %)	50.3 (14.3 %)	19.6 (4.78 %)
	Hill	-	5 (FIXED)	5 (FIXED)	2.46 (33.7 %)	5 (FIXED)	1.91 (19.1%)	5 (FIXED)
Colistin	E <sub>max</sub>	-	5.31 (1.51 %)	5.19 (2.00 %)	2.18 (14.3 %)	4.03 (1.48 %)	2.47 (2.75 %)	4.37 (6.97 %)
	E <sub>o</sub>	-	0.230 (0.694 %)	-0.208 (0.936 %)	-0.166 (2.08 %)	0.467 (0.467 %)	-0.028 (0.287%)	0.080 (0.313 %)
	IC <sub>50</sub>	mg/L	4.82 (2.02 %)	3.74 (2.23 %)	8.72 (24.6 %)	7.04 (2.64 %)	32.0 (3.50 %)	59.3 (12.3 %)
	Hill	-	3.93 (5.51 %)	5.99 (24.6 %)	2.63 (58.1 %)	4.77 (12.2 %)	2.89 (9.80 %)	1.20 (7.40 %)

## References

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2. **Bulitta JB, Ly NS, Yang JC, Forrest A, Jusko WJ, Tsuji BT.** 2009. Development and qualification of a pharmacodynamic model for the pronounced inoculum effect of ceftazidime against *Pseudomonas aeruginosa*. *Antimicrob Agents Chemother* **53**:46-56.
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5. **Bulitta JB, Bingolbali A, Shin BS, Landersdorfer CB.** 2011. Development of a new pre- and post-processing tool (SADAPT-TRAN) for nonlinear mixed-effects modeling in S-ADAPT. *The AAPS journal* **13**:201-211.