

Supplementary information

**Application of bacterial cytological profiling to crude natural product extracts
reveals the antibacterial arsenal of *Bacillus subtilis***

Poochit Nonejuie^{a*}, Rachelle M. Trial^{a*}, Gerald L. Newton^a, Anne Lamsa^a, Varahenage Ranmali Perera^a, Julieta Aguilar^a, Wei-Ting Liu^b, Pieter C. Dorrestein^{b, c}, Joe Pogliano^a and Kit Pogliano^{a**}

^a Division of Biological Sciences, University of California, San Diego

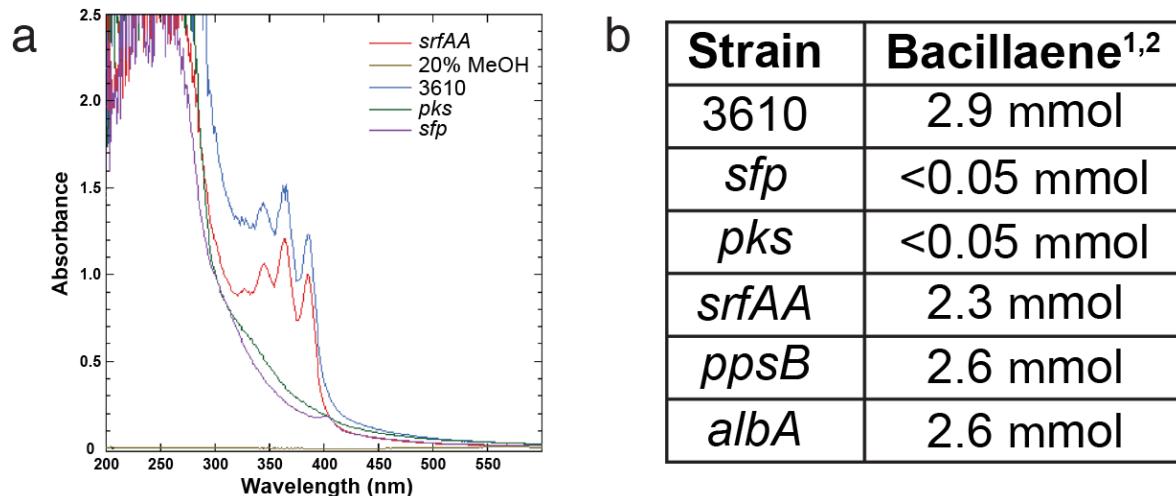
^b Skaggs School of Pharmacy and Pharmaceutical Sciences, University of California, San Diego

^c Center for Marine Biotechnology and Biomedicine, University of California, San Diego

*Indicates shared first author

**To whom correspondence should be directed: E-mail: kpogliano@ucsd.edu. Phone: (858) 822-1314 Mailing address: Natural Sciences Building 4113, 9500 Gilman Drive, La Jolla, CA 92093-0377

Supplementary figures



¹estimated extinction coefficient at 363 nm ($\epsilon_{363\text{nm}}$) is $80\text{mM}^{-1}\text{cm}^{-1}$

²mmol in 10 ml extract

Figure S1 Bacillaene signature spectrum and amount found in each extract

(a) Absorption spectrum of *B. subtilis* ethanolic crude extracts assayed for bacillaene (b)

Estimated bacillaene content in each crude extract based on 363 nm absorbance

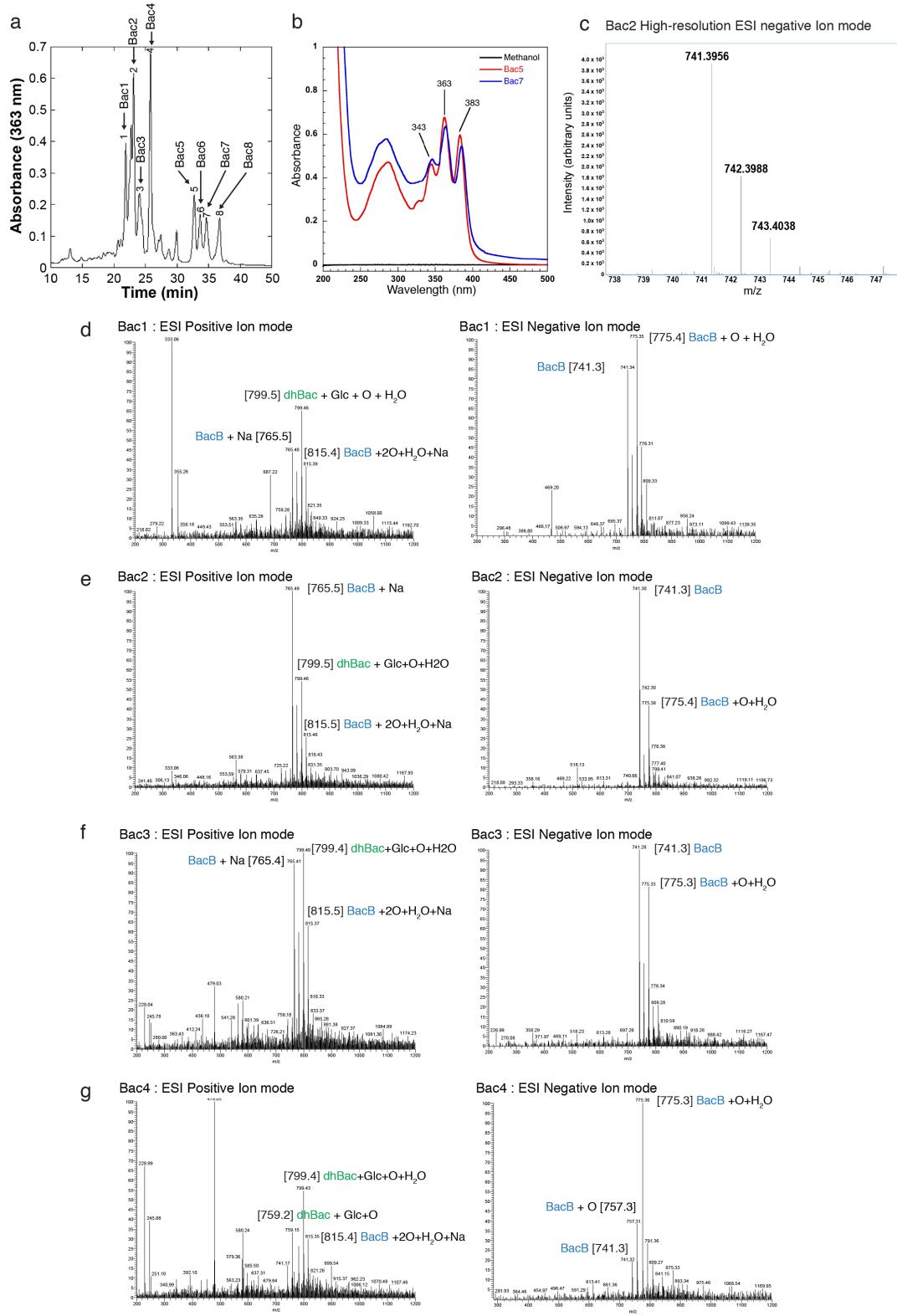


Figure S2

Figure S2 Bacillaene B dominated other bacillaene species in the early HPLC fractions (Bac1-Bac4 fractions) (a) HPLC separation of bacillaenes from *srfAA* SepPak fraction 5 showing 8 different major peaks (Bac1-Bac8) (b) Absorption spectrum of HPLC purified bacillaene fraction 5 and fraction 7 (c) High-resolution ESI-MS analysis in the negative ion mode of bacillaene fraction 2 (d) ESI-MS analysis in the positive (left) and negative (right) ion mode of fraction 1 from the bacillaene purification (e) ESI-MS analysis in the positive (left) and negative (right) ion mode of fraction 2 from the bacillaene purification (f) ESI-MS analysis in the positive (left) and negative (right) ion mode of fraction 3 from the bacillaene purification (g) ESI-MS analysis in the positive (left) and negative (right) ion mode of fraction 4 from the bacillaene purification

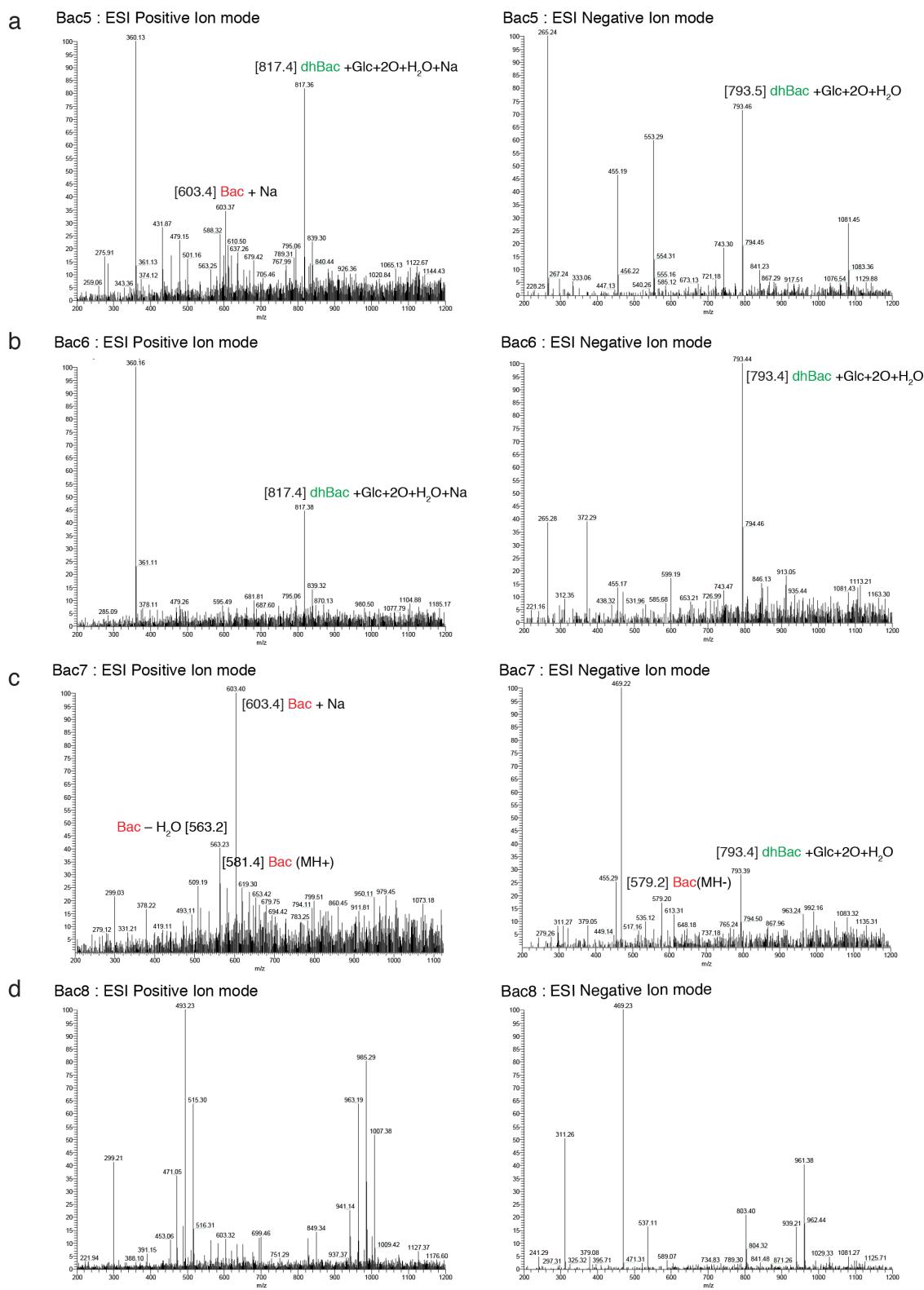


Figure S3

Figure S3 Bacillaene found in the later fractions of HPLC (Bac5 and Bac7)

(a) ESI-MS analysis in the positive (left) and negative (right) ion mode of fraction 5 from the bacillaene purification (b) ESI-MS analysis in the positive (left) and negative (right) ion mode of fraction 6 from the bacillaene purification (c) ESI-MS analysis in the positive (left) and negative (right) ion mode of fraction 7 from the bacillaene purification (d) ESI-MS analysis in the positive (left) and negative (right) ion mode of fraction 8 from the bacillaene purification

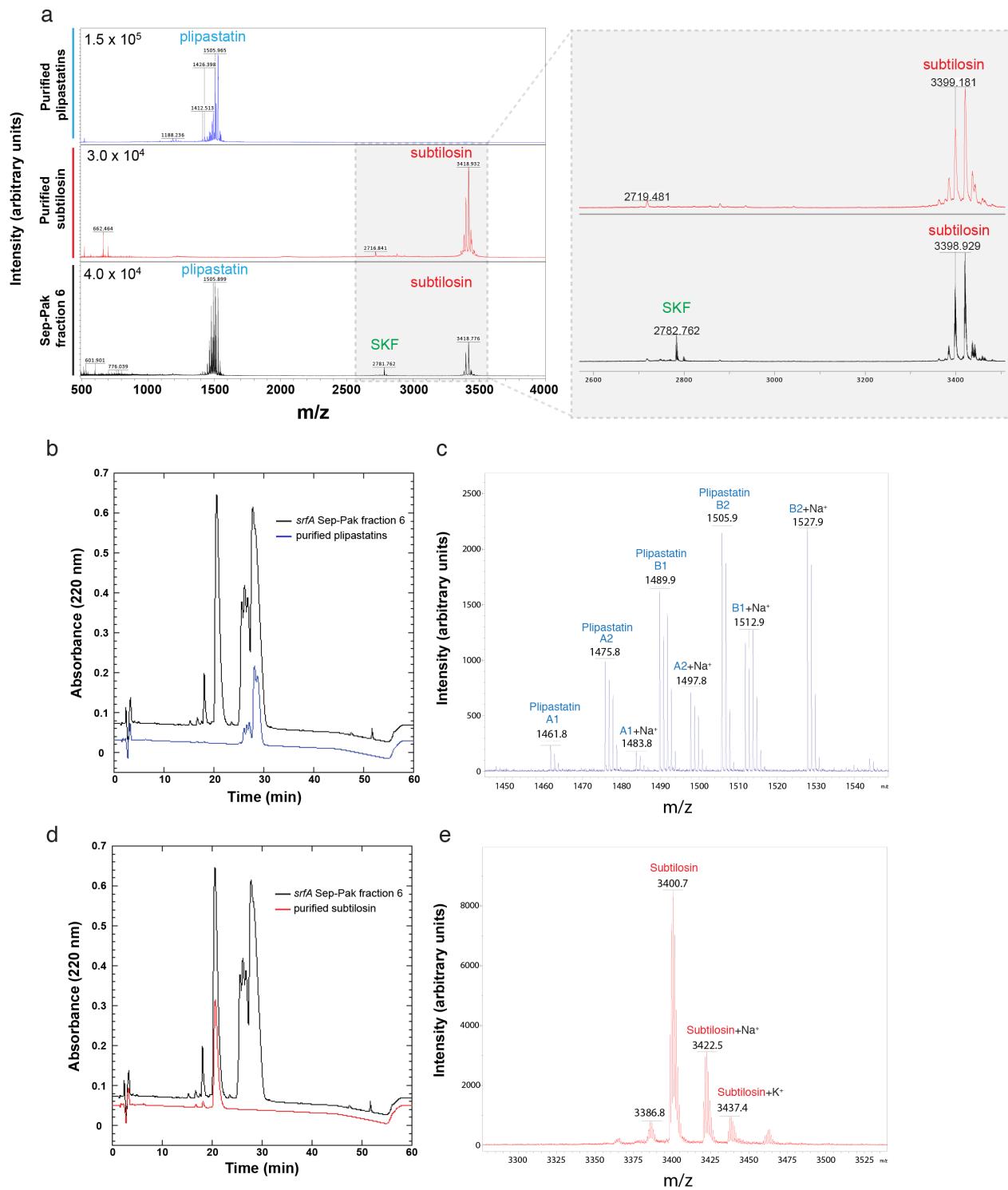


Figure S4

Figure S4 Plipastatin and subtilosin purification

(a) MALDI mass spectrometry analysis (positive mode) of purified plipastatins, purified subtilosin from *B. subtilis srfAA* SepPak fraction 6 (b) HPLC purification of plipastatins from *B. subtilis srfAA* SepPak fraction 6.(c) MALDI-TOF mass spectrometry analysis (positive mode) of HPLC purified and reconstituted *B. subtilis* plipastatins A1, A2, B1 and B2 (d) HPLC purification of subtilosin from *B. subtilis srfAA* SepPak fraction 6. (e) MALDI-TOF mass spectrometry analysis (positive mode) of HPLC purified subtilosin

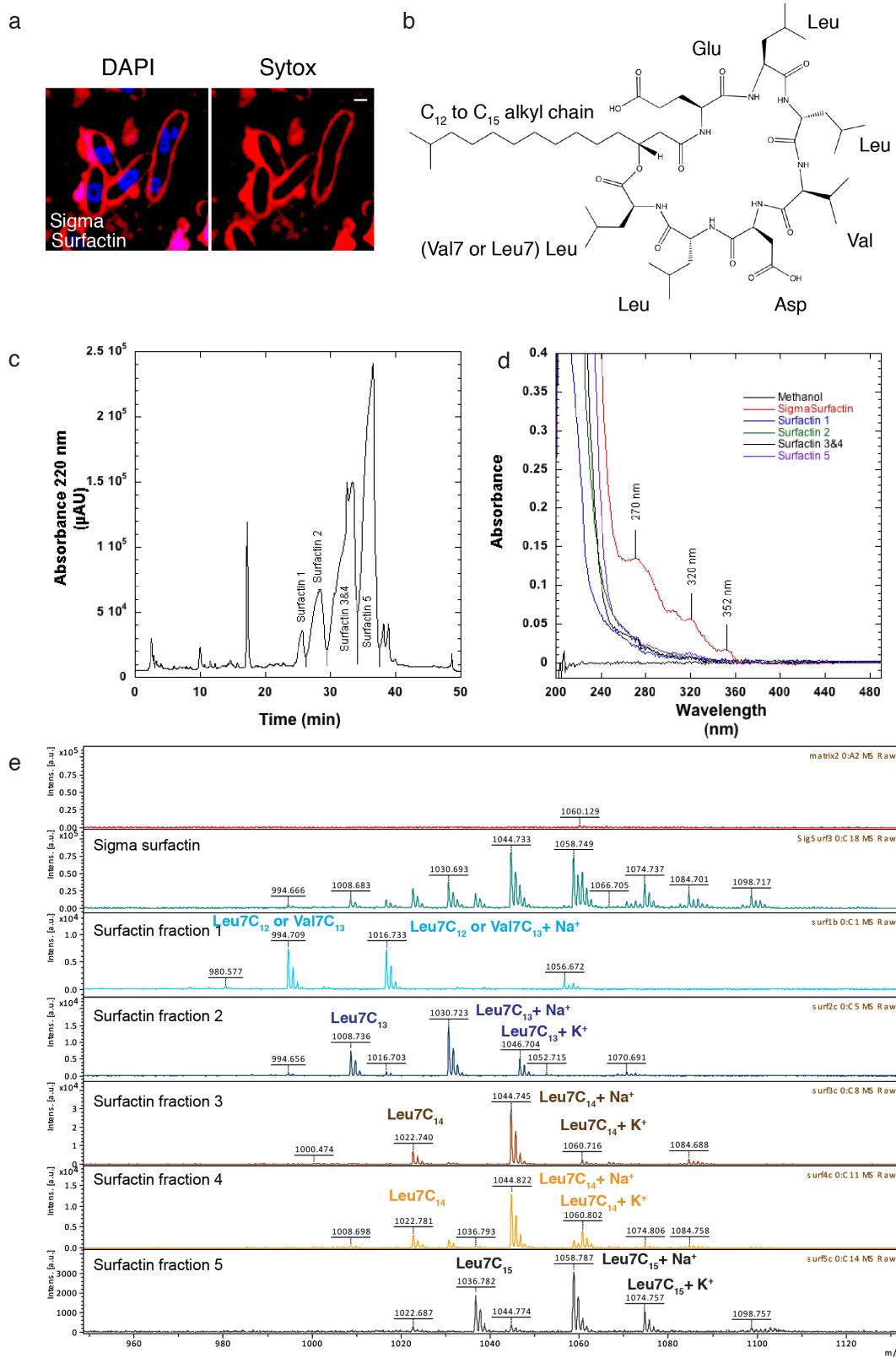


Figure S5

Figure S5 Surfactin purification

(a) *E. coli* cells treated for 2 hours with Sigma commercial surfactin 576 μ M. *E. coli* cells were stained with FM4-64 (Red), DAPI (Blue) and SYTOX-Green (Green). Scale bar, 1 μ m. (b) Surfactin chemical structure and annotation (c) HPLC purification of commercial *B. subtilis* surfactin (Sigma S3523) (d) UV-Visible absorption spectrum of commercial surfactin and HPLC purified surfactins assayed for bacillaene contamination. (e) MALDI-TOF mass spectrometry analysis (positive mode) of commercial surfactin and HPLC purified surfactin fractions.

Supplementary tables

Table S1 Strains used in this study

Strains	Genotype	References
<i>Bacillus subtilis</i>		
3610	undomesticated parent of 168	Previous study ¹
PSK0178	$\Delta pks::spc$	Previous study ²
PSK0060	$srfAA::mls$	Previous study ³
PSK0417	$\Delta albA::kan$	Previous study ⁴
EG220-1	$sfp::mls$	Previous studies ^{1,3}
PSK0156	$ppsB::spc$	Previous study ³
<i>Escherichia coli</i>		
NR698	$IptD4213$	Previous study ⁵

Table S2. Bacillaene fractions from HPLC of *srfAA* SepPak 5

Fraction (% of Total Bacillaenes)	Bacillaene concentration (μM) Assumes $E_{363} \sim 80 \text{ mM}^{-1}$	Killing activity*	Minimal Cytological Concentration (μM)
Fraction 1 : Bac1 (9.5%)	300	-	NA
Fraction 2 : Bac2 (26%)	353	+	20
Fraction 3 : Bac3 (11%)	188	-	NA
Fraction 4 : Bac4 (18%)	158	-	NA
Fraction 5 : Bac5 (7%)	61	-	NA
Fraction 6 : Bac6 (6%)	74	-	NA
Fraction 7 : Bac7 (6%)	68	+	2
Fraction 8 : Bac8 (6%)	44	-	NA

* Determined based on killing spot test

Table S3 Minimal inhibitory concentration of purified natural products from *B. subtilis*

Molecules	Minimal inhibitory concentration (μ M)
Bacillaene	NA
Bacillaene B	NA
Plipastatin	>128
Subtilosin	8
Surfactin	>512
SKF	NA

Supplementary references

1. Branda, S. S., González-Pastor, J. E., Ben-Yehuda, S., Losick, R. & Kolter, R. Fruiting body formation by *Bacillus subtilis*. *Proc. Natl. Acad. Sci.* **98**, 11621–11626 (2001).
2. Straight, P. D., Fischbach, M. A., Walsh, C. T., Rudner, D. Z. & Kolter, R. A singular enzymatic megacomplex from *Bacillus subtilis*. *Proc. Natl. Acad. Sci.* **104**, 305–310 (2007).
3. Straight, P. D., Willey, J. M. & Kolter, R. Interactions between *Streptomyces coelicolor* and *Bacillus subtilis*: Role of Surfactants in Raising Aerial Structures. *J. Bacteriol.* **188**, 4918–4925 (2006).
4. Butcher, R. A. et al. The identification of bacillaene, the product of the PksX megacomplex in *Bacillus subtilis*. *Proc. Natl. Acad. Sci.* **104**, 1506–1509 (2007).
5. Ruiz, N., Falcone, B., Kahne, D. & Silhavy, T. J. Chemical Conditionality: A GeneticStrategy to Probe Organelle Assembly. *Cell* **121**, 307–317 (2005).