

Fig S1. Genome map of StFRB508. Starting from the inside, circle 1 represents the mean of centered G + C content (bars facing outside, above mean; bars facing inside, below mean); circle 2 shows the GC skew (G - C)/(G + C); circle 3 represents comparison of gene content with *P. chlororaphis* subsp. *aurantiaca* JD37; the outermost ring highlights the regions of genetic elements. Prophage (SP) is shown in red and quorum sensing-related genes are shown in green. The inner black circle shows the scale line in kbps. The circular map that illustrates the general genomic feature as well as its comparison with another strain was plotted by using the BRIG program (http://brig.sourceforge.net/). The putative prophage regions were identified by PHAST program (http://phast.wishartlab.com/).



Fig. S2. Mass spectra of AHLs extracted from the cell-free supernatant of 508ACI for PhzI. After fractionation by reverse-phase HPLC, the ESI-MS/MS fragment peaks of AHLs were analyzed. All corresponding peaks for respective 3-OH-C6-HSL (A; m/z 216), 3-OH-C8-HSL (B; m/z 244), 3-OH-C10-HSL (C; m/z 272), and 3-OH-C12-HSL (D; m/z 300) along with the product ion peaks (m/z 102) are marked by arrows.



Fig. S3. Mass spectra of AHLs extracted from the cell-free supernatant of 508PCI for AurI. After fractionation by reverse-phase HPLC, the ESI-MS/MS fragment peaks of AHLs were analyzed. All corresponding peaks for respective C6-HSL (A; m/z 200) and 3-oxo-C6-HSL (B; m/z 214) along with the product ion peaks (m/z 102) are marked by arrows.

Fig. S4. Mass spectra of AHLs extracted from the cell-free supernatant of 508PAI for CsaI. After fractionation by reverse-phase HPLC, the ESI-MS/MS fragment peaks of AHLs were analyzed. All corresponding peaks for respective C6-HSL (A; m/z 200) and 3-oxo-C6-HSL (B; m/z 214) along with the product ion peaks (m/z 102) are marked by arrows.

Strains Description Source Escherichia coli DH5a F^{-} supE44 $\Delta lacU169$ ($\phi 80 \ lacZ\Delta M15$) hsdR17 recA1 Nippon Gene endA1 gyrA96 thi-1 relA1 19 S17-1 λ*pir* thi pro hsdR hsdM+ recA RP4 2Tc::Mu-Km::Tn7 Pseudomonas chlororaphis subsp. aurantiaca StFRB508 Phenazine and AHL producer isolated from potato 11 root surface 508∆PI StFRB508 derivative, $\Delta phzI$ This study 508ΔAI StFRB508 derivative, *\(\Delta aurI\)* This study 508∆CI StFRB508 derivative, $\Delta csaI$ This study 508∆PAI StFRB508 derivative, $\Delta phzI$ and $\Delta aurI$ This study 508∆ACI StFRB508 derivative, $\Delta aurI$ and $\Delta csaI$ This study 508∆PCI StFRB508 derivative, $\Delta phzI$ and $\Delta csaI$ This study 508∆PACI StFRB508 derivative, $\Delta phzI$, $\Delta aurI$, and $\Delta csaI$ This study StFRB508 derivative, $\Delta phzR$ This study 508∆PR 508∆AR StFRB508 derivative, $\Delta aurR$ This study 508∆CR StFRB508 derivative, $\Delta csaR$ This study $508 \Delta PAR$ StFRB508 derivative, $\Delta phzR$ and $\Delta aurR$ This study 508∆ACR StFRB508 derivative, $\Delta aurR$ and $\Delta csaR$ This study StFRB508 derivative, $\Delta phzR$ and $\Delta csaR$ $508 \Delta PCR$ This study 508∆PACR StFRB508 derivative, $\Delta phzR$, $\Delta aurR$, and $\Delta csaR$ This study 508∆PACR StFRB508 derivative, $\Delta phzR$, $\Delta aurR$, and $\Delta csaR$ This study 508∆PZ StFRB508 derivative, phzCDE::Kmr 11

Table S1. Bacterial strains used in the study

Description	Primers	Primer sequences (5'-3')*
Amplification of <i>phzIR</i>	phzIR-F	GCACTGGCGGCTACAGGAGGGAATTGGATAAACGG
	phzIR-R	GATCATTAAAGCCGCTAGGGGAAAGCGAAGCAGGC
Deletion of <i>phzI</i> internal sequence	phzIdel-F	TCT <u>AAGCTT</u> CCGACACATTATTGAAAGAGACAGC
	phzIdel-R	TCT <u>AAGCTT</u> CATCGCTCATTTCGTTCAGTGTGTGC
Deletion of <i>phzR</i> internal	phzRdel-F	TCT <u>AAGCTT</u> CGACAGCCGTAAACTCCTGCATATCC
sequence	phzRdel-R	TCT <u>AAGCTT</u> GGCAGTGTCTTACGCAGTCGCGATGG
Amplification of <i>aurIR</i>	aurIR-F	CGAGCAGATGGCGCCATAACCGGTTACAACTACCC
	aurIR-R	CAAGGTCTGGCAGGCGCTGCTGACCATTCCTTTCG
Deletion of <i>aurI</i> internal sequence	aurIdel-F	TCT <u>AAGCTT</u> GTGCTGTTGCTGAACATGCCGATCAG
	aurIdel-R	TCT <u>AAGCTT</u> GGCGTCGCACTATAGTCGAGCGTGTG
Deletion of <i>aurR</i> internal	aurRdel-F	TCT <u>AAGCTT</u> GCCAGAGTATTTGAGTCGACTATCC
sequence	aurRdel-R	TCT <u>AAGCTT</u> CATTTCCGGAAAGCTGCAGGTCTGC
Amplification of <i>csaIR</i>	csaIR-F	GCGCTTTCATATCGAACAGGCGCTCAGGCTCAACC
	csaIR-R	CCACCTGCTGATGGTGGCGATGGTGCTGTTCTTCG
Deletion of <i>csaI</i> internal sequence	csaIdel-F	TCT <u>AAGCTT</u> GCTTCGCTGAATCGGGTATTTCTGCG
	csaIdel-R	TCT <u>AAGCTT</u> GTGTCGGTGAAAGCTGGCTTTCATGC
Deletion of <i>csaR</i> internal sequence	csaRdel-F	TCTAAGCTTCTTCCACCTCAAGGTCATCCAGAAG
	csaRdel-R	TCT <u>AAGCTT</u> GGGAATCTAGTCCAGGGCAATACGC

 Table S2. Primers used in the study

**Hin*dIII sites are underlined.