

**Table S1A**

<i>Vibrio</i> spp. strains	NCBI reference sequence	Integrase gene position	Integrase gene length (bps)
<i>Aliivibrio salmonicida</i> LFI1238	NC_011312 NC_011313	593499-594482	984_reverse
<i>Vibrio alginolitycus</i> ATCC 17749	NC_022349 NC_022359	1009839-1010801	963_forward
<i>Vibrio cholerae</i> El Tor strain N16961	NC_002505 NC_002506	309750-310712	963_reverse
<i>Vibrio fischerii</i> MJ11	NC_011184 NC_011186	817245-818228	984_forward
<i>Vibrio parahaemolyticus</i> RIMD 2210633	NC_004603 NC_004605	1948281-1949243	963_forward
<i>Vibrio rotiferianus</i> DAT722	NZ_AFAJ00000000 NZ_AJZL00000000	2363443-2364405	936_reverse
<i>Vibrio vulnificus</i> CMCP6	NC_004459 NC_004460	2438314-2439276	963_reverse

**Table S1B**

Basic strains*		
Strain number	Relevant <i>Escherichia coli</i> genotypes and description	References
ω9420	<b>Π1</b> DH5α ΔthyA::erm-pir116) [Erm <sup>R</sup> ]	Demarre <i>et al</i> , 2005
ω8486	<b>MG1655</b> ΔdapA recA269::tn10 [Tc <sup>R</sup> ]	Loot <i>et al</i> , 2014
ω8488	ω8486 transformed with p3153 [Tc <sup>R</sup> , Carb <sup>R</sup> ]	Loot <i>et al</i> , 2014

Strain number	Relevant <i>Vibrio cholerae</i> genotype and description	References
ω8637	<b>N16961</b> [Str <sup>R</sup> ]	Heidelberg <i>et al</i> , 2000

Derivative MG1655ΔdapA recA269::tn10 strains*,**			
Cassette description	Plasmid number	Strain number (inserted plasmid)	Strain number (inserted plasmid+ pBADintII)
attC <sub>aadA7</sub> -60bp-attC <sub>ereA2</sub> (attP+)	pB322	C188	C448
attC <sub>aadA7</sub> -60+ <b>400bplacZ</b> -attC <sub>ereA2</sub> (attP+)	pB429	C189	C316
attC <sub>aadA7</sub> -60+ <b>750bplacZ</b> -attC <sub>ereA2</sub> (attP+)	pB254	D004	D066
attC <sub>aadA7</sub> -60+ <b>1500bplacZ</b> -attC <sub>ereA2</sub> (attP+)	pB255	B552	C480
attC <sub>aadA7</sub> -60+ <b>3000bplacZ</b> -attC <sub>ereA2</sub> (attP+)	pB321	E109	E156
attC <sub>aadA7</sub> -60bp-attC <sub>ereA2</sub> (attP-)	pB256	C186	C318
attC <sub>aadA7</sub> -60+ <b>400bplacZ</b> -attC <sub>ereA2</sub> (attP-)	pB428	B548	C319
attC <sub>aadA7</sub> -60+ <b>750bplacZ</b> -attC <sub>ereA2</sub> (attP-)	pB257	D005	D067
attC <sub>aadA7</sub> -60+ <b>1500bplacZ</b> -attC <sub>ereA2</sub> (attP-)	pB253	E108	E154
attC <sub>aadA7</sub> -60+ <b>3000bplacZ</b> -attC <sub>ereA2</sub> (attP-)	pB252	B550	C437
attC <sub>aadA7</sub> -60+ <b>750bphubP</b> -attC <sub>ereA2</sub> (attP+)	pE511	E906	F299
attC <sub>aadA7</sub> -60+ <b>1500bphubP</b> -attC <sub>ereA2</sub> (attP+)	pE512	F052	F300
attC <sub>aadA7</sub> -60+ <b>3000bphubP</b> -attC <sub>ereA2</sub> (attP+)	pE513	E908	F301
attC <sub>aadA7</sub> -60bp-VCR <sub>126</sub> (attP+)	pA621	D098	D115

$attC_{aadA7-60+400bplacZ-VCR_{126}}$ (attP+)	pB332	E288	E297
$attC_{aadA7-60+750bplacZ-VCR_{126}}$ (attP+)	pA696	E291	E300
$attC_{aadA7-60+1500bplacZ-VCR_{126}}$ (attP+)	pA703	B547	C449
$attC_{aadA7-60+3000bplacZ-VCR_{126}}$ (attP+)	pA705	C185	C310
$attC_{aadA7-60bp-VCR_{126}}$ (attP-)	pA672	E189	E310
$attC_{aadA7-60+400bplacZ-VCR_{126}}$ (attP-)	pB331	B541	C312
$attC_{aadA7-60+750bplacZ-VCR_{126}}$ (attP-)	pA669	E195	E316
$attC_{aadA7-60+1500bplacZ-VCR_{126}}$ (attP-)	pA675	E187	E308
$attC_{aadA7-60+3000bplacZ-VCR_{126}}$ (attP-)	pA670	B544	C315
$attC_{aadA7-60+750bphubP-VCR_{126}}$ (attP+)	pE507	E902	F302
$attC_{aadA7-60+1500bphubP-VCR_{126}}$ (attP+)	pE508	E903	F303
$attC_{aadA7-60+3000bphubP-VCR_{126}}$ (attP+)	pE509	E904	F304
$attC_{aadA7-60+750bphubP-VCR_{126}}$ (attP-)	pE503	E898	F305
$attC_{aadA7-60+1500bphubP-VCR_{126}}$ (attP-)	pE504	E899	F306
$attC_{aadA7-60+3000bphubP-VCR_{126}}$ (attP-)	pE505	F045	F307
$VCR_{16^*-60bp-VCR_{64}}$ (attP+)	pA702	D087	D107
$VCR_{16^*-60+400bplacZ-VCR_{64}}$ (attP+)	pB328	D090	D110
$VCR_{16^*-60+750bplacZ-VCR_{64}}$ (attP+)	pA717	D088	D108
$VCR_{16^*-60+1500bplacZ-VCR_{64}}$ (attP+)	pA701	D094	D114
$VCR_{16^*-60+3000bplacZ-VCR_{64}}$ (attP+)	pA697	D093	D113
$VCR_{16^*-60bp-VCR_{64}}$ (attP-)	pA671	D086	D106
$VCR_{16^*-60+400bplacZ-VCR_{64}}$ (attP-)	pB327	D089	D109
$VCR_{16^*-60+750bplacZ-VCR_{64}}$ (attP-)	pA667	D085	D105

VCR <sub>16</sub> *-60+ <b>1500bplacZ</b> -VCR <sub>64</sub> (attP-)	pA673	D092	D112
VCR <sub>16</sub> *-60+ <b>3000bplacZ</b> -VCR <sub>64</sub> (attP-)	pA663	D091	D111
VCR <sub>126</sub> *-60bp-VCR <sub>126</sub> ** (attP+)	pE894	F059	F294
VCR <sub>126</sub> *-60+ <b>400bplacZ</b> -VCR <sub>126</sub> ** (attP+)	pE978	F060	F295
VCR <sub>126</sub> *-60+ <b>750bplacZ</b> -VCR <sub>126</sub> ** (attP+)	pF308	F155	F296
VCR <sub>126</sub> *-60+ <b>1500bplacZ</b> -VCR <sub>126</sub> ** (attP+)	pE980	F062	F297
VCR <sub>126</sub> *-60+ <b>3000bplacZ</b> -VCR <sub>126</sub> ** (attP+)	pE922	F063	F298
VCR <sub>126</sub> *-60bp-VCR <sub>126</sub> ** (attP-)	pE893	F054	F289
VCR <sub>126</sub> *-60+ <b>400bplacZ</b> -VCR <sub>126</sub> ** (attP-)	pE975	F055	F290
VCR <sub>126</sub> *-60+ <b>750bplacZ</b> -VCR <sub>126</sub> ** (attP-)	pF309	F156	F291
VCR <sub>126</sub> *-60+ <b>1500bplacZ</b> -VCR <sub>126</sub> ** (attP-)	pE977	F057	F292
VCR <sub>126</sub> *-60+ <b>3000bplacZ</b> -VCR <sub>126</sub> ** (attP-)	pE921	F058	F293

\**Escherichia coli* and *Vibrio cholerae* strains were grown in Luria Bertani (LB) broth at 37°C or 30°C (when specified). Antibiotics were used at the following concentrations: carbenicillin (Carb), 100 µg/ml, spectinomycin (Sp), 40 µg/ml, erythromycin (Erm), 200 µg/ml, tetracyclin (Tet), 15 µg/ml, streptomycin (Str), 200 µg/ml, kanamycin (Kan), 25 µg/ml, Thymidine (Thy) and diaminopimelic acid (DAP) were supplemented when necessary to a final concentration of 0.3mM. To induce the P<sub>bad</sub> promoter, L-arabinose (Ara) was added to a final concentration of 2mg/ml; to repress it, glucose (Glc) was added to a final concentration of 10mg/ml. To induce the P<sub>lac</sub> promoter, isopropyl-β-D-thiogalactopyranoside (IPTG) was added to a final concentration of 200µg/ml.

\*\*All the constructed pSW derivative plasmids were inserted into the λ attB site of the ω8488

chromosome through lambda recombination mediated by plasmid pTSA29-CXI (p3153, Carb<sup>R</sup>, Valens & al., 2004). By inverting the *attP* site on the pSW plasmid, we inserted plasmids in both orientations: attP- corresponding to the bottom strand of *attC* sites carried by the lagging strand template, and attP+ to the bottom strand carried by the leading strand template. We confirmed plasmid insertions by performing PCR reactions with o1897/o1898 (presence of a monomeric inserted plasmid) and o361/o3725 (plasmid insertion in *attB* site) primers. We performed all the experiments in *recA* deleted strains to avoid homologous recombination between the *lacZ* fragment in the synthetic cassettes and the endogenous *lacZ* gene, as well as between *attC* sites having high degree of identity (i.e: VCR sites).

**Table S1C**

Plasmids used in this study*		
Plasmid name and description	Plasmid number	References
pBADIntI1, ori <sub>ColE1</sub> , [Carb <sup>R</sup> ]	p3938	Demarre <i>et al</i> , 2007
pTSA29-CXI, ori <sub>pSC101</sub> , [Carb <sup>R</sup> ]	p3153	Valens <i>et al</i> , 2004
pSW23T, ori <sub>R6K</sub> , [Sp <sup>R</sup> ]	p7988	Laboratory collection
pSU38Δ:: <i>lacZ</i> :: <i>attC<subaada7< sub=""></subaada7<></i> - <i>catT4t</i> -VCR <sub>2</sub> , ori <sub>p15A</sub> , [Kan] <sup>R</sup>	p1372	Demarre <i>et al</i> , 2007
pRS551, ori <sub>pMB1</sub> , [Carb <sup>R</sup> , Kan <sup>R</sup> ]	p923	Simons <i>et al</i> , 1987
pSW <i>attI</i> - <i>attC<subaada7< sub=""></subaada7<></i> - <i>dapA</i> , [Cm <sup>R</sup> ]	p7923	Baharoglu <i>et al</i> , 2010
pMA-VCR <sub>126*</sub> -term, ori <sub>ColE1</sub> , [Carb <sup>R</sup> ]	p8208	VCR <sub>126*</sub> -term GeneArt fragment (see Table III) in pMA plasmid

pTOPO-VCR <sub>16*</sub> -VCR <sub>64</sub> , ori <sub>PUC</sub> , [Kan <sup>R</sup> ]	pE884	<b>VCR<sub>16*</sub>-VCR<sub>64</sub> gBlocks® Gene Fragment from Integrated DNA Technologies (see Table III) in pCR™-Blunt II-TOPO® (Invitrogen)</b>
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<b>Derivative pSW plasmids constructed in this study</b>		
<b>Plasmid name and description</b>	<b>Plasmid number</b>	<b>Relevant properties and construction</b>
pSW23T, ori <sub>R6K</sub> , [Sp <sup>R</sup> ]		
pSWattC <sub>aadA7</sub> -VCR <sub>126</sub> (attP+)	p8146	<b>Plac-attC<sub>aadA7</sub></b> fragment (amplification of p1372 using o299 and o845 followed by EcoRI/BamHI digestion) and <b>VCR<sub>126</sub></b> fragment (annealing between o876, o877, o878 and o879) in EcoRI/XmaI digested p7988
pSWattC <sub>aadA7</sub> -VCR <sub>126**</sub> (attP+)	p8159	<b>attC<sub>aadA7</sub></b> fragment (p8146 digested by EcoRI/BglII) and <b>VCR<sub>126**</sub></b> fragment (annealing between o885 and o886) in EcoRI/XmaI digested p7988
pSWattC <sub>aadA7</sub> -60bp-VCR <sub>126</sub> (attP+)	p8161	<b>Terminator</b> fragment (annealing between o883 and o884) in BamHI digested p8146
pSWVCR <sub>126*</sub> -60bp-VCR <sub>126**</sub> (attP+)	p8241	<b>VCR<sub>126*</sub>-term</b> fragment (XbaI/BglIII digested p8208) in XbaI/BamHII digested p8159
pSWattC <sub>aadA7</sub> -60bp-VCR <sub>126</sub> (attP-)	p8257	Inversion of the <b>attP</b> region of p8161 by SacI

		digestion
pSWVCR <sub>126*</sub> -60bp-VCR <sub>126**</sub> (attP-)	p8256	Inversion of the <b>attP</b> region of p8241 by SacI digestion
pSWattC <sub>aadA7</sub> -60+ <b>3000bplacZ</b> -VCR <sub>126</sub> (attP+)	p8238	<b>3000bplacZ</b> fragment (PCR fragment (o863 and o864) from p923)) in BamHI digested p8161
pSWVCR <sub>126*</sub> -60+ <b>3000bplacZ</b> -VCR <sub>126**</sub> (attP+)	p8265	<b>3000bplacZ</b> fragment in BamHI digested p8241
pSWattC <sub>aadA7</sub> -60+ <b>3000bplacZ</b> -VCR <sub>126</sub> (attP-)	p8300	<b>3000bplacZ</b> fragment in BamHI digested p8257
pSWVCR <sub>126*</sub> -60+ <b>3000bplacZ</b> -VCR <sub>126**</sub> (attP-)	p8298	<b>3000bplacZ</b> fragment in BamHI digested p8256
pSWattC <sub>aadA7</sub> -60+ <b>750bplacZ</b> -VCR <sub>126</sub> (attP+)	p8255	<b>750bplacZ</b> fragment (PCR fragment (o863 and o868) from p923)) in BamHI/NheI digested p8238
pSWVCR <sub>126*</sub> -60+ <b>750bplacZ</b> -VCR <sub>126**</sub> (attP+)	p8338	<b>750bplacZ</b> fragment in BamHI/NheI digested p8265
pSWattC <sub>aadA7</sub> -60+ <b>750bplacZ</b> -VCR <sub>126</sub> (attP-)	p8341	<b>750bplacZ</b> fragment in BamHI/NheI digested p8300
pSWVCR <sub>126*</sub> -60+ <b>750bplacZ</b> -VCR <sub>126**</sub> (attP-)	p8339	<b>750bplacZ</b> fragment in BamHI/NheI digested p8298
pSWattC <sub>aadA7</sub> -60bp-VCR <sub>126</sub> (attP+)	p9935	<b>dapA</b> fragment (PCR fragment (o1069 and 1070) from p7923) in SmaI/PstI digested p8161
pSWVCR <sub>126*</sub> -60bp-VCR <sub>126**</sub> (attP+)	p9936	<b>dapA</b> fragment in SmaI/PstI digested p8241

pSWattC <sub>aadA7</sub> -60bp-VCR <sub>126</sub> (attP-)	p9938	<i>dapA</i> fragment in SmaI/PstI digested p8257
pSWVCR <sub>126*</sub> -60bp-VCR <sub>126**</sub> (attP-)	p9937	<i>dapA</i> fragment in SmaI/PstI digested p8256
pSWattC <sub>aadA7</sub> -60+ <b>3000bp</b> <i>lacZ</i> -VCR <sub>126</sub> (attP+)	p8456	<i>dapA</i> fragment in SmaI/PstI digested p8238
pSWVCR <sub>126*</sub> -60+ <b>3000bp</b> <i>lacZ</i> -VCR <sub>126**</sub> (attP+)	p8459	<i>dapA</i> fragment in SmaI/PstI digested p8265
pSWattC <sub>aadA7</sub> -60+ <b>3000bp</b> <i>lacZ</i> -VCR <sub>126</sub> (attP-)	p8469	<i>dapA</i> fragment in SmaI/PstI digested p8300
pSWVCR <sub>126*</sub> -60+ <b>3000bp</b> <i>lacZ</i> -VCR <sub>126**</sub> (attP-)	p8467	<i>dapA</i> fragment in SmaI/PstI digested p8298
pSWattC <sub>aadA7</sub> -60+ <b>750bp</b> <i>lacZ</i> -VCR <sub>126</sub> (attP+)	p8458	<i>dapA</i> fragment in SmaI/PstI digested p8255
pSWVCR <sub>126*</sub> -60+ <b>750bp</b> <i>lacZ</i> -VCR <sub>126**</sub> (attP+)	p8460	<i>dapA</i> fragment in SmaI/PstI digested p8338
pSWattC <sub>aadA7</sub> -60+ <b>750bp</b> <i>lacZ</i> -VCR <sub>126</sub> (attP-)	p8463	<i>dapA</i> fragment in SmaI/PstI digested p8341
pSWVCR <sub>126*</sub> -60+ <b>750bp</b> <i>lacZ</i> -VCR <sub>126**</sub> (attP-)	p8461	<i>dapA</i> fragment in SmaI/PstI digested p8339
pSWattC <sub>aadA7</sub> -60+ <b>1500bp</b> <i>lacZ</i> -VCR <sub>126</sub> (attP+)	pA350	<b>1500bp</b> <i>lacZ</i> fragment (PCR fragment (o863 and o866) from p923)) in BamHI/NheI digested p8458
pSWVCR <sub>126*</sub> -60+ <b>1500bp</b> <i>lacZ</i> -VCR <sub>126**</sub> (attP+)	pA351	<b>1500bp</b> <i>lacZ</i> fragment in BamHI/NheI digested p8460

pSWattC <sub>aadA7</sub> -60+ <b>1500bplacZ</b> -VCR <sub>126</sub> (attP-)	pA354	<b>1500bplacZ</b> fragment in BamHI/NheI digested p8463
pSWVCR <sub>126*</sub> -60+ <b>1500bplacZ</b> -VCR <sub>126**</sub> (attP-)	pA352	<b>1500bplacZ</b> fragment in BamHI/NheI digested p8461
pSWattC <sub>aadA7</sub> -60bp-VCR <sub>126</sub> (attP+)	pA621	<b>FRT</b> fragment (annealing between o2227 and o2228) in NotI/EcoRI digested p9935
pSWVCR <sub>126*</sub> -60bp-VCR <sub>126**</sub> (attP+)	pA702	<b>FRT</b> fragment in NotI/EcoRI digested p9936
pSWattC <sub>aadA7</sub> -60bp-VCR <sub>126</sub> (attP-)	pA672	<b>FRT</b> fragment in NotI/EcoRI digested p9938
pSWVCR <sub>126*</sub> -60bp-VCR <sub>126**</sub> (attP-)	pA671	<b>FRT</b> fragment in NotI/EcoRI digested p9937
pSWattC <sub>aadA7</sub> -60+ <b>3000bplacZ</b> -VCR <sub>126</sub> (attP+)	pA705	<b>FRT</b> fragment in NotI/EcoRI digested p8456
pSWVCR <sub>126*</sub> -60+ <b>3000bplacZ</b> -VCR <sub>126**</sub> (attP+)	pA697	<b>FRT</b> fragment in NotI/EcoRI digested p8459
pSWattC <sub>aadA7</sub> -60+ <b>3000bplacZ</b> -VCR <sub>126</sub> (attP-)	pA670	<b>FRT</b> fragment in NotI/EcoRI digested p8469
pSWVCR <sub>126*</sub> -60+ <b>3000bplacZ</b> -VCR <sub>126**</sub> (attP-)	pA663	<b>FRT</b> fragment in NotI/EcoRI digested p8467
pSWattC <sub>aadA7</sub> -60+ <b>750bplacZ</b> -VCR <sub>126</sub> (attP+)	pA696	<b>FRT</b> fragment in NotI/EcoRI digested p8458
pSWVCR <sub>126*</sub> -60+ <b>750bplacZ</b> -VCR <sub>126**</sub> (attP+)	pA717	<b>FRT</b> fragment in NotI/EcoRI digested p8460
pSWattC <sub>aadA7</sub> -60+ <b>750bplacZ</b> -VCR <sub>126</sub> (attP-)	pA669	<b>FRT</b> fragment in NotI/EcoRI digested p8463

pSWVCR <sub>126*</sub> -60+ <b>750bp</b> <i>lacZ</i> -VCR <sub>126**</sub> (attP-)	pA667	<b>FRT</b> fragment in NotI/EcoRI digested p8461
pSWattC <sub>aadA7</sub> -60+ <b>1500bp</b> <i>lacZ</i> -VCR <sub>126</sub> (attP+)	pA703	<b>FRT</b> fragment in NotI/EcoRI digested pA350
pSWVCR <sub>126*</sub> -60+ <b>1500bp</b> <i>lacZ</i> -VCR <sub>126**</sub> (attP+)	pA701	<b>FRT</b> fragment in NotI/EcoRI digested pA351
pSWattC <sub>aadA7</sub> -60+ <b>1500bp</b> <i>lacZ</i> -VCR <sub>126</sub> (attP-)	pA675	<b>FRT</b> fragment in NotI/EcoRI digested pA354
pSWVCR <sub>126*</sub> -60+ <b>1500bp</b> <i>lacZ</i> -VCR <sub>126**</sub> (attP-)	pA673	<b>FRT</b> fragment in NotI/EcoRI digested pA352
pSWattC <sub>aadA7</sub> -60bp- <i>attCereA2</i> (attP+)	pB322	<i>attCereA2</i> fragment (annealing between o2351, o2352, o2353 and o2354) in BamHI/SmaI digested pA621
pSWattC <sub>aadA7</sub> -60bp- <i>attCereA2</i> (attP-)	pB256	<i>attCereA2</i> fragment in BamHI/SmaI digested pA672
pSWattC <sub>aadA7</sub> -60+ <b>3000bp</b> <i>lacZ</i> - <i>attCereA2</i> (attP+)	pB321	<i>attCereA2</i> fragment in BamHI/SmaI digested pA705
pSWattC <sub>aadA7</sub> -60+ <b>3000bp</b> <i>lacZ</i> - <i>attCereA2</i> (attP-)	pB252	<i>attCereA2</i> fragment in BamHI/SmaI digested pA670
pSWattC <sub>aadA7</sub> -60+ <b>750bp</b> <i>lacZ</i> - <i>attCereA2</i> (attP+)	pB254	<i>attCereA2</i> fragment in BamHI/SmaI digested pA696
pSWattC <sub>aadA7</sub> -60+ <b>750bp</b> <i>lacZ</i> - <i>attCereA2</i> (attP-)	pB257	<i>attCereA2</i> fragment in BamHI/SmaI digested pA669

pSWattC <sub>aadA7</sub> -60+ <b>1500bplacZ</b> - attC <sub>ereA2</sub> (attP+)	pB255	attC <sub>ereA2</sub> fragment in BamHI/SmaI digested pA703
pSWattC <sub>aadA7</sub> -60+ <b>1500bplacZ</b> - attC <sub>ereA2</sub> (attP-)	pB253	attC <sub>ereA2</sub> fragment in BamHI/SmaI digested pA675
pSWattC <sub>aadA7</sub> -60+ <b>400bplacZ</b> -VCR <sub>126</sub> (attP+)	pB332	<b>400bplacZ</b> fragment (PCR fragment (o863 and o870) from p923)) in BamHI/NheI digested pA696
pSWVCR <sub>126*</sub> -60+ <b>400bplacZ</b> -VCR <sub>126**</sub> (attP+)	pB328	<b>400bplacZ</b> fragment in BamHI/NheI digested pA717
pSWattC <sub>aadA7</sub> -60+ <b>400bplacZ</b> - attC <sub>ereA2</sub> (attP+)	pB429	<b>400bplacZ</b> fragment in BamHI/SmaI digested pB254
pSWVCR <sub>126*</sub> -60+ <b>400bplacZ</b> -VCR <sub>126**</sub> (attP-)	pB327	<b>400bplacZ</b> fragment in BamHI/NheI digested pA667
pSWattC <sub>aadA7</sub> -60+ <b>400bplacZ</b> -VCR <sub>126</sub> (attP-)	pB331	<b>400bplacZ</b> fragment in BamHI/NheI digested pA669
pSWattC <sub>aadA7</sub> -60+ <b>400bplacZ</b> - attC <sub>ereA2</sub> (attP-)	pB428	<b>400bplacZ</b> fragment in BamHI/SmaI digested pB257
pSWattC <sub>aadA7</sub> -60+ <b>750bphubP</b> - attC <sub>ereA2</sub> (attP+)	pE511	<b>750bphubP</b> fragment (PCR fragment (o3771 and o3773) from N16961 <i>Vibrio cholerae</i> (Heidelberg <i>et al</i> , 2000))) in BamHI/NheI digested pB255
pSWattC <sub>aadA7</sub> -60+ <b>750bphubP</b> - VCR <sub>126</sub> (attP+)	pE507	<b>750bphubP</b> fragment in BamHI/NheI digested pA703

pSWattC <sub>aadA7</sub> -60+ <b>750bp</b> <i>hubP</i> - VCR <sub>126</sub> (attP-)	pE503	<b>750bp</b> <i>hubP</i> fragment in BamHI/NheI digested pA675
pSWattC <sub>aadA7</sub> -60+ <b>1500bp</b> <i>hubP</i> - attC <sub>ereA2</sub> (attP+)	pE512	<b>1500bp</b> <i>hubP</i> fragment (PCR fragment (o3771 and o3774) from N16961 <i>Vibrio cholerae</i> (Heidelberg <i>et al</i> , 2000))) in BamHI/NheI digested pB255
pSWattC <sub>aadA7</sub> -60+ <b>1500bp</b> <i>hubP</i> - VCR <sub>126</sub> (attP+)	pE508	<b>1500bp</b> <i>hubP</i> fragment in BamHI/NheI digested pA703
pSWattC <sub>aadA7</sub> -60+ <b>1500bp</b> <i>hubP</i> - VCR <sub>126</sub> (attP-)	pE504	<b>1500bp</b> <i>hubP</i> fragment in BamHI/NheI digested pA675
pSWattC <sub>aadA7</sub> -60+ <b>3000bp</b> <i>hubP</i> - attC <sub>ereA2</sub> (attP+)	pE513	<b>3000bp</b> <i>hubP</i> fragment (PCR fragment (o3771 and o3830) from N16961 <i>Vibrio cholerae</i> (Heidelberg <i>et al</i> , 2000))) in BamHI/NheI digested pB255
pSWattC <sub>aadA7</sub> -60+ <b>3000bp</b> <i>hubP</i> - VCR <sub>126</sub> (attP+)	pE509	<b>3000bp</b> <i>hubP</i> fragment in BamHI/NheI digested pA703
pSWattC <sub>aadA7</sub> -60+ <b>3000bp</b> <i>hubP</i> - VCR <sub>126</sub> (attP-)	pE505	<b>3000bp</b> <i>hubP</i> fragment in BamHI/NheI digested pA675
pSWVCR <sub>16*</sub> -60-VCR <sub>64</sub> (attP+)	pE894	<b>VCR<sub>16*</sub>-VCR<sub>64</sub></b> fragment (synthetic fragment) in NotI/NruI digested pA717
pSWVCR <sub>16*</sub> -60-VCR <sub>64</sub> (attP-)	pE893	<b>VCR<sub>16*</sub>-VCR<sub>64</sub></b> fragment in NotI/EcoRI digested pA667
pSWVCR <sub>16*</sub> -60+ <b>3000bp</b> <i>lacZ</i> - VCR <sub>64</sub>	pE922	<b>3000bp</b> <i>lacZ</i> fragment in BamHI digested pE894

(attP+)		
pSWVCR <sub>16*</sub> -60+ <b>3000bplacZ-</b> VCR <sub>64</sub> (attP-)	pE921	<b>3000bplacZ</b> fragment in BamHI digested pE893
pSWVCR <sub>16*</sub> -60+ <b>400bplacZ-</b> VCR <sub>64</sub> (attP+)	pE978	<b>400bplacZ</b> fragment in BamHI digested pE922
pSWVCR <sub>16*</sub> -60+ <b>750bplacZ-</b> VCR <sub>64</sub> (attP+)	pE979	<b>750bplacZ</b> fragment in BamHI digested pE922
pSWVCR <sub>16*</sub> -60+ <b>1500bplacZ-</b> VCR <sub>64</sub> (attP+)	pE980	<b>1500bplacZ</b> fragment in BamHI digested pE922
pSWVCR <sub>16*</sub> -60+ <b>400bplacZ-</b> VCR <sub>64</sub> (attP-)	pE975	<b>400bplacZ</b> fragment in BamHI digested pE921
pSWVCR <sub>16*</sub> -60+ <b>750bplacZ-</b> VCR <sub>64</sub> (attP-)	pE976	<b>750bplacZ</b> fragment in BamHI digested pE921
pSWVCR <sub>16*</sub> -60+ <b>1500bplacZ-</b> VCR <sub>64</sub> (attP-)	pE977	<b>1500bplacZ</b> fragment in BamHI digested pE921

\* pSW plasmids were constructed in the ΙΙΙ1 strain growing with Thy supplemented in the medium. We confirmed plasmid constructions by performing PCR reactions and sequencing using o1897 and o1962 primers.

**Table S1D**

<b>Primers used in this study*</b>
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Primer name	Primer number	Sequences
RecReportF	o299	TTTGAATTCGCGGCCGCTTAGAGTGAGCGAACGCAATTAA TG
BamHIattCR	o845	AGTGGATCCCATCTAACGCTTGAATTAGCCGCGC
5'BamHI-VCR-XmaI-1	o876	GATCCCATCTAACAAACGCCTCAAGAGGGACTGTCAACGCGTG GCGTTCCAGTCCCATTGAGC
5'BamHI-VCR-XmaI-2	o877	CGCGGTGGTTGCTGTTGTGTTGAGTTAGTGGTAGTGCG TTGTCAGCCCCTTAGGCGGCGTTATAC
3'BamHI-VCR-XmaI-1	o878	GAAACGCCACGCCTGACAGTCCCTCTGAGGCCTTGTAGA TGG
3'BamHI-VCR-XmaI-2	o879	CCGGGTATAACGCCCGCTAACGGGCTGACAACGCACTACCAC TAAACTCAAACACAACACAGCAACCACCGCGGCTCAATGGGA CTG
5term	o883	GATCCAAAAAAAAACCCGCCCTGACAGGGCGGGTTTTTT TA
3term	o884	GATCTAAAAAAAAACCCGCCCTGTCAGGGCGGGTTTTTT TG
modiVCR2-F	o885	AGGCAGGGCGTTATTAC
modiVCR2-R	o886	CCGGGTAAATAACGCCCGCTAACAG
5'BamHI-LacZ	o863	CGGGGATCCATGACCATGATTACGGATTCACTGG

3'NheI-BgLII-LacZ-3000	o864	GGCAGATCTGCTAGCCCAGGAGTCGTCGCCACCAATCC
3'NheI-LacZ-1500	o866	GGCGCTAGCAATAATATCGGTGGCCGTGGTGTGCG
3'NheI-LacZ-750	o868	GGCGCTAGCCTGCCGCACATCTGAACCTTCAGC
3'NheI-LacZ-400	o870	GGCGCTAGCAGCTTCATCAACATTAAATGTGAGC
XmaI-dapA-F	o1069	TTTCCCAGGTACTTCACGGGAAGTATTGT
PstI-dapA-R	o1070	CGGCTGCAGTTACAGCAAACCGGCATGCT
EcoRI-FRT-NotI	o2227	AATTCGAAGTTCCCTATACTttctagaGAATAGGAACCTCGGAATAGG AACTTCGC
NotI-FRT-EcoRI	o2228	GGCCGCGAAGTTCCCTATTCCGAAGTTCCCTATTCTctagaaaGTATAG GAACCTCG
5BamHIereA2	o2351	GATCCAAAAAAAACCCGCCCTGACAGGGCGGGTTTTTT TAGATCCCATCTAACCC
5ereA2SmaI	o2352	TGCCAATCCACCGGACGGTTTCAACCGCCGGTGATCAGCGCG TTATACCC
3ereA2SmaI	o2353	GGGTATAACCGCGCTGATCACCGGCCGGTTGAAAACCGTCCGGTG GATTGGCAGGTTAGATGGG
3BamHIereA2	o2354	ATCTAAAAAAAACCCGCCCTGTCAGGGCGGGTTTTTTTT

		G
5'-BamHI- <i>hubP</i>	o3771	CGGGGATCCCCAACGACTCCTGCTGCCAGTGGC
3'-750 NheI- <i>hubP</i>	o3773	GGCGCTAGCGAAGCTTCTTCGCTTACGGCGC
3'-1500 NheI- <i>hubP</i>	o3774	GGCGCTAGCGCCAGCTCATCATTGAGTAGCGTATCC
3'-3000 NheI- <i>hubP</i>	o3830	GGCGCTAGCACATTAGAAGGCGTACTTACTGCCATACCCGA GCGGAATCTTATTAAGCTCGGTAAAGGCTTCTCCTCGGCTGG C
Swbeg	o1897	CCGTCACAGGTATTATTGGCG
Swend	o1898	CCTCACTAAAGGGAACAAAAGCTG
DAP-A-R	o1962	GTGGTGCCAACAGAACGATCGC
Verif-lbd-attR	o361	GGCAAGCGCCTCGATTACTGCGATGTTAG
finDAPrev	o3725bis	GGCCTTCGTTTATCTGTTGTTGTCGG

**Synthetic fragments used in this study**

VCR <sub>126*</sub> -term	in p8208	TCTAGAGTGAGCGAACGCAATTAAATGTGAGTTAGCTCACTCATTAGGCAC CCCAGGCTTACACTTATGCTTCCGGCTCGTATGTTGTGGAATTGTGA GCGGATAACAATTTCACACAGGAAACAGCTATGAATGATTACGAGTCGGT TATAACAAACGCCTCAAGAGGGACTGTCAACCGTGGCGTTCCAGTCCA TTGGCCGCGGTGGTTGCTGTTGTTGAGTTAGTGGTAGTGCCTG TCAGCCCCTTAGGCCGGCGTTAGATGGGATCCAAAAAAAACCCCGCCCC GACAGGGCGGGTTTTTTAGATCT
VCR <sub>16*</sub> -VCR <sub>64</sub>	in pE884	GGCCCGCGGCCGCTTCTAGAGTGAGCGAACGCAATTAAATGTGAGTTAGCT CACTCATTAGGCACCCCAGGCTTACACTTATGCTTCCGGCTCGTATGTT GTGTGGAATTGTGAGCGGATAACAATTTCACACAGGAAACAGCTATGAATG ATTACGAGTTCGGTTATAACAAACGGCTCAAGAGGGACTGTCAACCGTGG CGTTTCCAGTCCCAAGAACTGCGCGTTACGGTTGCTGTGTTGAGTTA GTGTTAATGCGTTGCCAGCCCCCTAGCCGGCGTTAGATGGGATCCAAAAA AAAACCCGCCCTGACAGGGCGGGTTTTTTAGATCCCCTAACAAA CGCCTTAAGAGGGACTGCCAACCGTGGCATTCCAGTCCCAATGAGCCGT GGTGGTTACGGTTGTTGTTGAGTTGCTTGTATGCGTTGTCAGCCCC TTAGGCGGCGTTATTACCCGGGTACTTCACGGGAAGTATTGTCGCGACCG CG

\* Sequences are given in 5'→3' direction.