

Supplementary Figures

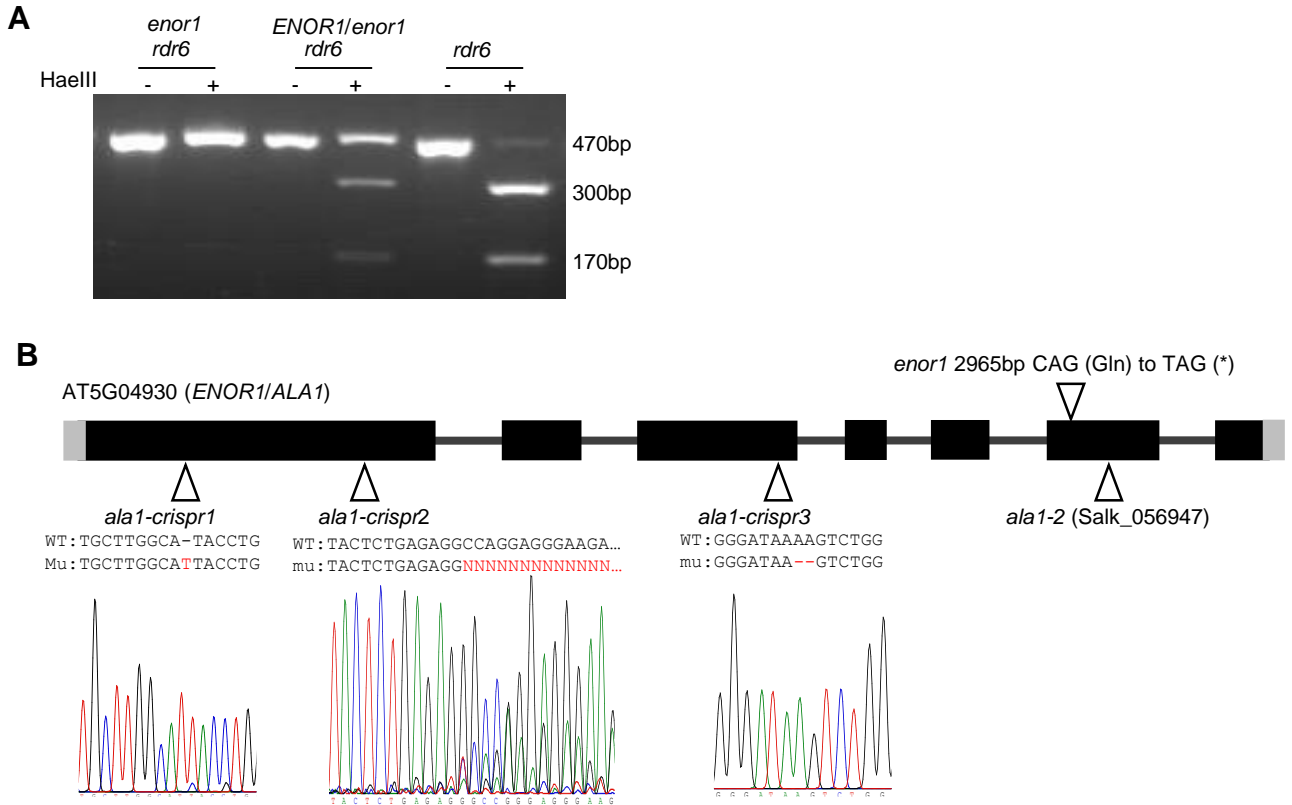


Figure S1. Molecular identification of *ENOR1*.

(A) The nucleotide mutation in *ENOR1* (*ALA1*) of *enor1 rdr6* led to a mutation (GGCC to GGCT) of the recognition sequence of HaeIII. The 470bp PCR product spanning the HaeIII site can be cleaved into fragments of 170bp and 300bp in *rdr6* but not in *enor1 rdr6*.

(B) Schematic diagram shows the structure of *ENOR1/ALA1* (AT5G04930) and the mutations of *ala1* mutants. The positions of mutation sites of *ALA1* in *enor1 rdr6*, *ala1-2* (Salk_056947) and *ala1* mutants (*ala1-crispr1*, *ala1-crispr2*, and *ala1-crispr3*) generated by CRISPR/Cas9 are indicated by open triangles. The *ALA1* gene in *enor1 rdr6* contains a C-to-T mutation at 2965th bp of CDS, converting a Gln codon (CAG) to a premature stop codon (TAG). The *ALA1* gene in *ala1-crispr1* contains a 1-bp insertion, and that in *ala1-crispr3* has a 2-bp deletion. The *ALA* gene in *ala1-crispr2* contains chimeric mutations represented with “NNN...” from the 1120th bp of *ALA1* CDS.

YFP-ALA1 + mCherry-ER-marker

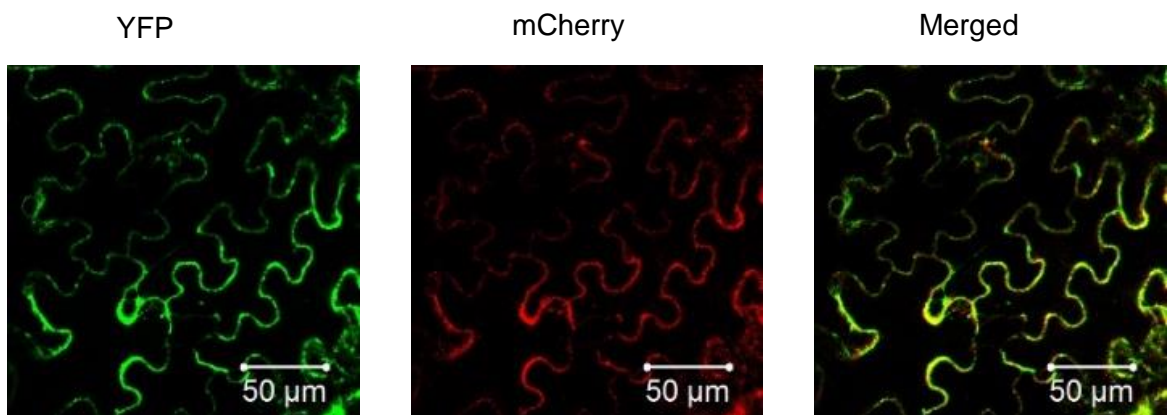


Figure S2. Subcellular localization of ALA1.

ALA1 with fusion of YFP (YFP-ALA1) was co-infiltrated with the mCherry-ER marker in leaves of *Nicotiana benthamiana*, and the fluorescence signals were detected 50h after co-infiltration.

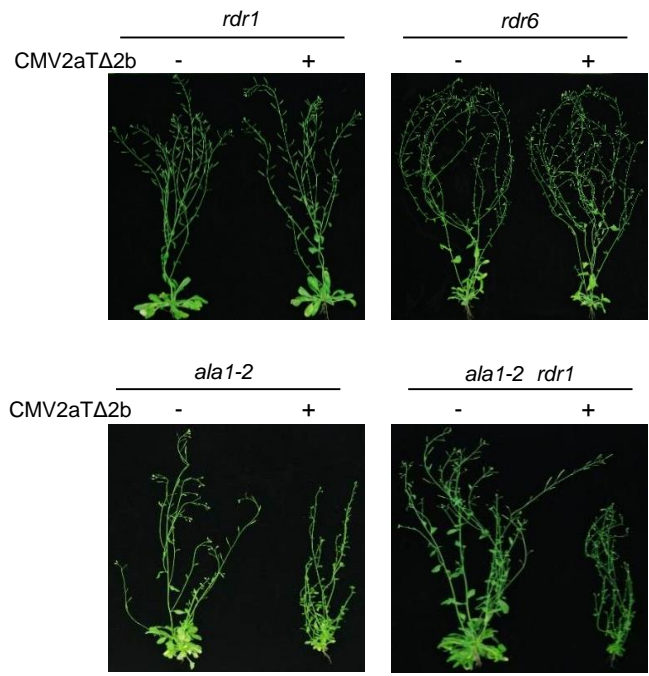


Figure S3. Pathogenic responses of *rdr1*, *rdr6*, *ala1-2* and *ala1-2 rdr1*.

The plants were photographed at 45 d after infection with mock or CMV2aTΔ2b.

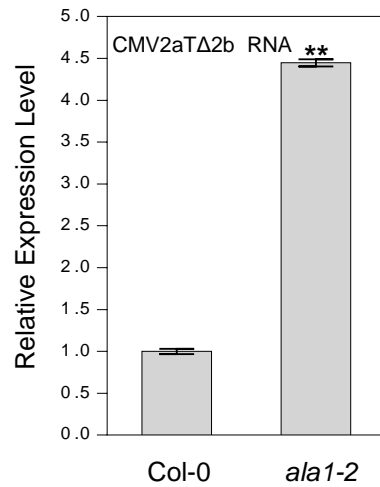
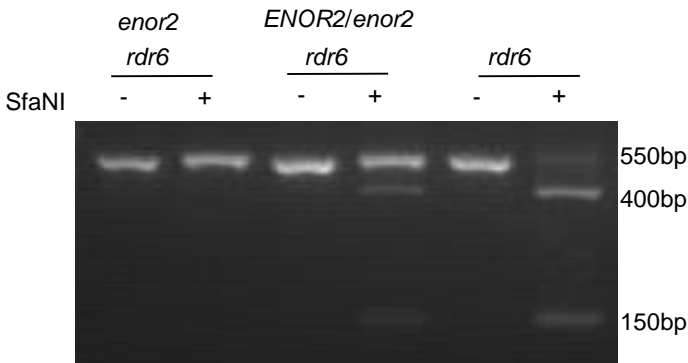
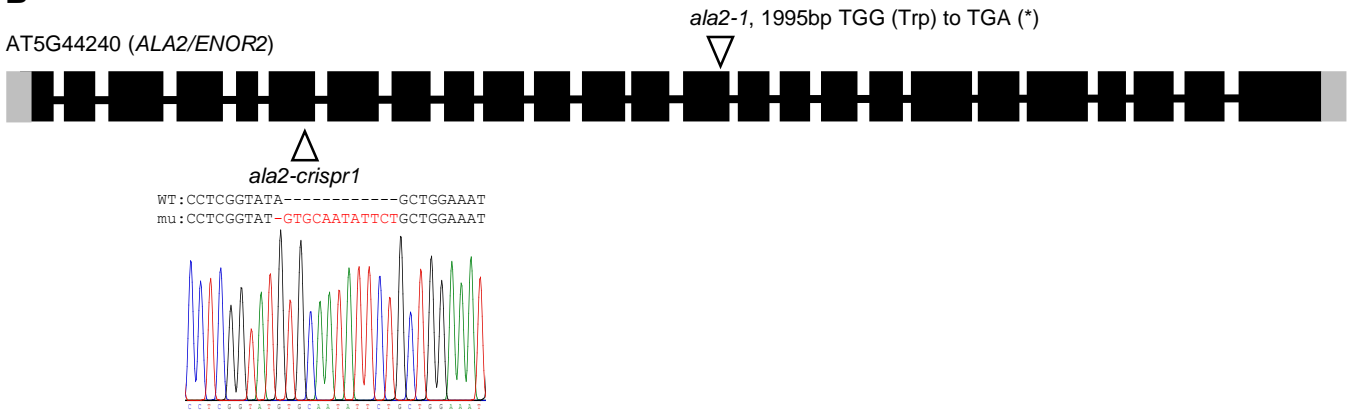


Figure S4. Accumulation of CMV2aTΔ2b genomic RNA in inoculated Col-0 and *ala1-2*. Quantitative real-time PCR analysis of the accumulation of CMV2aTΔ2b RNA in Col-0 and *ala1-2* at 21 d after inoculation. The data are means (\pm SE) from three biological repeats. The asterisks represent significant differences in mutants compared with Col-0 by Student's t-test, ** $P < 0.01$.

A**B****Figure S5. Molecular identification of *ENOR2*.**

(A) The nucleotide mutation in the *ENOR2* (*ALA2*) gene of *enor2 rdr6* led to a mutation (GATGC to AATGC) of the recognition sequence of SfaNI. The 550bp PCR product spanning the SfaNI site can be cleaved into fragments of 150bp and 400bp in *rdr6* but not in *enor2 rdr6*.

(B) Schematic diagram shows the structure of *ENOR2/ALA2* (AT5G44240) and the mutations of *ala2* mutants. The positions of mutation sites of *ALA2* are indicated by open triangles. The *ALA2* gene in *enor2 rdr6* contains a G-to-A mutation at 1995th bp of CDS, converting a Trp codon (TGG) to a premature stop codon (TGA). The *ALA2* gene in *ala2-crispr1* generated by CRISPR/Cas9 contains a 1-bp deletion and a 12-bp insertion.

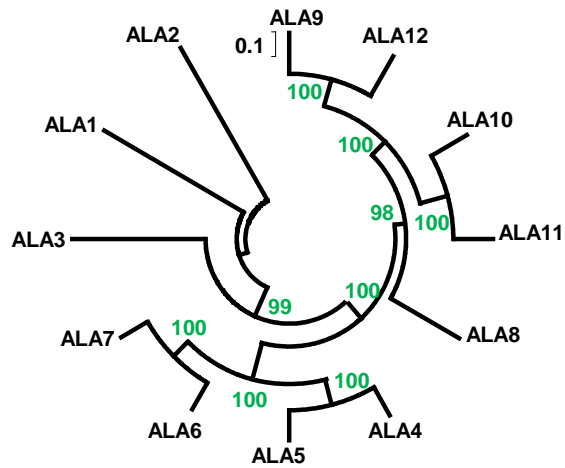


Figure S6. Phylogenetic analysis of Arabidopsis P4-type ATPase.

0.1 stands for distance scale. The numbers in green represent node statistics.

Supplementary Tables

Table S1. List of primers used in this work.

<i>rdr1-1</i> -TDNA	Forward	TGTCACCAGAAGAAGCTCCAG
<i>rdr1-1</i> -TDNA	Reverse	TTCCTTTCGTATAGTTGGGCC
<i>rdr6-15</i> -TDNA	Forward	TCTCCGCATTGCTTCTATTTC
<i>rdr6-15</i> -TDNA	Reverse	GCTGCAAATAAGCACAAAGC
Salk_056947	Forward	GCCATTGGTGATGGTAATGAC
Salk_056947	Reverse	CCCTGTTTAAGAAAAGCGATG
GK317H04-TDNA	Forward	TTTCTGATCGTGTCATTATTCAACTG
GK317H04-TDNA	Reverse	TGTTGTAGAAATCGAATTACCCTG
Salk_125598- TDNA	Forward	GGGGCGTCGTAGAATAAGATC
Salk_125598- TDNA	Reverse	GCTGATGAAGGAGACAAGCAC
Salk_024877- TDNA	Forward	GTACGGAGCTGTCGTTTTAC
Salk_024877- TDNA	Reverse	AGAACTCGAACCCGA ACTCTC
<i>alal-cripsr1-sg</i>	Forward	GGTTTCCTGCTTGGCATACCGTTTTAG AGCTAGAAATAGC
<i>alal-cripsr1-sg</i>	Reverse	GGTATGCCAAGCAGGAAACCAATCAC TACTTCGACTCTAG

<i>ala1-crispr1-sg-cas9</i>	Forward- HindIII	CCCTTAAGCTTCGTTGAACAACGGAA AC
<i>ala1-crispr1-sg-cas9</i>	Reverse-SalI	TATCAAGCTTATCGATACCGTCGAC
<i>ala1-crispr1-seq</i>	Forward	ACGGTTTGAGTTCACTGGGA
<i>ala1-crispr1-seq</i>	Reverse	TGGCCTCTCAGAGTAGTCCT
<i>ala1-crispr2-sg</i>	Forward	GAGGACTACTCTGAGAGGCCGTTTAA GAGCTAGAAATAGC
<i>ala1-crispr2-sg</i>	Reverse	GGCCTCTCAGAGTAGTCCTCAATCACT ACTTCGACTCTAG
<i>ala1-crispr2-sg-cas9</i>	Forward- HindIII	CCCTTAAGCTTCGTTGAACAACGGAA AC
<i>ala1-crispr2-sg-cas9</i>	Reverse-SalI	TATCAAGCTTATCGATACCGTCGAC
<i>ala1-crispr2-seq</i>	Forward	GGGCTTTAGGGGTTGTTGTG
<i>ala1-crispr2-seq</i>	Reverse	CTTCTGTTGCCTTGCCAGTT
<i>ala1-crispr3-sg</i>	Forward	GGATTGCAGGGATAAAAGTCGTTTAA GAGCTAGAAATAGC
<i>ala1-crispr3-sg</i>	Reverse	GACTTTTATCCCTGCAATCCAATCACT ACTTCGACTCTAG
<i>ala1-crispr3-sg-cas9</i>	Forward- HindIII	CCCTTAAGCTTCGTTGAACAACGGAA AC

<i>ala1-crispr3-sg-cas9</i>	Reverse-SalI	TATCAAGCTTATCGATACCGTCGAC
<i>ala1-crispr3-seq</i>	Forward	TACGGTGGCGTCATACATGA
<i>ala1-crispr3-seq</i>	Reverse	ATCCCTACCCCAACATCAGC
<i>ala2-crispr-1-sg</i>	Forward	GTTCTGGTCCTCGGTATAGCGTTTTAG AGCTAGAAATAGC
<i>ala2-crispr-1-sg</i>	Reverse	GCTATACCGAGGACCAGAACAATCAC TACTTCGACTCTAG
<i>ala2-crispr-1-sg-cas9</i>	Forward-HindIII	CCCTTAAGCTTCGTTGAACAACGGAA AC
<i>ala2-crispr-1-sg-cas9</i>	Reverse-SalI	TATCAAGCTTATCGATACCGTCGAC
<i>ala2-crispr1-seq</i>	Forward	GCAAACATGCGCTTATTTCCG
<i>ala2-crispr1-seq</i>	Reverse	GTCTGCCTCTGTCCTGAACT
<i>enor1-caps</i>	Forward	GGACACTGGAATTACCAAAGGATG
<i>enor1-caps</i>	Reverse	GTGAAGATTAACCACCACAACACTGC
<i>enor2-dcaps</i>	Forward	ATTTAAATGTCCAGGACTAGTGAG
<i>enor2-dcaps</i>	Reverse	TCAGAGCAATTTCTAGAGC
<i>ALA1-35SOE-3FLAG</i>	Forward-SmaI	GGGGGGCCCGGGATGGATCCCAGGAA ATCAATTG
<i>ALA1-35SOE-3FLAG</i>	Reverse-XbaI	GGGGGGTCTAGATCATCTCCGTGGAG GATCCTGAATC

<i>ALA1-PJG054-YFP</i>	Forward	CGACGACAAGACCGTGGATCCCAGGA AATCAATTGA
<i>ALA1-PJG054-YFP</i>	Reverse	GAGGAGAAGAGCCGTCATCATCTCCG TGGAGGATCCTGAATC
ALA1-qRT	Forward	ATGTTATACCCACACTCCCTGGT
ALA1-qRT	Reverse	ACATCCATGTCTTGCCCACT
ALA2-qRT	Forward	ACTTGTGGGAGCAATTCAGC
ALA2-qRT	Reverse	CACAGGGGTGATAAGTGACCA
ALA3-qRT	Forward	AGGCACTTGAAAGGACATGG
ALA3-qRT	Reverse	TGTTTGGTTGCTCACACTGAA
ALA4-qRT	Forward	GCCCTTGGAGTCTTTGAACA
ALA4-qRT	Reverse	TGTTGGTACAACGCTGGAAA
ALA5-qRT	Forward	ATGTTTCTTCCGAGATCTGCTT
ALA5-qRT	Reverse	GGTTCTTTGTGCCTTGTTGG
ALA6-qRT	Forward	GTAAAGGCGCTGACAGCAT
ALA6-qRT	Reverse	CCCGCTTCACCATATACATTC
ALA7-qRT	Forward	AGAAAGGGGTGCCTCAATG
ALA7-qRT	Reverse	ACCCTGTCTCAGCAAACCTGC
ALA8-qRT	Forward	AACCGGTTGTTGTTGCTTTC
ALA8-qRT	Reverse	TCCATGTTTCGCAAGTCTTTT
ALA9-qRT	Forward	GTGCGTCTGTCATTTGTTGC
ALA9-qRT	Reverse	TCCAGATTTTACTAGTCTTGTCACCA

ALA10-qRT	Forward	GGGAAGCTCTTGCTGTTGTC
ALA10-qRT	Reverse	ACGACCGTTCTTTGCAAGTC
ALA11-qRT	Forward	CGACGAAGATGGGAGACTTC
ALA11-qRT	Reverse	CCGTTTTTGGCTAGTCTTTCA
ALA12-qRT	Forward	AAAGATTTGGGTTTTGACTGGA
ALA12-qRT	Reverse	TTCATGTCTCGTCTGAGCAAA
ACTIN8-qRT	Forward	TCAGCACTTTCCAGCAGATG
ACTIN8-qRT	Reverse	CTGTGGACAATGCCTGGAC
CMV-RNA-qRT	Forward	GTGAACGGGTTGTCCATCCAGC
CMV-RNA-qRT	Reverse	TGAAACTAGCACGTTGTGCTAG