

Supplemental Figures

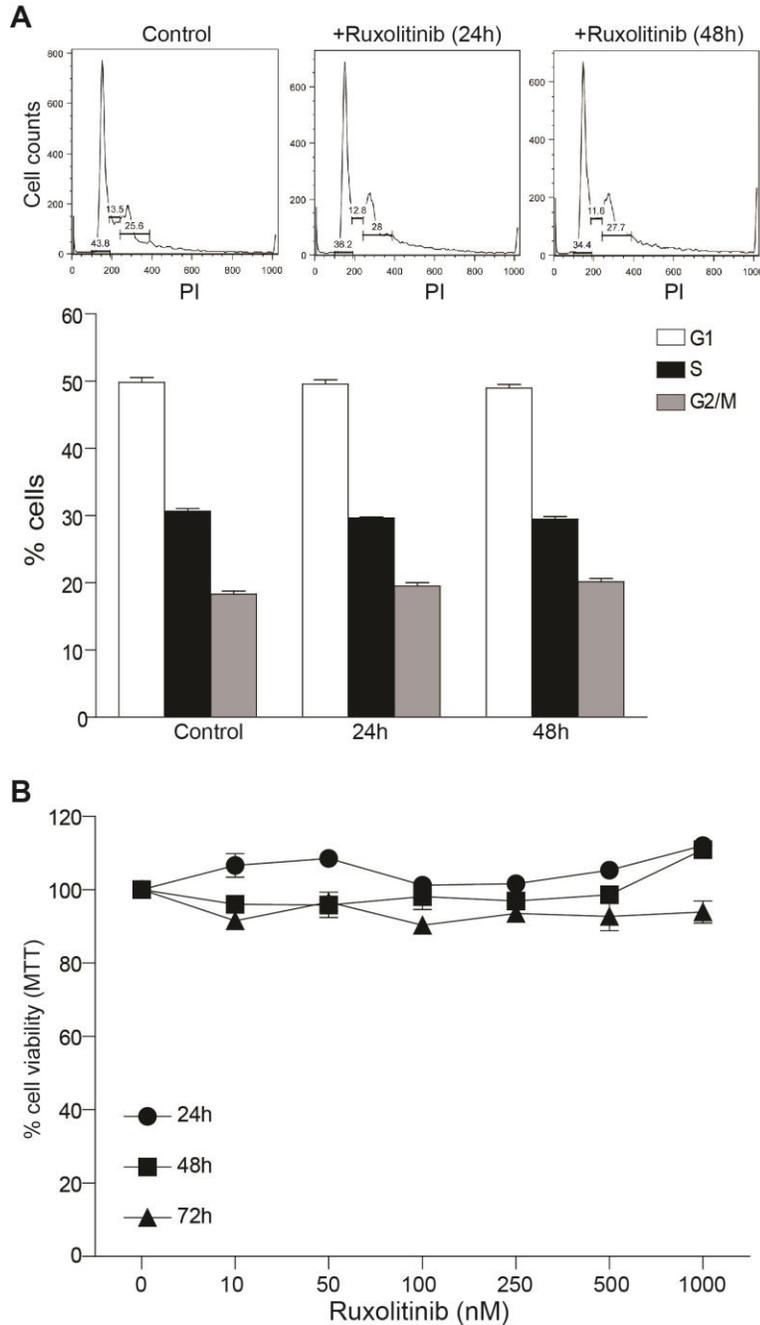


Figure S1. Ruxolitinib treatment does not alter cell cycle and proliferation of tumor cells. A. PANC02-H7 cells were cultured in the presence of ruxolitinib (250 nM). Cells were stained with PI and analyzed by flow cytometry at the indicated time points. The percentages of cells in the various cell cycle phases are indicated. Bottom panel: the percentage of cells in various stages of cell cycles as shown in the top panel is quantified. Column: mean; Bar: SD. B. PANC02-H7 cells were cultured in the presence of ruxolitinib at the indicated doses for the indicated time points and analyzed by MTT assays. The cell viability of untreated cells was set at 100%.

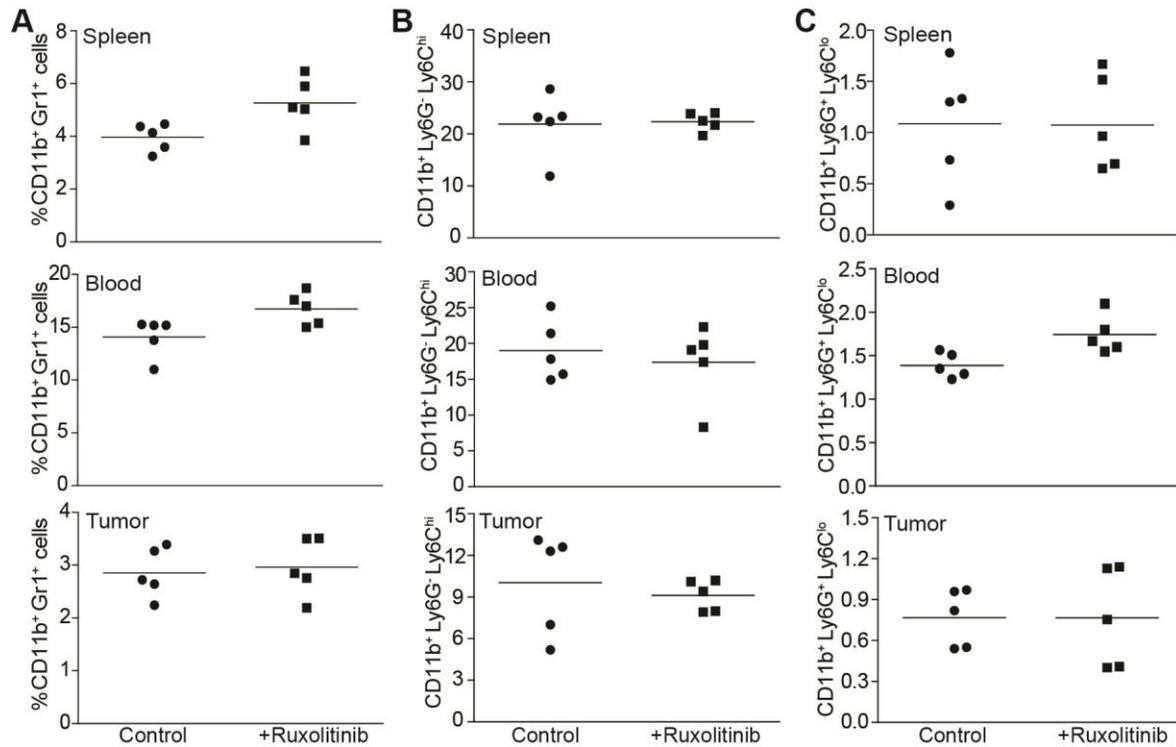


Figure S2. MDSC level in tumor-bearing mice. PANC02-H7 cells were orthotopically transplanted to mice. The tumor-bearing mice were treated daily with solvent (n=5) or ruxolitinib (n=5, 50 mg/kg body weight) for 10 days. Spleen, blood and tumors were collected to make single cells. The cell mixtures were stained with CD11b- and Gr1-specific mAbs (A), CD11b-Ly6G- and Ly6C-specific mAbs (B & C) and analyzed by flow cytometry. A: Quantification of CD11b⁺Gr1⁺ general MDSCs. B: CD11b⁺Ly6G⁺Ly6C^{hi} M-MDSCs. C: Quantification of CD11b⁺Ly6G⁺Ly6C^{lo} PMN-MDSCs. Each dot represents % MDSCs of one mouse.

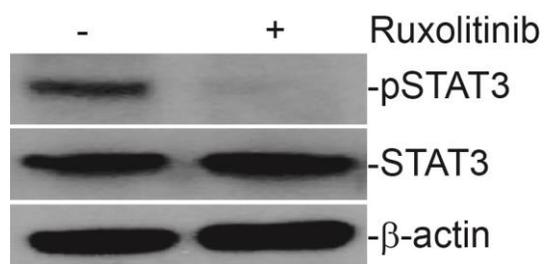


Figure S3. Ruxolitinib inhibits STAT3 activation in tumor-infiltrating CTLs. CD8⁺ T cells were purified from tumor tissues from control (n=5) and Ruxolitinib-treated (n=5) mice. The purified cells from the five control and five Ruxolitinib-treated mice were pooled and lysed for total protein, and analyzed by Western blotting using the indicated antibodies. β -actin is used as normalization control.

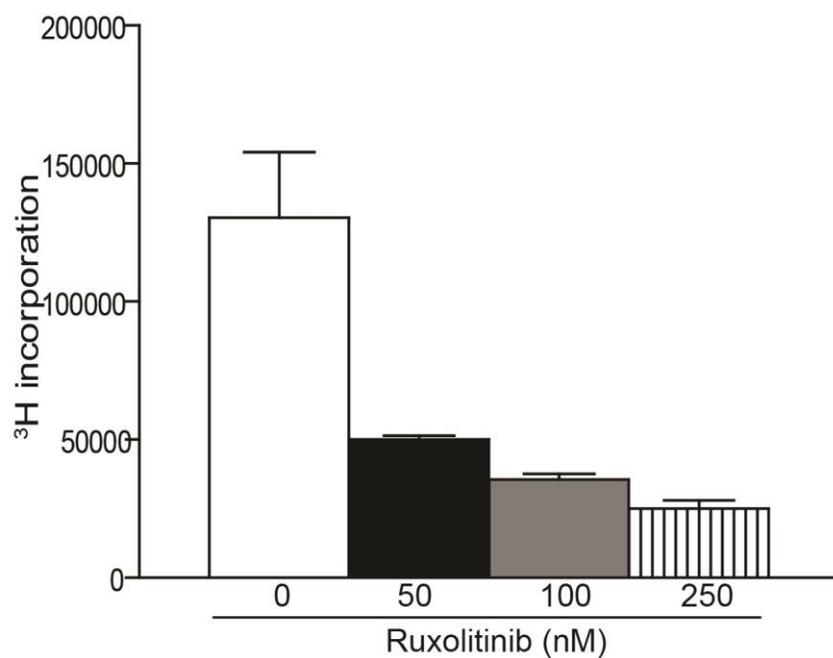


Figure S4. Ruxolitinib inhibits T cell proliferation in vitro. Purified CD3⁺ T cells were cultured in anti-CD3/CD28-coated plates in the presence of ruxolitinib for 3 days. T cell proliferation was measured by ³H-thymidine incorporation.

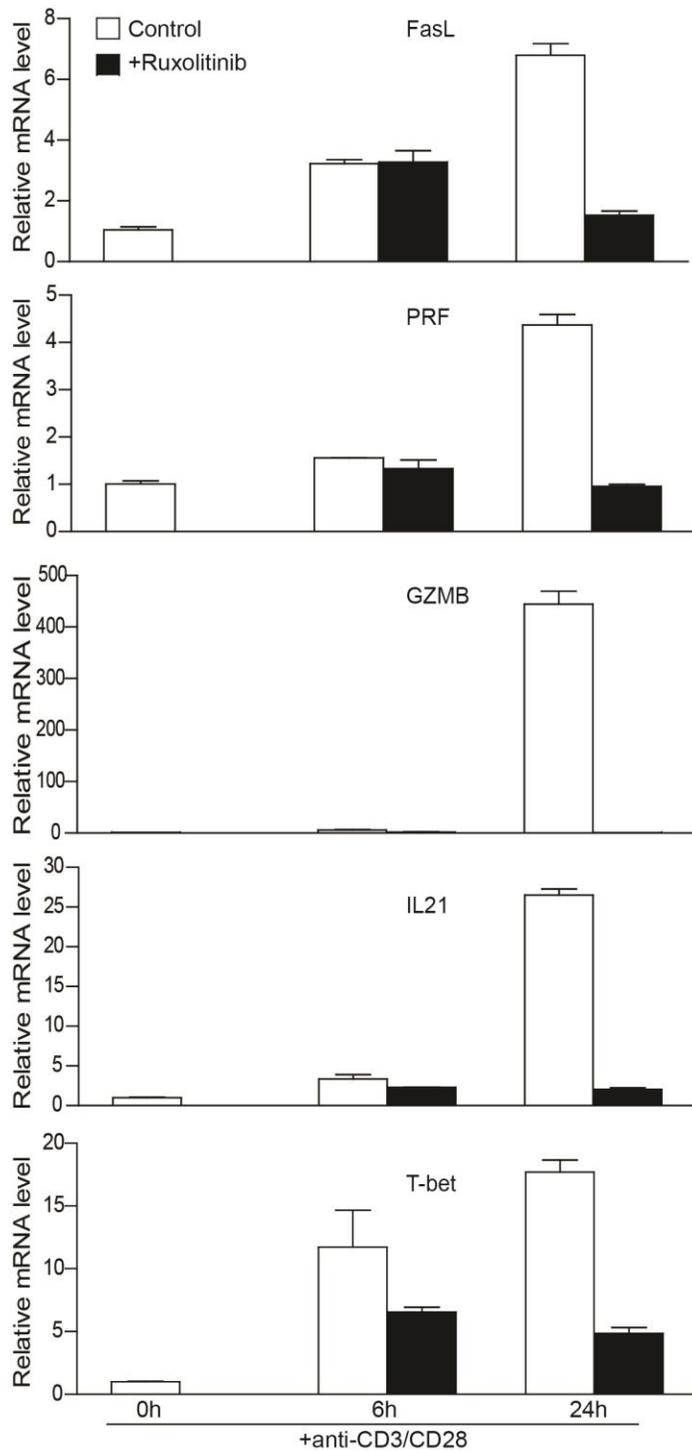


Figure S5. Ruxolitinib inhibits T cell effector expression. CD3⁺ T cells were purified from spleen of C57BL/6 mice and cultured in anti-CD3/CD28-coated plates in the absence or presence of ruxolitinib. RNA was isolated at the indicated time points and analyzed for expression levels of the indicated genes by real-time PCR.

Table S1. Reagents

Name	Company	Cat log#	Application
Pstat (Y701)	BD transduction laboratory	612133	WB
STST1 (C-terminus)	BD transduction laboratory	51-90002093	WB
pSTAT2 (Y690)	Abcam	ab53132	WB
STAT2 (H190)	Santa cruz	sc22816	WB
pSTAT3 (Y705)	Upstate	05-485	WB
STAT3	BD transduction laboratory	610190	WB
pSTAT4 (Y693)	Santa cruz	sc101804	WB
STAT4 (H119)	Santa cruz	sc7959	WB
pSTAT5 (Y641)	BD Bioscience Pharmigen	558242	WB
STAT 5	BD transduction laboratory	51-9002096	WB
pSTAT6 (Y641)	Santa cruz	sc11762	WB
STAT6	BD transduction laboratory	51-9002196	WB
β -actin	Sigma	A5441	WB
APC anti-mouse PD-L1	Biologend	124312	Flow
FITC anti-mouse CD11b	Biologend	101206	Flow
PE anti-mouse Gr1	Biologend	108408	Flow
PE anti-mouse Ly6G	BD Bioscience	551461	Flow
APC anti-mouse Ly6C	Biologend	128016	Flow
APC anti-human PD-L1	Biologend	329708	Flow
CD8	Dako	M103	IHC
PD-L1	Abcam	ab205921	IHC
Hamest anti-mouse CD3e	BD Pharmigen	553057	Coating
Hamest anti-mouse CD28	BD Pharmigen	553294	Coating
Recombinant mouse IL21	Biologend	574502	Coating
IgG	BioXCell	BP0091	In-vivo treatment
anti-IFN γ	BioXCell	BP0055	In-vivo treatment
Fludarabine	Santa cruz	sc204755	In-vitro treatment

Table S2.PCR Primers

mCTLA4-F	CCCTGCTCACTTCTTTTCATCC
mCTLA4-B	TTTGGTCATTGTCTGCCGC
mIL21-F	AAGAGGCAAGGGTGTAGTAAGAAGC
mIL21-B	GGAAAGGATGTGGGAGAGGAGAC
mRORC-F	AACTTGGGGAACCAGAACAGGG
mRORC-B	GCTTGGCAAACCTCCACCACATAC
mIL10-F	GCTGGACAACATACTGCTAACCGAC
mIL10-B	CTTGCTCTTATTTTACAGGGGAG
mTGF β -F	TTGAGTCCCTCGCATCCAG
mTGF β -B	TCCAAGGAAAGGTAGGTGATAGTC
mCD8-F	ACCTGGACATCAGAGCCCCTTG
mCD8-B	AATCCTACGCTTGGCCACC
mCD4-F	CCTCAAGATAACCCAGGTCTCG
mCD4-B	CAAGGAAACCCAGAAAGCCG
mIFN γ -F	CCATCAGCAACAACATAAGCGTC
mIFN γ -B	TCTCTCCCCACCCGAATCAGCAG
mPRF1-F	CCTATGGCACGCACTTATCACC
mPRF1-B	TTCCTGGAGACGCTGGCTTGG
mGZMB-F	GCCACAACATCAAAGAACAGG
mGZMB-B	CCAACCAGCCACATAGCACAC
mIL17A-F	CCCTCAGACTACCTCAACCGTTC
mIL17A-B	TCTCAGGCTCCCTCTTCAGGAC
mIL6-F	TCTGGGAAATCGTGGAAATGAG
mIL6-B	TCTCTGAAGACTCTGGCTTTGTC
mTbx21-F	TGTTCCCATTCCTGTCTTCAC
mTbx21-B	TGCTGCCTTCTGCCTTTC
mIL23 α -F	ATAATGTGCCCGTATCCAGTG
mIL23 α -B	GCTCCCTTTGAAGATGTCAGAG
mPD-L1-F	ATTGCTCCTTGACTGCTGGCTG
mPD-L1-B	TTCTGGGTTCCCTCCTCTTTC
mPD-1-F	CCGCCTTCTGTAATGGTTGAG
mPD-1-B	CGATTTTGCCTTGGGGTGC
mFasL-F	CTGGGGCTCCTCCAGGGTCAGT
mFasL-B	TCTCCTCCATTAGCACCAGATCC
mIL24-F	AATGAATGCTGACTGAGCCTGCC
mIL24-B	CCAAATCGGAACCTTGACCCTC
mIL27-F	TGCTTCCTCGCTACCACACTTC
mIL27-B	TCCTCTCCTCTTCCCTCTGTCC
mCSF1-F	CAACACCCCAATGCTAACG
mCSF1-B	CCCTCTGCCTCTGAGAATCATCC
mGM-CSF-F	TCCTGGGCATTGTGGTCTACAG
mGM-CSF-B	TGGGGGGCAGTATGTCTGG
mCXCL9-F1	TCATTGCTACACTGAAGAACGGAG
mCXCL9-B1	ACGACGACGACTTGGGGTG
mCXCL10-F1	TCTCTCCATCACTCCCCTTACC
mCXCL10-B1	CTTGCTTCGGCAGTTACTTTTGTCT
mIL9-F1	GCTTGTGTCTCTCCGTCCAAC
mIL9-B1	CACCCGATGGAAAACAGGC
mGATA3-F1	CCTCTACGTCCTTGCTACTCAGG
mGATA3-B1	CCCCCCCCAAAAAAAAGC
mIFN β -F1	CTGCGTTCCTGCTGTGCTTC
mIFN β -B1	TCTTCTCCGTCATCTCCATAGGG
mIFN α 1-F1	CTGAAGGACAGGAAGGACTTTGG
mIFN α 1-B1	CTGCTGGTGGAGGTCATTGC
m β -Actin-F	CTGGCACCACACCTTCTACAATG
m β -Actin-B	GGGTCACTTTTACGGTTGG

Table S3. Pancreatic cancer patient information

Patient*	Gender	Ethnicity	Age	Cancer Stage	Primary or Metastatic	Before or After Treatment
1	Male	Black	47	pT3N1M0 (Stage IIB)	Primary (G2 histology)	Patient with prior HNSCC treated with cisplatin and radiation. Last treatment 7 months prior to surgery to remove pancreatic tumor
2	Male	Black	62	pT3N1M0 (Stage IIB)	Primary (G2 histology)	none
3	Female	White	53	pT2N0M0 (Stage IB)	Primary (G2 histology)	none
4	Male	Black	67	pT4N0M0 (Stage III)	Primary (G2 histology)	none
5	Female	White	58	pT3N0M0 (Stage IIA)	Primary (G2 histology)	none

*Patient number as shown in Figure 7.