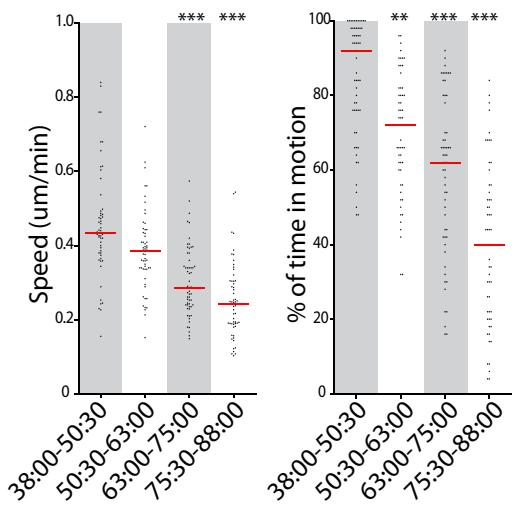


# **Supplemental Materials**

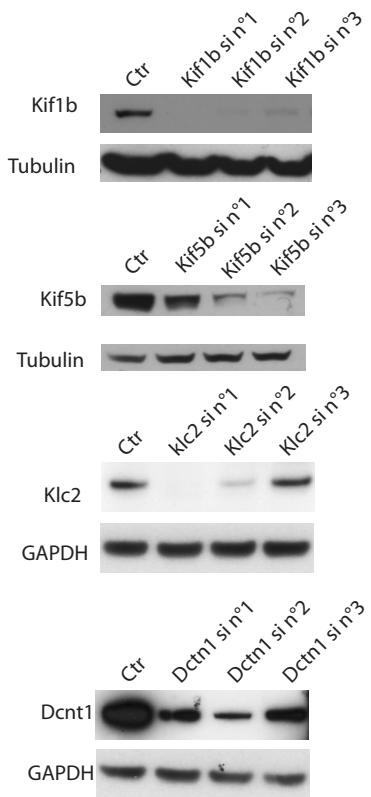
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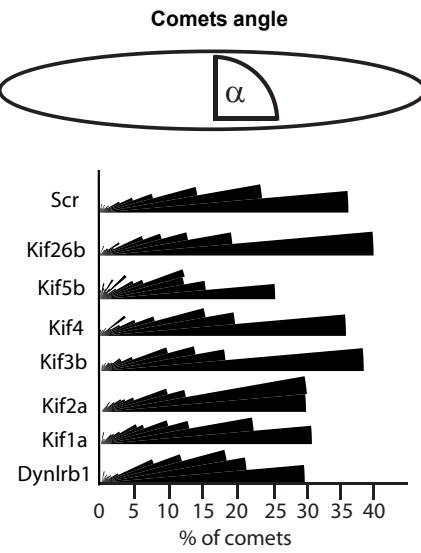
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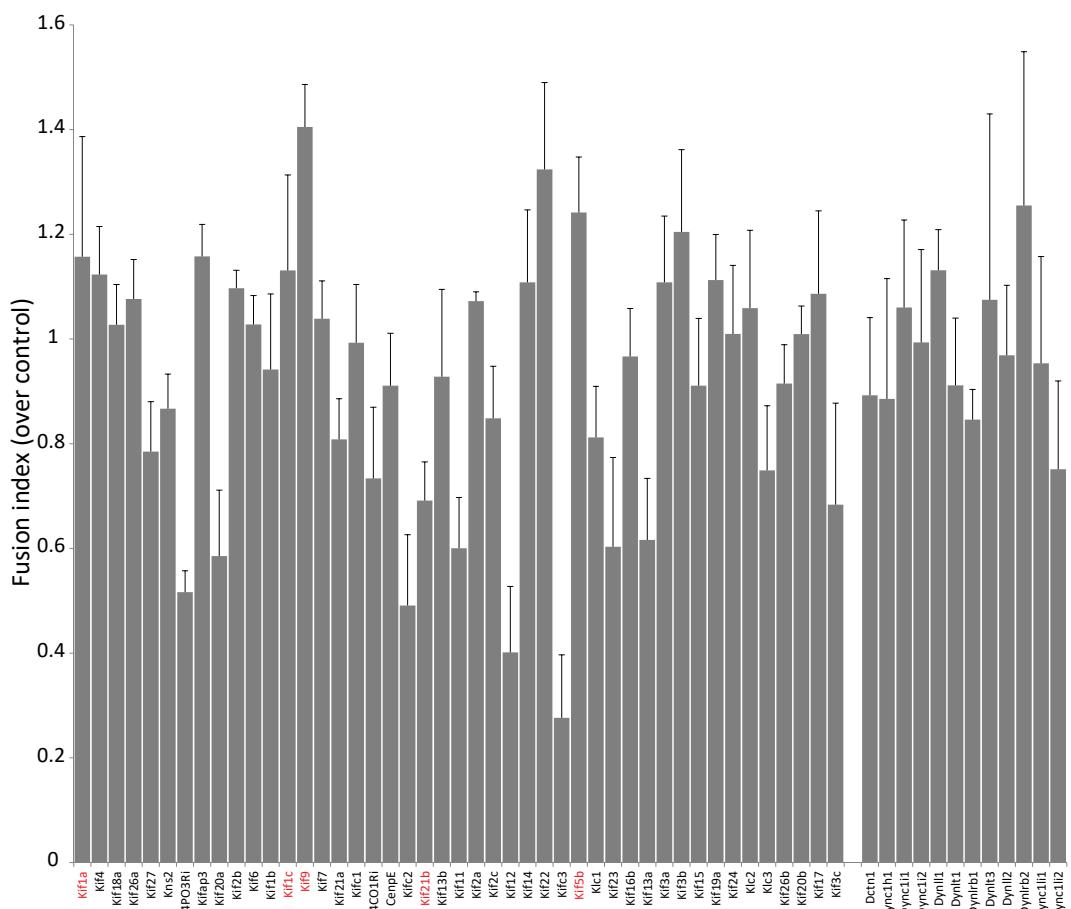
B



D



C



E

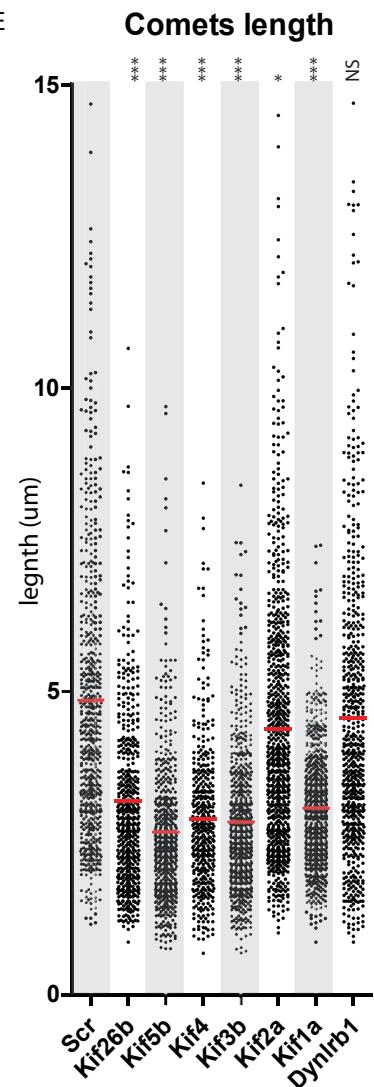


Fig S1

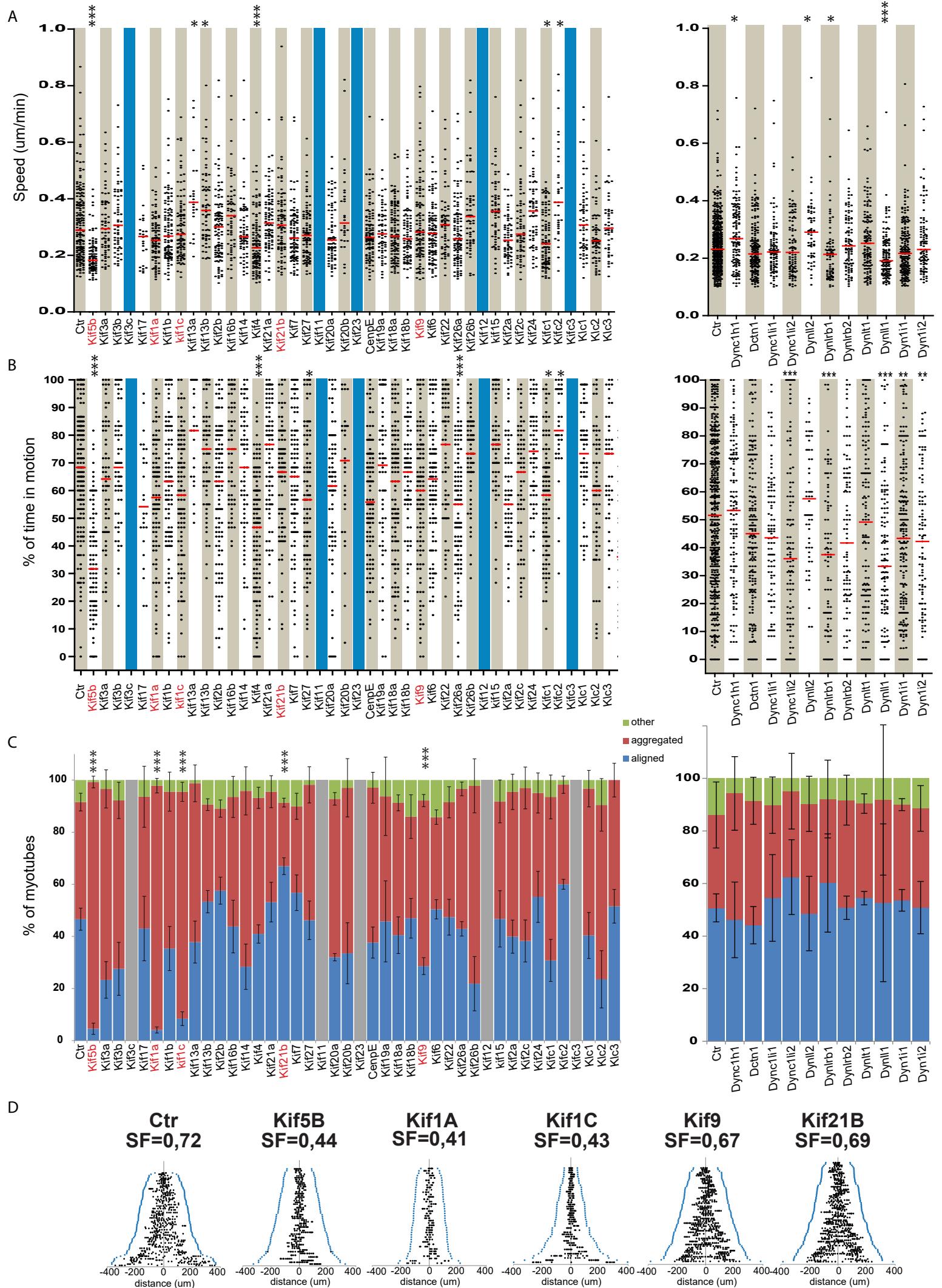


Fig S2

### **Supplementary Figure 1. Interfering with nuclear movement inside myotubes.**

**A:** Nuclear speed and percentage of time in motion during four successive intervals after differentiation. At least 57 nuclei were monitored from 3 different experiments. **B:** Expression levels after silencing of Kif1b, Dctn1, Kif5b and Klc2. **C:** Fusion index over control after depletion of the indicated motors. Average of three independent experiments is represented. **D:** Angles of EB1 comets were measured compared to the longest myotube axis. All angles were averaged on a 90° quadrant to facilitate reading, and sampled every 5°. **E:** The length of EB1 comets was measured over a period of 200s.

### **Supplementary Figure 2. Screening for nuclei behavior inside myotubes**

**Speed (A), Time in motion (B)** and nuclear distribution **(C)** of nuclei inside myotubes from differentiated GFP-H1-C2 cells in non-treated (ctrl), myotubes treated with 50 nM siRNA targeting kinesin and dynein members. Red line indicates the median. In average 110 nuclei were monitored from three different experiments. Nuclear distribution is quantified as “aligned” if >70% of nuclei are aligned along the same axis; “aggregated” if >70% of nuclei do not align along the same axis; “other” if nuclei are both aggregated and aligned in the same myotube. Silencing of kinesins required for cell cycle progression was not analyzed as their depletion induced cell death before differentiation was induced (blue bars). **D:** Representation of nuclei spreading in several myotubes after siRNA for the indicated proteins with the corresponding spreading factor. Each horizontal line represents a myotube, with the extremities depicted as blue dots. Each nucleus is a black dot.

### **Supplementary movie 1**

Phase contrast time-lapse movies of myotubes in presence of 75 nM Nocodazole, or 100 nM Taxol or 200nM Jasplakinolide or 10 nM cytochalasine D as indicated.

**Table1** Listing of siRNA sequences and Taqman probes used for each molecular motor.

RefSeq Accession Number	Gene Symbol	Sense siRNA Sequence	Antisense siRNA Sequence	Taqman Assay ID
NM_008440	Kif1a	GGACAUCAACUAUGCCUCUtt	AGAGGCAUAGUUGAUGUCCtC	Mm00492863_m1
		GGAAACAGAGAAGGAUCAUtt	AAUGAUCCUUUCUCUGUUUCtC	
		CCAAGGUCCUUCAUCGAUAtt	UAUUCGAUGAAGGCACUUGGtC	
NM_207682	Kif1b	CGGGCUGAUUCAACUGGUGtt	CACCAGUUGAACAGCCCCGtt	Mm00801827_m1
		CCUCAAUGAAGACCCAUUAtt	UAAUGGGUCUUCAUGAGGtt	

		GGAUGGAAUUACAAGGGUtt	AACCCUUGUAUUCCAUCCtt	
NM_153103	Kif1c	CCUUCGACUAUUCUUACUGtt	CAGUAAGAAUAGCGAAGGtg	Mm00462184_m1
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		CCAUGUUUCCGCUUCAAUtt	AUUGAAGCGGAAAACAUGGtt	
		GGGAUUUUUAUGCAUUAGCatt	UGCUAUAGCAAAAAUCCCtt	
NM_008442	Kif2a	CGCAGAUCAUUUUCAUAGtt	CUAUGAAAUAUGAUCUGCGtt	Mm00515233_m1
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