

Unraveling Biophysical Interactions of Radiation Pneumonitis in Non-Small-Cell Lung Cancer via Bayesian Network Analysis

Yi Luo, Issam El Naqa, Daniel L. McShan, Dipankar Ray, Ines Lohse, Martha M. Matuszak, Dawn Owen, Shruti Jolly, Theodore S. Lawrence, Feng-Ming Kong, Randall K. Ten Haken

Affiliations of authors: Department of Radiation Oncology, The University of Michigan, Ann Arbor, MI (YL, IEN, DM, DR, IL, MS, MM, DO, SJ, TL, RTH); Department of Radiation Oncology, Indiana University, Indianapolis, IN (FMK)

Correspondence to: Randall K. Ten Haken, PhD, University of Michigan, Department of Radiation Oncology, UH B2C432, SPC 5010, 1500 East Medical Center Dr., Ann Arbor, MI 48109-5010 (e-mail: rth@med.umich.edu); phone: (734) 936-8695; Fax (734) 936-7859)

Appendix E: Mechanistic Biological Interpretation of the Results

Our study of genetic variants identified four SNPs: Rs1800469 (TGF- β 1), Rs4803455 (TGF- β 1), Rs1799983 (NOS3), and Rs6494633 (SMAD3) that were specifically prominent in our dataset. Rs1800469 (-509 C/T) is a SNP within the promoter of polymorphisms of the transforming growth factor beta1 (TGF- β 1), a multifunctional growth factor involved in wound healing and radiation pneumonitis [1, 2]. All 49 SNPs in the SMAD3 gene were examined to identify eight haplotype blocks within the gene, and significant associations between haplotypes and overall survival were observed for haplotype block 5 of SMAD3 including Rs6494633 (intron G/A) [3]. SMAD3 is a TGF- β 1-dependent nuclear transcription factor and has been recently described as a key down-stream mediator in fibrotic signaling pathways [4]. In addition to activating extracellular matrix synthesis, SMAD3 appears to mediate the inhibitory activity of TGF- β 1 on matrix-degrading enzymes and increase expression of matrix metalloproteinase inhibitors [5]. Additionally, it has been proposed that SMAD3 is up-regulated through a TGF- β 1-dependent pathway and acts as an important transcription factor during the fibrotic process following radiation injury [4]. Moreover, Flanders et al. proposed that SMAD3 plays a unique role in the cellular and tissue responses to wounding, and attenuation of SMAD3 signaling may improve wound healing in previously irradiated skin [6]. Hildebrandt et al. found that Rs1799983 in NOS3 genotype was associated with a 70% reduction in risk of pneumonitis, and their study also demonstrate a dose-related effect in inflammation-related SNPs [7]. Based on the pre-treatment BN data, all the above identified SNPs were primarily negatively associated with dosimetric parameters (Fig. 3a), whose intra-relationships minimally altered even during treatment (Fig. 5a). Additionally, during treatment, we have identified a negative association between smad3_Rs6494633 and Interferon-gamma (IFN γ), which impacted RP2 prediction via regulating TGF- β 1 (Fig. 5a). Previous study identified a negative correlation between SMAD3 and IFN γ , where loss of SMAD3 upon genetic ablation caused increased IFN γ production by natural killers (NK) cells following

treatments with CD-16 and IL-12 [8]. Taken together, these findings support the relationships between SNPs and RP2 identified in our BNs (Figs. 3a and 5a).

Damage caused by radiotherapy, such as RP, likely results from the interaction between pro- and anti-inflammatory cytokines, whether the inflammation occurs and to what severity depends on the inter-balance between these two categories of cytokines [9]. The role of IL-2 in the inflammatory process is complex, and the balance between the IL-2 pro- and anti-inflammatory effects is critical for an appropriate mounting and resolution of immune responses [10]. There is increasing evidence to suggest that IL-15 may play an important role in protective immune responses such as allograft rejection and the pathogenesis of autoimmune diseases, where mononuclear cell infiltration is its hallmark feature [11]. IFN γ is a cytokine that is produced primarily by activated CD4+ or CD8+ T cells and NK cells and is recognized as chief mediator of innate as well as adaptive immunity[12]. Among the biological activities of IFN γ , activation of macrophages is of key importance and IFN γ upregulates a variety of pro-inflammatory parameters such as IL-15[13]. Desai et al. have evaluated the cytokines secretion profile of human lung tumor cells, to compare their cytokine profile either before or after acute (6 Gy) and fractionated doses (3 \times 2 Gy), and they observed that the secretion of certain cytokines was cell line-specific and that TGF- β 1 was highly represented in irradiated conditioned medium rather than IFN γ . In addition, in all the cell lines studied, they showed that TGF- β 1 increased markedly in a dose dependent manner[14, 15]. These findings corroborate the interactions found among dosimetric information, pre- and during cytokines shown in our BN (Fig. 5a). Moreover, our previous work demonstrated that patients who developed RP2 following radiotherapy had higher plasma TGF- β 1 levels compared with those prior to radiotherapy, unlike patients who did not develop RP2 [16], confirming the relationship between SLP_TGF_ β 1 and RP2 observed in the BN at mid-treatment timepoint.

Furthermore, our study of genetic expression parameters identified two microRNAs (miR-223-3p and miR-191-5p) as upstream regulators of various cytokines. Both of them are among the top 25 microRNA up-regulated in non-muscle-invasive Bladder Cancer according to white blood cells (WBC)-

derived bio-specimen source[17]. Liang et al. showed that miR-223 overexpression down-regulates ATM expression and sensitizes U87 cells to radiation in vitro and in vivo[18]. In contrast, miR-223 is upregulated in T cell acute lymphocytic leukemia (T-ALL), Epstein–Barr virus (EBV)-positive diffuse large B-cell lymphoma, and metastatic gastric cancer [19]. Mir-191 is one of the highly expressed and stable miRNA in human serum or saliva [20]. It has been found to be differentially expressed in lung cancer patients; however, it was shown to not alter cell cycle, proliferation or chemo-sensitivity of lung cancer cell lines[21, 22]. In the context of inflammatory response, miR-191 has been shown to be involved in human NK cell activation through IL-2 and IL-15 stimulations[23]; and this relationship is also captured in the BNs (Figs. 3a and 5a).

References

1. Azhar M, Yin M, Bommireddy R, *et al.* Generation of mice with a conditional allele for transforming growth factor beta 1 gene. *Genesis* 2009;47(6):423-31.
2. Lu Y, Boer JM, Barsova RM, *et al.* TGFβ1 genetic polymorphisms and coronary heart disease risk: a meta-analysis. *BMC Med Genet* 2012;13:39.
3. Lin M, Stewart DJ, Spitz MR, *et al.* Genetic variations in the transforming growth factor-beta pathway as predictors of survival in advanced non-small cell lung cancer. *Carcinogenesis* 2011;32(7):1050-1056.
4. Lee JW, Zoumalan RA, Valenzuela CD, *et al.* Regulators and mediators of radiation-induced fibrosis: Gene expression profiles and a rationale for Smad3 inhibition. *Otolaryngol Head Neck Surg* 2010;143(4):525-30.
5. Flanders KC, Major CD, Arabshahi A, *et al.* Interference with transforming growth factor-beta/Smad3 signaling results in accelerated healing of wounds in previously irradiated skin. *Am J Pathol* 2003;163(6):2247-57.
6. Flanders KC, Major CD, Arabshahi A, *et al.* Interference with Transforming Growth Factor-β/Smad3 Signaling Results in Accelerated Healing of Wounds in Previously Irradiated Skin *The American Journal of Pathology* 2003;163(6):2247-57.
7. Hildebrandt MA, Komaki R, Liao Z, *et al.* Genetic variants in inflammation-related genes are associated with radiation-induced toxicity following treatment for non-small cell lung cancer. *PLoS One* 2010;5(8):e12402.
8. Trotta R, Dal Col J, Yu J, *et al.* TGF-beta utilizes SMAD3 to inhibit CD16-mediated IFN-gamma production and antibody-dependent cellular cytotoxicity in human NK cells. *J Immunol* 2008;181(6):3784-92.
9. Guo CX, Wang J, Huang LH, *et al.* Impact of single-nucleotide polymorphisms on radiation pneumonitis in cancer patients. *Mol Clin Oncol* 2016;4(1):3-10.
10. Lan RY, Selmi C, Gershwin ME. The regulatory, inflammatory, and T cell programming roles of interleukin-2 (IL-2). *J Autoimmun* 2008;31(1):7-12.

11. Perera LP. Interleukin 15: its role in inflammation and immunity. *Arch Immunol Ther Exp (Warsz)* 2000;48(6):457-64.
12. Muhl H, Pfeilschifter J. Anti-inflammatory properties of pro-inflammatory interferon-gamma. *Int Immunopharmacol* 2003;3(9):1247-55.
13. Doherty TM, Seder RA, Sher A. Induction and regulation of IL-15 expression in murine macrophages. *J Immunol* 1996;156(2):735-41.
14. Desai S, Kumar A, Laskar S, *et al.* Cytokine profile of conditioned medium from human tumor cell lines after acute and fractionated doses of gamma radiation and its effect on survival of bystander tumor cells. *Cytokine* 2013;61(1):54-62.
15. Di Maggio FM, Minafra L, Forte GI, *et al.* Portrait of inflammatory response to ionizing radiation treatment. *Journal of Inflammation-London* 2015;12.
16. Anscher MS, Kong FM, Andrews K., *et al.* Plasma transforming growth factor beta1 as a predictor of radiation pneumonitis. *Int J Radiat Oncol Biol Phys* 1998;41:1029-1035.
17. Armstrong DA, Green BB, Seigne JD, *et al.* MicroRNA molecular profiling from matched tumor and bio-fluids in bladder cancer. *Mol Cancer* 2015;14(1):194.
18. Liang L, Zhu J, Zaorsky NG, *et al.* MicroRNA-223 enhances radiation sensitivity of U87MG cells in vitro and in vivo by targeting ataxia telangiectasia mutated. *Int J Radiat Oncol Biol Phys* 2014;88(4):955-60.
19. R. Moles MB, C Nicot. STAT1: A Novel Target of miR-150 and miR-223 Is Involved in the Proliferation of HTLV-I-Transformed and ATL Cells. *Neoplasia* 2015;17:449-462.
20. Patel RS, Jakymiw A, Yao B, *et al.* High resolution of microRNA signatures in human whole saliva. *Archives of Oral Biology* 2011;56(12):1506-1513.
21. Volinia S, Calin GA, Liu CG, *et al.* A microRNA expression signature of human solid tumors defines cancer gene targets. *Proc Natl Acad Sci U S A* 2006;103(7):2257-61.
22. Patnaik SK, Kannisto E, Yendamuri S. Overexpression of MicroRNA miR-30a or miR-191 in A549 Lung Cancer or BEAS-2B Normal Lung Cell Lines Does Not Alter Phenotype. *Plos One* 2010;5(2).
23. Liu X, Wang Y, Sun Q, *et al.* Identification of microRNA transcriptome involved in human natural killer cell activation. *Immunol Lett* 2012;143(2):208-17.