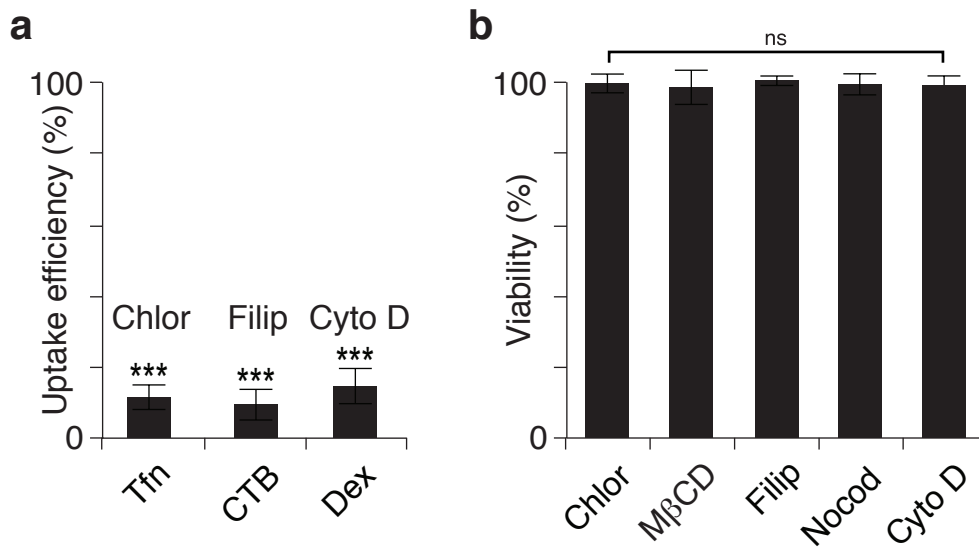
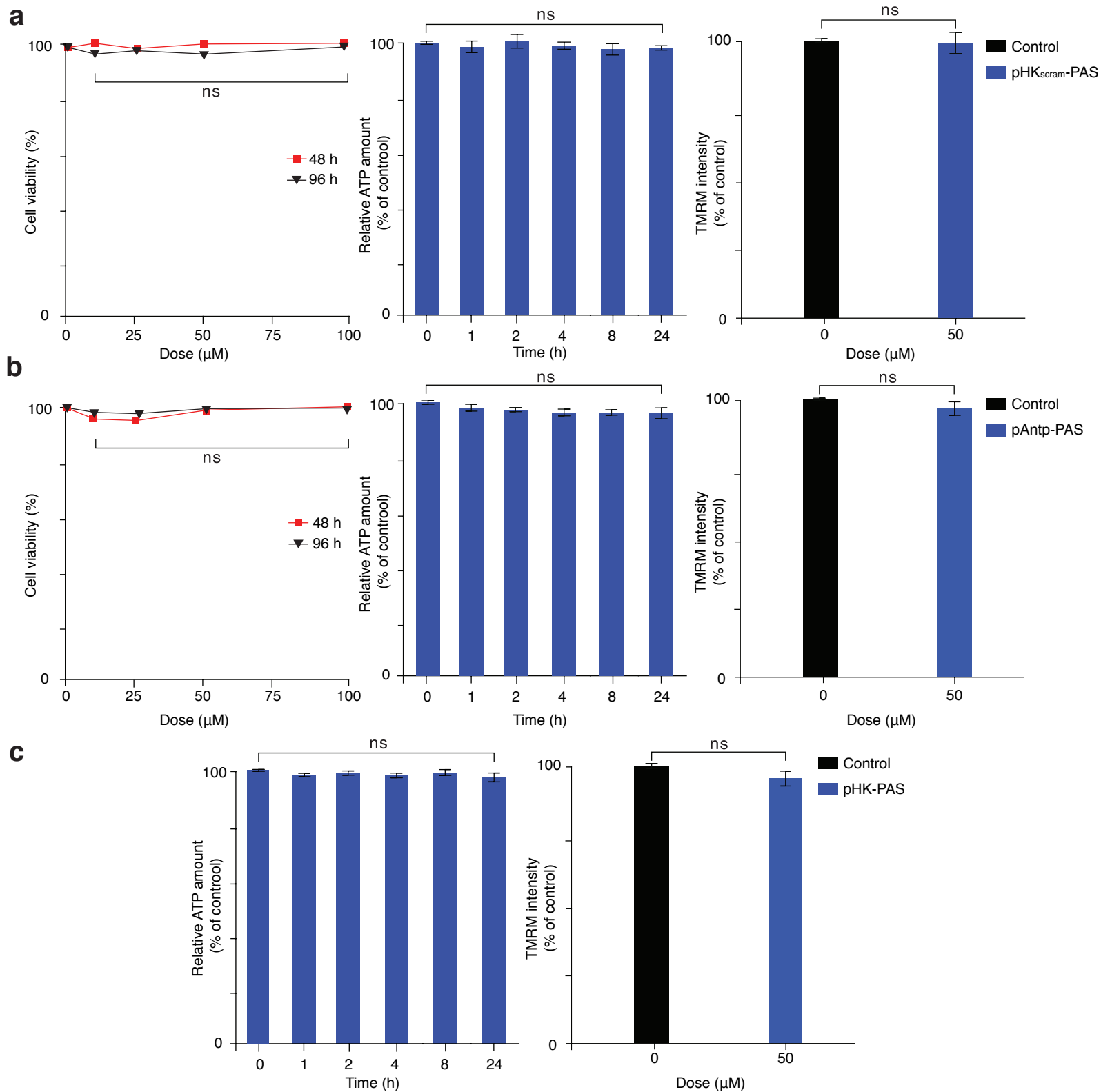


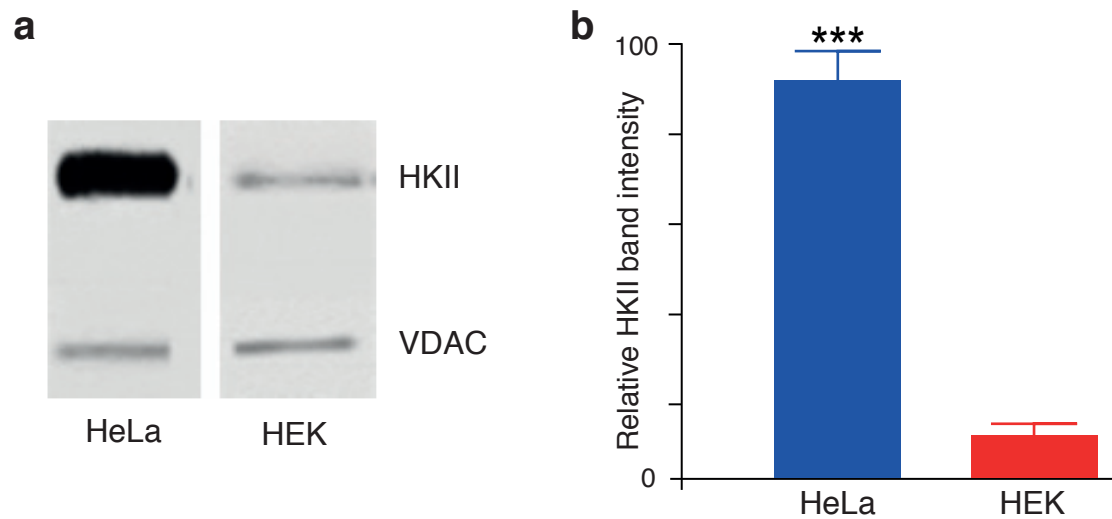
Supplemental Figure S1. Confocal microscopy control experiments. (a) Verification of removal of membrane-bound peptide. Confocal fluorescence microscopy image of HeLa cells following trypsinization. The image shows intact cells with no colocalization between pHK-PASA488 (*green*) and the cell membrane (WGA-Alexa 594; *red*), clearly illustrating that the trypsin treatment removes all membrane-bound peptide. The round shape is due to the fact that cells have not yet adhered to the substrate. Scale bar = 10 μ m. (b – d) Confocal imaging of peptide uptake at low temperature, in ATP-depleted cells, or in the presence of endocytosis inhibitor. HeLa cells were pre-incubated for 1 h at 4 °C in serum-free DMEM (b), pre-treated for 1 h at 37 °C with 10 mM sodium azide/6 mM 2-deoxy-D-glucose in serum- and glucose-free DMEM (c), or pre-treated for 30 min at 37 °C with 10 μ M cytochalasin D in serum-free DMEM (d). 25 μ M pHKA488 (*top panels*) or pHK-PASA488 (*lower panels*) was then added, and the cells were maintained for 2 h at 4 °C (b), or 2 h at 37 °C (c, d). Finally, the cells were stained with organelle markers (5 μ g/mL Hoechst 33342 and 50 nM MitoTracker Red) for 30 min prior to confocal imaging. Scale bar = 10 μ m.



Supplemental Figure S2. Effects of the endocytosis inhibitors on uptake of endocytosis markers and cell viability. (a) HeLa cells were pretreated for 30 min at 37 °C in serum-free DMEM with the inhibitors: 10 μM chlorpromazine (Chlor), 4 μM filipin (Filip) or 10 μM cytochalasin D (Cyto D). Subsequently, the following endocytosis markers were added: 25 μg/ml transferrin-Alexa 546 (Tfn; clathrin-mediated endocytosis), 1 μg/ml cholera toxin B subunit-Alexa 555 (CTB; caveolae-dependent endocytosis), or 100 μg/ml 70 kDa neutral dextran-tetramethylrhodamine (Dex; macropinocytosis). The cells were then maintained for 2 h at 37 °C before the fluorescence was measured by FACS. (b) HeLa cells were treated for 2.5 h with the inhibitors: 10 μM chlorpromazine (Chlor), 5 mM methyl-β-cyclodextrin (MβCD), 4 μM filipin (Filip), 10 μM nocodazole (Nocod), or 10 μM cytochalasin D (Cyto D). 20 μL MTS reagent was then added and the cells were incubated for 4 h at 37 °C. The absorbance of the soluble formazan product ($\lambda = 490$ nm) of MTS reduction was measured on a Synergy H1MF Multi-Mode microplate-reader, with a reference wavelength of 650 nm to subtract background. Cells treated with peptide-free carrier were used as control. The % viability was determined from the ratio of the absorbance of the treated cells to the control cells. ***, $p < 0.0001$; non-significant (ns), $p > 0.05$ compared with controls.



Supplemental Fig. S3. Peptide control experiments. (a,b) Cytotoxic effects of the pHK-PAS variants in HeLa cells. Effects of pHK_{scram}-PAS (a) and pAntp-PAS (b) on MTS reduction (left panels), intracellular ATP levels (middle panels) and mitochondrial inner membrane potential ($\Delta\Psi_m$) (right panels). HeLa cells were treated with the indicated concentrations of peptides for the indicated durations (left panels), or 50 μM peptide for the indicated durations (middle panels) or 24 h (right panels). Cells treated with peptide-free carrier were used as control. The % cell viability (MTS reduction), intracellular ATP, or tetramethylrhodamine methyl ester (TMRM) signal ($\Delta\Psi_m$) was determined from the ratio of the absorbance, luminescence or fluorescence, respectively, of the treated cells to the control cells. (c) Effects of pHK-PAS on intracellular ATP levels (left panel) and $\Delta\Psi_m$ (right panel) in HEK-293 cells. HEK-293 cells were treated with 50 μM pHK-PAS for the indicated durations (left panel) or 24 h (right panel). Cells treated with peptide-free carrier were used as control. The % intracellular ATP or TMRM signal ($\Delta\Psi_m$) was determined from the ratio of the luminescence or fluorescence, respectively, of the treated cells to the control cells. Note: for some data points, the error bars lie within the symbol. Non-significant (ns), $p > 0.05$ compared with controls.



Supplemental Fig. S4. Comparison of the expression levels of HKII in HeLa and HEK-293 cells. HeLa and HEK-293 cells were harvested and the mitochondria-enriched fractions were isolated. Samples were normalized for protein content and electrophoresed, then transferred to a nitrocellulose membrane and incubated overnight with mouse anti-human HKII antibody, followed by 3 h with horseradish peroxidase-conjugated mouse IgG antibody, and finally visualized using SuperSignal West Pico Chemiluminescent Substrate. **(a)** Immunoblots of HKII in the mitochondria-enriched fractions of HeLa and HEK-293 cells. **(b)** Densitometric quantification of the HKII band intensities normalized against the VDAC control. **(a)**. ***, $p < 0.0001$ compared with HEK-293 cells.