

Supporting Information

Harnessing human ADAR2 for RNA repair – Recoding a PINK1 mutation rescues mitophagy

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wt hADAR2 protein production from yeast

Gene and protein sequence of the produced wt hADAR2-His6, the expression was controlled by pGal promotor and cyc1 terminator, similar as described before for SNAP-ADAR(1-3). A His₆-tag was directly cloned after ADAR2.

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          10          20          30          40          50          60
1      GAGAAAAAACCCCGGATTCTAGAAATGGATATAGAAGATGAAGAAAACATGAGTTCAGCA
1
          Xba1      M D I E D E E N M S S S
          70          80          90          100         110         120
61     GCACTGATGTGAAGGAAAACCGCAATCTGGACAACGTGTCCCCAAGGATGGCAGCACAC
20     S T D V K E N R N L D N V S P K D G S T
          130         140         150         160         170         180
121    CTGGGCCTGGCGAGGGCTCTCAGCTCTCCAATGGGGGTGGTGGTGGCCCCGGCAGAAAAGC
40     P G P G E G S Q L S N G G G G G P G R K
          190         200         210         220         230         240
181    GGCCCCTGGAGGAGGGCAGCAATGGCCACTCCAAGTACCGCCTGAAGAAAAGGAGGAAAA
60     R P L E E G S N G H S K Y R L K K R R K
          250         260         270         280         290         300
241    CACCAGGGCCCGTCCTCCCCAAGAACGCCCTGATGCAGCTGAATGAGATCAAGCCTGGTT
80     T P G P V L P K N A L M Q L N E I K P G
          310         320         330         340         350         360
301    TGCAGTACACACTCCTGTCCCAGACTGGGCCCCGTGCACGCGCCTTTGTTTGTTCATGTCTG
100    L Q Y T L L S Q T G P V H A P L F V M S
          370         380         390         400         410         420
361    TGGAGGTGAATGGCCAGGTTTTTTGAGGGCTCTGGTCCCACAAAGAAAAAGGCAAAACTCC
120    V E V N G Q V F E G S G P T K K K A K L
          430         440         450         460         470         480
421    ATGCTGCTGAGAAGGCCTTGAGGTCTTTTCGTTTCAGTTTCTAATGCCCTCTGAGGCCACC
140    H A A E K A L R S F V Q F P N A S E A H
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481 490 500 510 520 530 540
 TGGCCATGGGGAGGACCCTGTCTGTCAACACGGACTTCACATCTGACCAGGCCGACTTCC
 160 L A M G R T L S V N T D F T S D Q A D F

541 550 560 570 580 590 600
 CTGACACGCTCTTCAATGGTTTTGAAACTCCTGACAAGGCGGAGCCTCCCTTTTACGTGG
 180 P D T L F N G F E T P D K A E P P F Y V

601 610 620 630 640 650 660
 GCTCCAATGGGGATGACTCCTTCAGTTCCAGCGGGGACCTCAGCTTGTCTGCTTCCCCGG
 200 G S N G D D S F S S S G D L S L S A S P

661 670 680 690 700 710 720
 TGCCTGCCAGCCTAGCCCAGCCTCCTCTCCCTGCCTTACCACCATTCCCACCCCGAGTG
 220 V P A S L A Q P P L P A L P P F P P P S

721 730 740 750 760 770 780
 GGAAGAATCCCGTGATGATCTTGAACGAACTGCGCCAGGACTCAAGTATGACTTCCTCT
 240 G K N P V M I L N E L R P G L K Y D F L

781 790 800 810 820 830 840
 CCGAGAGCGGGGAGAGCCATGCCAAGAGCTTCGTCATGTCTGTGGTCTGGATGGTTCAGT
 260 S E S G E S H A K S F V M S V V V D G Q

841 850 860 870 880 890 900
 TCTTTGAAGGCTCGGGGAGAAACAAGAAGCTTGCCAAGGCCCGGGCTGCGCAGTCTGCCC
 280 F F E G S G R N K K L A K A R A A Q S A

901 910 920 930 940 950 960
 TGGCCGCCATTTTTAACTTGCACCTTGATCAGACGCCATCTCGCCAGCCTATTCCCAGTG
 300 L A A I F N L H L D Q T P S R Q P I P S

961 970 980 990 1000 1010 1020
 AGGGTCTTCAGCTGCATTTACCGCAGGTTTTAGCTGACGCTGTCTCACGCCTGGTCTCGG
 320 E G L Q L H L P Q V L A D A V S R L V L

1021 1030 1040 1050 1060 1070 1080
 GTAAGTTTTGGTGACCTGACCCGACAACCTTCTCCTCCCCTCACGCTCGCAGAAAAGTGCTGG
 340 G K F G D L T D N F S S P H A R R K V L

1081 1090 1100 1110 1120 1130 1140
 CTGGAGTCGTATGACAACAGGCACAGATGTTAAAGATGCCAAGGTGATAAGTGTTCCTA
 360 A G V V M T T G T D V K D A K V I S V S

1141 1150 1160 1170 1180 1190 1200
 CAGGAACAAAATGTATTAATGGTGAATACATGAGTGATCGTGGCCTTGCAATTAATGACT
 380 T G T K C I N G E Y M S D R G L A L N D

1201 1210 1220 1230 1240 1250 1260
 GCCATGCAGAAATAATATCTCGGAGATCCTTGCTCAGATTTCTTTATACACAACCTTGAGC
 400 C H A E I I S R R S L L R F L Y T Q L E

1261 1270 1280 1290 1300 1310 1320
 TTTACTTAAATAACAAAGATGATCAAAAAAGATCCATCTTTCAGAAATCAGAGCGAGGGG
 420 L Y L N N K D D Q K R S I F Q K S E R G

1321 1330 1340 1350 1360 1370 1380
 GGTTTAGGCTGAAGGAGAATGTCCAGTTTCATCTGTACATCAGCACCTCTCCCTGTGGAG
 440 G F R L K E N V Q F H L Y I S T S P C G

1390 1400 1410 1420 1430 1440

1381 ATGCCAGAATCTTCTCACCACATGAGCCAATCCTGGAAGAACCAGCAGATAGACACCCAA
 460 D A R I F S P H E P I L E E P A D R H P

1441 1450 1460 1470 1480 1490 1500
 ATCGTAAAGCAAGAGGACAGCTACGGACCAAAATAGAGTCTGGTGAGGGGACGATTCCAG
 480 N R K A R G Q L R T K I E S G E G T I P

1501 1510 1520 1530 1540 1550 1560
 TGCGCTCCAATGCGAGCATCCAAACGTGGGACGGGGTGCTGCAAGGGGAGCGGCTGCTCA
 500 V R S N A S I Q T W D G V L Q G E R L L

1561 1570 1580 1590 1600 1610 1620
 CCATGTCTCAGTGCAGTACAAGATTGCACGCTGGAACGTGGTGGGCATCCAGGGATCCCTGC
 520 T M S C S D K I A R W N V V G I Q G S L

1621 1630 1640 1650 1660 1670 1680
 TCAGCATTTTTCGTGGAGCCCATTTACTTCTCGAGCATCATCCTGGGCAGCCTTTACCACG
 540 L S I F V E P I Y F S S I I L G S L Y H

1681 1690 1700 1710 1720 1730 1740
 GGGACCACCTTTCCAGGGCCATGTACCAGCGGATCTCCAACATAGAGGACCTGCCACCTC
 560 G D H L S R A M Y Q R I S N I E D L P P

1741 1750 1760 1770 1780 1790 1800
 TCTACACCCTCAACAAGCCTTTGCTCAGTGGCATCAGCAATGCAGAAGCACGGCAGCCAG
 580 L Y T L N K P L L S G I S N A E A R Q P

1801 1810 1820 1830 1840 1850 1860
 GGAAGGCCCCCAACTTCAGTGTCAACTGGACGGTAGGCGACTCCGCTATTGAGGTCATCA
 600 G K A P N F S V N W T V G D S A I E V I

1861 1870 1880 1890 1900 1910 1920
 ACGCCACGACTGGGAAGGATGAGCTGGGCCGCGCTCCCGCCTGTGTAAGCACGCGTTGT
 620 N A T T G K D E L G R A S R L C K H A L

1921 1930 1940 1950 1960 1970 1980
 ACTGTGCGTGGATGCGTGTGCACGGCAAGGTTCCCTCCCCTTACTACGCTCCAAGATTA
 640 Y C R W M R V H G K V P S H L L R S K I

1981 1990 2000 2010 2020 2030 2040
 CCAAACCAACGTGTACCATGAGTCCAAGCTGGCGGCAAAGGAGTACCAGGCCGCAAGG
 660 T K P N V Y H E S K L A A K E Y Q A A K

2041 2050 2060 2070 2080 2090 2100
 CGCGTCTGTTACAGCCTTCATCAAGGCGGGGCTGGGGGCTGGGTGGAGAAGCCACCG
 680 A R L F T A F I K A G L G A W V E K P T

2101 2110 2120 2130 2140 2150 2160
 AGCAGGACCAGTTCTCACTCACGCCCCACCATCACCATCACCATTAATAGTCGACCTCGA
 700 E Q D Q F S L T P H H H H H H * * Sall

2161 2170 2180
 720 GTCATGTAATTAGTTATGTCACGC

R/G-guideRNA synthesis

Templates for *in vitro* transcription were obtained by Phusion PCR templated with a pMG211 vector that contains the guideRNA downstream of the hammer head cassette. The forward primer (5'-GGT CAGGCCAGGTTCTCCG) was chosen in a way that additional 280 bp were included before the T7 promoter to improve subsequent agarose gel work-up. The backward primer (5'-ACTCTGTGCTGGGGTGGTGGG) was chosen such that the guideRNA ended cleanly with no additional overhanging nucleotides. Shown is the PCR template from the T7 promoter until its 3'-end:

```

1      GCGAAAT TAA TACGACTCAC TATAG GGGAA TTGTGAGCGG ATAACAATTC CCCTCTAGAA
          T7 promotor
61     ATAATTTTGT TAACTTTTAA GAAGGAGATA TACATATGGC TAGCTATTCC ACCTGATGAG
                                   HH-cassette
121    TTTTTACGAA ACGTTCCCGT GAGGGAACGT C*GTGGAATAG TATAACAATA TGCTAAATGT
                                   *=cut R/G-guideRNA
181    TGTTATAGTA TCCCACCACC CCAGCACAGA GT
          R/G-guideRNA
  
```

After urea PAGE-purification the following 61 nt guideRNA results from iv-T7 transcription of the above construct:

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1      GUGGAAUAGU AUAACAAUUAU GCUAAAUGUU GUUAUAGUAU CCCACCACCC
*C* AGCACAGAG U
*C* = counter base
  
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Urea/TBE 8%-PAGE-separation of a guideRNA synthesis

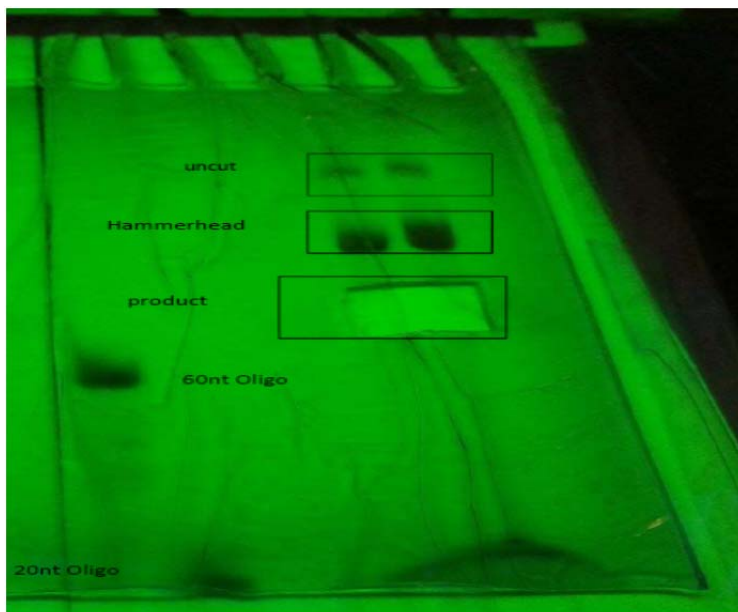


Figure S1. Preparative urea/TBE 8%-PAGE gel for the purification of the *in-vitro* transcribed R/G-guideRNA from the hammer head ribozyme. Indicated are the position of the uncut transcript (156 nt), the hammer head ribozyme (95 nt), the product (61 nt, cut out), and a 60 nt ssDNA and a 20 nt ssDNA as markers. The signal comes from UV shadowing on a TLC plate.

***In-vitro* editing**

RNA sequencing traces of the full eCFP ORF revealing different levels of off-target editing

Figure S2. Full sequencing trace corresponding to the trace shown in Figure 1B, a)

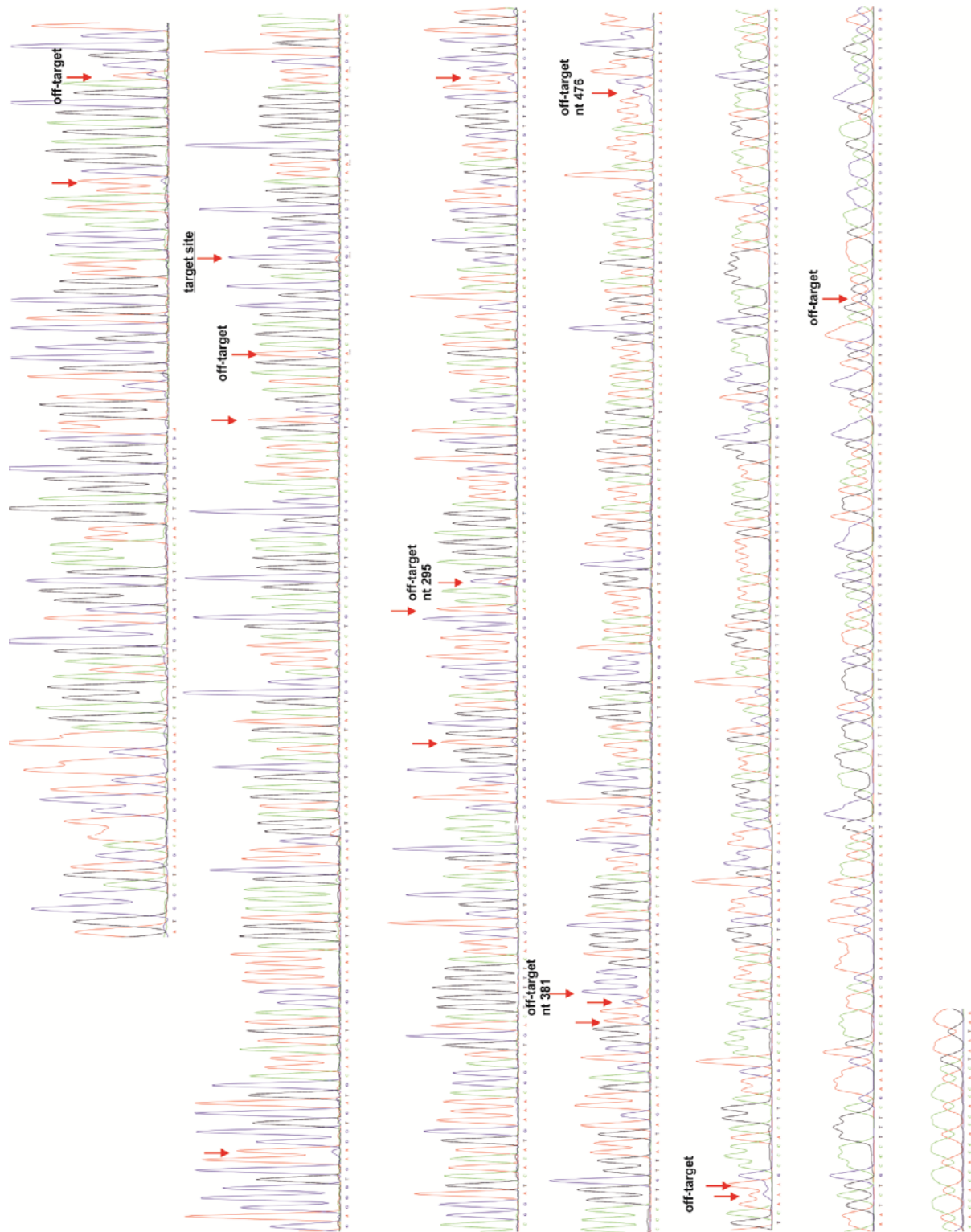


Figure S3. Full sequencing trace corresponding to the trace shown in Figure 1B, c)

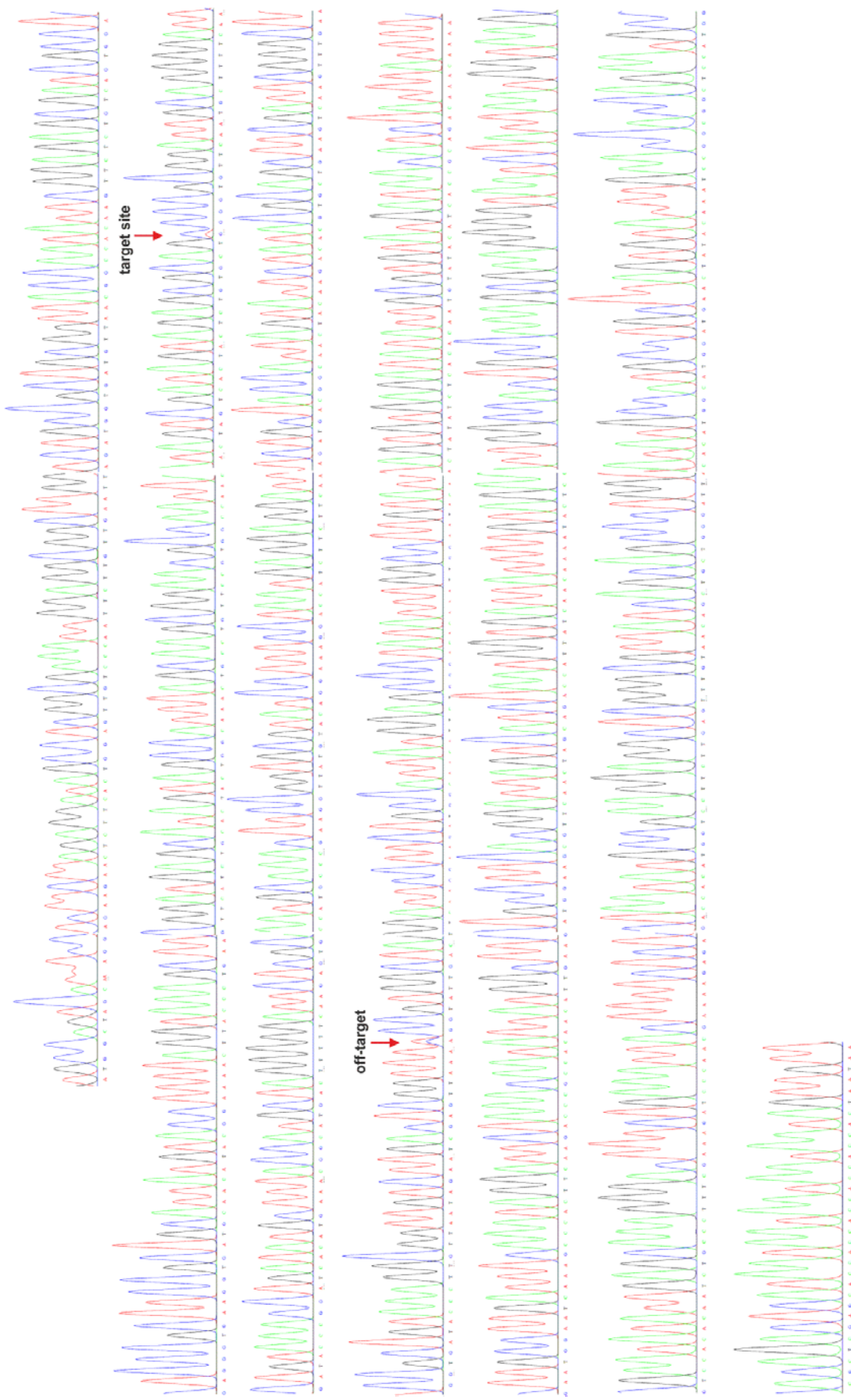


Figure S4. Full sequencing trace corresponding to the trace shown in Figure 1C, c)

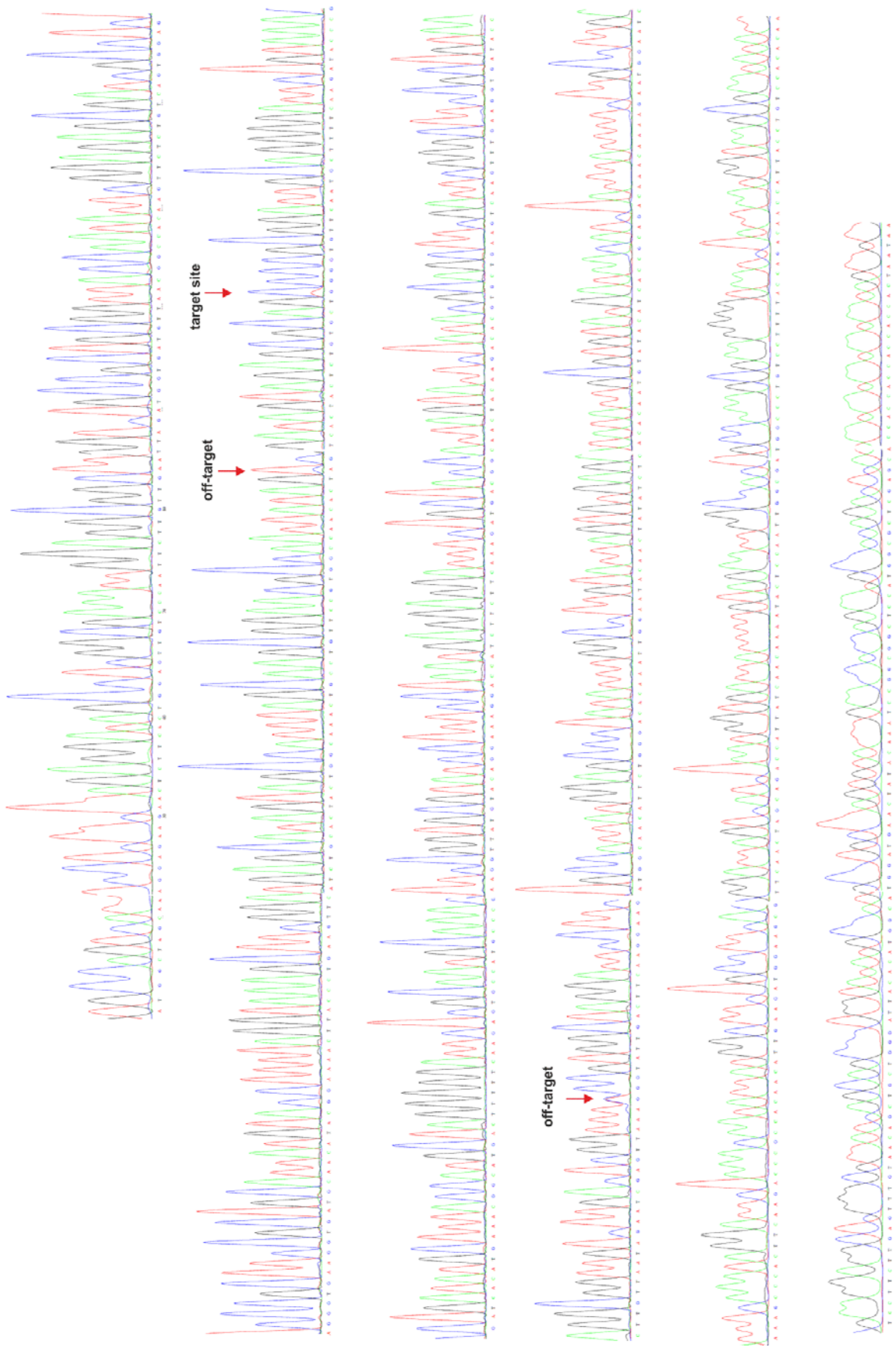
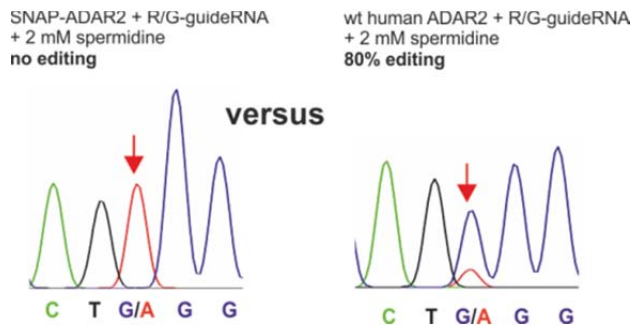


Figure S5. Control experiment to experiments shown in Figure 1B. The R/G-guideRNA recruits only wt human ADAR2 but not SNAP-ADAR2 for editing. Conditions are identical to Figure 1B.



Cellular editing

Gene & protein sequence of wt hADAR2 in the context of the pcDNA 3.1 vector, under control of the CMV promoter and BGH terminator:

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      10      20      30      40      50      60
1  CTCGGATCCACCATGGATATAGAAGATGAAGAAAACATGAGTTCCAGCAGCACTGATGTG
1  BamHI      M D I E D E E N M S S S S T D V

      70      80      90      100     110     120
61  AAGGAAAACCGCAATCTGGACAACGTGTCCCCAAGGATGGCAGCACACCTGGGCCTGGC
21  K E N R N L D N V S P K D G S T P G P G

      130     140     150     160     170     180
121 GAGGGCTCTCAGCTCTCCAATGGGGGTGGTGGTGGCCCCGGCAGAAAGCGGCCCTGGAG
41  E G S Q L S N G G G G G P G R K R P L E

      190     200     210     220     230     240
181 GAGGGCAGCAATGGCCACTCCAAGTACCGCCTGAAGAAAAGGAGGAAAACACCAGGGCCC
61  E G S N G H S K Y R L K K R R K T P G P

      250     260     270     280     290     300
241 GTCTCCCCAAGAACGCCCTGATGCAGCTGAATGAGATCAAGCCTGGTTTGCAGTACACA
81  V L P K N A L M Q L N E I K P G L Q Y T

      310     320     330     340     350     360
301 CTCCTGTCCCAGACTGGGCCCCGTGCACGCGCCTTTGTTTGCATGTCTGTGGAGGTGAAT
101 L L S Q T G P V H A P L F V M S V E V N

      370     380     390     400     410     420
361 GGCCAGGTTTTTTGAGGGCTCTGGTCCCACAAAAGAAAAGGCAAACTCCATGCTGCTGAG
121 G Q V F E G S G P T K K K A K L H A A E

      430     440     450     460     470     480
421 AAGGCCTTGAGGTCTTTTCGTTTCAGTTTCCTAATGCCTCTGAGGCCACCTGGCCATGGGG
141 K A L R S F V Q F P N A S E A H L A M G

      490     500     510     520     530     540
481 AGGACCCTGTCTGTCAACACGGACTTCACATCTGACCAGGCCGACTTCCCTGACACGCTC
161 R T L S V N T D F T S D Q A D F P D T L

      550     560     570     580     590     600
541 TTCAATGGTTTTTGAAACTCCTGACAAGGCGGAGCCTCCCTTTTACGTGGGCTCCAATGGG
181 F N G F E T P D K A E P P F Y V G S N G

      610     620     630     640     650     660
601 GATGACTCCTTCAGTTCCAGCGGGGACCTCAGCTTGTCTGCTTCCCCGGTGCCTGCCAGC
201 D D S F S S S G D L S L S A S P V P A S

      670     680     690     700     710     720
661 CTAGCCCAGCCTCCTCTCCCTGCCTTACCACCATTCCCACCCCCGAGTGGGAAGAATCCC
221 L A Q P P L P A L P P F P P P S G K N P

      730     740     750     760     770     780
721 GTGATGATCTTGAACGAACTGCGCCAGGACTCAAGTATGACTTCTCTCCGAGAGCGGG
241 V M I L N E L R P G L K Y D F L S E S G

      790     800     810     820     830     840
781 GAGAGCCATGCCAAGAGCTTCGTTCATGTCTGTGGTCCGTGGATGGTTCAGTTCTTTGAAGGC
261 E S H A K S F V M S V V V D G Q F F E G
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841 850 860 870 880 890 900
 TCGGGGAGAAAACAAGAAGCTTGCCAAGGCCCGGGCTGCGCAGTCTGCCCTGGCCGCCATT
 281 S G R N K K L A K A R A A Q S A L A A I

901 910 920 930 940 950 960
 TTAACTTGCACCTTGGATCAGACGCCATCTCGCCAGCCTATTTCCCAGTGAGGGTCTTCAG
 301 F N L H L D Q T P S R Q P I P S E G L Q

961 970 980 990 1000 1010 1020
 CTGCATTTACCGCAGGTTTTAGCTGACGCTGTCTCACGCCTGGTCCTGGGTAAGTTTGGT
 321 L H L P Q V L A D A V S R L V L G K F G

1021 1030 1040 1050 1060 1070 1080
 GACCTGACCGACAACCTTCTCCTCCCTCACGCTCGCAGAAAAGTGCTGGCTGGAGTCGTC
 341 D L T D N F S S P H A R R K V L A G V V

1081 1090 1100 1110 1120 1130 1140
 ATGACAACAGGCACAGATGTTAAAGATGCCAAGGTGATAAGTGTCTTCTACAGGAACAAAA
 361 M T T G T D V K D A K V I S V S T G T K

1141 1150 1160 1170 1180 1190 1200
 TGTATTAATGGTGAATACATGAGTGATCGTGGCCTTGCATTAAATGACTGCCATGCAGAA
 381 C I N G E Y M S D R G L A L N D C H A E

1201 1210 1220 1230 1240 1250 1260
 ATAATATCTCGGAGATCCTTGTCTCAGATTTCTTTATACACAACCTTGAGCTTTACTTAAAT
 401 I I S R R S L L R F L Y T Q L E L Y L N

1261 1270 1280 1290 1300 1310 1320
 AACAAAGATGATCAAAAAAGATCCATCTTTTCAGAAATCAGAGCGAGGGGGTTTTAGGCTG
 421 N K D D Q K R S I F Q K S E R G G F R L

1321 1330 1340 1350 1360 1370 1380
 AAGGAGAATGTCCAGTTTTCATCTGTACATCAGCACCTCTCCCTGTGGAGATGCCAGAATC
 441 K E N V Q F H L Y I S T S P C G D A R I

1381 1390 1400 1410 1420 1430 1440
 TTCTCACCATGATGAGCCAATCCTGGAAGAACCAGCAGATAGACACCCAAATCGTAAAGCA
 461 F S P H E P I L E E P A D R H P N R K A

1441 1450 1460 1470 1480 1490 1500
 AGAGGACAGCTACGGACCAAAATAGAGTCTGGTGAGGGGACGATTCCAGTGCGCTCCAAT
 481 R G Q L R T K I E S G E G T I P V R S N

1501 1510 1520 1530 1540 1550 1560
 GCGAGCATCCAAACGTGGGACGGGGTGTCTGCAAGGGGAGCGGCTGCTCACCATGTCCTGC
 501 A S I Q T W D G V L Q G E R L L T M S C

1561 1570 1580 1590 1600 1610 1620
 AGTGACAAGATTGCACGCTGGAACGTGGTGGGCATCCAGGGTTCCCTGCTCAGCATTTTC
 521 S D K I A R W N V V G I Q G S L L S I F

1621 1630 1640 1650 1660 1670 1680
 GTGGAGCCCATTTACTTCTCGAGCATCATCCTGGGCAGCCTTTACCACGGGGACCACCTT
 541 V E P I Y F S S I I L G S L Y H G D H L

1681 1690 1700 1710 1720 1730 1740
 TCCAGGGCCATGTACCAGCGGATCTCCAACATAGAGGACCTGCCACCTCTCTACACCCTC
 561 S R A M Y Q R I S N I E D L P P L Y T L

1741 1750 1760 1770 1780 1790 1800
 581 AACAAAGCCTTTGCTCAGTGGCATCAGCAATGCAGAAGCACGGCAGCCAGGGAAGGCCCC
 N K P L L S G I S N A E A R Q P G K A P

 1801 1810 1820 1830 1840 1850 1860
 601 AACTTCAGTGTCAACTGGACGGTAGGCGACTCCGCTATTGAGGTCATCAACGCCACGACT
 N F S V N W T V G D S A I E V I N A T T

 1861 1870 1880 1890 1900 1910 1920
 621 GGGAAGGATGAGCTGGGCCGCGCTCCCGCCTGTGTAAGCACGCGTTGTACTGTTCGCTGG
 G K D E L G R A S R L C K H A L Y C R W

 1921 1930 1940 1950 1960 1970 1980
 641 ATGCGTGTGCACGGCAAGGTTCCCTCCCCTTACTACGCTCCAAGATTACCAAACCCAAC
 M R V H G K V P S H L L R S K I T K P N

 1981 1990 2000 2010 2020 2030 2040
 661 GTGTACCATGAGTCCAAGCTGGCGGCAAAGGAGTACCAGGCCGCAAGGCCGCTGTGTT
 V Y H E S K L A A K E Y Q A A K A R L F

 2041 2050 2060 2070 2080 2090 2100
 681 ACAGCCTTCATCAAGGCGGGGCTGGGGGCTGGGTGGAGAAGCCCACCGAGCAGGACCAG
 T A F I K A G L G A W V E K P T E Q D Q

 2101 2110 2120 2130 2140 2150
 701 TTCTCACTCAGCCCCTCTAGAGGGCCCTATTCTATAGTGTACCTAAATGCTAG
 F S L T P S R G P Y S I V S P K C *
 Xba-I

Gene and protein sequence of W58X eGFP in the context of the pcDNA 3.1 vector

1 10 20 30 40 50 60
 1 CTCGGATCCACCATGGCTAGCAAAGGAGAAGAACTCTTCACTGGAGTTGTCCCAATTCTT
 BamH1 M A S K G E E L F T G V V P I L

 61 70 80 90 100 110 120
 21 GTTGAATTAGATGGTGATGTTAACGGCCACAAGTTCTCTGTCTCAGTGGAGAGGGTGAAGGT
 V E L D G D V N G H K F S V S G E G E G

 121 130 140 150 160 170 180
 41 GATGCAACATACGGAAAACCTTACCCTGAAGTTCATCTGCCTACTGGCAAACCTGCCTGTT
 D A T Y G K L T L K F I C T T G K L P V

 181 190 200 210 220 230 240
 61 CCGTAGCCGACACTAGTGACGACGCTCTGCTATGGCGTCCAGTGCTTTTCAAGATACCCG
 P * P T L V T T L C Y G V Q C F S R Y P
 W58x

 241 250 260 270 280 290 300
 81 GATCACATGAAACGGCATGACTTTTTTCAAGAGTGCCATGCCCGAAGGTTATGTACAGGAA
 D H M K R H D F F K S A M P E G Y V Q E

 301 310 320 330 340 350 360
 101 AGGACCATCTTCTTCAAAGATGACGGCAACTACAAGACACGTGCTGAAAGTCAAGTTTGAA
 R T I F F K D D G N Y K T R A E V K F E

 361 370 380 390 400 410 420
 121 GGTGATACCCTTGTTAATAGAATCGAGTTAAAAAGGTATTGACTTCAAGGAAGATGGCAAC
 G D T L V N R I E L K G I D F K E D G N

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          430      440      450      460      470      480
421  ATTCTGGGACACAAATTGGAATACAAC TATAACTCACACAATGTATACATCATGGCAGAC
141    I L G H K L E Y N Y N S H N V Y I M A D

          490      500      510      520      530      540
481  AAACAAAAGAATGGAATCAAAGTGAAC TTCAAGACCCGCCACAACATTGAAGATGGAAGC
161    K Q K N G I K V N F K T R H N I E D G S

          550      560      570      580      590      600
541  GTTCAACTAGCAGACCATTATCAACAAA AACTCTCCAATTGGCGATGGCCCTGTCCTTTTA
181    V Q L A D H Y Q Q N T P I G D G P V L L

          610      620      630      640      650      660
601  CCAGACAACCATTACCTGTCCACACAAT CTGCCCTTTTCGAAAAGATCCCAACGAAAAGAGA
201    P D N H Y L S T Q S A L S K D P N E K R

          670      680      690      700      710      720
661  GACCACATGGTCCTTCTTTGAGTTTGT AAACAGCTGCTGGGATTACACATGGCATGGATGAA
221    D H M V L L E F V T A A G I T H G M D E

          730      740      750      760      770      780
721  CTATACAAATCCGGCTCTCTAGAGGGG CCCTTCGAACAAAAACTCATCTCAGAAGAGGATCTG
241    L Y K S G S R G P F E Q K L I S E E D L

          790      800      810      820      830      840
781  AATATGCATACCGGTCATCATCACCATC ACCATTGAGTTTAAACCCGCTGATCAGCCTCG
261    N M H T G H H H H H H * V *

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Gene sequence of a 16 nt R/G-guideRNA against eGFP W58X in the context of the pSilencer vector

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1      CGGAAGAGCG CCCAATACGC AAACCGCCTC TCCC GCGCG TTGGCCGATT CATTAAATGCA
      SapI

61     GCTGGCACGA CAGGTTTCCC GACTGGAAAG CGGGCAGTGA GCGCAACGCA ATTAATGTGA
121    GTTAGCTCAC TCATTAGGCA CCCCAGGCTT TACACTTTAT GCTTCCGGCT CGTATGTTGT
181    GTGGAATTGT GAGCGGATAA CAATTT CACA CAGGAAACAG CTATGACATG ATTACGAATT
241    GCAACGATTT AGGTGACACT ATAGAAGAGA AGGAATTAAT ACGACTCACT ATAGGGAGAG
301    AGAGAGAATT ACCCTCACTA AAGGGAGGAG AAGCATGAAT TCCCCAGTGG AAAGACGCGC
361    AGGCAAAACG CACCACGTGA CGGAGCGTGA CCGCGCGCCG AGCGCGCGCC AAGGTCGGGC
421    AGGAAGAGGG CCTATTTCCC ATGATTCCTT CATATTTGCA TATACGATAC AAGGCTGTTA
481    GAGAGATAAT TAGAATTAAT TTGACTGTAA ACACAAAGAT ATTAGTACAA AATACGTGAC
541    GTAGAAAGTA ATAATTTCTT GGGTAGTTTG CAGTTT TAAA ATTATGTTTT AAAATGGACT
601    ATCATATGCT TACCGTAACT TGAAAGTAT TCGATTTCTT GGTTTATAT ATCTTGTGGA
661    AAGGACGCGG GATCC *GTGGA ATAGTATAAC AATATGCTAA ATGTTGTTAT
AGTATCCCAC

      * = transcription start R/G-motif

721    TCGGCCACGG AACAGGTTTT TTGGAAAGCT TGG
      mRNA template U6-term. HindIII

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Table S1. List of all R/G-gRNAs used in cell culture. Given is the mRNA binding site and the 3'-terminal hairpin (highlighted in gray) if applicable. The invariant R/G-motif is omitted for clarity.

Name of R/G-gRNA	Sequence of the mRNA binding site 5'→3'	Experimental number
W58X GFP P6 16nt	UCGG CCA CGGAACAGG	Fig. 2B
W58X GFP P6 18nt + boxB	UCGG CCA CGGAACAGGCA UCUAGAGGGCCCUGAAGAGGGCC	Fig. 2C / Fig. S10
W58X GFP P6 20nt + boxB	UCGG CCA CGGAACAGGCAGU UCUAGAGGGCCCUGAAGAGGGCCC	Fig. 2C / Fig. S10
W58X GFP P6 25nt + boxB	UCGG CCA CGGAACAGGCAGUUUGCC UCUAGAGGGCCCUGAAGAGGGCCC	Fig. 2C / Fig. S10
W58X GFP P6 29nt + boxB	UCGG CCA CGGAACAGGCAGUUUGCCAGUA UCUAGAGGGCCCUGAAGAGGGCCC	Fig. 2C / Fig. S10
W58X GFP P3 16 nt + boxB	GG CCA CGGAACAGGCA UCUAGAGGGCCCUGAAGAGGGCCC	Fig. 2D / Fig. S11
W58X GFP P4 16 nt + boxB	CGG CCA CGGAACAGGC UCUAGAGGGCCCUGAAGAGGGCCC	Fig. 2D / Fig. S11
W58X GFP P5 16 nt + boxB	UCGG CCA CGGAACAGG UCUAGAGGGCCCUGAAGAGGGCCC	Fig. 2D / Fig. S11
W58X GFP P7 16 nt + boxB	GUCGG CCA CGGAACAG UCUAGAGGGCCCUGAAGAGGGCCC	Fig. 2D / Fig. S11
W58X GFP P8 16 nt + boxB	UGUCGG CCA CGGAACA UCUAGAGGGCCCUGAAGAGGGCCC	Fig. 2D / Fig. S11
W58X GFP P9 16 nt + boxB	GUGUCGG CCA CGGAAC UCUAGAGGGCCCUGAAGAGGGCCC	Fig. 2D / Fig. S11
W58X GFP P10 16 nt + boxB	AGUGUCGG CCA CGGAA UCUAGAGGGCCCUGAAGAGGGCCC	Fig. 2D / Fig. S11
R407Q PINK-1 P4 16 nt + boxB	CC CCG AUCCACGUACC UCUAGAGGGCCCUGAAGAGGGCCC	Fig. S15
R407Q PINK-1 P5 16 nt + boxB	GCC CCG AUCCACGUAC UCUAGAGGGCCCUGAAGAGGGCCC	Fig. S15
R407Q PINK-1 P6 16 nt + boxB	CGCC CCG AUCCACGUA UCUAGAGGGCCCUGAAGAGGGCCC	Fig. S15
R407Q PINK-1 P7 16 nt + boxB	CCG CCG AUCCACGU UCUAGAGGGCCCUGAAGAGGGCCC	Fig. S15
R407Q PINK-1 P8 16 nt + boxB	UCCGCC CCG AUCCACG UCUAGAGGGCCCUGAAGAGGGCCC	Fig. S15
R407Q PINK-1 P9 16 nt + boxB	UUCCGCC CCG AUCCAC UCUAGAGGGCCCUGAAGAGGGCCC	Fig. S15
R407Q PINK-1 P10 16 nt + boxB	UUUCCGCC CCG AUCCA UCUAGAGGGCCCUGAAGAGGGCCC	Fig. S15
W437X Amber PINK-1 P8 16 nt + boxB	CACUGC CCA GGGAUCA GGGCCCUCUUCAGGGCCC	Fig. 3C / Fig. 3D / Fig. 4
3'UTR TAG#1 Actin P8 16 nt + boxB	ACGCA CCA AGUCAUA UCUAGAGGGCCCUGAAGAGGGCCC	Fig. 3B
3'UTR TAG#2 Actin P8 16 nt + boxB	GAAUG CCA UUAAAA UCUAGAGGGCCCUGAAGAGGGCCC	Fig. 3B
3'UTR TAG#3 Actin P8 16 nt + boxB	GCAAUG CCA UACCCUC UCUAGAGGGCCCUGAAGAGGGCCC	Fig. 3B
3'UTR TAG#1 GAPDH P8 16 nt + boxB	AGGGGU CCA CAUGGCA UCUAGAGGGCCCUGAAGAGGGCCC	Fig. 3B
3'UTR TAG#2 GAPDH P8 16 nt	GGCUCC CCA GGCCCCU UCUAGAGGGCCCUGAAGAGGGCCC	Fig. 3B

+ boxB		
V627V TAG#1 GPI P8 16 nt + boxB	UGCCGU CC ACCAGGAUUCUAGAGGGCCCUGAAGAGGGCCC	Fig. 3B
L456L TAG#1 GusB P8 16 nt + boxB	CAGAUU CC AGGUGGGAUCUAGAGGGCCCUGAAGAGGGCCC	Fig. 3B
3'UTR TAG#2 GusB P8 16 nt + boxB	UCCUG CC AGAAUAGAUCUAGAGGGCCCUGAAGAGGGCCC	Fig. 3B
3'UTR TAG#1 VCP P8 16 nt + boxB	CUCCG CC ACCAAAUGUCUAGAGGGCCCUGAAGAGGGCCC	Fig. 3B
3'UTR TAG#2 VCP P8 16 nt + boxB	CCCAA CC CAACAGAUUCUAGAGGGCCCUGAAGAGGGCCC	Fig. 3B
3'UTR TAG#3 VCP P8 16 nt + boxB	ACCCAC CC ACCAGGUUCUAGAGGGCCCUGAAGAGGGCCC	Fig. 3B
3'UTR TAG#1 RAB7A P8 16 nt + boxB	CUGCCG CC AGCUGGAUUCUAGAGGGCCCUGAAGAGGGCCC	Fig. 3B
3'UTR TAG#2 RAB7A P8 16 nt + boxB	AGGGAA CC AGACAGUUUCUAGAGGGCCCUGAAGAGGGCCC	Fig. 3B

RNA sequencing traces of the full eGFP ORF

Figure S6. Full sequencing trace corresponding to the trace shown in Figure 2B, e), first experiment



Figure S7 Full sequencing trace corresponding to the trace shown in Figure 2E, 50 ng experiment



Figure S8: The prolongation of the editing time from 24 hours to 48 hours increases the editing yield in cell culture. All editing samples included the amount of 300 ng W58X eGFP plasmid, 300 ng of ADAR2 plasmid and 1300 ng / 1600 ng of R/G-gRNA plasmid (c-f). The positive controls contained 300 ng eGFP plasmid, 300 ng of ADAR2 plasmid and 1300 ng of R/G-gRNA (a, b). An increment of the fluorescent signal is obtained for the positive controls by prolonging the incubation time up to 48 hours (a-b), as well as for the two editing samples (c->e, d->f). The amount of fluorescing cells and the editing yields were increased for both editing samples. Total magnification: 100x, GFP exposure time: 50 ms.

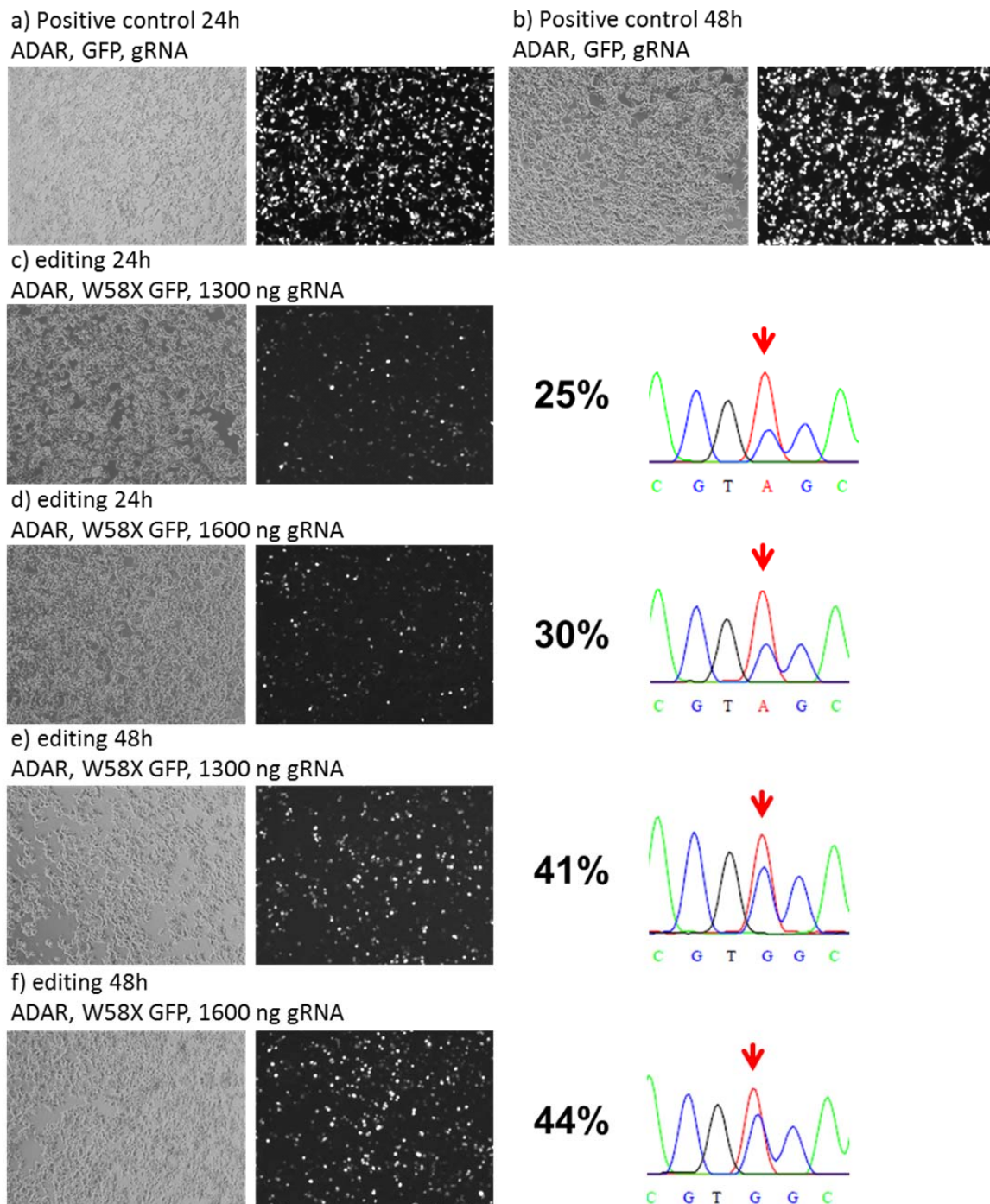
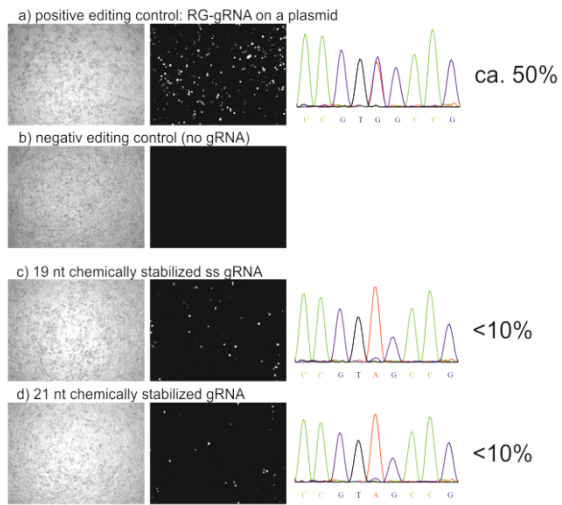


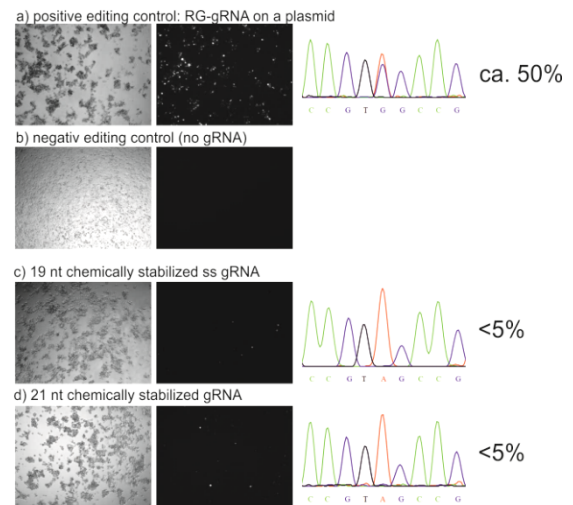
Figure S9: Comparing the editing efficiency using chemically stabilized single-stranded (ss) gRNA and U6-driven unstructured ss guideRNAs, and replacement of ADAR2 by SNAP-ADAR2. A) & B) Replacement of the R/G-gRNA plasmid by a ss-gRNA transfection: A) cotransfection of 500 ng W58X eGFP and 100 ng ADAR2 plasmid; B) transfection of 500 ng W58X eGFP plasmid plus induction of ADAR2 with 10 ng/mL doxycycline. 24 hours after plasmid transfection in 24-wells, the cells were detached and reseeded in a 96-well format and directly reverse transfected with 10 pmol (A) or 20 pmol (B) ss-gRNA. After 48h, fluorescence images were taken and RNA was isolated. In the controls a), 260 ng R/G-guideRNA have been transfected. The chemically stabilized guideRNAs contain 2'-O-methyl groups globally apart from a 3 nt gap around the adenosine to be edited, and terminal phosphorothioates. The full guideRNA sequences are given in Hanswillemenke et al., *JACS* **2015**. **C) & D)**: U6-driven expression of unstructured guideRNAs. Cotransfection in 24-well format: 300 ng W58x eGFP and 1300 ng respective U6-driven guideRNA-vector, in case of C) 100 ng ADAR2 plasmid, in case of D) 10 ng/ml doxycycline. Fluorescence imaging was taken 48 hrs post transfection. The placement of the unstructured guideRNAs (C) and D)) relative to the mRNA is given, 1* stands for the edited adenosine. Negative control z) was like all four positive controls a), but with SNAP-ADAR2 instead of human ADAR2. Total magnification: 100x, GFP exposure time: 50 ms.

Editing with chemically stabilized ss guideRNAs

A) transient ADAR2 expression in 293T cells

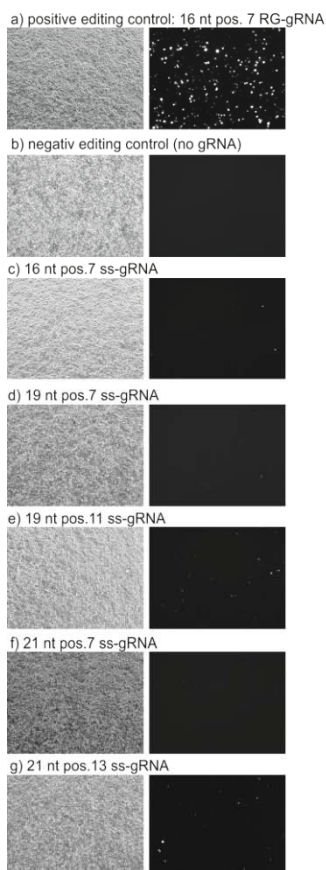


B) genomic ADAR2 dox-induced expression in 293T-FlipIN cells

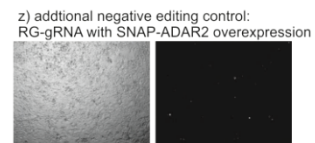
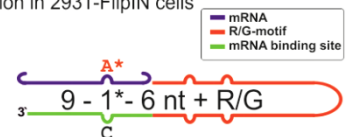
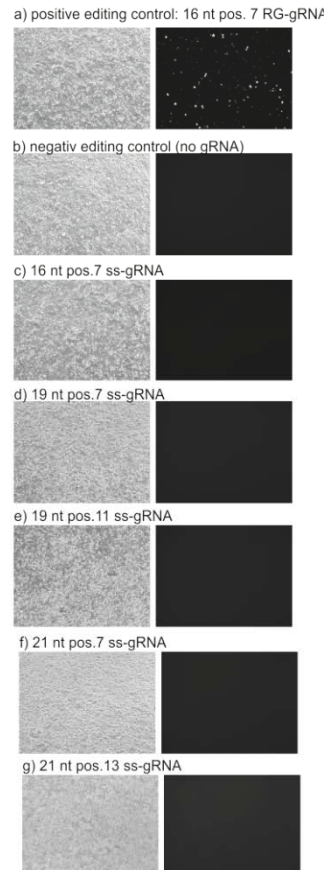


Editing with U6-driven, unstructured ss guideRNAs

C) transient ADAR2 expression in 293T cells



D) genomic ADAR2 dox-induced expression in 293T-FlipIN cells



12 - 1* - 6 nt

8 - 1* - 10 nt

14 - 1* - 6 nt

8 - 1* - 12 nt

Figure S10: Microscopy analysis and editing yields of R/G-gRNAs with varying length of the flexible part and off-target editing at position 53. The strongest fluorescent signal and highest editing yield was obtained for a R/G-gRNA with 16 or 18 nt length of the mRNA binding site. With the prolongation of the mRNA binding site of the R/G-gRNA less fluorescent signal and lower editing yields were observed. The off-target adenosine at position 53 is edited up to 10% if a 25 nt and 29 nt long R/G-gRNA is used for the editing reaction. Total magnification: 100x, GFP exposure time: 50 ms.

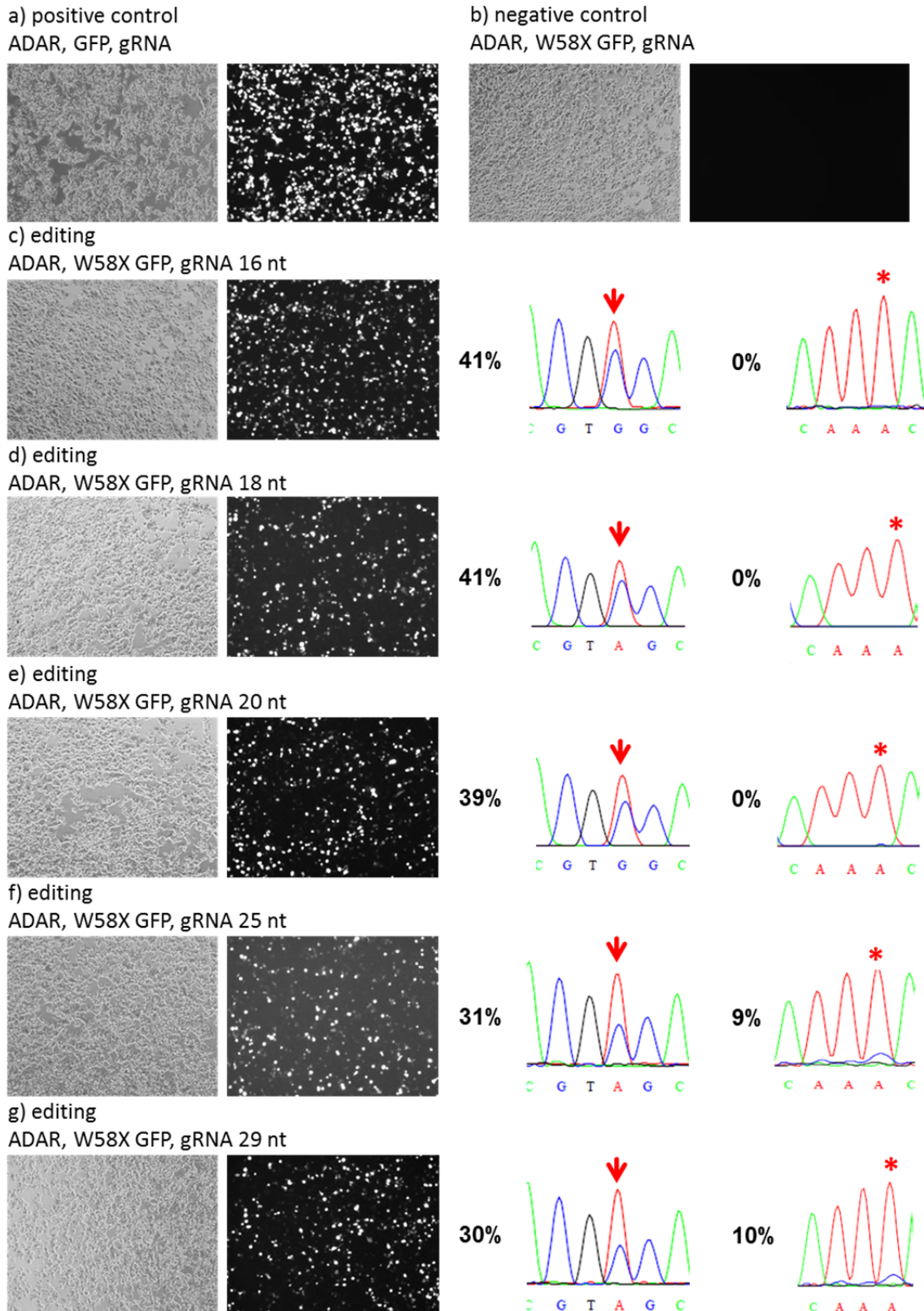
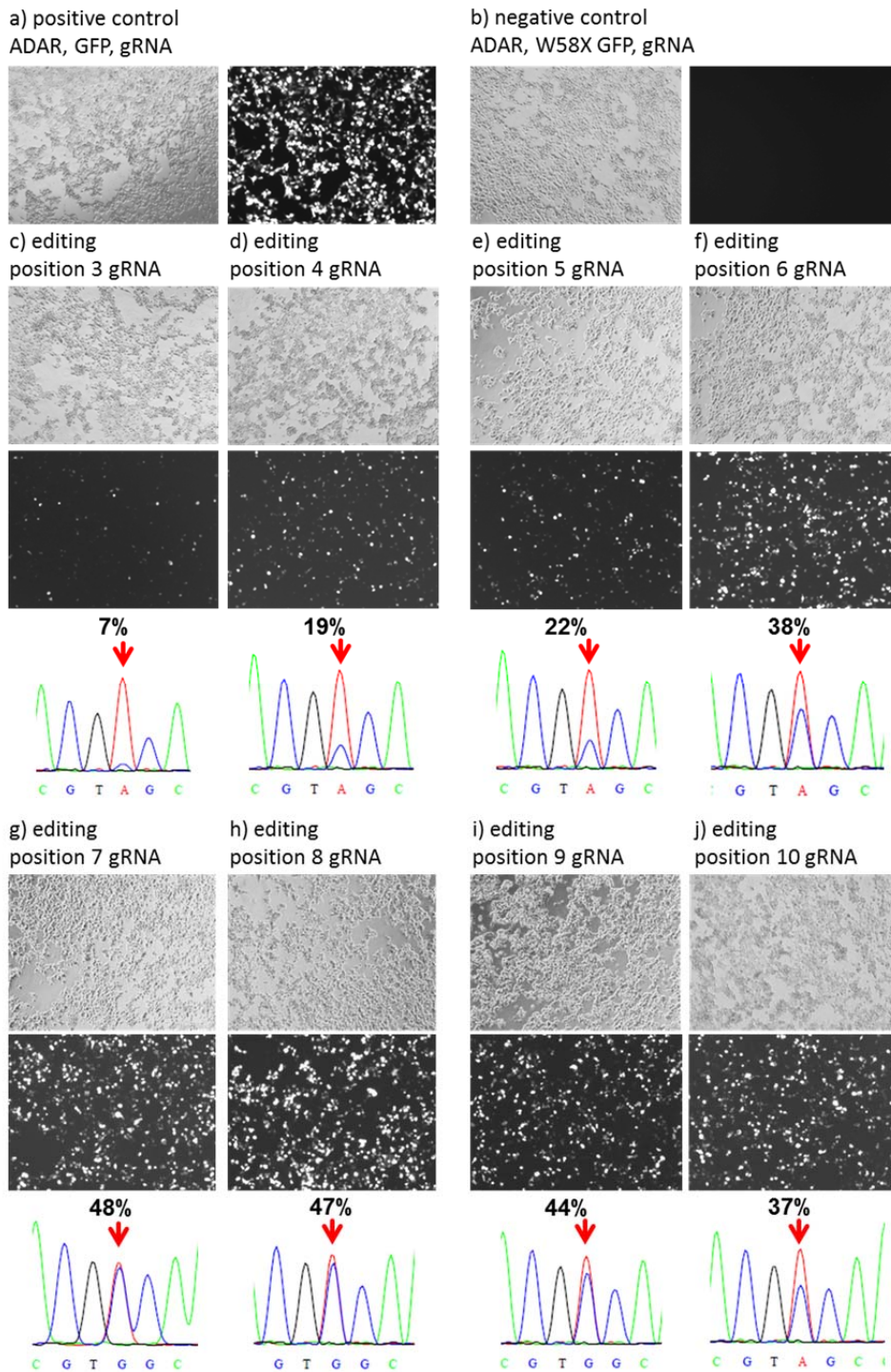
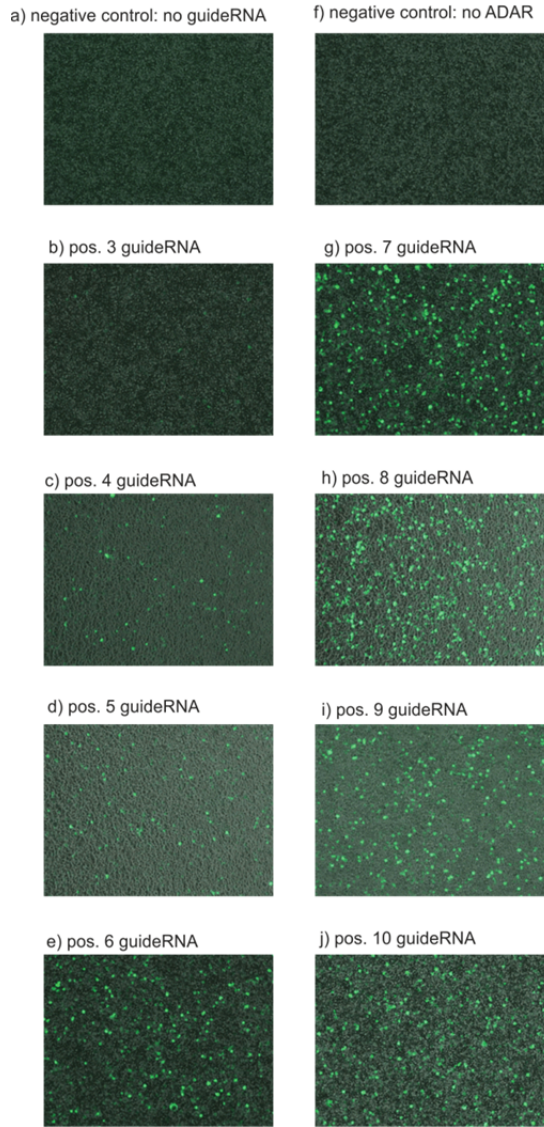


Figure S11: Effect of variable positions of the W58X eGFP R/G-gRNA towards the target adenosine in cell culture. Co-transfection of 300 ng W58X eGFP and ADAR2 plasmid together with 1300 ng of the tested position of the R/G-gRNA was performed in a 24-well plate format. The microscopic analysis and RNA isolation for sequence analysis was performed 48 hours post transfection. The R/G-gRNAs positions 3 until 10 are abbreviated by P3 – P10 (this equals 2-9 intervening nucleotides). Starting from the R/G-gRNA position 3 an increasing fluorescent signal and amount is visible until R/G-gRNA position 8. The R/G-gRNA position 9 and 10 showed a dropping fluorescent signal. The microscopic results are confirmed by the sequence analysis and demonstrate that R/G-gRNA position 8 was the most successful guideRNA to achieve maximum editing yields. Total magnification: 100x, GFP exposure time: 50 ms.



continuation Figure S11, replication of the positional effect.

Replication 1: 24-well format, 300 ng ADAR2 plasmid, 300 ng W58X plasmid, 1600 ng R/G-guideRNA plasmid, imaging 48 hrs post transfection, total magnification 100x, 50 ms exposure



Replication 2: exactly as replication 1, but 1300 ng R/G-guideRNA

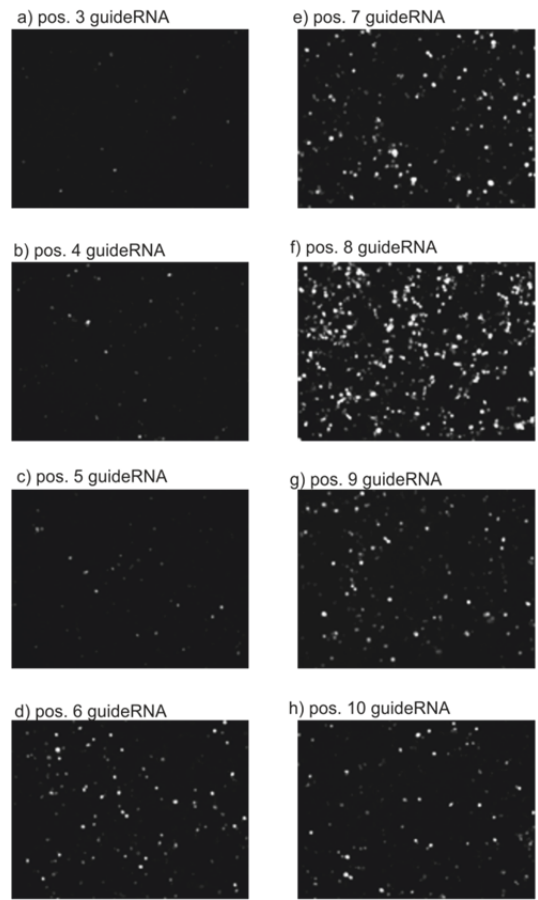
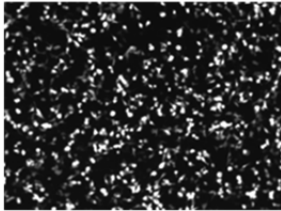
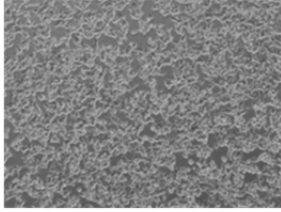
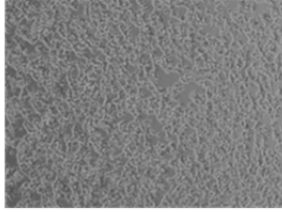


Figure S12: Effect of variable amounts of ADAR2 and R/G-gRNA plasmids in cell culture. Different amounts of R/G-gRNA P8 plasmid were transfected in 24-well plate format together with a constant amount of 300 ng W58X eGFP and ADAR2 plasmid (P8 equals 7 intervening nucleotides). Higher amounts of R/G-gRNA plasmid resulted in more and stronger fluorescence, as well as in higher editing levels. Total magnification: 100x, GFP exposure time: 50 ms.

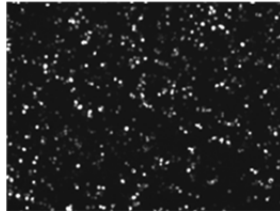
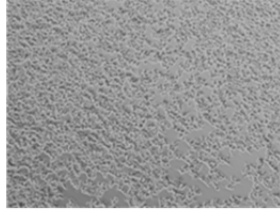
a) positive control
ADAR, GFP, gRNA



b) negative control
ADAR, W58X GFP, gRNA



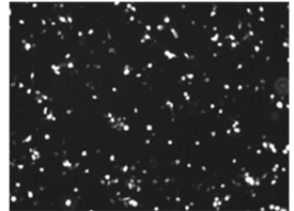
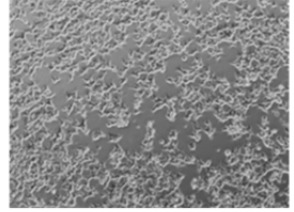
c) editing
650 ng gRNA



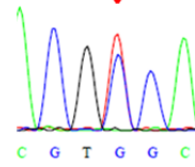
38%



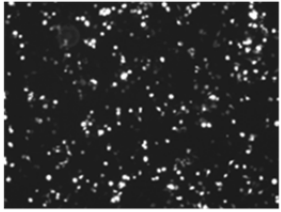
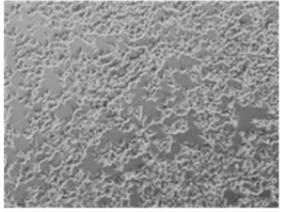
d) editing
750 ng gRNA



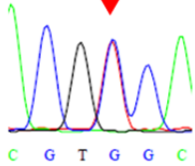
43%



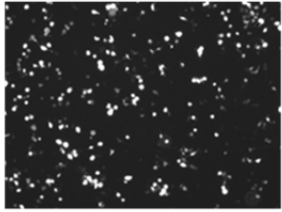
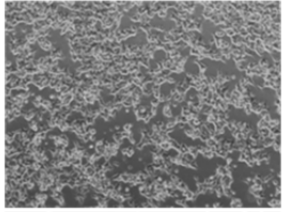
e) editing
1000 ng gRNA



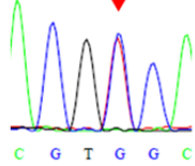
50%



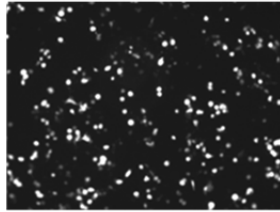
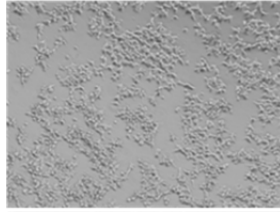
f) editing
1300 ng gRNA



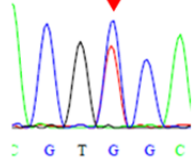
52%



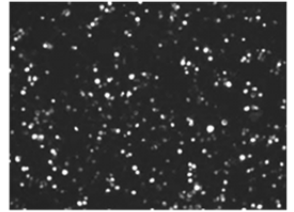
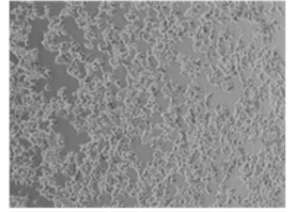
g) editing
1600 ng gRNA



57%



h) editing
2000 ng gRNA



52%

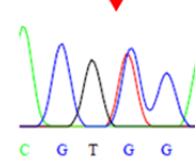


Figure S13: Prolongation of the editing time for the R/G-gRNA position 8 (position 8 equals 7 intervening nucleotides). The co-transfection experiment of 300 ng W58X eGFP plasmid, 300 ng of ADAR2 plasmid and 1300 ng or 1600 ng R/G-gRNA P8 plasmid was performed in a 24-well format. The editing efficiency was analyzed 24h, 48h, 72h and 96h post transfection. The positive control showed the strongest fluorescent signal for 48 hours of incubation. Shorter and longer incubation led to a reduced eGFP signal. For both chosen R/G-gRNA P8 plasmid amounts an increasing fluorescent signal and amount until 72h of incubation was visible. The eGFP intensity and amount of cells was declining after 96h of incubation. The sequence analysis confirmed the fluorescent microscopy: the editing yields increased until 72 hours of editing time and decreased after 96h again. Total magnification: 100x, GFP exposure time: 50 ms.

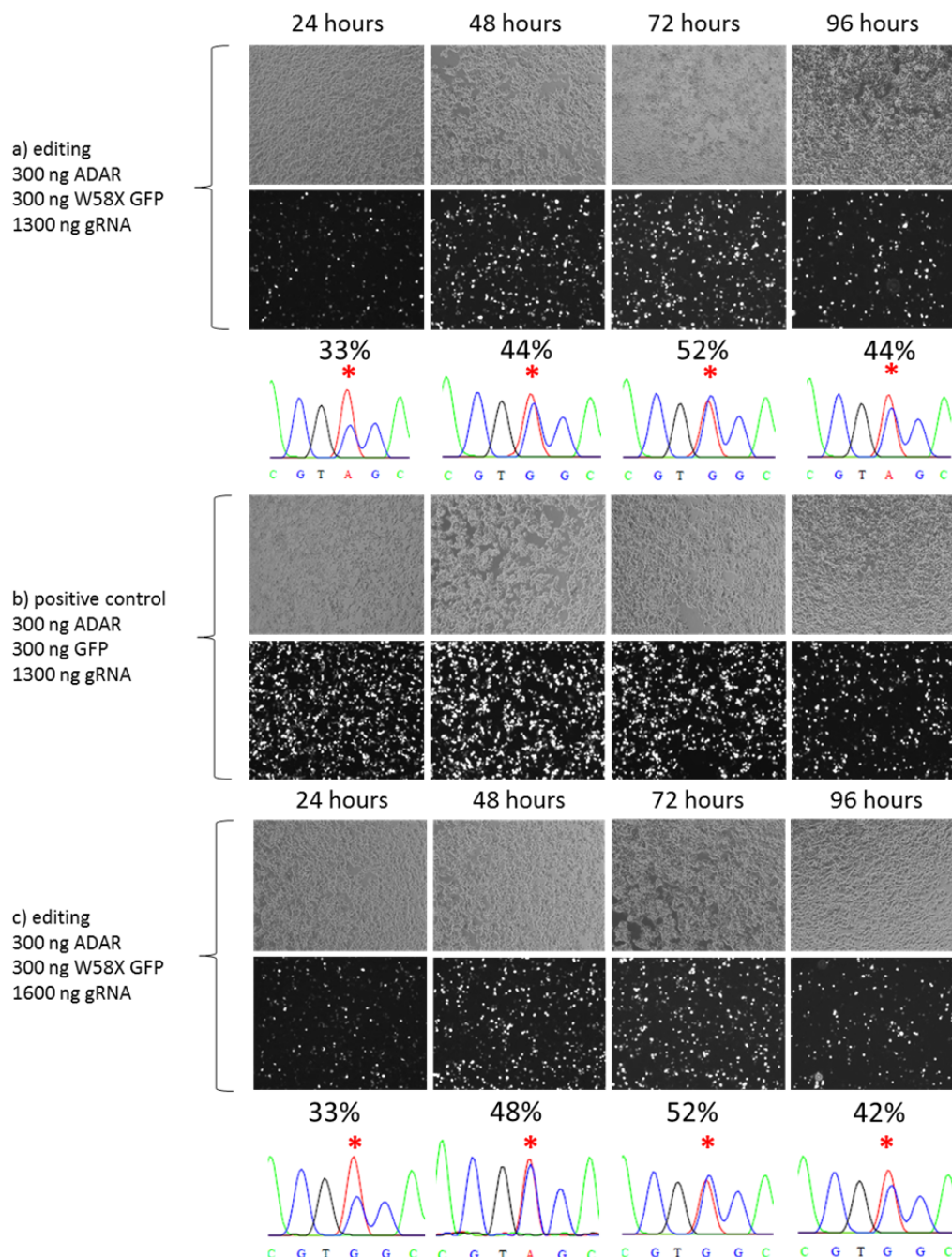


Figure S14: Effect of decreasing amounts of transfected ADAR2 plasmid on the editing yield and the off-target editing at position A381. In a 24-well format cells were co-transfected with 300 ng W58X eGFP and 1300 ng R/G-gRNA-P8 (P8 equals 7 intervening nucleotides) together with varying amounts of ADAR2 plasmid. The microscopic analysis and RNA isolation was carried out 48 h post transfection. The decrease of the ADAR2 plasmid down to 50 ng reduces the editing yield by 5 % compared to the starting concentration of 300 ng. The usage of 25 ng of ADAR2 plasmid markedly lowers the editing level down to 36% compared to 52% editing yield for 300 ng of ADAR2 plasmid. The reduction of the transfected ADAR2 plasmid amount led to a decrease at the off-target site A381 eGFP. Transfection of 100 ng or lower ADAR2 plasmid amount completely prevents the off-target editing. Total magnification: 100x, GFP exposure time: 50 ms.

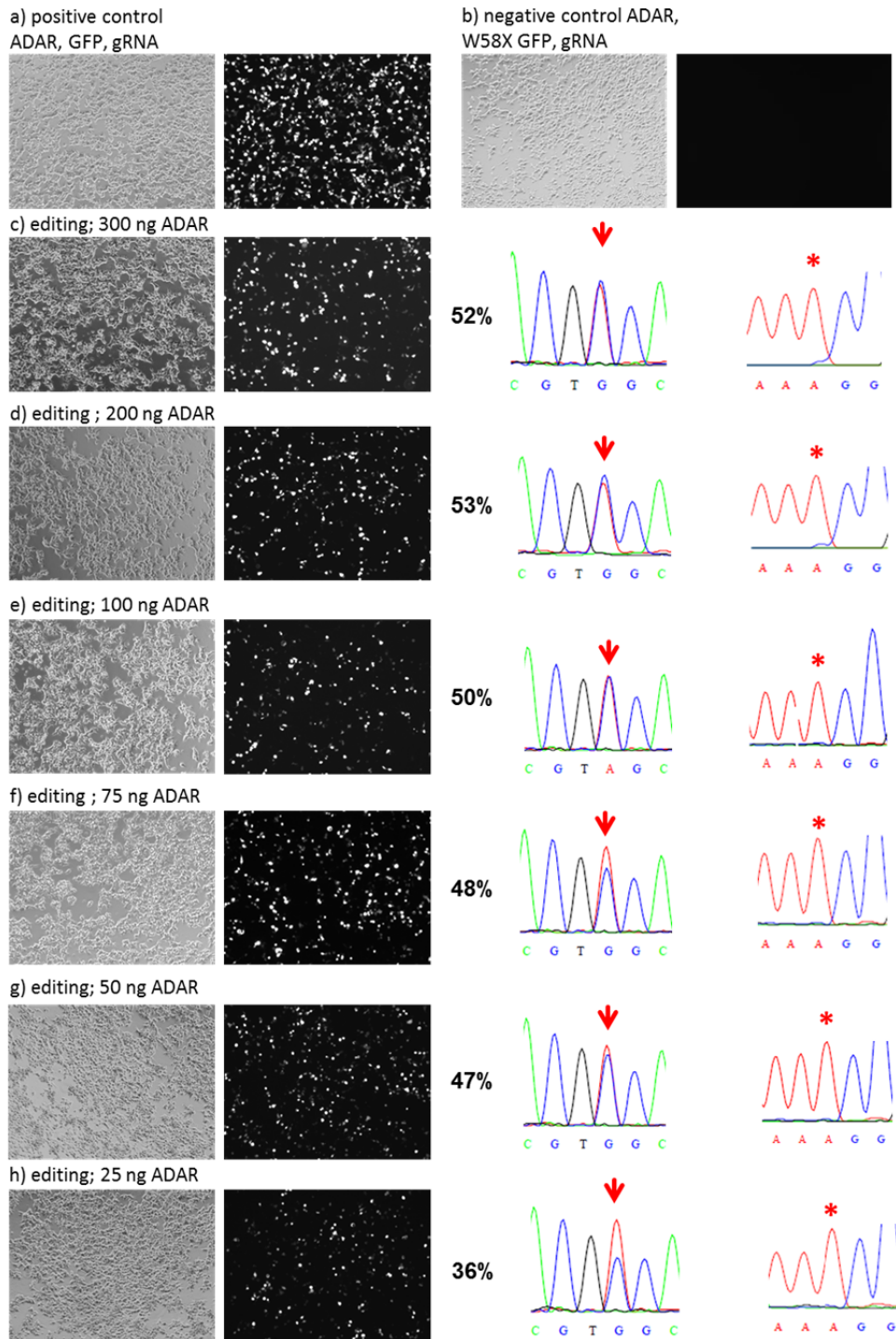


Figure S15 Editing depends on the position of the targeted adenosine to the R/G-motif. This was also found for editing of the R407Q site in PINK1 in 293 cells analog to the experiments shown in Figure 2D. (n = 3)

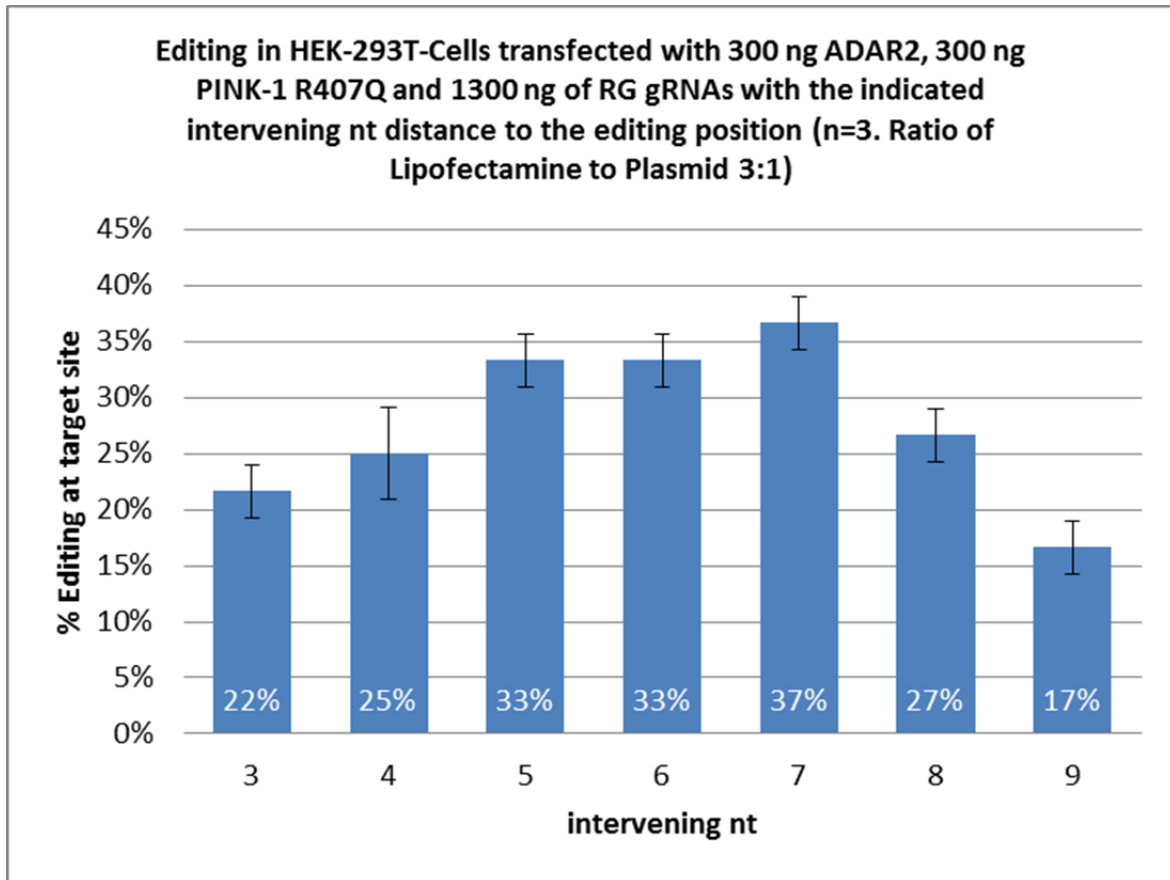
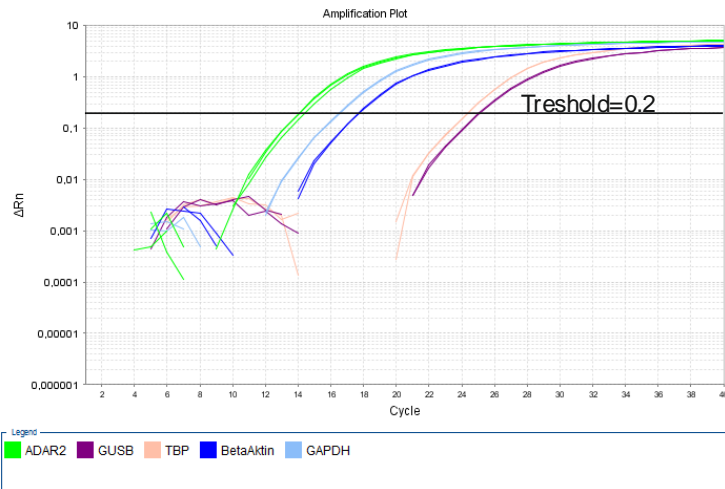


Figure S16. qPCR analysis of ADAR2 expression. The relative ADAR2 mRNA amount in 293T cells transiently transfected (trans.) with ADAR2 and 293T cells with a genomically integrated copy of ADAR2 controlled by a CMV tet-on promoter (integr.) was determined by quantitative real-time PCR (qPCR) after 24h (doxycycline induced expression of integr. ADAR2) and 48h (expression of trans. and integr. ADAR2). For this, RNA was extracted from cell lysates (RNeasy MinElute Kit, Qiagen). After DNaseI digestion (NEB) and reverse transcription (high capacity cDNA reverse transcription kit, Applied Biosystems), 20 ng cDNA was mixed with Fast SYBR Green Master Mix (Applied Biosystems) and analyzed by the 7500 Fast Real-Time PCR System (Applied Biosystems). **(A)** For determining gene expression, primers were designed for targeting ADAR2 and the housekeeping genes β -actin, glyceraldehyde-3-phosphate dehydrogenase (GAPDH), β -glucuronidase (GUSB) and TATA-box binding protein (TBP), **(B)** shows an example of the sybr green traces during qPCR. **(C)** qPCR of ADAR2 and the housekeeping gene was performed in triplicates and duplicates, respectively. The table displays the mean values of the cycles where the fluorescence crosses the threshold of 0.2 (ct values). **(D)** Based on these ct values, the expression of ADAR2 compared to housekeeping gene expression was determined in 293T cells after transient ADAR2 transfection or doxycycline induction by the delta ct equation. **(E)** To compare ADAR2 expression of ADAR2 transiently transfected 293T cells and ADAR2 genomically integrated 293T cells after 48h of expression, two methods were applied. In **method 1**, a calibration curve was generated from 1:5 dilutions of 20 ng cDNA of transiently transfected 293T cells (mean values from triplicates with standard derivation). For normalization, the corresponding ct (ADAR2) values were divided by the ct-value of β -actin for 20 ng cDNA. In **method 2**, the delta-delta ct method was used to calculate difference in ADAR2 expression.

(A) Primers for qPCR

Gene	Sequence (5' to 3')	Product size
ADAR2	fw.: CGGAGATCCTTGCTCAGATT	99 bp
	rev.: CCCTCGCTCTGATTTCTGAA	
β -actin	fw.: CGGGACCTGACTGACTAC	91 bp
	rev.: TAATGTCACGCACGATTTCC	
GAPDH	fw.: CAACAGCCTCAAGATCATCAG	96 bp
	rev.: CCTCCACGATACCAAAGTTG	
GUSB	fw.: ACCTGTTCAAGTTGGAAGTG	93 bp
	rev.: CACCTGGCACCTTAAGTTG	
TBP	fw.: CGGAGAGTTCTGGGATTGTA	90 bp
	rev.: GAAGTGCAATGGTCTTTAGGT	

(B) An example of the raw data for transient ADAR2 expression (300 ng) in 293T cells.



(C) Measured ct-values of all experiments. Values are averaged from three technical replicates for ADAR2 and two technical replicates for the housekeeping genes

	Sample	ct (ADAR2)	ct (β-actin)	ct (GAPDH)	ct (GUSB)	ct (TBP)
Tra	293T	25.061	18.089	16.88	24.829	24.174
	293T + 300ng ADAR2, 48h	14.195	17.710	16.532	25.08	24.356
Integr.	293-pcDNA5 + Dox, 24h	23.807	17.143	16.617	23.935	23.086
	293-ADAR2 without Dox	20.916	16.733	16.199	23.696	22.72
	293-ADAR2 + Dox, 24h	18.710	17.483	16.941	24.093	23.546
	293-ADAR2 + Dox, 48h	18.633	17.654	16.766	24.651	23.784

(D) Calculation of relative expression levels from the Δct values for ADAR2 versus four housekeeping genes

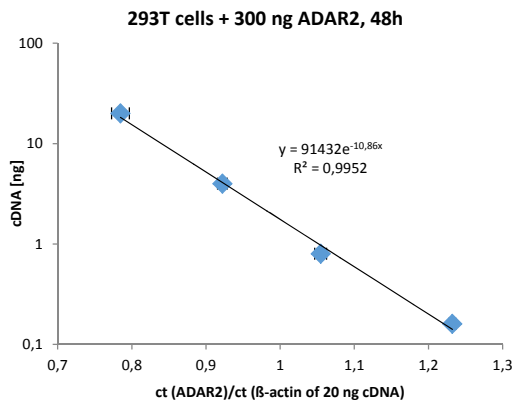
	Sample	β-actin	GAPDH	GUSB	TPB
Tra	293T	0.008	0.003	0.851	0.541
	293T + 300ng ADAR2, 48h	11.432	5.053	1891.087	1144.895
Integr.	293-pcDNA5	0.009	0.007	1.093	0.607
	293-ADAR2 without Dox	0.055	0.038	6.869	3.492
	293-ADAR2 + 10 ng/ml Dox, 24h	0.427	0.293	41.730	28.562
	293-ADAR2 + 10ng/ml Dox, 48h	0.507	0.274	64.804	35.531

relative expression = $2^{-\Delta ct}$, with $\Delta ct = ct(ADAR2) - ct(\text{housekeeping gene})$

(E) Comparison of transient versus genomic ADAR2 expression

Method 1

A calibration curve was taken for four different ADAR2 (transient expression) dilutions (1x, 5x, 25x, 125x). Plotted is the ADAR2 ct values normalized by the ct value for beta-actin of the undiluted sample versus the amount of cDNA.



From the regression curve of the calibration plot, the relative expression of genomically expressed ADAR2 was calculated. For this genomically expressed ADAR2 was normalized to beta-actin.

$ct(ADAR2)/ct(\beta\text{-actin}) = 1.055$ [for the experiment with 293-ADAR2 + 10 ng/ml Dox, 48h] (20 ng cDNA)

from $x = 1.055$ one can calculate $y = 0.967$, and the genomic expression to **be 20fold** below that of the transient expression

Method 2

Here we estimated the relative expression level of ADAR2 transient versus genomic by the delta-delta ct method applying the following equation.

$$\Delta\Delta ct = (ct(ADAR2) - ct(\beta\text{actin}))_{\text{transient}} - ((ct(ADAR2) - ct(\beta\text{actin}))_{\text{genomic}})$$

$$\text{relative expression} = 2^{-\Delta\Delta ct}$$

relative expression = 25.795, meaning genomic expression is approx. **26fold** below transient expression

Figure S17 Dependency of the editing yield with genomically encoded ADAR2 with varying amounts of guideRNA. Shown are the averaged yields and standard deviation for n=3.

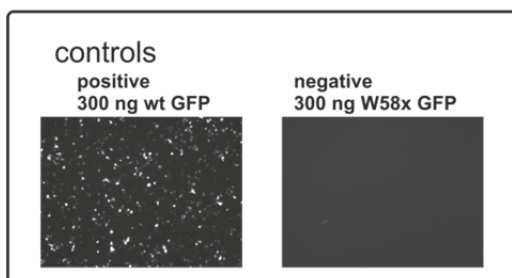
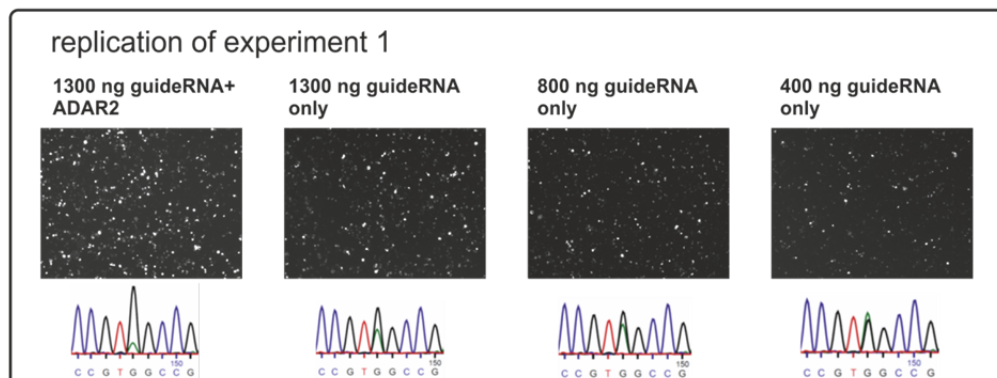
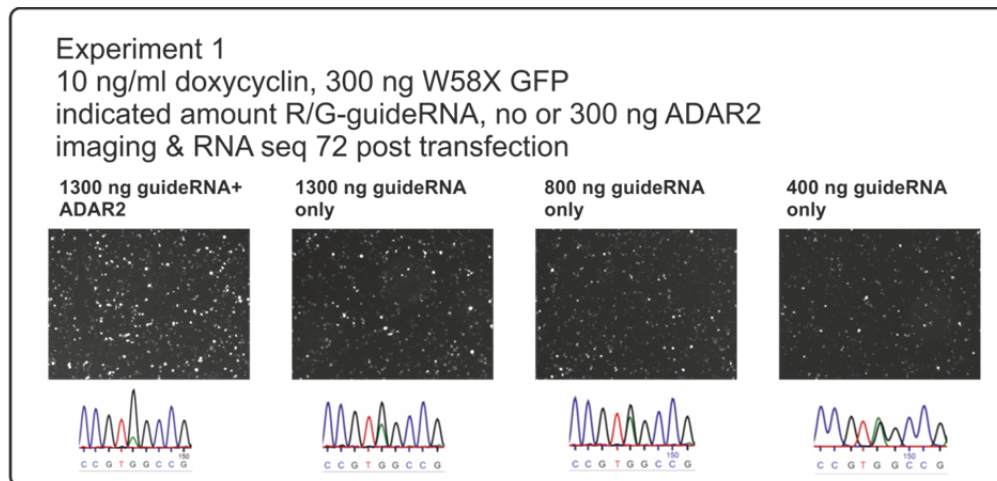
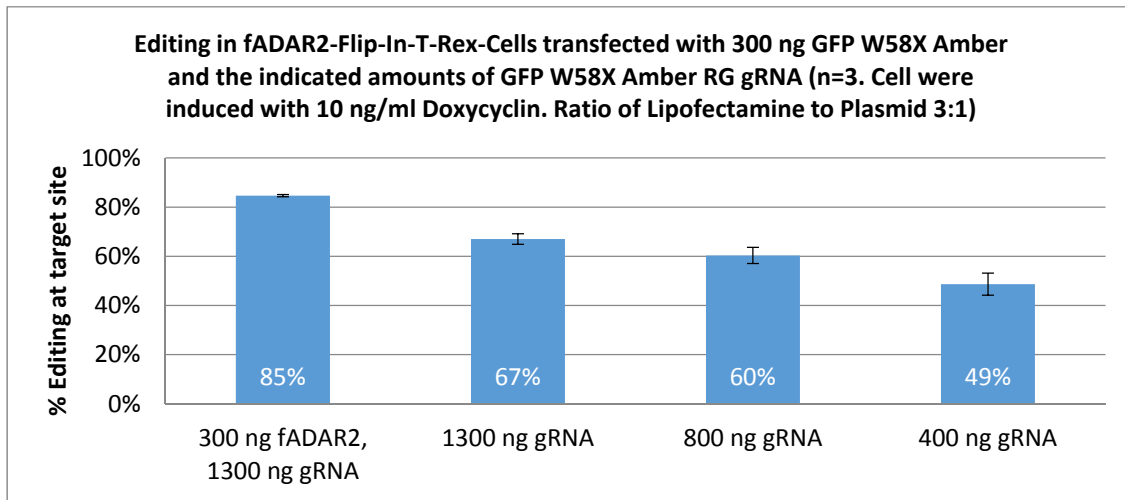


Figure S18. Full sequencing trace corresponding to the trace shown in Figure 3A, b), 1300 ng guideRNA



Editing of endogenous transcripts

Editing experiments have been carried out in duplicates exactly as described before (without further optimization), but without co-transfection of a target or reporter gene. 293 cells have been transfected with 300 ng ADAR2 and of the respective 1300 ng R/G-guideRNA in 24 well format with lipofectamine 2000, and were harvested 48h post transfection. 293-ADAR2 flip-in cells have been induced with doxycycline (10 ng/ml), then 1300 ng of the respective R/G-guideRNA was transfected with lipofectamine 2000. 72 h after transfection of the guideRNA total RNA was isolated, RT-PCR with transcript-specific primers have been done to obtain the RNA sequencing traces. The respective guideRNA sequences are listed in Table S1. Primers for RT-PCR are given in Table S2.

Target sites for site-directed RNA editing on six genes:

cDNA sequence of β -actin

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      10      20      30      40      50      60
1      ACCGCCGAGACCGCGTCCGCCCCGCGAGCACAGAGCCTCGCCTTTGCCGATCCGCCGCC
1
      70      80      90      100     110     120
61     GTCCACACCCCGCCGAGCTCACCATGGATGATGATATCGCCGCGCTCGTCGTCGACAAC
21                                     M D D D I A A L V V D N
      130     140     150     160     170     180
121    GGCTCCGGCATGTGCAAGGCCGGCTTCGCGGGCGACGATGCCCCCGGGCCGTCTTCCCC
41    G S G M C K A G F A G D D A P R A V F P
      190     200     210     220     230     240
181    TCCATCGTGGGGCGCCCCAGGCACCAGGGCGTGATGGTGGGCATGGGTTCAGAAGGATTCC
61    S I V G R P R H Q G V M V G M G Q K D S
      250     260     270     280     290     300
241    TATGTGGGCGACGAGGCCAGAGCAAGAGAGGCATCCTCACCTGAAGTACCCCATCGAG
81    Y V G D E A Q S K R G I L T L K Y P I E
      310     320     330     340     350     360
301    CACGGCATCGTCACCAACTGGGACGACATGGAGAAAATCTGGCACCACACCTTCTACAAT
101   H G I V T N W D D M E K I W H H T F Y N
      370     380     390     400     410     420
361    GAGCTGCGTGTGGCTCCCGAGGAGCACCCCGTGCTGCTGACCGAGGCCCCCTGAACCCC
121   E L R V A P E E H P V L L T E A P L N P
      430     440     450     460     470     480
421    AAGGCCAACCGCGAGAAGATGACCCAGATCATGTTTGAGACCTTCAACACCCCAGCCATG
141   K A N R E K M T Q I M F E T F N T P A M
      490     500     510     520     530     540
481    TACGTTGCTATCCAGGCTGTGCTATCCCTGTACGCTCTGGCCGTACCACTGGCATCGTG
161   Y V A I Q A V L S L Y A S G R T T G I V
      550     560     570     580     590     600
541    ATGGACTCCGGTGACGGGGTCACCCACACTGTGCCCATCTACGAGGGGTATGCCCTCCCC
181   M D S G D G V T H T V P I Y E G Y A L P
      610     620     630     640     650     660
601    CATGCCATCCTGCGTCTGGACCTGGCTGGCCGGGACCTGACTGACTACCTCATGAAGATC
201   H A I L R L D L A G R D L T D Y L M K I
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661          670          680          690          700          710          720
221 CTCACCGAGCGCGGCTACAGCTTCACCACCACGGCCGAGCGGAAAATCGTGCGTGACATT
    L T E R G Y S F T T T A E R E I V R D I

          730          740          750          760          770          780
721 AAGGAGAAGCTGTGCTACGTGCGCCCTGGACTTCGAGCAAGAGATGGCCACGGCTGCTTCC
241 K E K L C Y V A L D F E Q E M A T A A S

          790          800          810          820          830          840
781 AGCTCCTCCCTGGAGAAGAGCTACGAGCTGCCTGACGGCCAGGTCATCACCATTGGCAAT
261 S S S L E K S Y E L P D G Q V I T I G N

          850          860          870          880          890          900
841 GAGCGGTTCCGCTGCCCTGAGGCACTCTTCCAGCCTTCCTTCCTGGGCATGGAGTCTGT
281 E R F R C P E A L F Q P S F L G M E S C

          910          920          930          940          950          960
901 GGCATCCACGAAACTACCTTCAACTCCATCATGAAGTGTGACGTGGACATCCGCAAAGAC
301 G I H E T T F N S I M K C D V D I R K D

          970          980          990          1000          1010          1020
961 CTGTACGCCAACACAGTGCTGTCTGGCGGCACCACCATGTACCCTGGCATTGCCGACAGG
321 L Y A N T V L S G G T T M Y P G I A D R

          1030          1040          1050          1060          1070          1080
1021 ATGCAGAAGGAGATCACTGCCCTGGCACCCAGCACAAATGAAGATCAAGATCATTGCTCCT
341 M Q K E I T A L A P S T M K I K I I A P

          1090          1100          1110          1120          1130          1140
1081 CCTGAGCGCAAGTACTCCGTGTGGATCGGCGGCTCCATCCTGGCCTCGCTGTCCACCTTC
361 P E R K Y S V W I G G S I L A S L S T F

          1150          1160          1170          1180          1190          1200
1141 CAGCAGATGTGGATCAGCAAGCAGGAGTATGACGAGTCCGGCCCCTCCATCGTCCACCGC
381 Q Q M W I S K Q E Y D E S G P S I V H R

          1210          1220          1230          1240          1250          1260
1201 AAATGCTTCTAGGCGGACTATGACTTAGTTGCGTTACACCCTTTCTTGACAAAACCTAAC
401 K C F *          target 1

          1270          1280          1290          1300          1310          1320
1261 TTGCGCAGAAAACAAGATGAGATTGGCATGGCTTTATTTGTTTTTTTTGTTTTGTTTTGG
421

          1330          1340          1350          1360          1370          1380
1321 TTTTTTTTTTTTTTTTTTGGCTTGACTCAGGATTTAAAAACTGGAACGGTGAAGGTGACAGC
441

          1390          1400          1410          1420          1430          1440
1381 AGTCGGTTGGAGCGAGCATCCCCAAAGTTCACAATGTGGCCGAGGACTTTGATTGCACA
461

          1450          1460          1470          1480          1490          1500
1441 TTGTTGTTTTTTTTTAATAGTCATTCCAAATATGAGATGCGTTGTTACAGGAAGTCCCTTGC
481          target 2

          1510          1520          1530          1540          1550          1560
1501 CATCCTAAAAGCCACCCCACTTCTCTCTAAGGAGAATGGCCCAGTCCCTCCTCCCAAGTCCA
501

```

1561 1570 1580 1590 1600 1610 1620
 521 CACAGGGGAGGTGATAGCATTGCTTTCGTGTAATAATGTAATGCAAAATTTTTTTAATC
 target 3

 1621 1630 1640 1650 1660 1670 1680
 541 TTCGCCTTAATACTTTTTTATTTTTGTTTTATTTTGAATGATGAGCCTTCGTGCCCCCCT

 1681 1690 1700 1710 1720 1730 1740
 561 TCCCCCTTTTTTGTCCCCAACTTGAGATGTATGAAGGCTTTTGGTCTCCCTGGGAGTGG

 1741 1750 1760 1770 1780 1790 1800
 581 GTGGAGGCAGCCAGGGCTTACCTGTACTGACTTGAGACCAGTTGAATAAAAAGTGCACA

 1801 1810 1820 1830 1840 1850
 601 CCTTAAAAATGAAA

cDNA sequence of GAPDH

1 10 20 30 40 50 60
 1 GCCTCAAGACCTTGGGCTGGGACTGGCTGAGCCTGGCGGGAGGCGGGTCCGAGTCACCG
 1

 61 70 80 90 100 110 120
 20 CCTGCCGCGCGCCCCGGTTTCTATAAATGAGCCCGCAGCCTCCCGCTTCGCTCTCTG

 121 130 140 150 160 170 180
 40 CTCCTCCTGTTTCGACAGTCAGCCGCATCTTCTTTTGCCTCGCCAGCCGAGCCACATCGCT

 181 190 200 210 220 230 240
 60 CAGACACCATGGGGAAGGTGAAGGTGCGGAGTCAACGGATTTGGTTCGTATTGGGCGCCTGG
 M G K V K V G V N G F G R I G R L

 241 250 260 270 280 290 300
 80 TCACCAGGGCTGCTTTTAACTCTGGTAAAGTGGATATTGTTGCCATCAATGACCCCTTCA
 V T R A A F N S G K V D I V A I N D P F

 301 310 320 330 340 350 360
 100 TTGACCTCAACTACATGGTTTACATGTTCCAATATGATTCCACCCATGGCAAATTCATG
 I D L N Y M V Y M F Q Y D S T H G K F H

 361 370 380 390 400 410 420
 120 GCACCGTCAAGGCTGAGAACGGGAAGCTTGTCAATGGAAATCCCATCACCATCTTCC
 G T V K A E N G K L V I N G N P I T I F

 421 430 440 450 460 470 480
 140 AGGAGCGAGATCCCTCCAAAATCAAGTGGGGCGATGCTGGCGCTGAGTACGTCGTGGAGT
 Q E R D P S K I K W G D A G A E Y V V E

 481 490 500 510 520 530 540
 160 CCACTGGCGTCTTCACCACCATGGAGAAGGCTGGGGCTCATTTGCAGGGGGAGCCAAAA
 S T G V F T T M E K A G A H L Q G G A K

 541 550 560 570 580 590 600
 180 GGGTCATCATCTCTGCCCCCTCTGCTGATGCCCCATGTTTCGTTCATGGGTGTGAACCATG
 R V I I S A P S A D A P M F V M G V N H

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610      620      630      640      650      660
601  AGAAGTATGACAACAGCCTCAAGATCATCAGCAATGCCTCCTGCACCACCAACTGCTTAG
200  E K Y D N S L K I I S N A S C T T N C L

670      680      690      700      710      720
661  CACCCCTGGCCAAGGTCATCCATGACAACCTTTGGTATCGTGGAAGGACTCATGACCACAG
220  A P L A K V I H D N F G I V E G L M T T

730      740      750      760      770      780
721  TCCATGCCATCACTGCCACCCAGAAGACTGTGGATGGCCCCTCCGGGAAACTGTGGCGTG
240  V H A I T A T Q K T V D G P S G K L W R

790      800      810      820      830      840
781  ATGGCCGCGGGGCTCTCCAGAACATCATCCCTGCCTCTACTGGCGCTGCCAAGGCTGTGG
260  D G R G A L Q N I I P A S T G A A K A V

850      860      870      880      890      900
841  GCAAGGTCATCCCTGAGCTGAACGGGAAGCTCACTGGCATGGCCTTCCGTGTCCCCACTG
280  G K V I P E L N G K L T G M A F R V P T

910      920      930      940      950      960
901  CCAACGTGTCAAGTGGTGGACCTGACCTGCCGTCTAGAAAAACCTGCCAAATATGATGACA
300  A N V S V V D L T C R L E K P A K Y D D

970      980      990      1000      1010      1020
961  TCAAGAAGGTGGTGAAGCAGGCGTCGGAGGGCCCCCTCAAGGGCATCCTGGGCTACACTG
320  I K K V V K Q A S E G P L K G I L G Y T

1030      1040      1050      1060      1070      1080
1021  AGCACCAGGTGGTCTCCTCTGACTTCAACAGCGACACCCACTCCTCCACCTTTGACGCTG
340  E H Q V V S S D F N S D T H S S T F D A

1090      1100      1110      1120      1130      1140
1081  GGGCTGGCATTGCCCTCAACGACCACTTTGTCAAGCTCATTTCCTGGTATGACAACGAAT
360  G A G I A L N D H F V K L I S W Y D N E

1150      1160      1170      1180      1190      1200
1141  TTGGCTACAGCAACAGGGTGGTGGACCTCATGGCCCACATGGCCTCCAAGGAGTAAGACC
380  F G Y S N R V V D L M A H M A S K E *

1210      1220      1230      1240      1250      1260
1201  CCTGGACCACCAGCCCCAGCAAGAGCACAAGAGGAAGAGAGACCCTCACTGCTGGGGA
400

1270      1280      1290      1300      1310      1320
1261  GTCCCTGCCCACTCAGTCCCCCACCACACTGAATCTCCCCTCCTCACAGTTGCCATGTA
420                                     target 1

1330      1340      1350      1360      1370      1380
1321  GACCCCTTGAAGAGGGGAGGGGCCTAGGGAGCCGCACCTTGTCATGTACCATCAATAAAG
440                                     target 2

1390      1400      1410      1420
1381  TACCCTGTGCTCAACCAGTTAAAAAAAAAAAAAAAAAAAAAAAAAAAA
460

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cDNA sequence of GPI

10 20 30 40 50 60

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1      AATAGCCCTTACCACCAGCAGACACACATCATCTGTTGTA CTTGCTTATTTGGCACATAT
1
          70          80          90          100          110          120
61     GTATCCACAGCGCCTAGAACACTGCCTGTAACGTGGAAGGTGTTTCGATCTATAGAGTTTT
20
          130          140          150          160          170          180
121    GTCGAATGAATGAATGAAGCCGACTAGTGCACAGGGAGTGCAGCGGCGGATGGTAGCTC
40                                          M V A
          190          200          210          220          230          240
181    TCTGCAGCCTCCAACACCTGGGCTCCAGTGATCCCCGGGCTCTGCCCACCTCCCCACTG
60    L C S L Q H L G S S D P R A L P T L P T
          250          260          270          280          290          300
241    CCACTTCCGGGCAGAGGCCAGCAAAGCGGCGGCAAGAGTCCCGCCATGGCCGCTCTCA
80    A T S G Q R P A K R R R K S P A M A A L
          310          320          330          340          350          360
301    CCCGGGACCCCCAGTTCCAGAAGCTGCAGCAATGGTACCGCGAGCACCGCTCCGAGCTGA
100   T R D P Q F Q K L Q Q W Y R E H R S E L
          370          380          390          400          410          420
361    ACCTGCGCCGCTCTTCGATGCCAACAAGGACCGCTTCAACCACTTCAGCTTGACCCTCA
120   N L R R L F D A N K D R F N H F S L T L
          430          440          450          460          470          480
421    ACACCAACCATGGGCATATCCTGGTGGATTACTCCAAGAACCTGGTGACGGAGGACGTGA
140   N T N H G H I L V D Y S K N L V T E D V
          490          500          510          520          530          540
481    TCGGATGCTGGTGGACTTGGCCAAGTCCAGGGGCGTGGAGGCCCGCCGGGAGCGGATGT
160   M R M L V D L A K S R G V E A A R E R M
          550          560          570          580          590          600
541    TCAATGGTGAGAAGATCAACTACACCGAGGGTCGAGCCGTGCTGCACGTGGCTCTGCGGA
180   F N G E K I N Y T E G R A V L H V A L R
          610          620          630          640          650          660
601    ACCGGTCAAACACACCCATCCTGGTGGACCGGCAAGGATGTGATGCCAGAGGTCAACAAGG
200   N R S N T P I L V D G K D V M P E V N K
          670          680          690          700          710          720
661    TTCTGGACAAGATGAAGTCTTTCTGCCAGGGACCCCTCATGGTGA C TGAAGCCCTTAAGC
220   V L D K M K S F C Q G P L M V T E A L K
          730          740          750          760          770          780
721    CATACTCTTCAGGAGGTCCCCGCGTCTGGTATGTCTCCAACATTGATGGA A CT CACAT TG
240   P Y S S G G P R V W Y V S N I D G T H I
          790          800          810          820          830          840
781    CCAAAACCCCTGGCCCAGCTGAACCCCGAGTCCCTCCCTGTTCA TCAT TGCCTCCAAGACCT
260   A K T L A Q L N P E S S L F I I A S K T
          850          860          870          880          890          900
841    TTACTACCCAGGAGACCATCACGAATGCAGAGACGGCGAAGGAGTGGTTTCTCCAGGCGG
280   F T T Q E T I T N A E T A K E W F L Q A
          910          920          930          940          950          960
901    CCAAGGATCCTTCTGCAGTGGCGAAGCACTTTGTTGCCCTGTCTACTA ACACAACCAAAG

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300 A K D P S A V A K H F V A L S T N T T K
 970 980 990 1000 1010 1020
 961 TGAAGGAGTTTGGAAATTGACCCCTCAAACATGTTTCGAGTTCTGGGATTGGGTGGGAGGAC
 320 V K E F G I D P Q N M F E F W D W V G G
 1030 1040 1050 1060 1070 1080
 1021 GCTACTCGCTGTGGTCGGCCATCGGACTCTCCATTGCCCTGCACGTGGGTTTTGACAAC
 340 R Y S L W S A I G L S I A L H V G F D N
 1090 1100 1110 1120 1130 1140
 1081 TCGAGCAGCTGCTCTCGGGGGCTCACTGGATGGACCAGCACTTCCGCACGACGCCCTGG
 360 F E Q L L S G A H W M D Q H F R T T P L
 1150 1160 1170 1180 1190 1200
 1141 AGAAGAACGCCCCGTCTTGCTGGCCCTGCTGGGTATCTGGTACATCAACTGCTTTGGGT
 380 E K N A P V L L A L L G I W Y I N C F G
 1210 1220 1230 1240 1250 1260
 1201 GTGAGACACACGCCATGCTGCCCTATGACCAGTACCTGCACCGCTTTGCTGCGTACTTCC
 400 C E T H A M L P Y D Q Y L H R F A A Y F
 1270 1280 1290 1300 1310 1320
 1261 AGCAGGGCGACATGGAGTCCAATGGGAAATACATCACCAAATCTGGAACCCGTGTGGACC
 420 Q Q G D M E S N G K Y I T K S G T R V D
 1330 1340 1350 1360 1370 1380
 1321 ACCAGACAGGCCCCATTGTGTGGGGGGAGCCAGGGACCAATGGCCAGCATGCTTTTTACC
 440 H Q T G P I V W G E P G T N G Q H A F Y
 1390 1400 1410 1420 1430 1440
 1381 AGCTCATCCACCAAGGCACCAAGATGATACCCTGTGACTTCCTCATCCCGGTCCAGACCC
 460 Q L I H Q G T K M I P C D F L I P V Q T
 1450 1460 1470 1480 1490 1500
 1441 AGCACCCCATACGGAAGGGTCTGCATCACAAGATCCTCCTGGCCAACTTCTTGCCCA
 480 Q H P I R K G L H H K I L L A N F L A Q
 1510 1520 1530 1540 1550 1560
 1501 CAGAGGCCCTGATGAGGGGAAAATCGACGGAGGAGGCCGAAAGGAGCTCCAGGCTGCGG
 500 T E A L M R G K S T E E A R K E L Q A A
 1570 1580 1590 1600 1610 1620
 1561 GCAAGAGTCCAGAGGACCTTGAGAGGCTGCTGCCACATAAGGTCTTTGAAGGAAATCGCC
 520 G K S P E D L E R L L P H K V F E G N R
 1630 1640 1650 1660 1670 1680
 1621 CAACCAACTCTATTGTGTTACCAAGCTCACACCATTTCATGCTTGGAGCCTTGGTCGCCA
 540 P T N S I V F T K L T P F M L G A L V A
 1690 1700 1710 1720 1730 1740
 1681 TGTATGAGCACAAGATCTTCGTTTCAGGGCATCATCTGGGACATCAACAGCTTTGACCAGT
 560 M Y E H K I F V Q G I I W D I N S F D Q
 1750 1760 1770 1780 1790 1800
 1741 GGGGAGTGGAGCTGGGAAAGCAGCTGGCTAAGAAAATAGAGCCTGAGCTTGATGGCAGTG
 580 W G V E L G K Q L A K K I E P E L D G S
 1810 1820 1830 1840 1850 1860
 1801 CTCAAGTGACCTCTCACGACGCTTCTACCAATGGGCTCATCAACTTCATCAAGCAGCAGC
 600 A Q V T S H D A S T N G L I N F I K Q Q

1861 1870 1880
 GCGAGGCCAGAGTCCAATAA
 620 R E A R V Q *

cDNA sequence of GusB

1 10 20 30 40 50 60
 GTCTCTCAACCAAGATGGCGCGGATGGCTTCAGGCGCATCACGACACCGGCGCGTTCACGCG
 1

61 70 80 90 100 110 120
 ACCCGCCCTACGGGCACCTCCC CGCTTTTCTTAGCGCCGCAGACGGTGGCCGAGCGGGG
 20

121 130 140 150 160 170 180
 GACCGGGAAGCATGGCCCCGGGGTTCGGCGGTTGCCTGGGCGGCGCTCGGGCCGTTGTTGT
 40 M A R G S A V A W A A L G P L L

181 190 200 210 220 230 240
 GGGGCTGCGCGCTGGGGCTGCAGGGCGGGATGCTGTACCCCCAGGAGAGCCCGTTCGCGGG
 60 W G C A L G L Q G G M L Y P Q E S P S R

241 250 260 270 280 290 300
 AGTGCAAGGAGCTGGACGGCCTCTGGAGCTTCCGCGCCGACTTCTCTGACAACCGACGCC
 80 E C K E L D G L W S F R A D F S D N R R

301 310 320 330 340 350 360
 GGGGCTTCGAGGAGCAGTGGTACCGGCGGCCGCTGTGGGAGTCAGGCCCCACCGTGGACA
 100 R G F E E Q W Y R R P L W E S G P T V D

361 370 380 390 400 410 420
 TGCCAGTTCCTCCAGCTTCAATGACATCAGCCAGGACTGGCGTCTGCGGCATTTTGTTCG
 120 M P V P S S F N D I S Q D W R L R H F V

421 430 440 450 460 470 480
 GCTGGGTGTGGTACGAACGGGAGGTGATCCTGCCGAGCGATGGACCCAGGACCTGCGCA
 140 G W V W Y E R E V I L P E R W T Q D L R

481 490 500 510 520 530 540
 CAAGAGTGGTGTGAGGATTGGCAGTGCCCATTCCTATGCCATCGTGTGGGTGAATGGGG
 160 T R V V L R I G S A H S Y A I V W V N G

541 550 560 570 580 590 600
 TCGACACGCTAGAGCATGAGGGGGGCTACCTCCCCTTCGAGGCCGACATCAGCAACCTGG
 180 V D T L E H E G G Y L P F E A D I S N L

601 610 620 630 640 650 660
 TCCAGGTGGGGCCCCTGCCCTCCCGGCTCCGAATCACTATCGCCATCAACAACACACTCA
 200 V Q V G P L P S R L R I T I A I N N T L

661 670 680 690 700 710 720
 CCCCCACCACCTGCCACCAGGGACCATCCAATACCTGACTGACACCTCCAAGTATCCCA
 220 T P T T L P P G T I Q Y L T D T S K Y P

721 730 740 750 760 770 780
 AGGGTTACTTTGTCCAGAACACATATTTTGACTTTTTCAACTACGCTGGACTGCAGCGGT
 240 K G Y F V Q N T Y F D F F N Y A G L Q R

781 790 800 810 820 830 840
 CTGTACTTCTGTACACGACACCCACCACCTACATCGATGACATCACCGTCCACCACCGCG

260 S V L L Y T T P T T Y I D D I T V T T S
 850 860 870 880 890 900
 841 TGGAGCAAGACAGTGGGCTGGTGAATTACCAGATCTCTGTCAAGGGCAGTAACCTGTTCA
 280 V E Q D S G L V N Y Q I S V K G S N L F
 910 920 930 940 950 960
 901 AGTTGGAAGTGCCTCTTTTGGATGCAGAAAACAAAGTCGTGGCGAATGGGACTGGGACCC
 300 K L E V R L L D A E N K V V A N G T G T
 970 980 990 1000 1010 1020
 961 AGGGCCAACCTTAAGGTGCCAGGTGTCAGCCTCTGGTGGCCGTACCTGATGCACGAACGCC
 320 Q G Q L K V P G V S L W W P Y L M H E R
 1030 1040 1050 1060 1070 1080
 1021 CTGCCTATCTGTATTTCATTGGAGGTGCAGCTGACTGCACAGACGTCACCTGGGGCCTGTGT
 340 P A Y L Y S L E V Q L T A Q T S L G P V
 1090 1100 1110 1120 1130 1140
 1081 CTGACTTCTACACACTCCCTGTGGGGATCCGCACTGTGGCTGTCACCAAGAGCCAGTTCC
 360 S D F Y T L P V G I R T V A V T K S Q F
 1150 1160 1170 1180 1190 1200
 1141 TCATCAATGGGAAACCTTTTCTATTTCCACGGTGTCAACAAGCATGAGGATGCGGACATCC
 380 L I N G K P F Y F H G V N K H E D A D I
 1210 1220 1230 1240 1250 1260
 1201 GAGGGAAGGGCTTCGACTGGCCGCTGCTGGTGAAGGACTTCAACCTGCTTCGCTGGCTTG
 400 R G K G F D W P L L V K D F N L L R W L
 1270 1280 1290 1300 1310 1320
 1261 GTGCCAACGCTTTCCGTACCAGCCACTACCCCTATGCAGAGGAAGTGATGCAGATGTGTG
 420 G A N A F R T S H Y P Y A E E V M Q M C
 1330 1340 1350 1360 1370 1380
 1321 ACCGCTATGGGATTGTGGTCATCGATGAGTGTCCCGCGTGGGCCTGGCGCTGCCGAGT
 440 D R Y G I V V I D E C P G V G L A L P Q
 1390 1400 1410 1420 1430 1440
 1381 TCTTCAACAACGTTTCTCTGCATCACCACATGCAGGTGATGGAAGAAGTGGTGCCTAGGG
 460 F F N N V S L H H H M Q V M E E V V R R
 1450 1460 1470 1480 1490 1500
 1441 ACAAGAACCACCCCGGGTCGTGATGTGGTCTGTGGCCAACGAGCCTGCGTCCCACCTAG
 480 D K N H P A V V M W S V A N E P A S H L
 1510 1520 1530 1540 1550 1560
 1501 AATCTGCTGGCTACTACTTTGAAGATGGTGTGATCGCTCACACCAAATCCTTGGACCCCTCCC
 500 E S A G Y Y L K M V I A H T K S L D P S
 1570 1580 1590 1600 1610 1620
 1561 GGCCTGTGACCTTTGTGAGCAACTCTAACTATGCAGCAGACAAGGGGGCTCCGTATGTGG
 520 R P V T F V S N S N Y A A D K G A P Y V
 1630 1640 1650 1660 1670 1680
 1621 ATGTGATCTGTTTGAACAGCTACTACTCTTGGTATCACGACTACGGGCACCTGGAGTTGA
 540 D V I C L N S Y Y S W Y H D Y G H L E L
 1690 1700 1710 1720 1730 1740
 1681 TTCAGCTGCAGCTGGCCACCCAGTTTGTGAACTGGTATAAGAAGTATCAGAAGCCCATTA
 560 I Q L Q L A T Q F E N W Y K K Y Q K P I

1741 1750 1760 1770 1780 1790 1800
 580 TTCAGAGCGAGTATGGAGCAGAAAACGATTGCAGGGTTTCACCAGGATCCACCTCTGATGT
 I Q S E Y G A E T I A G F H Q D P P L M

 1801 1810 1820 1830 1840 1850 1860
 600 TCACTGAAGAGTACCAGAAAAGTCTGCTAGAGCAGTACCATCTGGGTCTGGATCAAAAAC
 F T E E Y Q K S L L E Q Y H L G L D Q K

 1861 1870 1880 1890 1900 1910 1920
 620 GCAGAAAATACGTGGTTGGAGAGCTCATTTGGAATTTTGCCGATTTTCATGACTGAACAGT
 R R K Y V V G E L I W N F A D F M T E Q

 1921 1930 1940 1950 1960 1970 1980
 640 CACCGACGAGAGTGCTGGGGAATAAAAAAGGGGATCTTCACTCGGCAGAGACAACCAAAAA
 S P T R V L G N K K G I F T R Q R Q P K

 1981 1990 2000 2010 2020 2030 2040
 660 GTGCAGCGTTCCTTTTGCGAGAGAGATACTGGAAGATTGCCAATGAAACCAGGTATCCCC
 S A A F L L R E R Y W K I A N E T R Y P

 2041 2050 2060 2070 2080 2090 2100
 680 ACTCAGTAGCCAAGTCACAATGTTTGGAAAAACAGCCTGTTTACTTGAGCAAGACTGATAC
 H S V A K S Q C L E N S L F T *

 2101 2110 2120 2130 2140 2150 2160
 700 CACCTGCGTGTCCCTTCCCTCCCCGAGTCAGGGCGACTTCCACAGCAGCAGAACAAGTGCC

 2161 2170 2180 2190 2200 2210 2220
 720 TCCTGGACTGTTTACGGCAGACCAGAACGTTTCTGGCCTGGGTTTTGTGGTTCATCTATTC

 2221 2230 2240 2250 2260 2270 2280
 740 TAGCAGGGAACACTAAAGGTGAAAATAAAAGATTTTCTATTATGAAAATAAAGAGTTGGC
 target 2

 2281 2290 2300 2310 2320
 760 ATGAAAGTGGCTACTGAAAAAAAAAAAAAAAAAAAAAAAAAAAAA

cDNA sequence of RAB7A

1 10 20 30 40 50 60
 1 GTCTCGTGACAGGTA CTTCCGCTCGGGGCGGCGGGTGGCGGAAGTGGGAGCGGGCCTG

 61 70 80 90 100 110 120
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 41 GGGAGAGTTCCCTGGAACCAGAACTTGGACCTTCTCGCTTCTGTCCCTCCGTTTGTCTCC

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 61 TCCTCGGCGGGAGCCCTCGCGACGCGCCCGGCCGAGCCCCAGCGCAGCGGCCGCGTT

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301 GTCGGGAAGACATCACTCATGAACCAGTATGTGAATAAGAAATTCAGCAATCAGTACAAA
101 V G K T S L M N Q Y V N K K F S N Q Y K
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361 GCCACAATAGGAGCTGACTTTCTGACCAAGGAGGTGATGGTGGATGACAGGCTAGTCACA
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430 440 450 460 470 480
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141 M Q I W D T A G Q E R F Q S L G V A F Y
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161 R G A D C C V L V F D V T A P N T F K T
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730 740 750 760 770 780
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841 AAGGCCTCGGCAGAAAAGCTGCAGTTGCTGAGGGGGCAGTGAGAGTTGAGCACAGAGTCTC
281 K A S A E S C S C *
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901 TCACAAACCAAGAACACACGTAGGCCTTCAACACAATTCCTCTCCTCTTCCAAACAAA
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1090 1100 1110 1120 1130 1140
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361
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1141 CGCGAGTATGGCAGCAGGACAAGCCAGCGGTGGAAGTCATTTCTGATATGGAGTTGGCATT
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1261 1270 1280 1290 1300 1310 1320
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 441

1381 1390 1400 1410 1420 1430 1440
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 501

1561 1570 1580 1590 1600 1610 1620
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1621 1630 1640 1650 1660 1670 1680
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 target 1

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 561

1741 1750 1760 1770 1780 1790 1800
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 581
 target 2

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 601

1861 1870 1880 1890 1900 1910 1920
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 621

1921 1930 1940 1950 1960 1970 1980
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 641
 target 3

1981 1990 2000 2010 2020 2030 2040
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2041 2050 2060 2070 2080 2090 2100
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 681

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                2170      2180      2190      2200      2210      2220
2161  CCAAAGGTCTACATTCTAGGGTGTGGGCTGAGTTCTTCTGTAAAGAGATGAACGCAATGC
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                2230      2240
2221  CAATAAAATTGAACAAGAACAATG
741

```

Only 3' UTR sequence of VCP (very long transcript, thus only this part is shown)

```

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21

                130     140     150     160     170     180
121 CGTGCAGTGAGCTGGCCTGCCTGGACCTTGTTCCTGGGGGTGGGGGCGCTTGCCAGGA
41

                190     200     210     220     230     240
181  GAGGGACCAGGGGTGCGCCACAGCCTGCTCCATTCTCCAGTCTGAACAGTTCAGCTACA
61

                250     260     270     280     290     300
241  GTCTGACTCTGGACAGGGGGTTCCTGTTGCAAAAAATACAAAACAAAAGCGATAAAAAATAA
81

                310     320     330     340     350     360
301  AGCGATTTTCATTTGGTAGGCGGAGAGTGAATTACCAACAGGGAATTGGGCCCTTGGGCCCT
101
                target 1

                370     380     390     400     410     420
361  ATGCCATTTCTGTTGAGTTTGGGGCAGTGCAGGGGACCTGTGTGGGGTGTGAACCAAGG
121
                target 2

                430     440     450     460     470     480
421  CACTACTGCCACCTGCCACAGTAAAGCATCTGCACCTTGACTCAATGCTGCCCGAGCCCTC
141

                490     500     510     520     530     540
481  CCTTCCCCCTATCCAACCTGGGAGGTGGGTAGGGGCCACAGTTGCTGGATGTTTATATA
161
                target 3

                550     560     570     580     590     600
541  GAGAGTAGGTTGATTTATTTTACATGCTTTTGAGTTAATGTTGGAAAATAATCACAAGC
181

                610     620     630     640     650     660
601  AGTTTCTAAACCAAAAAATGACATGTTGTAAAAGGACAATAAACGTTGGGTCAAAATGGA
201

                670     680     690     700     710     720
661  GCCTGAGTCTGGGCCCTGTGCCTGCTTCTTTTCTGGGAACAGCCTTGGGCTACCCACC
221

                730     740     750     760     770     780
721  ACTCCCAAGGCATTCTTCCAAATGTGAAATCCTGGAAGTAAGATTGCACCTTCTTCTCT
241

                790     800     810     820     830     840
781  CCTGATCAACATCGGTATGATGTCTCCTGTTGCCCTCACCCCTTGTCTGCAGTATCACTGG
261

                850     860     870     880     890     900
841  ATAGGACTGGTGGAAAGGGAGCAGCCTGACAGAGCTCCAAATGTGGAGAATATGGCATCC
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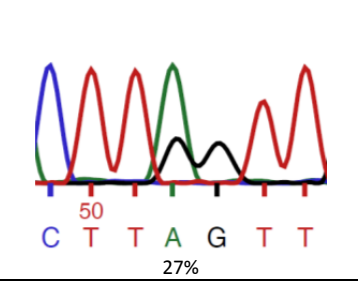
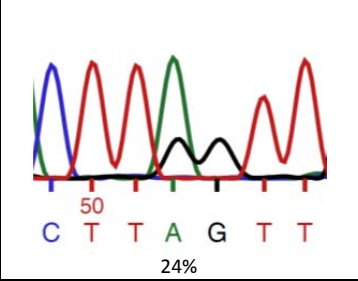
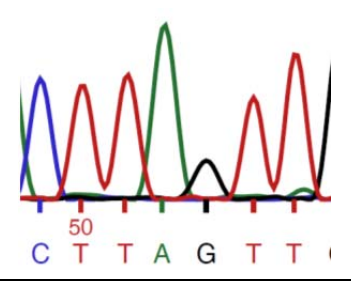
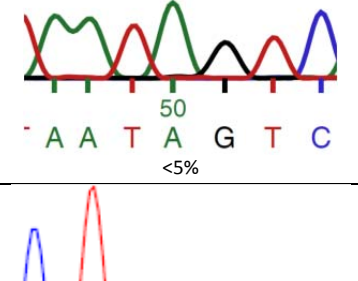
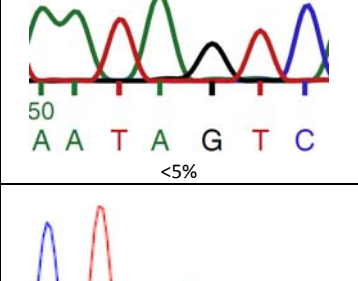
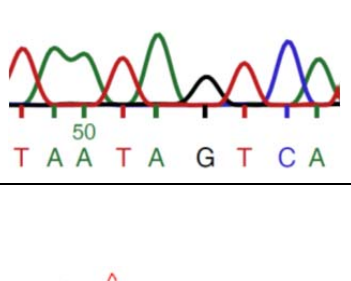
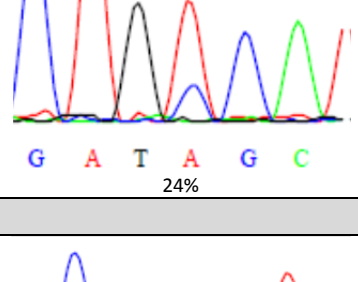
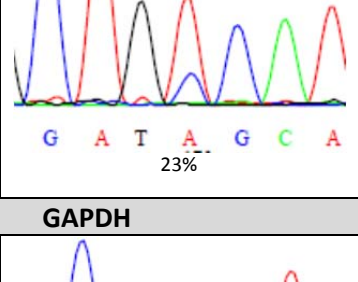
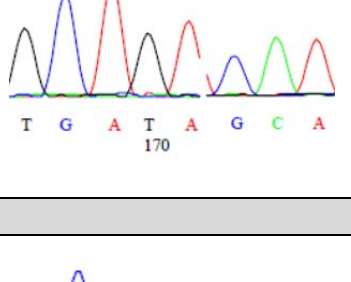
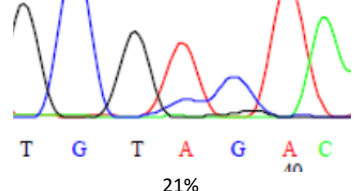
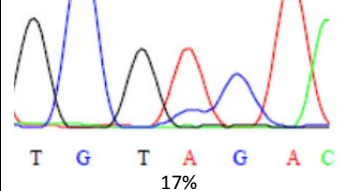
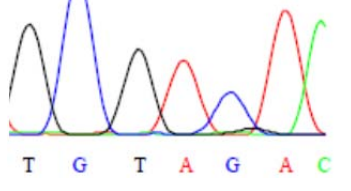
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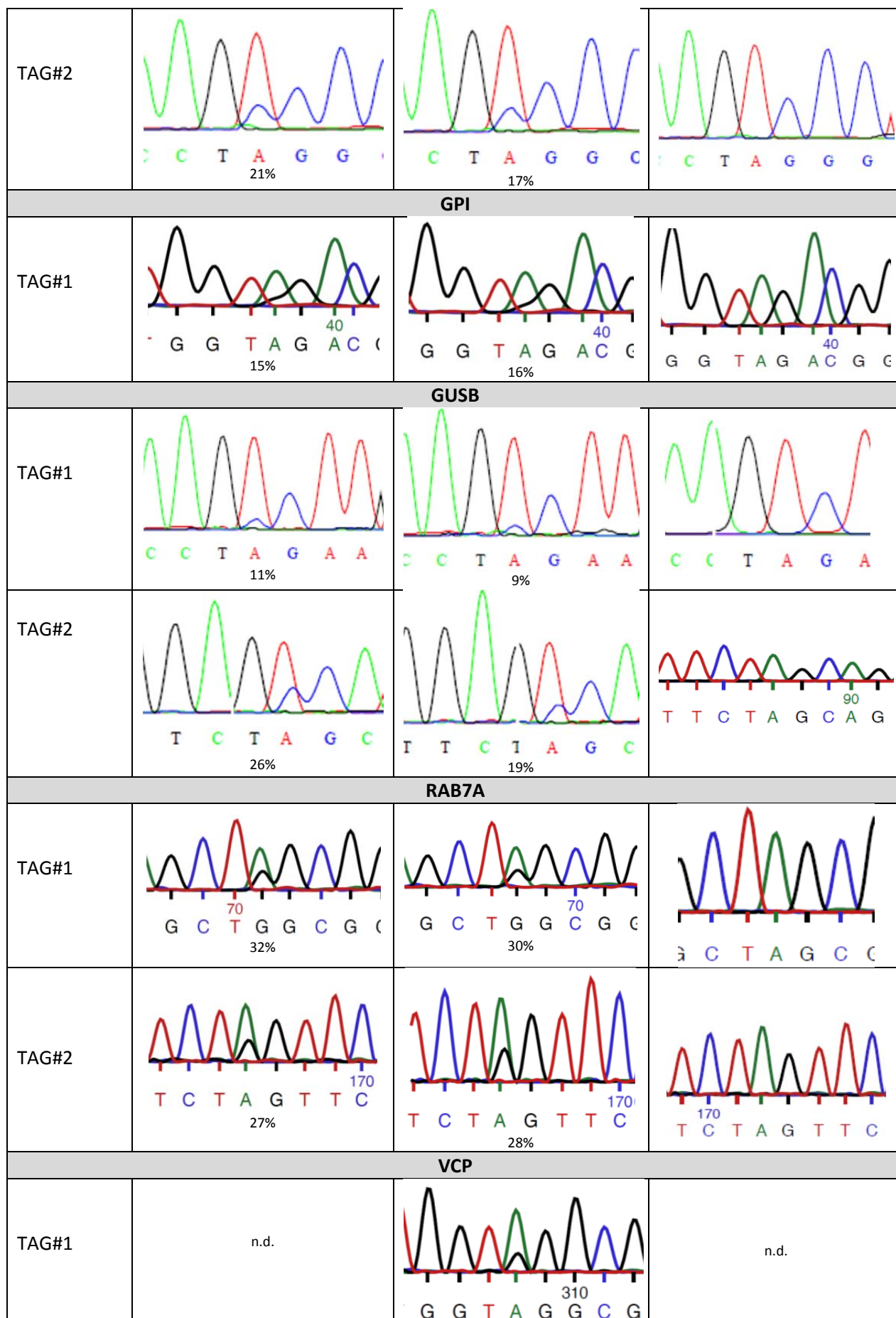
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1021 GCAGGTGGATATTCTCTATACTCTCTTTTAATGCATCTAAAAATCCCAAACATCCCCTGG
341
          1090          1100          1110          1120          1130          1140
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361
          1150
1141 ACTTTTTTGTTAATATAAAA
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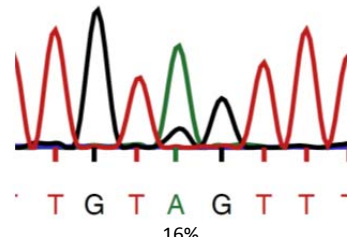
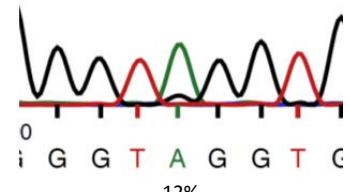
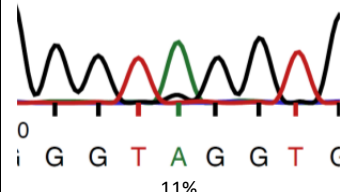
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Figure S19. Sequencing results. Not all RT-PCR reactions were successful or gave sufficient sequencing quality. Those are indicated here with n.d. Primers used in RT-PCR are given on Table S2.

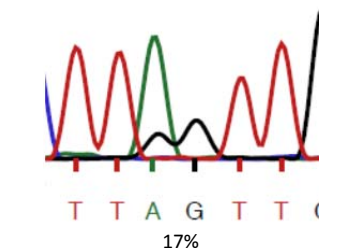
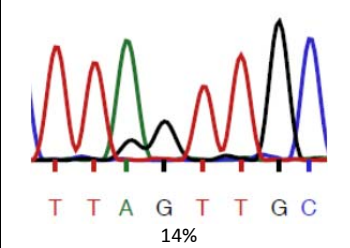
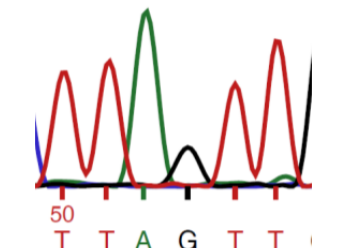
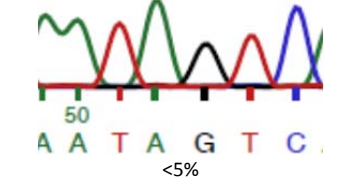
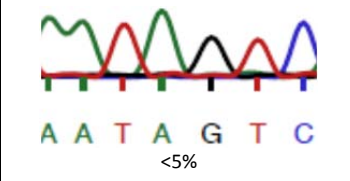
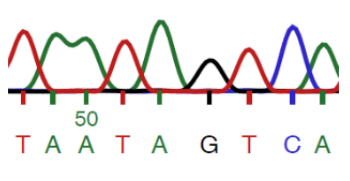
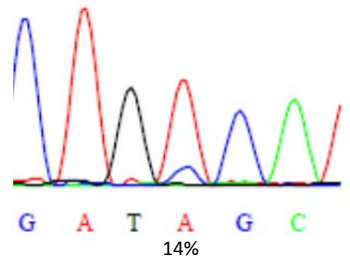
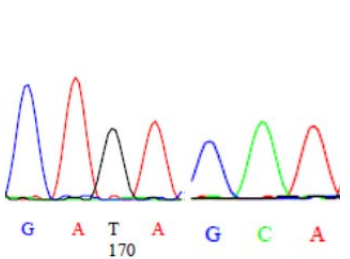
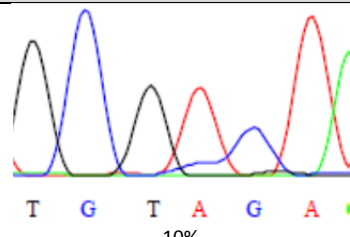
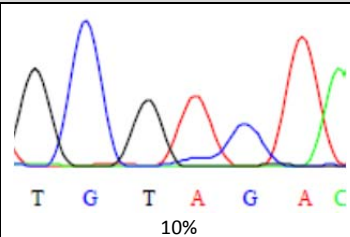
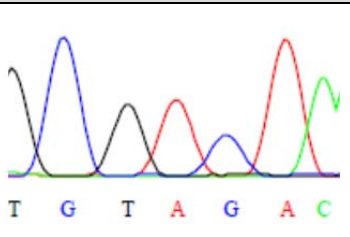
a) Editing in 293T-Cells with ADAR2 overexpression

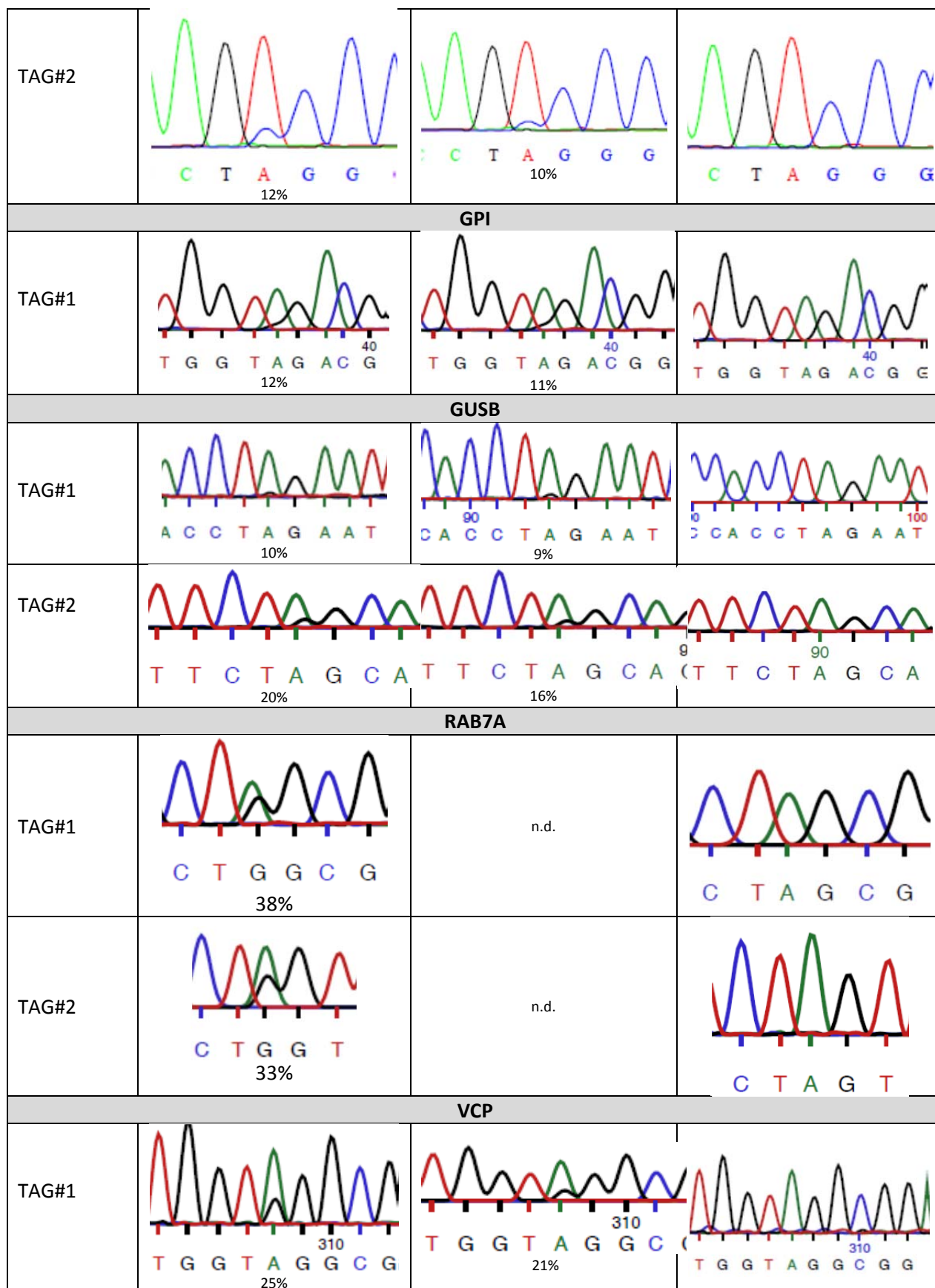
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General Settings: 300 ng pTS57 (ADAR2), 1300 ng gRNA, DNA to Lipofectamine Ratio 1:3, negative control just without guideRNA the rest is treated equally			
B-actin			
TAG#1			
TAG#2			
TAG#3			
GAPDH			
TAG#1			



		25%	
TAG#2	 T G T A G T T 16%	n.d.	n.d.
TAG#3	 G G T A G G T C 12%	 G G T A G G T C 11%	n.d.

cont. Figure S19 b) Editing in 293-ADAR2-Flip-In-Cells

settings	experiment A:	experiment B:	negative control:
General Settings: 1300 ng gRNA, DNA to Lipofectamine Ratio 1:3; negative control just empty transfection			
B-actin			
TAG#1	 T T A G T T C 17%	 T T A G T T G C 14%	 T T A G T T C 50
TAG#2	 A A T A G T C <5%	 A A T A G T C <5%	 T A A T A G T C A 50
TAG#3	 G A T A G C 14%	n.d.	 G A T A G C A 170
GAPDH			
TAG#1	 T G T A G A C 10%	 T G T A G A C 10%	 T G T A G A C



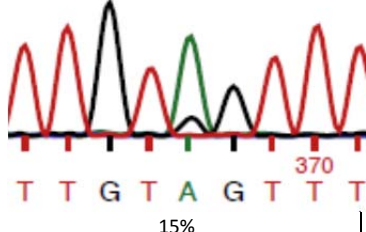
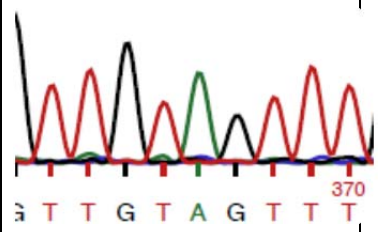
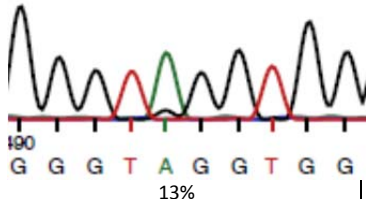
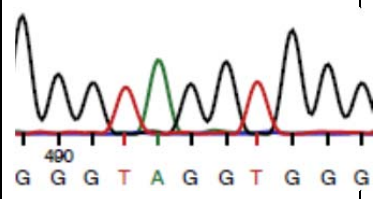
TAG#2	 <p>T T G T A G T T T 370 15%</p>	n.d.	 <p>T T G T A G T T T 370</p>
TAG#3	 <p>190 G G G T A G G T G G 13%</p>	n.d.	 <p>490 G G G T A G G T G G</p>

Table S2.

Primers used for reverse-transcription and PCR:	Sequence 5'->3':
Beta-Aktin_fw	CAGCAGATGTGGATCAGCAAGCAGGAG
Beta-Aktin_bw	GGAAGGGGGGGCACGAAGGCTCATC
Beta-Aktin_Seq_fw	GGTGACAGCAGTCGGTTGGAGCGAGC
GAPDH_fw	CTCAAGATCATCAGCAATGCCTCCTGC
GAPDH_bw	GAGCACAGGGTACTTTATTGATGGTACATGACAAGG
GAPDH_Seq_fw	CACTGCTGGGGAGTCCCTGCCAC
GPI_fw_1	CTACACCGAGGGTTCGAGCCGTGCTG
GPI_fw_2	CCTGGACACCACCCAGAGCACCCCTC
GPI_fw_3	GGGAGTACAGGCACCTGCCACCATG
GPI_bw_1	CAGGGCAACAAAGTGCTTCGCCACTGC
GPI_bw_2	CAGAGGTGAGGAGTGGAAAACAGTCTTGGG
GPI_bw_3	CAGCTATGATTGTATCACTGCAGTCCAGCCTG
GUSB_fw_1	CAACCAAGATGGCGCGGATGGCTTCAGG
GUSB_fw_2	CGCTGCCCGAGTTCTTCAACAACGTTTCTCTG
GUSB_bw_1	GTTGATGGCGATAGTGATTTCGGAGCCGGG
GUSB_bw_2	CAGTAGCCACTTTCATGCCAACTCTTTATTTCCATAATAG
GUSB_seq_fw1	CCTGCGTGTCCCTTCCTCCCCG
GUSB_seq_fw2	CCGGCCTGTGACCTTTGTGAGCAACTC
RAB7A_fw	CTTGGATTATGTGTTTAAGTCCTGTAATGCAGGCC
RAB7A_bw	GGAGCAGAAGTCCAGGGTTCCAACC
RAB7A_Seq_fw	CAGTGGTATGTGCAATGGATAAATTGCCG
VCP_3'UTR_Exon-Junction_fw	GTCGGGGCTTTGGCAGCTTCAGATTCC
VCP_3'UTR_bw	CCTACTCTCTATATAAACATCCAGCAACTGTGGCC

PINK1 Editing

Gene sequence of **wt PINK1** with a C-terminal V5 and His₆-tag in the context of the pcDNA3.1 vector. PINK1 is controlled by the CMV promoter and the BGH terminator:

```

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1  Nhe-I M A V R Q A L G R G L Q L G R A L

      70      80      90      100     110     120
61 CTGCTGCGCTTACGGGCAAGCCCGGCCGGGCTACGGCTTGGGGCGGCCGGGCCCGGCG
21  L L R F T G K P G R A Y G L G R P G P A

      130     140     150     160     170     180
121 GCGGGCTGTGTCCGCGGGGAGCGTCCAGGCTGGGCGCAGGACCGGGCGCGGAGCCTCGC
41  A G C V R G E R P G W A A G P G A E P R

      190     200     210     220     230     240
181 AGGGTCGGGCTCGGGCTCCCTAACCGTCTCCGCTTCTTCCGCCAGTCGGTGGCCGGGCTG
61  R V G L G L P N R L R F F R Q S V A G L

      250     260     270     280     290     300
241 GCGGCGCGGTTGCAGCGGCAGTTTCGTGGTGCGGGCTGGGGCTGCGCGGGCCCTTGCGGC
81  A A R L Q R Q F V V R A W G C A G P C G

      310     320     330     340     350     360
301 CGGGCAGTCTTTCTGGCCTTCGGGCTAGGGCTGGGCCTCATCGAGAAAAACAGGCGGAG
101 R A V F L A F G L G L G L I E E K Q A E

      370     380     390     400     410     420
361 AGCCGGCGGGCGGTCTCGGCCTGTCAGGAGATCCAGGCAATTTTACCAGAAAAGCAAG
121 S R R A V S A C Q E I Q A I F T Q K S K

      430     440     450     460     470     480
421 CCGGGGCTGACCCGTTGGACACGAGACGCTTGCAGGGCTTTCGGCTGGAGGAGTATCTG
141 P G P D P L D T R R L Q G F R L E E Y L

      490     500     510     520     530     540
481 ATAGGGCAGTCCATTGGTAAGGGCTGCAGTGTGCTGTGTATGAAGCCACCATGCCTACA
161 I G Q S I G K G C S A A V Y E A T M P T

      550     560     570     580     590     600
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181 L P Q N L E V T K S T G L L P G R G P G

      610     620     630     640     650     660
601 ACCAGTGCACCAGGAGAAGGGCAGGAGCGAGCTCCGGGGGCCCTGCCTTCCCCTTGCC
201 T S A P G E G Q E R A P G A P A F P L A

      670     680     690     700     710     720
661 ATCAAGATGATGTGGAACATCTCGGCAGGTTCCCTCCAGCGAAGCCATCTTGAACACAATG
221 I K M M W N I S A G S S S E A I L N T M

      730     740     750     760     770     780
721 AGCCAGGAGCTGGTCCCAGCGAGCCGAGTGGCCTTGGCTGGGGAGTATGGAGCAGTCACT
241 S Q E L V P A S R V A L A G E Y G A V T

      790     800     810     820     830     840
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 281 L R A F T S S V P L L P G A L V D Y P D
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 901 GTGCTGCCCTCACGCCTCCACCCTGAAGGCCTGGGCCATGGCCGGACGCTCTTTCTAGTC
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 970 980 990 1000 1010 1020
 961 ATGAAGAACTATCCCTGTACCCTGCGCCAGTACCTTTGTGTGAACACACCCAGCCCCGCGC
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 1021 CTCGCCGCATGATGCTGCTGCAGCTGCTGGAAGGCCTGGACCATCTGGTTCAACAGGGC
 341 L A A M M L L Q L L E G V D H L V Q Q G
 1090 1100 1110 1120 1130 1140
 1081 ATCGCGCACAGAGACCTGAAATCCGACAACATCCTTGTGGAGCTGGACCCAGACGGCTGC
 361 I A H R D L K S D N I L V E L D P D G C
 1150 1160 1170 1180 1190 1200
 1141 CCCTGGCTGGTGATCGCAGATTTTGGCTGCTGCCTGGCTGATGAGAGCATCGGCCTGCAG
 381 P W L V I A D F G C C L A D E S I G L Q
 1210 1220 1230 1240 1250 1260
 1201 TTGCCCTTCAGCAGCTGGTACGTGGATCGGGGCGAAACGGCTGTCTGATGGCCCCAGAG
 401 L P F S S W Y V D R G G N G C L M A P E
 1270 1280 1290 1300 1310 1320
 1261 GTGTCCACGGCCCGTCCTGGCCCCAGGGCAGTGATTGACTACAGCAAGGCTGATGCCTGG
 421 V S T A R P G P R A V I D Y S K A D A W
 1330 1340 1350 1360 1370 1380
 1321 GCAGTGGGAGCCATCGCCTATGAAATCTTCGGGCTTGTCAATCCCTTCTACGGCCAGGGC
 441 A V G A I A Y E I F G L V N P F Y G Q G
 1390 1400 1410 1420 1430 1440
 1381 AAGGCCACCTTGAAAGCCGAGCTACCAAGAGGCTCAGCTACCTGCACTGCCCCAGTCA
 461 K A H L E S R S Y Q E A Q L P A L P E S
 1450 1460 1470 1480 1490 1500
 1441 GTGCCTCCAGACGTGAGACAGTTGGTGAGGGCACTGCTCCAGCGAGAGGCCAGCAAGAGA
 481 V P P D V R Q L V R A L L Q R E A S K R
 1510 1520 1530 1540 1550 1560
 1501 CCATCTGCCCCGAGTAGCCGAAAATGTGCTTCATCTAAGCCTCTGGGGTGAACATATTCTA
 501 P S A R V A A N V L H L S L W G E H I L
 1570 1580 1590 1600 1610 1620
 1561 GCCCTGAAGAATCTGAAGTTAGACAAGATGGTTGGCTGGCTCCTCCAACAATCGGCCGCC
 521 A L K N L K L D K M V G W L L Q Q S A A
 1630 1640 1650 1660 1670 1680
 1621 ACTTTGTTGGCCAACAGGCTCACAGAGAAGTGTGTGTGGAAACAAAAATGAAGATGCTC
 541 T L L A N R L T E K C C V E T K M K M L
 1690 1700 1710 1720 1730 1740
 1681 TTTCTGGCTAACCTGGAGTGTGAAACGCTCTGCCAGGCAGCCCTCCTCCTCTGCTCATGG
 561 F L A N L E C E T L C Q A A L L L C S W

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1741      1750      1760      1770      1780      1790      1800
581      AGGGCAGCCCTGCTCGAGTCTAGAGGGCCCTTTCGAAGGTAAGCCTATCCCTAACCCCTCTC
          R A A L L E S R G P F E G K P I P N P L

1801      1810      1820      1830      1840      1850
601      CTCGGTCTCGATTCTACGCGTACCGGTCATCATCACCATCACCATTTGAGTTTAAACCCG
          L G L D S T R T G H H H H H H * Pme-I

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Gene & protein sequence of PINK1 W437X amber with a C-terminal V5- and His₆-tag in the context of the pcDNA 3.1 vector, under control of the the CMV promoter and the BGH terminator. The amber Stop codon is highlighted in yellow.:

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1          10          20          30          40          50          60
1      CTGGCTAGCATGGCGGTTGCGACAGGCGCTGGGCCGCGGCTGCAGCTGGGTCGAGCGCTG
1          Nhe-I M A V R Q A L G R G L Q L G R A L

61          70          80          90          100         110         120
21     CTGCTGCGCTTACGGGCAAGCCCGGCCGGCCTACGGCTTGGGGCGGCGGGCCCGGCG
          L L R F T G K P G R A Y G L G R P G P A

121        130        140        150        160        170        180
41     GCGGGCTGTGTCCGCGGGGAGCGTCCAGGCTGGGCCGAGGACCGGGCGGAGCCTCGC
          A G C V R G E R P G W A A G P G A E P R

181        190        200        210        220        230        240
61     AGGGTCGGGCTCGGGCTCCCTAACCGTCTCCGCTTCTTCCGCCAGTTCGGTGGCCGGGCTG
          R V G L G L P N R L R F F R Q S V A G L

241        250        260        270        280        290        300
81     GCGGCGCGGTTGCAGCGGCAGTTTCGTGGTGCGGGCTGGGGCTGCGCGGGCCCTTGCGGC
          A A R L Q R Q F V V R A W G C A G P C G

301        310        320        330        340        350        360
101    CGGGCAGTCTTTCTGGCCTTCGGGCTAGGGCTGGGCCTCATCGAGGAAAAACAGGCGGAG
          R A V F L A F G L G L G L I E E K Q A E

361        370        380        390        400        410        420
121    AGCCGGCGGGCGGTCTCGGCCTGTCAGGAGATCCAGGCAATTTTACCAGAAAAGCAAG
          S R R A V S A C Q E I Q A I F T Q K S K

421        430        440        450        460        470        480
141    CCGGGGCTGACCCGTTGGACACGAGACGCTTGCAGGGCTTTCGGCTGGAGGAGTATCTG
          P G P D P L D T R R L Q G F R L E E Y L

481        490        500        510        520        530        540
161    ATAGGGCAGTCCATTGGTAAGGGCTGCAGTGCTGCTGTGTATGAAGCCACCATGCCTACA
          I G Q S I G K G C S A A V Y E A T M P T

541        550        560        570        580        590        600
181    TTGCCCCAGAACCTGGAGGTGACAAAGAGCACCGGGTTGCTTCCAGGGAGAGGCCAGGT
          L P Q N L E V T K S T G L L P G R G P G

601        610        620        630        640        650        660
201    ACCAGTGCACCAGGAGAAGGGCAGGAGCGAGCTCCGGGGGCCCTGCCTTCCCCTTGGCC
          T S A P G E G Q E R A P G A P A F P L A

          670        680        690        700        710        720

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661 ATCAAGATGATGTGGAACATCTCGGCAGGTTCTCCAGCGAAGCCATCTTGAACACAATG
221 I K M M W N I S A G S S S E A I L N T M

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721 AGCCAGGAGCTGGTCCCAGCGAGCCGAGTGGCCTTGGCTGGGGAGTATGGAGCAGTCACT
241 S Q E L V P A S R V A L A G E Y G A V T

790 800 810 820 830 840
781 TACAGAAAATCCAAGAGAGGTCCAAGCAACTAGCCCTCACCCCAACATCATCCGGGTT
261 Y R K S K R G P K Q L A P H P N I I R V

850 860 870 880 890 900
841 CTCCGCGCCTTACCTCTTCCGTGCCGCTGCTGCCAGGGGCCCTGGTCTGACTACCCTGAT
281 L R A F T S S V P L L P G A L V D Y P D

910 920 930 940 950 960
901 GTGCTGCCCTCACGCCTCCACCCTGAAGGCCTGGGCCATGGCCGGACGCTCTTTCTAGTC
301 V L P S R L H P E G L G H G R T L F L V

970 980 990 1000 1010 1020
961 ATGAAGAACTATCCCTGTACCCTGCGCCAGTACCTTTGTGTGAACACACCCAGCCCCCGC
321 M K N Y P C T L R Q Y L C V N T P S P R

1030 1040 1050 1060 1070 1080
1021 CTCGCCGCCATGATGCTGCTGCAGCTGCTGGAAGGCGTGGACCATCTGGTTCAACAGGGC
341 L A A M M L L Q L L E G V D H L V Q Q G

1090 1100 1110 1120 1130 1140
1081 ATCGCGCACAGAGACCTGAAATCCGACAACATCCTTGTGGAGCTGGACCCAGACGGCTGC
361 I A H R D L K S D N I L V E L D P D G C

1150 1160 1170 1180 1190 1200
1141 CCCTGGCTGGTGATCGCAGATTTTGGCTGCTGCCTGGCTGATGAGAGCATCGGCCCTGCAG
381 P W L V I A D F G C C L A D E S I G L Q

1210 1220 1230 1240 1250 1260
1201 TTGCCCTTCAGCAGCTGGTACGTGGATCGGGCGGAAACGGCTGTCTGATGGCCCCAGAG
401 L P F S S W Y V D R G G N G C L M A P E

1270 1280 1290 1300 1310 1320
1261 GTGTCCACGGCCCGTCTGGCCCCAGGGCAGTGATTGACTACAGCAAGGCTGATGCC**TAG**
421 V S T A R P G P R A V I D Y S K A D A *****

1330 1340 1350 1360 1370 1380
1321 GCAGTGGGAGCCATCGCCTATGAAATCTTCGGGCTTGTCAATCCCTTCTACGGCCAGGGC
441 A V G A I A Y E I F G L V N P F Y G Q G

1390 1400 1410 1420 1430 1440
1381 AAGGCCACCTTGAAAGCCGCGAGCTACCAAGAGGCTCAGCTACCTGCACTGCCCGAGTCA
461 K A H L E S R S Y Q E A Q L P A L P E S

1450 1460 1470 1480 1490 1500
1441 GTGCCTCCAGACGTGAGACAGTTGGTGGAGGGCACTGCTCCAGCGAGAGGCCAGCAAGAGA
481 V P P D V R Q L V R A L L Q R E A S K R

1510 1520 1530 1540 1550 1560
1501 CCATCTGCCCCGAGTAGCCGAAATGTGCTTCATCTAAGCCTCTGGGGTGAACATATTCTA
501 P S A R V A A N V L H L S L W G E H I L

1570 1580 1590 1600 1610 1620
1561 GCCCTGAAGAATCTGAAGTTAGACAAGATGGTTGGCTGGCTCCTCCAACAATCGGCCGCC

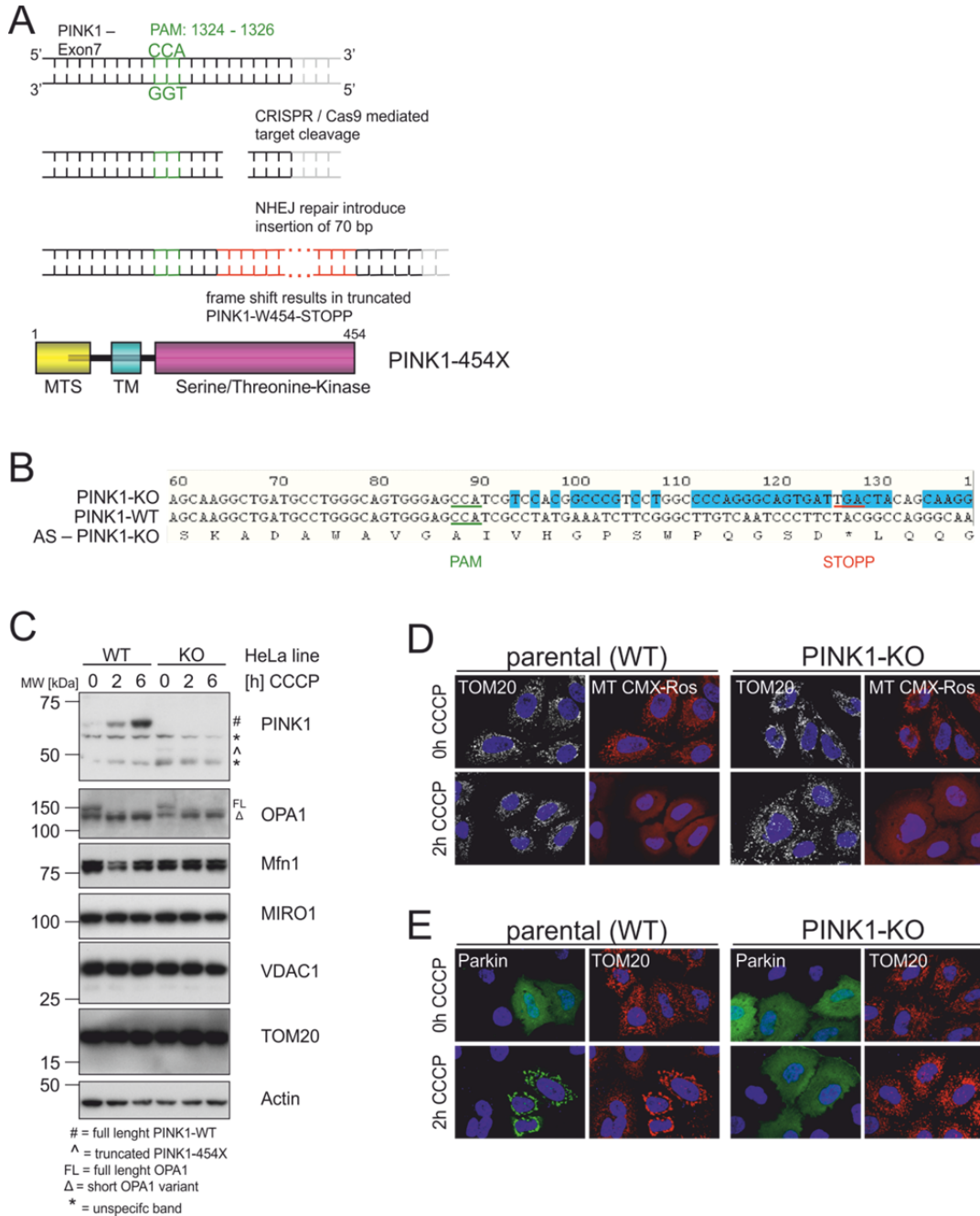
521 A L K N L K L D K M V G W L L Q Q S A A
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 1621 ACTTTGTTGGCCAACAGGCTCACAGAGAAGTGTGTTGTGTGGAAACAAAAATGAAGATGCTC
 541 T L L A N R L T E K C C V E T K M K M L
 1690 1700 1710 1720 1730 1740
 1681 TTTCTGGCTAACCTGGAGTGTGAAACGCTCTGCCAGGCAGCCCTCCTCCTCTGCTCATGG
 561 F L A N L E C E T L C Q A A L L L C S W
 1750 1760 1770 1780 1790 1800
 1741 AGGGCAGCCCTGCTCGAGTCTAGAGGGCCCTTCGAAGGTAAGCCTATCCCTAACCCCTCTC
 581 R A A L L E S R G P F E G K P I P N P L
 1810 1820 1830 1840 1850
 1801 CTCGGTCTCGATTCTACGCGTACCGGTCATCATCACCATCACCATTGAGGTTTAAACCCG
 601 L G L D S T R T G H H H H H H * Pme-I

Generation and Characterization of the PINK1-Knock out HeLa cell line

To genomically engineer a PINK1-knock out HeLa cell line, the CRISPR/Cas9 system was used. Therefore, the guideRNA sequence 5'-**CCATCGCCTATGAAATC**-3' (with the PAM (protospacer adjacent motif) in bold) within exon 7 of PINK1, was inserted into the Cas9 expression plasmid pSpCas9(BB)-2A-Puro (F. A. Ran et al. Genome engineering using the CRISPR-Cas9 system. Nat. Protocols, 2013, doi:10.1038/nprot.2013.143) to yield pSpCas9(PINK1-KO)-2A-PURO. The parental HeLa cells were transfected with 2 µg of the pSpCas9(PINK1-KO)-2A-PURO plasmid using the Human Stem Cell Nucleofector Kit (Lonza) and a nucleofector device (Amaxa). 48h after transfection, the cells were selected using 2 µg/ml puromycin in DMEM+10% FCS for 48h. The genomic DNA from clones was isolated using the ReliaPrep gDNA Tissue Miniprep System (Promega). The PINK1 exon 7 was amplified by flanking intronic primers (forward: 5' TGGATCAGGTGATGTGCAGGA 3' and reverse: 5' AGGATCTGTCACTGTG GCTCT 3'). The CRISPR/Cas9 genomic editing resulted in an introduction of a 70 bp DNA fragment into PINK1 exon 7, 4 bp upstream of the PAM sequence. This disrupts the functional PINK1 allele by introducing a premature Stop codon at position 454 inside the kinase domain (A, B). Due to the premature Stop codon, the resulting PINK1 protein is non-functional. While in parental wt HeLa cells a stabilization of full length PINK1 is seen after CCCP treatment (#, Fig. S21 C), no PINK1 protein expression in mutant HeLa cells was detected (Fig. S21 C). Indeed, only faint amounts of the unstable truncated PINK1-454X were detected (^, Fig. S21 C). We thus call this cell line "PINK1 knock-out (KO)" even though a truncated PINK1 protein of similar length like the pathogenic W437X mutant could potentially be expressed. The characterization of the PINK1-KO cells highlights no differences in mitochondrial protein levels compared to wt HeLa cells (Fig. S21 C). In addition, PINK1 functional null did not interfere with the disruption of the mitochondrial membrane potential by CCCP, since the loss of the long OPA1 isoform (Fig. S21 C) as well as the loss of mitochondrial staining, with the membrane potential sensitive MitoTracker Red CMX-ROS was observed (Fig. S21 D). The mutated PINK1 is in addition also unable to induce perinuclear co-localization of Parkin and mitochondria (see microscopy pictures, Fig. S21 D and E). Thus, we considered that the PINK1-KO HeLa cells are functional null regarding PINK1. Important, due to the insertion mutation this non-functional transcript is not repaired by site-directed RNA editing.

Figure S20. Generation and characterization of the PINK1-KO HeLa cell line

Besides the antibodies described on page 61-62, the following additional antibodies were used:
 rabbit anti-OPA1 (BD Transduction Laboratories, #612606)
 mouse anti-Mfn1 (Abnova, H00055669-M04)
 rabbit anti-VDAC1 (Millipore, Ab10527)
 mouse anti-MIRO1 (Sigma, WH0055288M1).



PINK1 Functional Assay

HeLa cells (PINK1 wt or KO) were cultured under standard conditions (DMEM + 10% FBS, 37°C, 5% CO₂). The mitophagy assay was performed in 24-wells. Each well contained a cover-slip coated with poly-D-lysine (Sigma Aldrich). The cells were seeded at 2.5x10⁴/well. After 24h the cells were transfected with the indicated plasmids using FuGene6 (Promega). If the editing vector was transfected a 1-to-6 ratio was used. If ADAR2 or guideRNA alone were transfected, a 1-to-3 ratio was applied. If not indicated, the plasmid amounts/well has been 300 ng for EGFP-Parkin, 300 ng for PINK1 wt or PINK1 W437amber, 300 ng editing vector. In control experiment d) 1300 ng of a guideRNA plasmid based on pSilencer lacking ADAR2 was co-transfected instead of the editing vector. In control e) 200 ng of an editing vector lacking any guideRNAs but containing ADAR2 was co-transfected instead of the original editing vector. Treatment with 10 μM CCCP (in DMEM + 10% FBS) was either performed 46h after transfection for 2h or 24h after transfection for 24 h. The depolarization of the mitochondrial membrane potential with 10 μM CCCP (in DMEM + 10% FBS) was either performed 46h after transfection for 2h or 24h after transfection for 24 h. To visualize the mitochondria with a membrane potential sensitive dye, like MitoTracker Red CMXRos, a CCCP wash out was performed. For this, the depolarizing agent CCCP was washed out by changing the media twice every 15 min. Then the cells were incubated with 100 nM MitoTracker Red CMXRos (Invitrogen, M7512) in DMEM for 30 min at 37°C directly prior fixation or harvesting. Then 48 h after transfection, the cells were washed once with PBS and then either fixated for immunocytochemical staining (A) or harvested for RNA isolation (B).

(A) After fixation with 4% PFA/PBS for 20 minutes at RT and three wash steps with PBS, the cells were permeabilized with 1% Triton X-100/PBS for 5 minutes at RT followed by three wash steps with PBS. Then, the cells were blocked with 10% FCS/PBS for 1h at RT and stained with following antibodies diluted in 5% FCS/PBS for 2h at RT: mouse anti-ADAR2 (Santa Cruz, SC-73409, 1:1000), and rabbit anti-PINK1 (Novus Biologicals, BC 100-949, 1:750). The cells were then washed three times with PBS and incubated with the following secondary antibodies diluted in 10% FCS/PBS: goat anti-mouse or rabbit Alexa Fluor-488, 568 or 647 (Invitrogen, 1:1000). After two washing steps with PBS the nuclei of the cells where stained with Hoechst33342 (Thermo Fisher, 1:5000) for 5 minutes at RT. The cover-slips where mounted onto glass-slides using the Dako fluorescent mounting medium. Confocal images with a slice thickness of 0,7 μm were obtained with an AxioImager microscope equipped with an ApoTome imaging system (Carl Zeiss) using a 63x objective. The images were processed using the AxioVision software 4.8.1. For the quantification of Parkin clustering, double- (EGFP-Parkin and PINK1) and triple- (EGFP-Parkin, PINK1 and hADAR2) positive cells were evaluated. More than 100 cells per cover slip and condition were analyzed for quantification.

(B) The cells were harvested by trypsination using 60 μl Trypsin-EDTA. After inactivation with 440 μl DMEM+10%FBS the cells were pelleted at 300 g for 5 minutes at 4°C. After removing the supernatant the cells where washed once in 500 μl ice-cold PBS and centrifuged again at 300 g for 5 minutes at 4°C. The cell pellet was snap frozen in liquid nitrogen prior RNA isolation using the RNeasy Mini Kit (Qiagen). Remaining Plasmid DNA that could interfere with following PCRs was removed by DNase-I (NEB) digestion for 10 min at 37°C. Then, the RNA was reverse transcribed using 1 μl M-MuLV reverse transcriptase (NEB), 0,25 μl murine RNase inhibitor (NEB) and a BHG backward primer (5'-CTAGAAGGCACAGTCGAGGC). The cDNA was isolated using the nucleospin gel and PCR-cleanup kit (Machery-Nagel). The following Phusion-PCR was performed with 4% DMSO, 0.8 M betaine and a PINK1 specific primer pair 5'-CTAGA AGGCACAGTCGAGGC and 5'-GAGCCAGGAGCTGGTCCCAGCGAGCCGAG". The 1167 bp

PINK1 DNA fragment was isolated from the 1,4% agarose gel using the nucleospin gel and PCR-cleanup kit (Machery-Nagel). The sequencing was performed by the company Eurofins using the sequencing primer 5'-GGTACGTGGATCAGGGCG GAAACGGC.

Figure S21. Western Blot analysis of PINK1-W437X-V5 editing. HeLa WT or PINK1-KO cells were transfected with plasmids for wt PINK1-V5 or PINK1-W437X-V5, and with or without the editing vector for 48h as indicated. The cells were treated with 10 μ M CCCP 6h prior lysis with 8M Urea buffer (10 mM Tris (pH 8.0), 100 mM NaH₂PO₄, 8M urea). 10 μ g of total lysates were separated by SDS-PAGE and transferred onto PVDF membrane using the wet blotting method. The following primary antibodies were used: mouse anti-ADAR2 (Santa Cruz, sc-73409), rabbit anti-PINK1 (Novus Biologicals, BC 100-494), mouse anti-V5 (Invitrogen, R960-25), rabbit anti-TOM20 (Santa Cruz, sc-11415), and mouse anti beta-actin (Sigma, clone AC-15). In panel **A**, the potential PINK1 variants are shown. In panel **B**, the PINK1 antibody stains all processed and truncated versions of the wt and the overexpressed PINK1 variants. Upon editing a small fraction of fulllength PINK1 (FL & Δ FL) appears beside an excess of truncated PINK1 (X & Δ X) which is in accordance with an editing yield of 10%. The V5 antibody detects fulllength V5-tagged PINK1 (FL & Δ FL) only in presence of the editing vector, which again indicates the successful editing of the transfected PINK1-W437X-V5. After editing and CCCP treatment TOM20 ubiquitinylation (Ub-1, Ub-2, Ub-3) is visible similar to wt PINK1.

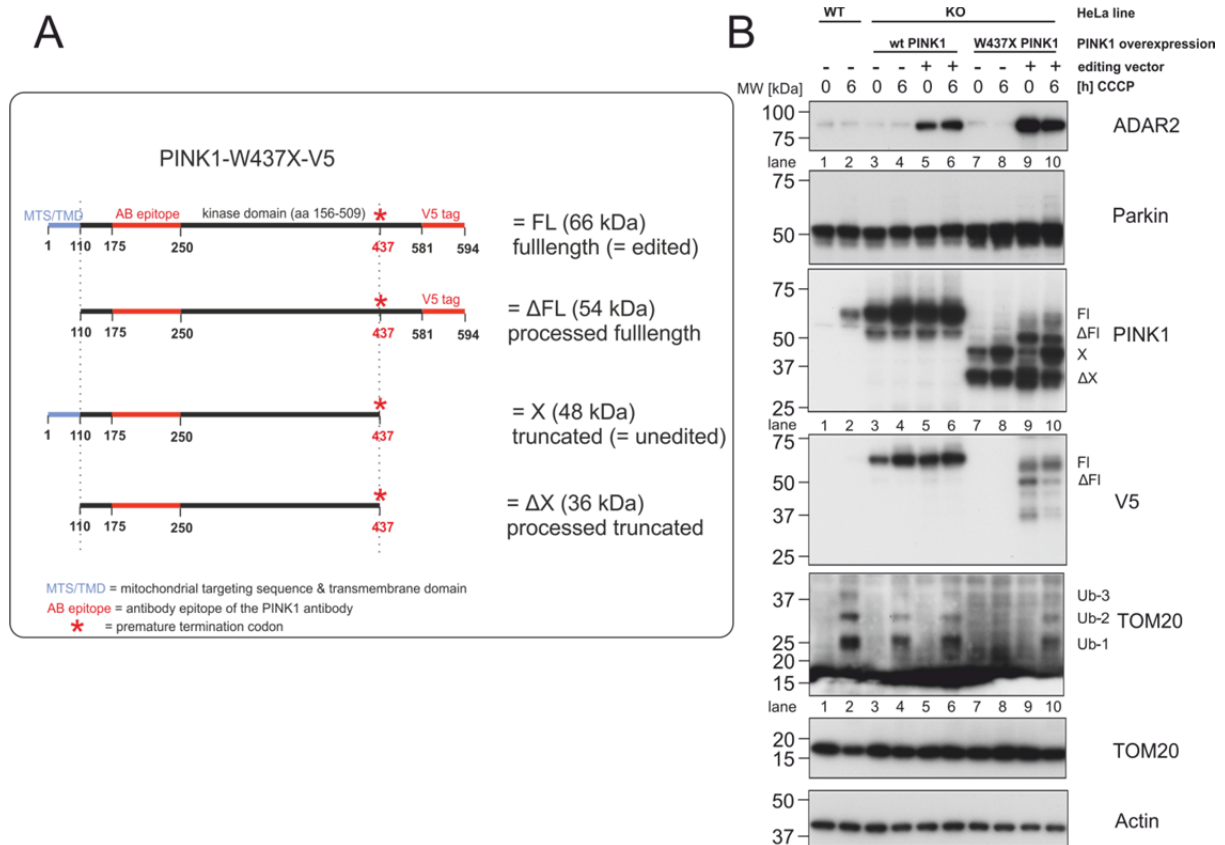


Figure S22. Full sequencing trace of the PINK1 W437amber editing experiment in HeLa cells, corresponding to the experiment shown in Figure 4A, 4B c).

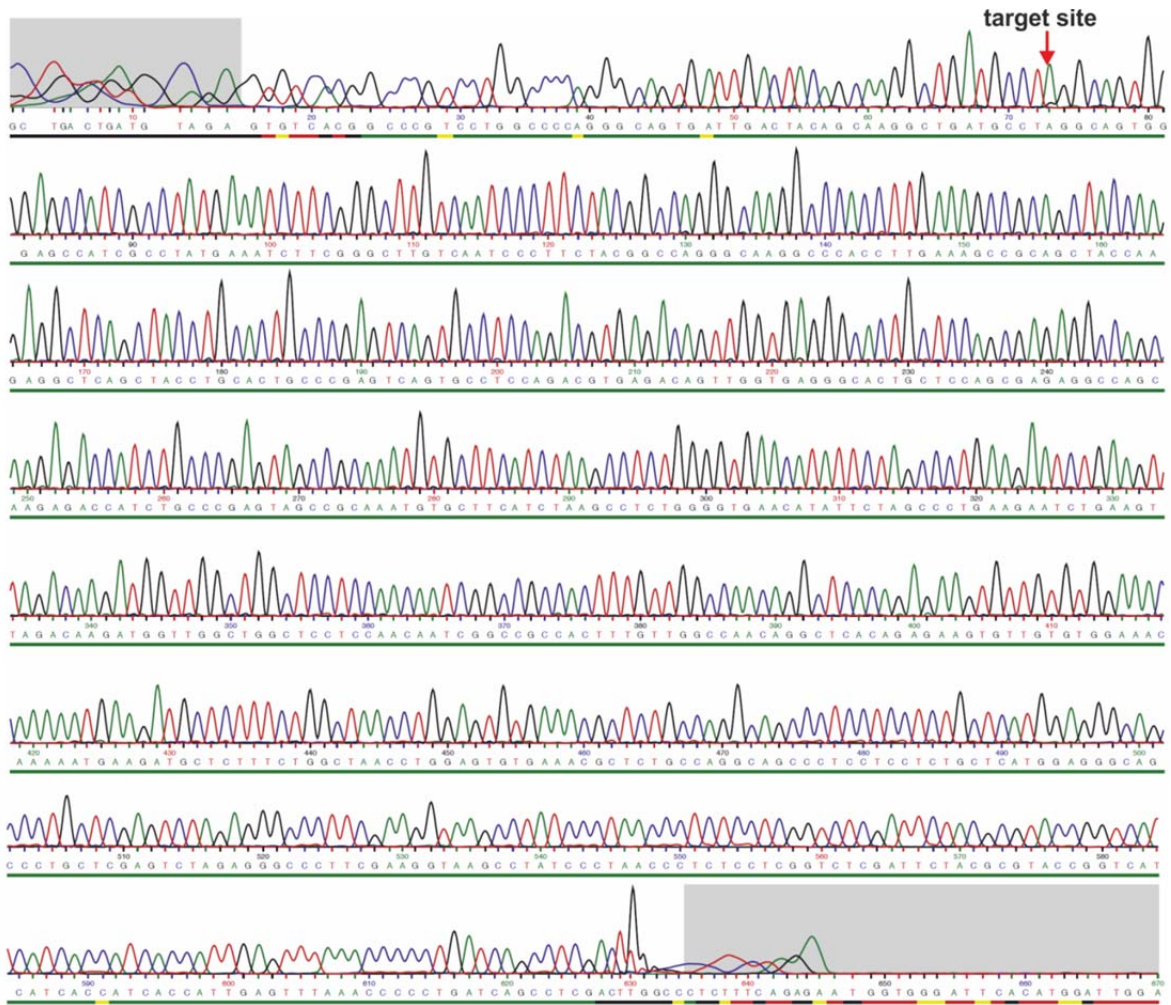
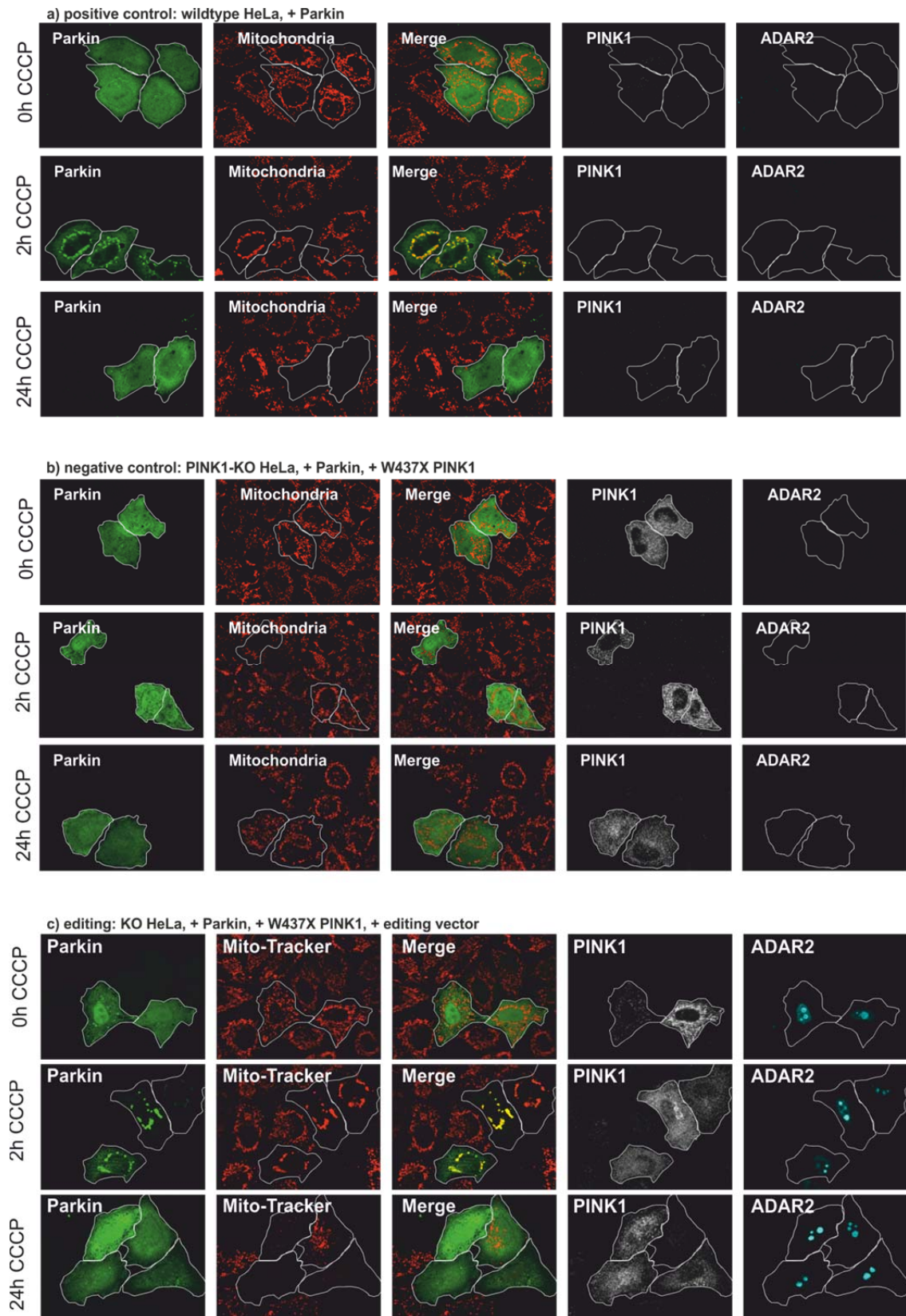
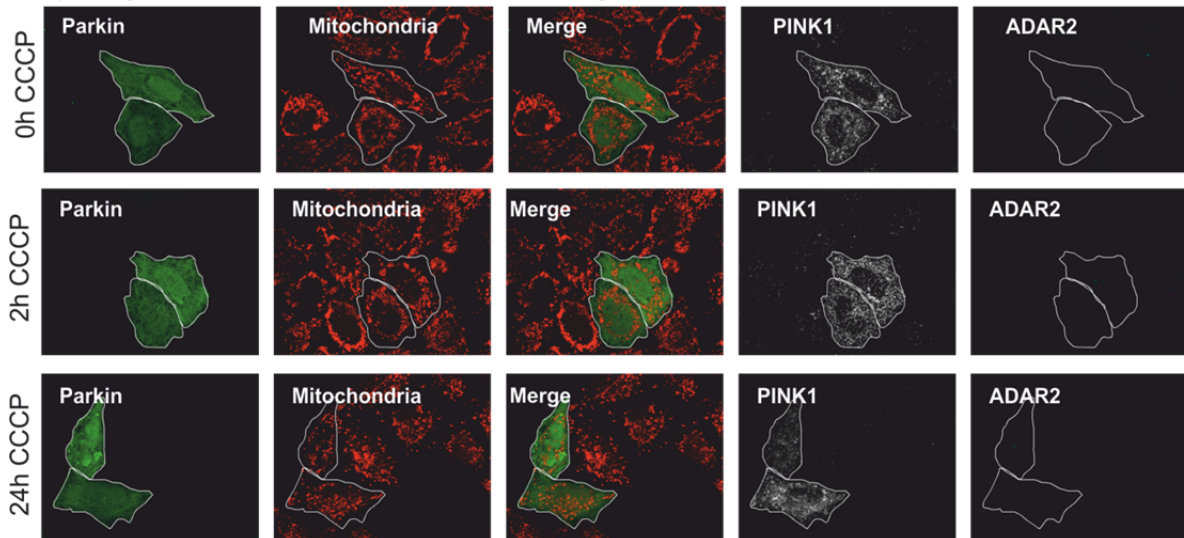


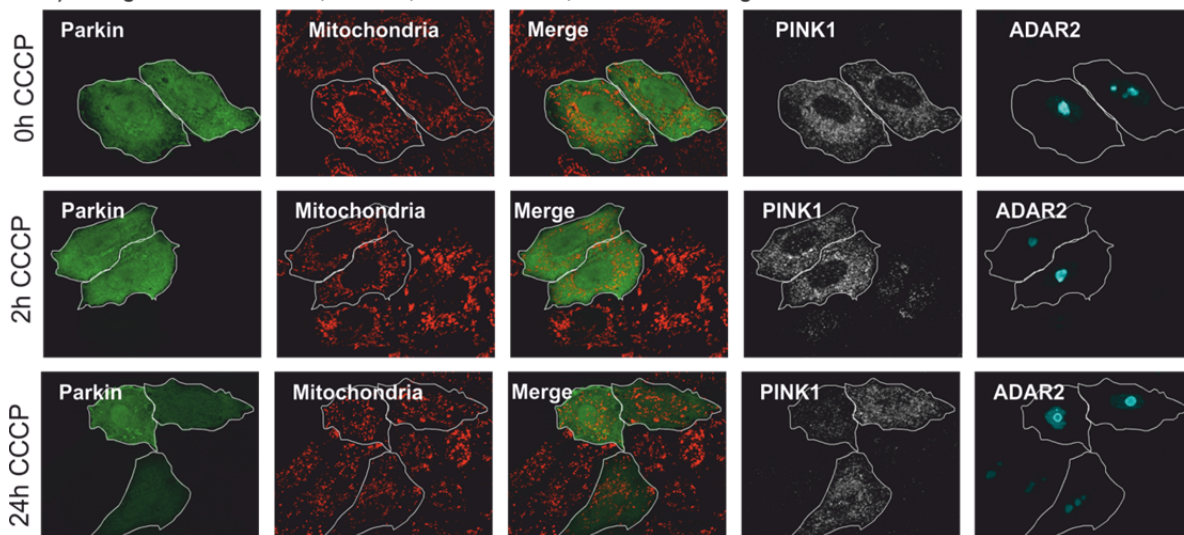
Figure S23. Mitophagy assay. Shown are the complete mitophagy assays that relate to the Parkin-clustering experiments a) – f) shown on Figure 4A in the main text. The mitophagy assay shown in c) is the same as shown in the main text Figure 4C. For better visualization of mitophagy, the Parkin-positive cells were encircled.



d) editing control 1: KO HeLa, + Parkin, + W437X PINK1, + guideRNA --> no ADAR2



e) editing control 2: KO HeLa, + Parkin, + W437X PINK1, + ADAR2 --> no guideRNA



f) editing control 3: KO HeLa, + Parkin, + editing vector --> no PINK1 W437X substrate

