

Supplementary data to

**Restriction endonuclease Agel is a monomer which dimerizes to cleave DNA**

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**Supplementary Table S1.** Oligoduplexes used in this study

Substrate	Used for	Sequence
SP11 (specific)	crystallization	5' - TCG <u>ACCGGT</u> CG - 3' 3' - GCT <u>TGGCCA</u> AGCT - 5'
SP13 (specific)	crystallization	5' - TTTCG <u>ACCGGT</u> CGA - 3' 3' - AGCT <u>TGGCCA</u> AGCTT - 5'
SP (specific)	DNA binding, hydrolysis and gelfiltration	5' - AGACCCACGCTC <u>ACCGGT</u> TCCAGATTTATC - 3' 3' - TCTGGGTGCGAG <u>TGGCCA</u> AGGTCTAAATAG - 5'
NC (non- canonical)	DNA binding	5' - AGACCCACGCTC <u>ACCGG</u> ATCCAGATTTATC - 3' 3' - TCTGGGTGCGAG <u>TGGCC</u> TAGGTCTAAATAG - 5'
NSP (non-specific)	DNA binding	5' - CAGCACAGTTCAGCAGCCCAGTGCTACGCT - 3' 3' - GTCGTGTCAAGTCGTCGGGTCACGATGCCA - 5'
<b>Mutant</b>		
Q86A	mutagenesis	5'-CGACCCAAGCGCTAGCTGGGTTCGCACGTCGGGCG-3' 5'-GCGAACCAGCTAGCGCTTGGGTTCGGTCCGATCTTC-3'
S138A	mutagenesis	5'-CGCGTCGGCGCCCGCAAGGTTGACGTGATGATCG-3' 5'-CAACCTTGCGGGCGCCGACGCGGTTCGGCCACAC-3'
D142A	mutagenesis	5' - CTCGCAAGGTTGCCGTCATGATCGAGAAGCAGGGAGGCGG - 3' 5' - CTTCTCGATCATGACGGCAACCTTGCAGAGCCGACGCGG - 3'
D177A	mutagenesis	5' - GGGTCTCGGCCGACATACCCGCCAGCAGGATC - 3' 5' - CGGGTATGTCCGCCGAGACCCTCTCGGCTAG - 3'
D178A	mutagenesis	5' - GTCTCGGACGCCATACCCGCCAGCAGGATCATG - 3' 5' - CTGGCGGGTATGGCGTCCGAGACCCTTTTCGGCTAGGCTC - 3'
D223A	mutagenesis	5' - CCGGCCCTCGGCCGAAGAGGAATTACATTGAGGGACAC - 3' 5' - GTAATTCCTCTTCGCCGAGGGCCGGTTCGGGCGTGCC - 3'

Agel recognition sequence (5'-ACCGGT-3') is underlined. In NC only nucleotides, which coincide with the recognition sequence, are underlined.

**Supplementary Table S2.** Crystallization and cryo-protection conditions

<b>Crystal form</b>	<b>Space group</b>	<b>Crystallization conditions</b>	<b>Cryo-protection conditions</b>
Agel+SP11	Form I P6 <sub>1</sub> 22	Na Malonate 7.0 0.2M, PEG3500 19% protein concentration 8.3 mg/ml, Agel:DNA ratio 1:1.2	Mother liquor supplemented with 20% PEG400
Agel+SP11	Form II P2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>	0.2 M NaCl, 0.1 M BisTRIS 5.5, 17- 19% PEG3500 (protein concentration 8.3 mg/ml, Agel:DNA ratio 1:1.2)	stepwise transfer into mother liquor containing 2-5-10-15% ethylene glycol
Agel+SP13	Form III P2 <sub>1</sub>	0.2M NaCl, 0.1M BisTRIS 5.5, 17-19% PEG3500 protein concentration 7.2 mg/ml, Agel:DNA ratio 1:1.2	stepwise transfer into mother liquor containing 2-5-10-15% ethylene glycol
Agel	P2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>	0.1 M Na-Hepes (pH 7.5), 0.2 M NaCl, 25% PEG3350, 3% ethylene glycol (protein concentration 5.7 mg/ml)	stepwise transfer into mother liquor containing 5-10-15% ethylene glycol

**Supplementary Table S3.** SAXS data collection parameters and shape models description.

<b>Data collection parameters</b>	
X-ray wave length, nm	0.124
Sample to detector distance, m	3.1
Detector	Pilatus 2M
s range, nm <sup>-1</sup>	0.0290642-4.488440
Exposure time, s	0.05
Collected frames	20
Sample storage temperature, °C	10
Data collection temperature, °C	20

<b>Structural parameters of models</b>		
<b>Parameter</b>	<b>Agel</b>	<b>Agel+DNA</b>
Protein concentration, mg/ml	1.3 and 3.1	2.8
s range of P(r) calculated by GNOM, Å <sup>-1</sup>	0.0229 – 0.2986	0.0111 – 0.3435
Real space Rg calculated by GNOM, Å	21.29 ± 0.050	24.00 ± 0.022
D <sub>max</sub> used by GNOM, Å	67	74
Porod volume calculated by GNOM, Å <sup>3</sup>	65894	53576
Averaged volume of DAMMIN models, Å <sup>3</sup>	56078 ± 1247	73930 ± 2012

**Supplementary Table S4.** Comparison DNA recognition and active site residues of SP- and preSP-complexes

	SP-complex residue subunits A/B	preSP-complex residue subunits A/B
DNA	13 bp	11 bp
Dimer <i>buried surface area</i> , Å <sup>2</sup>	630	400
DNA binding <i>buried surface area</i> , Å <sup>2</sup>	5823	5215
Major groove contacts	E173 <b>+/+</b> R174 <b>+/+</b> K200 <b>+/+</b> E214 <b>+/+</b> K224 <b>+/+</b>	<b>E173</b> <b>-/-</b> <b>R174</b> <b>+/+</b> K200 <b>+/+</b> E214 <b>+/+</b> <b>K224</b> <b>+/+</b>
Minor groove contacts	Q86 <b>+/+</b> R90 <b>+/+</b>	Q86 <b>+/+</b> R90 <b>+/+</b>
Active site	E97 <b>+/+</b> D142 <b>+/+</b> K168 <b>+/+</b> D178 <b>+/+</b>	E97 <b>+/+</b> D142 <b>+/+</b> <b>K168</b> <b>+/+</b> <b>D178</b> <b>-/-</b>

**+/+** Identical contacts to DNA are made in both subunits of both complexes or the conformation of residues in both subunits are similar to the SP-complex

**+/–** Only one subunit of a dimer makes contacts similar to the SP-complex

**-/-** None of the subunits of preSP-complex makes contacts similar to the SP-complex

Residues, making different contacts to DNA in the SP-complex and at least in one subunit of the preSP-complex are shown in boldface

**Supplementary Table S5.** Agel contacts with phosphates in SP- and preSP-complexes

Phosphate # (base of Agel recognition site)	Interacting residues from SP- complex*	Interacting residues from preSP-complex		
		both subunits	only A	only B
-2	–			
-1	–			
0	K224 D223(H <sub>2</sub> O)			
1 (A)	K224(H <sub>2</sub> O)			R139
2 (C)		K140		
3 (C)	V169 R174		V169(H <sub>2</sub> O)	V169
4 (G)	H68 S170 W88	H68 S170 W88		
5 (G)	Y72 R73(H <sub>2</sub> O) S201(H <sub>2</sub> O)	Y72 R73(H <sub>2</sub> O) S201(H <sub>2</sub> O)		
6 (T)	D83 T82	D83 T82	H206	
7	T82 S87 K79(H <sub>2</sub> O)	T82 S87 K79(H <sub>2</sub> O)		
8	S43(H <sub>2</sub> O) R44(H <sub>2</sub> O)	S43(H <sub>2</sub> O) R44(H <sub>2</sub> O)		
9	R44 K42	R44**		

\*(H<sub>2</sub>O) – the residue makes water-mediated hydrogen bond. Similar contacts are shaded in grey.

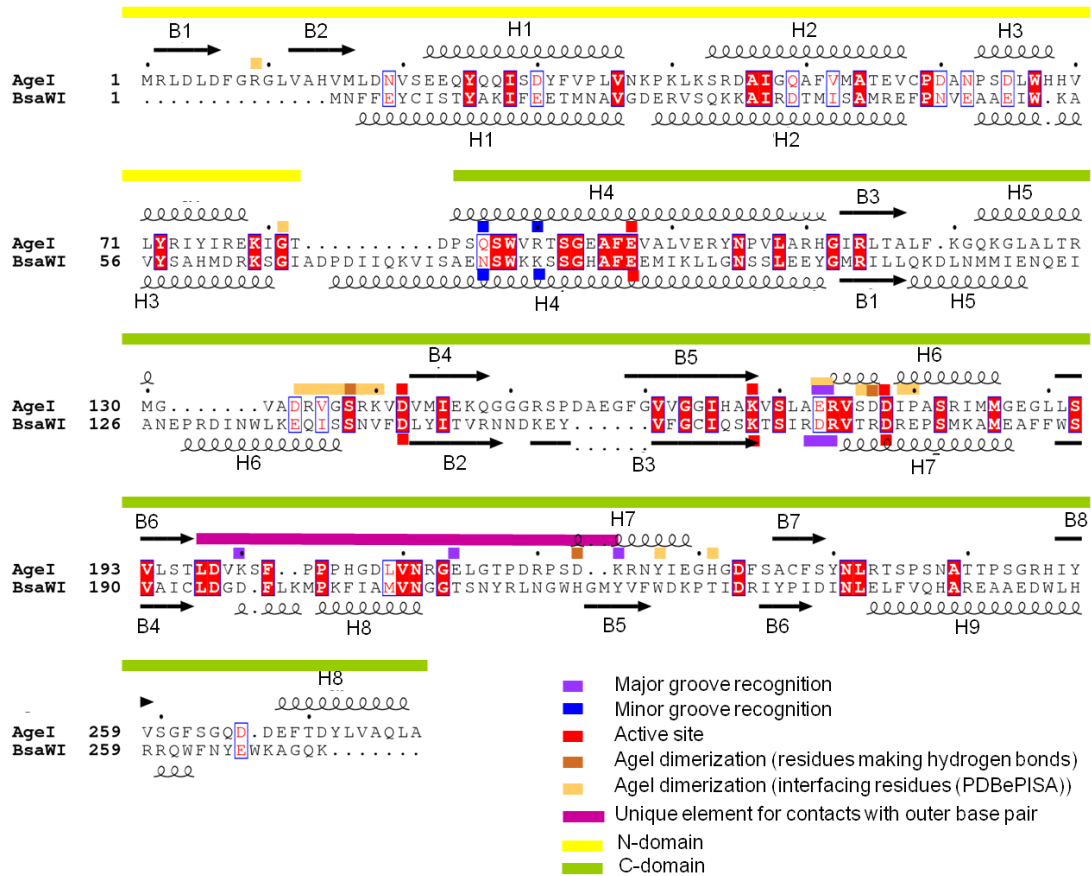
\*\*R44 makes hydrogen bond to O3\* atom of G8.

**Supplementary Table S6.** Gel filtration of Agel.

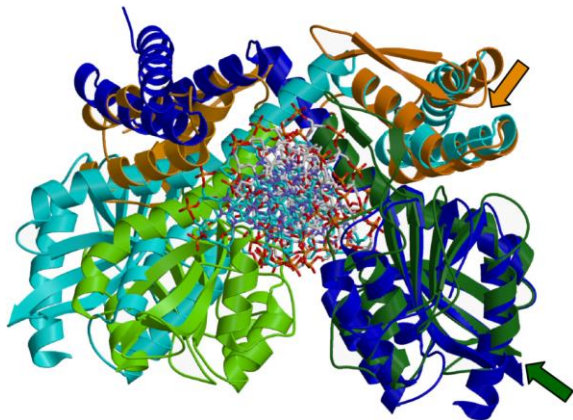
<b>Apo Agel concentration, <math>\mu\text{M}</math></b>	<b>Estimated <math>M_w</math>, kDa</b>
35	24.9
20	28.8
10	24.9
7	26.3
5	26.0
<b>Agel:DNA ratio</b>	<b>Estimated <math>M_w</math>, kDa</b>
1:0.5	65.3
1:1	64.3
1:5	67.1
1:10	66.3
1:20	67.2
<b>DNA</b>	<b>Estimated <math>M_w</math>, kDa</b>
SP	44

Calculated theoretical  $M_w$  for Agel monomer 32.5 kDa, dimer 65 kDa, DNA 18.4 kDa.

A

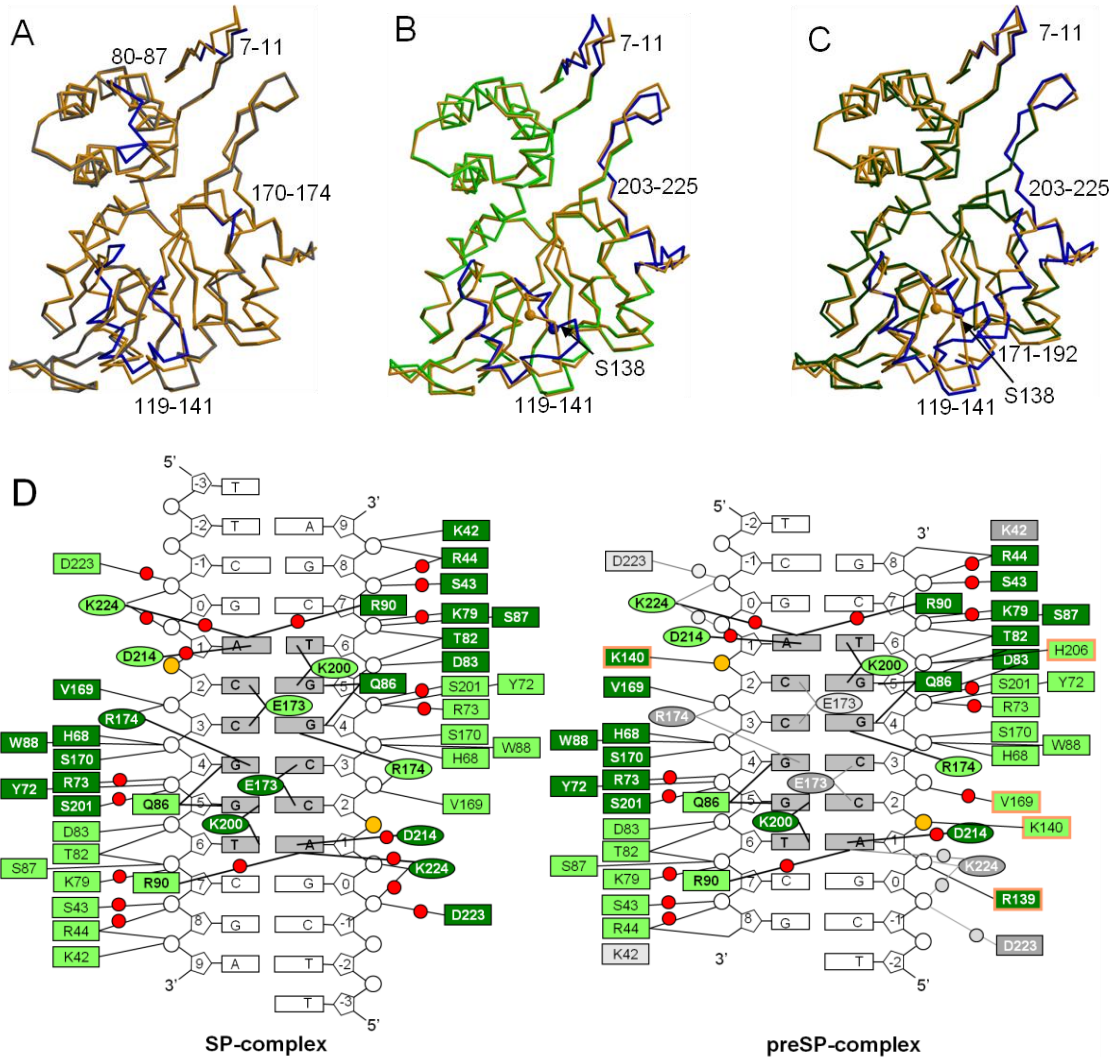


B

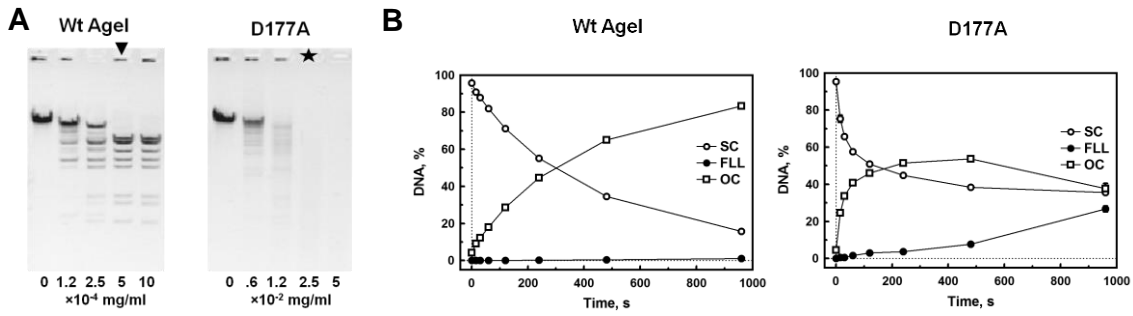


**Supplementary Figure S1.** Similarity between Agel and BsaWI. (A) Pairwise alignment of Agel and BsaWI sequences. Secondary structure elements of Agel and BsaWI are shown above and below the alignment, respectively. DNA recognition, active site and Agel dimerization residues are marked. The figure was prepared with ESPript (<http://esprpt.ibcp.fr>). (B) Structural overlay of Agel and BsaWI dimers. BsaWI subunits are coloured blue, N-domains of Agel are orange, C-domains are green. Orange and green arrows show overlay of the same Agel subunit with the domains of BsaWI from different subunits.

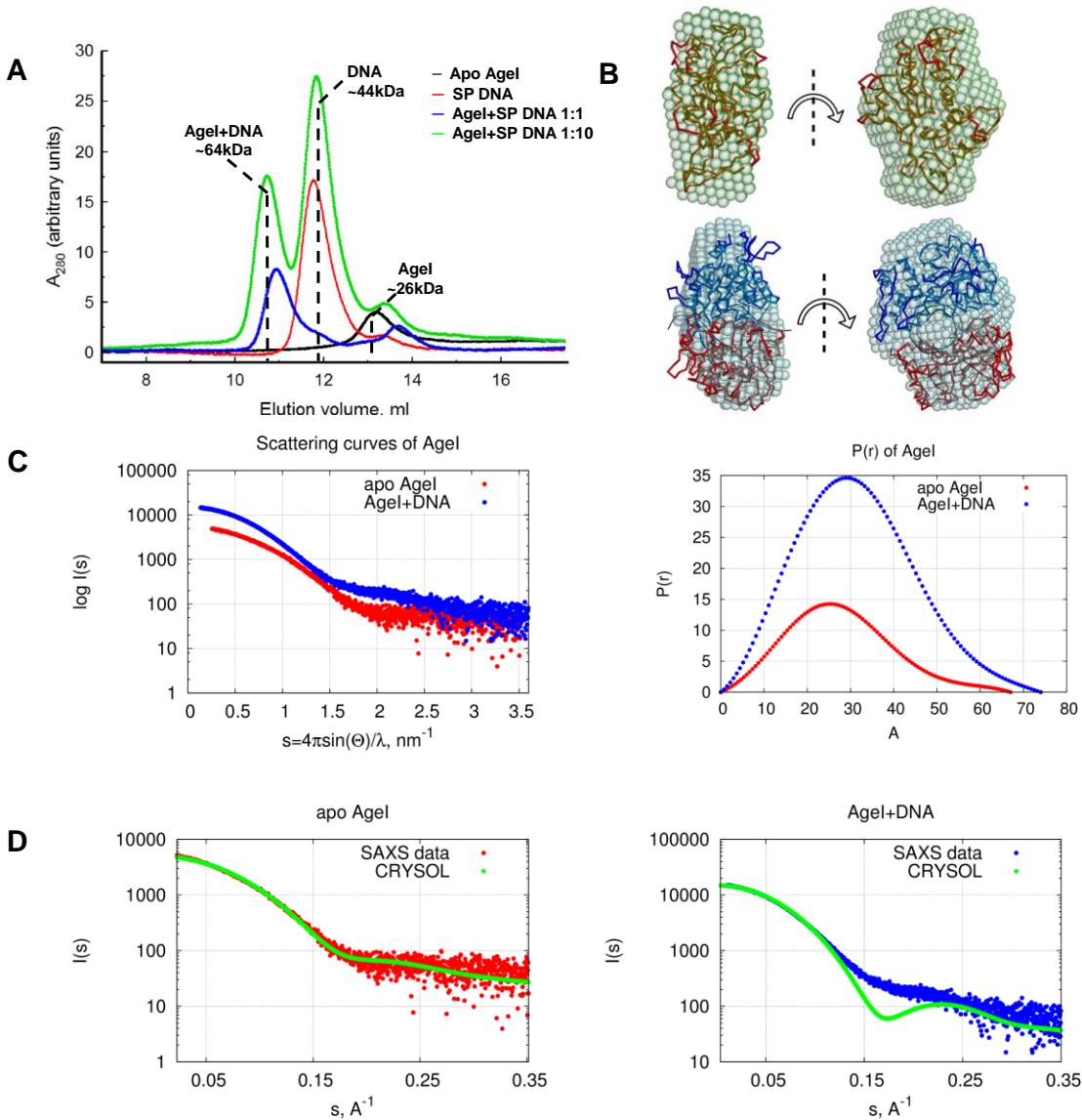




**Supplementary Figure S2.** Comparison of Agel structures. (A-C) Conformational differences of Agel in different structures. (A) Overlay of apo-Agel (grey) and A subunit of SP-complex (orange). Parts of the apo-subunit which do not overlay with the subunit of SP-complex are blue. Numbers indicate non-overlapping residues. (B-C) Overlay of A subunit of SP-complex (orange) and A subunit of preSP-complex (green, B) or B subunit of preSP-complex (dark green, C). Parts of the preSP-complex subunit which do not overlay with the subunit of SP-complex are blue. Ca atoms of S138 from the SP-complex subunit (orange) and the preSP-complex subunit (blue) are shown. (D) Comparison of Agel contacts with DNA in the SP- and preSP-complexes. Schematic representation of the Agel-DNA contacts: residues from the subunit A are coloured light green, from the subunit B - dark green; residues making contacts in the major groove are shown in ellipses; red spheres represent water molecules. Scissile phosphates are coloured orange. Missing contacts in the preSP-complex are shaded in grey, new contacts are boxed in orange.

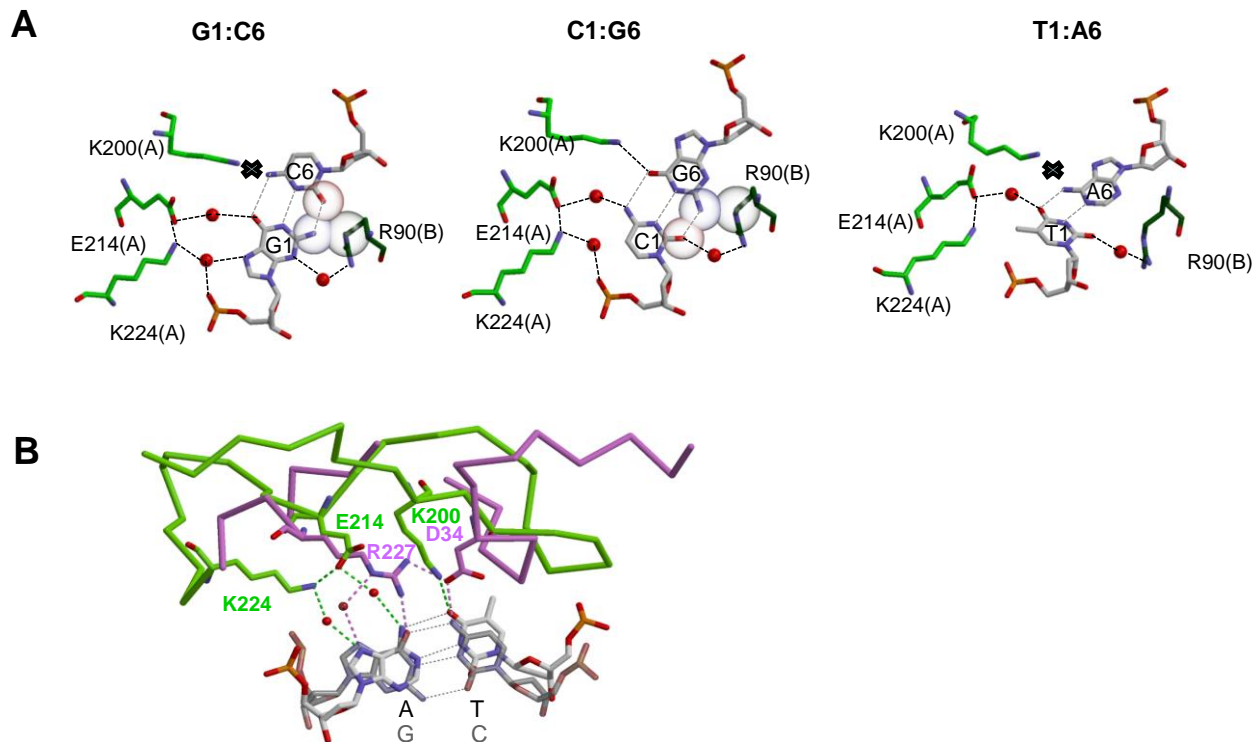


**Supplementary Figure S3.** Relaxed specificity of the D177A mutant analyzed using (A) phage  $\lambda$  DNA (13 Agel sites) and (B) plasmid pUC18 DNA (0 Agel site). The DNA cleavage reactions were performed and analyzed as described in 'Materials and Methods'. In (A) the final protein concentrations are indicated below the gels; a downward-facing triangle indicates the minimal enzyme concentration needed for complete digestion at the cognate recognition sites; a star indicates the minimal enzyme concentration needed for complete DNA digestion. In (B) the rate constant values ( $2.6 \pm 0.1 \times 10^{-3} \text{ s}^{-1}$  for Wt Agel and  $2.0 \pm 0.1 \times 10^{-2} \text{ s}^{-1}$  for D177A) were obtained by fitting a single exponential to the time-courses of supercoiled form depletion.



**Supplementary Figure S4.** Agel structure in solution. (A) Gel filtration analysis of Agel. Apo Agel (black line) elutes from the column as a 26 kDa protein with a molecular weight close to the calculated Mw of the monomer (calculated Mw of apo Agel 32kDa). SP DNA (red) elutes from the column at a volume corresponding to 44 kDa possibly due to the elongated form of the molecule (calculated Mw is 18.4 kDa). Agel-DNA complex elutes from the column as a 64 kDa species (blue and green). Experimentally determined Mw's are shown on the graph. Gel filtration was carried out as described in 'Materials and Methods'. (B) Averaged DAMMIN models of apo-Agel (transparent green spheres) and Agel-SP13 complex (light blue transparent spheres) complex were superimposed with the apo (monomer) and SP-complex (dimer) crystal structures, respectively (shown as backbone with different monomer colours). (C). SAXS data of apo-Agel (red curves) and Agel-SP13 complex (blue curves). (D). Theoretical scattering curves (green) calculated from crystal structures of apo-Agel (left panel) and SP-complex (right panel) by CRY SOL (35) compared with the experimental scattering curves.





**Supplementary Figure S7.** Outer bp recognition by Agel. (A) *In silico* models of Agel interaction with the non-cognate C1:G6, G1:C6 and T1:A6 base pairs. X represents no possibility to form hydrogen bonds between two donors. (B) Outer bp recognition elements of Agel (green) and NgoMIV (violet) in the overlaid structures. DNA from NgoMIV complex is transparent.