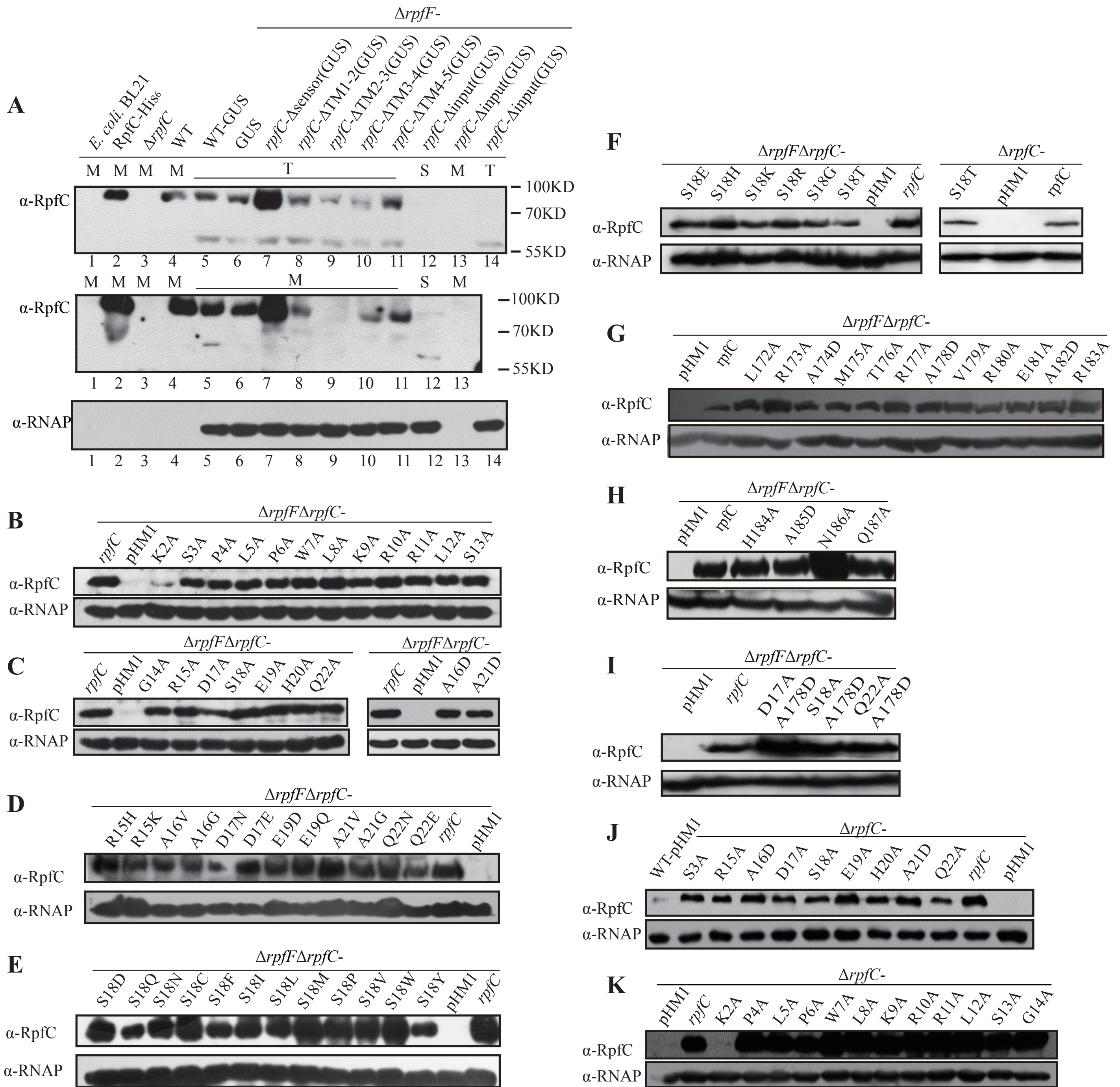


**S2 Fig. Stability of RpfC in various bacterial mutants.** (A) Stability of RpfC in strains with mutations in both *rpfF* and various input regions of *rpfC*. All strain names containing GUS show that the strain contains a P*engXcc-GUS* biosensor provided *in trans* by the pHM2 vector. Full-length RpfC with C-terminal $His_6$ tag embedded in inverted membrane vesicles (IMV) (upper panel, lane 2) was used as a positive control. Total proteins (T), membrane proteins (M), and soluble cytoplasmic proteins (S) from various bacterial strains were extracted, fractionated, and separated by 12% SDS-PAGE. Upper and middle panels: RpfC in different cellular fractions. Lower panel: loading control. (B-I) Stability of various RpfC proteins in Δ*rpfF*Δ*rpfC* double mutant mutant background. *rpfC* and its various point-mutated forms were provided *in trans* by recombinant pHM1 vectors. Total proteins were extracted for western blotting. (J-K) Stability of various RpfC proteins in an Δ*rpfC* mutant background. *rpfC* and its various point-mutated forms were provided *in trans* by recombinant pHM1 vectors to complement the Δ*rpfC* mutant. Total proteins were extracted for western blotting. Western blotting was used to measure the amount of RpfC. Polyclonal antibodies against RpfC (α-RpfC) and RNA polymerase (α-RNAP) from *X. campestris* pv. *campestris* were used in western blotting to measure the amount of total proteins, and to compare sample loading. Each experiment was repeated independently three times. Samples were separated on a 12% SDS-PAGE gel and signals were detected using an EZ-ECL enhanced chemiluminescence kit.