

Use of bacteria to stabilize archaeological iron

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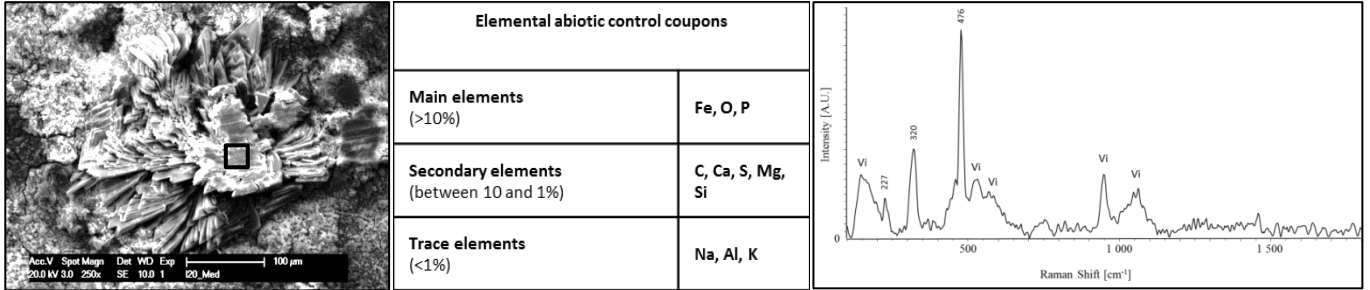
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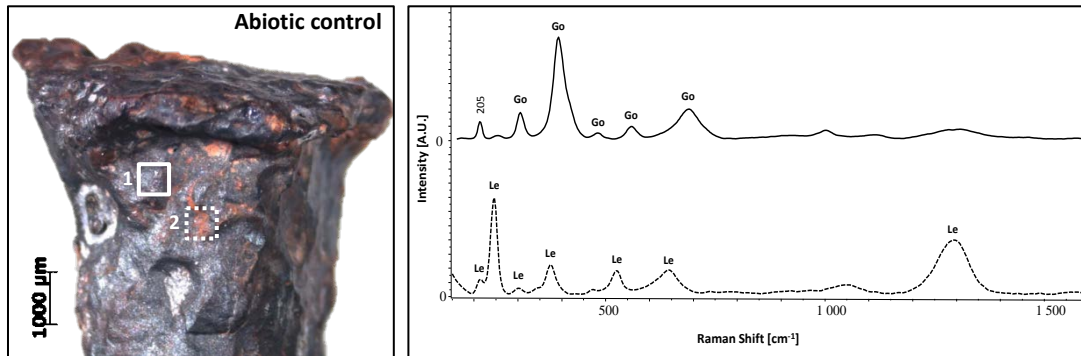
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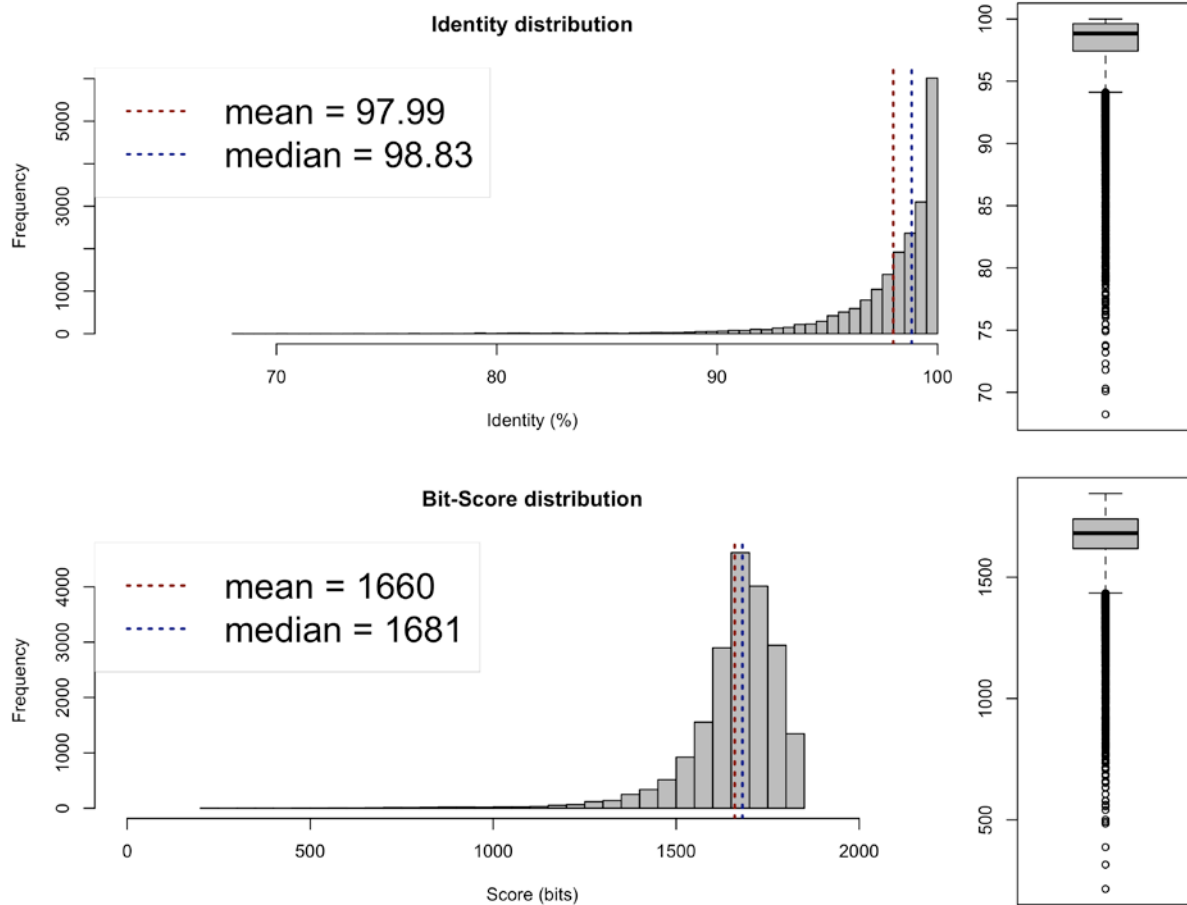
Supplementary material



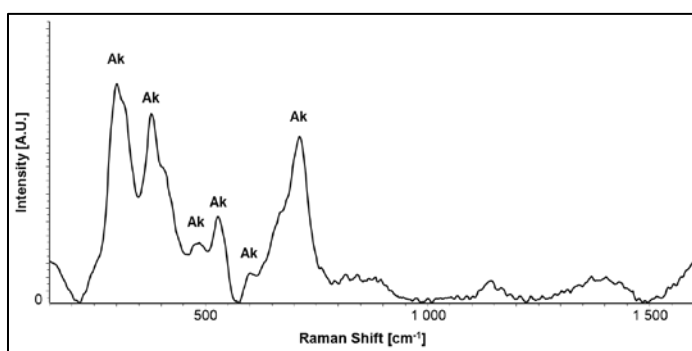
Supplementary Figure S1. Secondary elemental image of the crystals found on the abiotic control coupons, elemental composition, and corresponding Raman spectra identified as vivianite (Vi).



Supplementary Figure S2. Raman analysis of the abiotic control nail after the reduction test. On the left, the stereoscope images of the area sampled for the analysis; and, on the right, the corresponding Raman spectra identified as; 1: Goethite (Go), 2: Lepidocrocite (Le).



Supplementary Figure S3. Results of the average nucleotide identity (ANI) analysis performed between *Desulfitobacterium hafniense* strain LBE and the reference genome of *D. hafniense* strain DCB-2 (GenBank accession number NC_011830.1). The analysis was performed using the on-line ANI calculator tool available at <http://enve-omics.ce.gatech.edu>.



Supplementary Figure S4. Raman spectrum of the synthetic akaganeite. The analysis was carried out with a Horiba-Jobin Yvon Labram Aramis microscope equipped with a Nd:YAG laser of 532 nm at power lower than 1 mW (600 gr/mm). The spectral interval was 100 and 1600 cm⁻¹ and the measurement conditions were 1000 μm hole, 100 μm slit and 5 accumulations of 100 s.

Supplementary material: Anaerobic standard medium for *Desulfitobacterium hafniense*

Standard minimal medium (MM) according to Prat L, Maillard J, Grimaud R, Holliger C. 2011.

Physiological adaptation of *Desulfitobacterium hafniense* strain TCE1 to tetrachloroethene respiration. Applied and environmental microbiology **77**:3853-3859.

Mixture for 50 mL of culture, add aseptically with syringes the following solutions to 45 mL of **solution A**:

- 1.25 mL solution B
- 2.50 mL solution C
- 1.25 mL solution D
- 0.5 mL sodium lactate 40% (v/v)
- 1 mL disodium fumarate 16% (v/v)

Solutions

Solution A :

- $K_2HPO_4 \cdot 3 H_2O$ 0.958 g/L
- $NaH_2PO_4 \cdot 2H_2O$ 0.218 g/L
- Peptone 0.1 g/L
- Resazurin 0.5g/L 1 mL

➔ Boil, cool down under N_2/CO_2 , distribute to anaerobic flasks, gas exchange for N_2/CO_2 , autoclave.

Solution B :

To 20 mL of anaerobic sterile H_2O , add the following solutions:

- 1 mL solution IV, filter sterilize
- 1 mL solution V
- 1 mL solution VI
- 1 mL solution VII
- 1 mL solution VIII

Solution C :

To 49 mL of solution IX, add :

- 1 mL solution X

Solution D :

- $CaCl_2 \cdot 2 H_2O$ 4.40 g/L
- $MgCl_2 \cdot 6 H_2O$ 4.06 g/L

→ Gas exchange for N₂, autoclave.

Solution IV : Trace elements

- EDTA 500 mg/L, dissolve in 900 mL H₂O, adjust the pH to 7.0 with HCl, then add:
- FeCl₂·4 H₂O 2000 mg/L
- MnCl₂·4 H₂O 100 mg/L
- CoCl₂·6 H₂O 190 mg/L
- ZnCl₂ 70 mg/L
- CuCl₂·2 H₂O 2.55 mg/L
- AlCl₃ 5.52 mg/L
- H₃BO₃ 6 mg/L
- Na₂MoO₄·2 H₂O 41.4 mg/L
- NiCl₂·6 H₂O 24 mg/L, add to 1 L with H₂O

Solution V : Vitamins-1

- Biotin 50 mg/L
 - P-aminobenzoate (sodium salt) 250mg/L
 - Pantothenate (sodium salt) 50mg/L
 - Folic acid·2 H₂O 20mg/L
 - Lipoic acid 50mg/L
 - Pyridoxine 100mg/L
 - Nicotinic acid 550mg/L
- Filter sterilize in sterile anaerobic flasks, gas exchange for N₂.

Solution VI : Vitamins-2

- Thiamine-HCl 100 mg/L
- Filter sterilize in sterile anaerobic flasks, gas exchange for N₂.

Solution VII : Vitamins-3

- Riboflavine 50 mg/L
- Filter sterilize in sterile anaerobic flasks, gas exchange for N₂.

Solution VIII : Vitamins-4

- Cyanocobalamin 250 mg/L
- Filter sterilize in sterile anaerobic flasks, gas exchange for N₂.

Solution IX :

- NH₄HCO₃ 9.01g
 - NaHCO₃ 76.11 g/L
- Boil, cool down under N₂/CO₂, distribute 49 mL to anaerobic flasks, gas exchange for N₂/CO₂, autoclave.

Solution X :

- $\text{Na}_2\text{S}\cdot 9\text{H}_2\text{O}$ 24.02 g in 100 mL
- ➔ Wash crystals with N_2 -degassed H_2O , weight, dissolve in degassed H_2O , filter sterilize into anaerobic flasks, gas exchange for N_2 .