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Vaccines against human diarrheal pathogens: Current status and perspectives

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Vaccines against human diarrheal pathogens: current status and perspectives

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Further important bacterial pathogens causing diarrhea

Enteropathogenic *Escherichia coli* **(EPEC)**

With more than one million estimated deaths of children every year, EPEC is the main cause of diarrhea in children $¹$ [.](#page-10-0) By means of a type III secretion system (T3SS) encoding by the</sup> pathogenicity island LEE (locus of enterocyte effacement)^{[2](#page-10-1)}, EPEC can attach to intestinal epithelial cells and initiate the formation of pedestal ^{[3](#page-10-2)}. The T3SS translocates effector TIR molecule into host cells, that acts as a receptor for the non-fimbrial adhesin intimin expressed on the surface of EPEC^{[4](#page-10-3)}. Additionally, some EPEC strains harbor the adherence factor (EAF) plasmid, encoding for bundle-forming pili that contribute to formation of microcolonies and subsequent assembly of biofilms.

Campylobacter **spp.**

Campylobacter spp., a microaerophil ε-proteobacterium, currently causes more cases of diarrhea than foodborne *Salmonella* and is a major cause of acute gastroenteritis throughout the world. Campylobacteriosis caused by *Campylobacter* spp. is a zoonotic infection. The predominant natural host for this pathogen is poultry and the pathogen can be transmit by undercook meat and raw animal products from poultry and other livestock animals (WHO October 2011). Because of their high motility, *Campylobacter* easily penetrates the viscous intestinal mucus and then travel to the deeper intestinal crypts in which it comes to a ulcus inflammation and cryptabscess [5](#page-10-4) . A further complication of the recurrent *C. jejuni* is a neuropathy, the Guillain-Barré syndrome ^{[6](#page-10-5)}. It is initiated through the production of autoantibodies induced by the hyper-variable sequences in the genome, which encode the genes for the biosynthesis or modification of surface structures with lipopolysaccharides and capsule polysaccharides ^{[7](#page-10-6)}. Most symptoms of Campylobacteriosis are treated with substitution of water and electrolytes and, in more complicated courses of disease, by antibiotic therapy. Currently there is no vaccine against *Campylobacter* infections available.

Non-typhoidal *Salmonella enterica*

Non-typhoidal Salmonellosis is a zoonosis and can be transmitted by oral ingestion of contaminated food and water or by direct contact to pets or livestock animals. The Gram-negative rod-shaped and motile γ-proteobacteria are armed with a large array of virulence factors 8 . Following the adhesion to enterocytes, *Salmonella* Pathogenicity Island 1 (SPI1)-encoded T3SS enables an active invasion by the trigger mechanism, similar to *Shigella* spp*.* SPI1- T3SS effector induce n actin remodeling leading to macropinocytosis and uptake of the pathogen. The intracellular lifestyle of *Salmonella* is characterized by formation of the *Salmonella-containing vacuole* (SCV), a modified phagosome with a maturation process manipulated by SPI2-T3SS, leading to intracellular replication of the pathogen ^{[9](#page-10-8)}. Because of the selflimiting gastroenteritis in the intestine, where the stimulated inflammatory response contributes to diarrhea, an antibiotic treatment is unnecessary but the symptoms can be treated.

Aeromonas **spp.**

Infections with *Aeromonas* spp. cause gastrointestinal diseases and nosocomial complication, for example the 'flesh-eating bacteria syndrome' as worst outcome of *A. hydrophila* infections. Although causing these kind of infections, the Gram-negative rods of the *Vibrionaceae* family belong to bacteria with a low pathogenicity and are classical opportunists like many Enterobacteriaceae. The *Aeromonas* cytotoxic enterotoxins result in degeneration of crypts and villi of the small intestine. This is supported by another virulence factor of some strains, secreted aerolysin, which permeabilizes susceptible membranes by channel formation, similar to the α-toxin of *Staphylococcus aureus* [1](#page-10-0) . Gastroenteritis triggered by these toxins ranges from a mild, self-limiting watery diarrhea to a dysenteric form 10 . Because of the self-limiting course of disease, treatment will only address the resolution of symptoms without application of antibiotics.

Proteus **spp.**

The genus *Proteus* describes Gram-negative rods, which occur as putrefactive agents in water, soil, on the surface of carcasses, in some food and in the natural intestinal flora. Typical characteristic for these opportunistic bacteria is the rapid motility with swarming growth and the production of the enzyme urease (reviewed in $\frac{11}{1}$), which hydrolyzes urea in ammonia and carbon dioxide. This leads to an increased pH value and the precipitation of magnesium and calcium salts. Because of this, *P. mirabelis* is the most frequent cause of infection-related kidney stones. The production of ammonia in close proximity to the gastric epithelium causes cell damage and inflammation reactions [12.](#page-10-11) Further areas can be infected by *Proteus* bacteria, like the respiratory tract, skin and urinary tract. It is also a nosocomial pathogen like *Clostrid-* *ium difficile* and can be treated with antibiotics and prevented with higher hygiene standards combined with other control methods for enteric pathogens.

Enterobacter **spp.**

Enterobacter spp. are Gram-negative, rod-shaped, motile and opportunistic members of Enterobacteriaceae and can cause various disease patterns such as urinary tract and abdominal infections. One of the four pathogenic strains, *E. sakazakii,* has been proposed for reclassification as new genus *Cronobacter* by Iversen *et al*. [13.](#page-10-12) *Cronobacter* can contaminate powdered milk and infant formulas 14 , leading to necrotizing enterocolitis (NEC) in infants 15 . In vitro experiments have shown that *C. sakazakii* disrupt tight junctions, induce monolayer permeability and cause apoptosis by binding to intestinal epithelial cells ¹⁶. Due to the lack of knowledge on the action cause of NEC, the treatment involves different approaches like the administration of probiotics, antibiotics, surgical examination, Epidermal Growth Factor (EGF) and more 17 .

Yersinia **spp.**

Yersinia enterocolitica, Y. pseudotuberculosis and *Y. pestis* are the three only human pathogen species of this genus of Gram-negative, rod-shaped, and zoonotic bacteria. The two first named species cause enterocolitis, mesenteric lypmphadenitis, septicemia and reactive arthritis. *Yersinia* spp. use for invading and colonization the Peyer's patches and the M cells for transport ¹⁸. Furthermore, they can resist in phagocytic cells, which transfer the pathogens in the spleen. The virulence factors for adhesion to and invasion of host cells are encoded chromosomally (outer membrane proteins Inv and Ail) and the virulence plasmid (YadA). Further plasmid-encoded antiphagocytic factors (Yop proteins) and a T3SS (Ysc) lead to inactivation of phagocytes [19.](#page-10-18) Because of the similarity in contamination of food between *Salmonella* and *Yersinia* the prevention is the same. Only in serious and chronic cases an antibiotic therapy is necessary.

Staphylococcus aureus

One of the most common and clinical important pathogen is *S. aureus.* It synthesizes an array of different virulence factors, for example the staphylococcal enterotoxins (SEs) that trigger food-mediated intoxications and lead to diarrhea and vomiting 20 . The SEs are often found in dairy products and meats, if improperly stored or processed 20 . The targets of these toxins are the major histocompatibility complex class II (MHCII) on antigen-presenting cells and the Tcell receptor (TCR) on T-cells, where they act as super antigens $2¹$. Characteristic SEs are high thermostability and low degradation in the stomach and intestinal tract 22 . Food poisoning caused by SE is not lethal 20 . Depending on the toxin, rapamycin is effective after an SE B intoxication 2^0 . The generation of clinically efficacious vaccines has failed so far 2^3 .

Clostridium difficile

Clostridium difficile are Gram-positive, spore-forming, thick rod-shaped bacteria and the major cause for diseases in hospitals associated with broad-spectrum antibiotic treatment. Antibiotic therapy causes alterations of the normal colonic microflora 24 and induces stress responses in bacteria such as *C. difficile*. This pathogen produces two toxins, the enterotoxin A (TcdA) ^{[25](#page-11-3)} and the cytotoxin B (TcdB), which provoke diarrhea and pseudomembranous colitis 26 . Traditionally, the treatment consists of curing the symptoms and discontinuing the initial antibiotic therapy. Then, the administration of oral metronidazole or oral vancomycin is reasonable. Metronidazole is the first choice therapy in order to prevent proliferation of vancomycin-resistant nosocomial pathogens 24 . Several new approaches for treatment are under investigation, for example vaccines, toxin-binding agents, flora-sparing antibiotics and antibodies [27.](#page-11-5)

Bacillus cereus

The Gram-positive, spore-forming species *B. cereus* also produce a variety of toxins that causes gastrointestinal diseases 28 . It is present in nature, and in rice, dairy and meat products and vegetables 29 . Normally, the induced diarrhea is self-limiting and heals quickly 30 . There are three enterotoxins, i) the hemolytic enterotoxin hemolysin BL (HBL), ii) the nonhemolytic enterotoxin (Nhe) and iii) cytotoxin K (CytK), that play major roles in diarrheal infection, but there are others causing vomiting (emetic type) 31 . It is assumed that the three toxins have an destructive effect on epithelial cell membrane integrity in the small intestine ³¹. Similar to *S. aureus, B. cereus* is resistant against many antibiotics ²⁹. A specific vaccine has not been reported.

Protozoans causing diarrheal diseases

Entamoeba histolytica

The protozoan parasite *Entamoeba histolytica* causes invasive amoebic dysentery and can lead to death if not treated [32,](#page-11-10) [33.](#page-11-11) It is a cosmopolitan pathogen, which is most common in tropic regions [34,](#page-11-12) where poor sanitary standards prevail and water and food are often contaminated with feces ³². Annually, 50 million people worldwide contract an amoebic dysentery and about 40,000 to 100,000 cases are fatal [35.](#page-11-13) *E. histolytica* secretes proteases that degrade red blood cells and tissues [33.](#page-11-11) For treating amoebiasis, metronidazole and nitazoxanide are the most widely used drugs [36.](#page-11-14)

Giardia lamblia

Giardia lamblia is a flagellated human pathogenic protozoan and causative agent of giardiasis and noninvasive diarrhea [37,](#page-11-15) [38.](#page-11-16) *G. lamblia* exist in two stages, an inactive form as cyst found in polluted water and food, and in vegetative form as trophozoites in human ³⁸. Inside the intestine, the parasite changes its surface molecules continuously, allowing to escape immune responses and repeating infections [39.](#page-11-17) Worldwide, more than 280 million people suffer from giardiasis every year [40.](#page-11-18) For treatment, metronidazole is prescribed, but there are already metronidazole-resistant strains. New drugs are required, one could be auranofin, which blocks the thioredoxin oxidoreductase activity of *G. lamblia* [41.](#page-11-19)

Cryptosporidia

The water-borne, obligate intracellular pathogen *Cryptosporidium* is part of the phylum Apicomplexa and causes diarrhea in humans [42-44.](#page-11-20) Especially *C. parvum* invades gastrointestinal epithelial cells [45.](#page-11-21) An infection with Cryptosporidia is self-limiting in healthy humans, but can be fatal for persons with reduced immune functions ⁴⁶. The infective form are oocysts and only a small amount is required 44 . In form of oocysts, the protozoans are tolerant against most of the chemicals used of water disinfection, such as chloride, and they survive for several months in nature [44.](#page-11-23) Invasion is mediated via glycoproteins and circumsporozoite surface ligand (CSL). After infection, the microvilli are destroyed and loss of enzymes and ions is detected [44.](#page-11-23) For treating a *Cryptosporidium* spp*.* infection, paromomycin and nitazoxanide were proved to be effective 47 . A vaccine against Cryptosporidia is pending.

Further viruses causing diarrheal diseases

Norovirus

Another pandemic virus is norovirus, which is also found in feces and contaminated water and food $48, 49$ $48, 49$. The RNA virus is non-enveloped and contains a capsid 50 . GII4 is the most widespread human-specific *norovirus* strain and antigen and receptor variation lead to recurrent infections [51.](#page-12-1) Additionally, it is not cultivable, most of the information are based on RT-PCR [48.](#page-11-25) Mortality of estimated 200,000 children under 5 years was reported for developing countries ⁵². Hitherto, there are no vaccines available, but there is a new approach based on blocking the histo-blood group antigens that norovirus uses as receptor**.**

Table S1. Important microbial pathogens causing diarrhea

Suppl. References

- 1. Wilson M, McNab R, Henderson B. Bacterial disease mechanisms : an introduction to cellular microbiology. Cambridge ; New York, NY: Cambridge University Press, 2002.
- 2. McDaniel TK, Jarvis KG, Donnenberg MS, Kaper JB. A genetic locus of enterocyte effacement conserved among diverse enterobacterial pathogens. ProcNatlAcadSciU S A 1995; 92:1664-8.
- 3. Croxen MA, Finlay BB. Molecular mechanisms of *Escherichia coli* pathogenicity. Nat Rev Microbiol 2010; 8:26-38.
- 4. Kenny B, Jepson M. Targeting of an enteropathogenic *Escherichia coli* (EPEC) effector protein to host mitochondria. Cell Microbiol 2000; 2:579-90.
- 5. Janssen R, Krogfelt KA, Cawthraw SA, van Pelt W, Wagenaar JA, Owen RJ. Host-pathogen interactions in Campylobacter infections: the host perspective. Clin Microbiol Rev 2008; 21:505-18.
- 6. Vucic S, Kiernan MC, Cornblath DR. Guillain-Barre syndrome: an update. J Clin Neurosci 2009; 16:733-41.
- 7. Parkhill J, Wren BW, Mungall K, Ketley JM, Churcher C, Basham D, et al. The genome sequence of the food-borne pathogen *Campylobacter jejuni* reveals hypervariable sequences. Nature 2000; 403:665-8.
- 8. Haraga A, Ohlson MB, Miller SI. Salmonellae interplay with host cells. Nat Rev Microbiol 2008; 6:53-66.
- 9. Kuhle V, Hensel M. Cellular microbiology of intracellular *Salmonella enterica*: functions of the type III secretion system encoded by Salmonella pathogenicity island 2. Cell Mol Life Sci 2004; 61:2812-26.
- 10. Janda JM, Duffey PS. Mesophilic aeromonads in human disease: current taxonomy, laboratory identification, and infectious disease spectrum. Rev Infect Dis 1988; 10:980-97.
- 11. Rozalski A, Sidorczyk Z, Kotelko K. Potential virulence factors of *Proteus* bacilli. Microbiol Mol Biol Rev 1997; 61:65-89.
- 12. Follmer C. Ureases as a target for the treatment of gastric and urinary infections. J Clin Pathol 2010; 63:424-30.
- 13. Iversen C, Lehner A, Mullane N, Bidlas E, Cleenwerck I, Marugg J, et al. The taxonomy of Enterobacter sakazakii: proposal of a new genus Cronobacter gen. nov. and descriptions of Cronobacter sakazakii comb. nov. Cronobacter sakazakii subsp. sakazakii, comb. nov., Cronobacter sakazakii subsp. malonaticus subsp. nov., Cronobacter turicensis sp. nov., Cronobacter muytjensii sp. nov., Cronobacter dublinensis sp. nov. and Cronobacter genomospecies 1. BMC Evol Biol 2007; 7:64.
- 14. van Acker J, de Smet F, Muyldermans G, Bougatef A, Naessens A, Lauwers S. Outbreak of necrotizing enterocolitis associated with *Enterobacter sakazakii* in powdered milk formula. J Clin Microbiol 2001; 39:293-7.
- 15. Yan QQ, Condell O, Power K, Butler F, Tall BD, Fanning S. *Cronobacter* species (formerly known as *Enterobacter sakazakii*) in powdered infant formula: a review of our current understanding of the biology of this bacterium. J Appl Microbiol 2012; 113:1-15.
- 16. Liu Q, Mittal R, Emami CN, Iversen C, Ford HR, Prasadarao NV. Human isolates of *Cronobacter sakazakii* bind efficiently to intestinal epithelial cells in vitro to induce monolayer permeability and apoptosis. J Surg Res 2012; 176:437-47.
- 17. Henry MC, Moss RL. Necrotizing enterocolitis. Annu Rev Med 2009; 60:111-24.
- 18. Grutzkau A, Hanski C, Hahn H, Riecken EO. Involvement of M cells in the bacterial invasion of Peyer's patches: a common mechanism shared by *Yersinia enterocolitica* and other enteroinvasive bacteria. Gut 1990; 31:1011-5.
- 19. Viboud GI, Bliska JB. *Yersinia* outer proteins: role in modulation of host cell signaling responses and pathogenesis. Annu Rev Microbiol 2005; 59:69-89.
- 20. Krakauer T, Stiles BG. The staphylococcal enterotoxin (SE) family: SEB and siblings. Virulence 2013; 4.
- 21. Krakauer T. Update on staphylococcal superantigen-induced signaling pathways and therapeutic interventions. Toxins (Basel) 2013; 5:1629-54.
- 22. Argudin MA, Mendoza MC, Rodicio MR. Food poisoning and *Staphylococcus aureus* enterotoxins. Toxins (Basel) 2010; 2:1751-73.
- 23. Kobayashi SD, Deleo FR. *Staphylococcus aureus* Protein A Promotes Immune Suppression. MBio 2013; 4.
- 24. Kelly CP, LaMont JT. *Clostridium difficile* infection. Annu Rev Med 1998; 49:375-90.
- 25. Kyne L, Warny M, Qamar A, Kelly CP. Asymptomatic carriage of *Clostridium difficile* and serum levels of IgG antibody against toxin A. N Engl J Med 2000; 342:390-7.
- 26. Drudy D, Fanning S, Kyne L. Toxin A-negative, toxin B-positive *Clostridium difficile*. Int J Infect Dis 2007; 11:5-10.
- 27. Rupnik M, Wilcox MH, Gerding DN. *Clostridium difficile* infection: new developments in epidemiology and pathogenesis. Nat Rev Microbiol 2009; 7:526-36.
- 28. Ramarao N, Sanchis V. The pore-forming haemolysins of *Bacillus cereus*: a review. Toxins (Basel) 2013; 5:1119-39.
- 29. Bottone EJ. Bacillus cereus, a volatile human pathogen. Clin Microbiol Rev 2010; 23:382-98.
- 30. CDC CfDCaP. Bacillus cereus food poisoning associated with fried rice at two child day care centers--Virginia, 1993. MMWR Morb Mortal Wkly Rep 1994; 43:177-8.
- 31. Senesi S, Ghelardi E. Production, secretion and biological activity of *Bacillus cereus* enterotoxins. Toxins (Basel) 2010; 2:1690-703.
- 32. Marie C, Petri WA, Jr. Amoebic dysentery. Clin Evid (Online) 2013; 2013.
- 33. Stanley SL, Jr. Amoebiasis. Lancet 2003; 361:1025-34.
- 34. Ho G, Sanchez MM, Leonard P, Van Esbroeck M, Hayette MP. [Amoebic liver abscess contracted in India with diagnosis confirmed by PCR]. Rev Med Liege 2013; 68:428-32.
- 35. Ocadiz-Ruiz R, Fonseca W, Martinez MB, Ocadiz-Quintanar R, Orozco E, Rodriguez MA. Effect of the silencing of the Ehcp112 gene on the in vitro virulence of *Entamoeba histolytica*. Parasit Vectors 2013; 6:248.
- 36. Chacin-Bonilla L. [Current pharmacotherapy of amebiasis, advances in new drugs, and design of a vaccine]. Invest Clin 2012; 53:301-14.
- 37. Moreno-Galindo EG, Rodriguez-Elias JC, Ramirez-Herrera MA, Sanchez-Chapula JA, Navarro-Polanco RA. The principal conductance in Giardia lamblia trophozoites possesses functional properties similar to the mammalian ClC-2 current. Pflugers Arch 2013.
- 38. Adam RD. Biology of *Giardia lamblia*. Clin Microbiol Rev 2001; 14:447-75.
- 39. Lujan HD. Mechanisms of adaptation in the intestinal parasite Giardia lamblia. Essays Biochem 2011; 51:177-91.
- 40. Lane S, Lloyd D. Current trends in research into the waterborne parasite Giardia. Crit Rev Microbiol 2002; 28:123-47.
- 41. Tejman-Yarden N, Miyamoto Y, Leitsch D, Santini J, Debnath A, Gut J, et al. A reprofiled drug, auranofin, is effective against metronidazole-resistant Giardia lamblia. Antimicrob Agents Chemother 2013; 57:2029-35.
- 42. Mazurie AJ, Alves JM, Ozaki LS, Zhou S, Schwartz DC, Buck GA. Comparative genomics of cryptosporidium. Int J Genomics 2013; 2013:832756.
- 43. Bhalchandra S, Ludington J, Coppens I, Ward HD. Identification and characterization of *Cryptosporidium parvum* Clec, a novel C-type lectin domain-containing mucin-like glycoprotein. Infect Immun 2013; 81:3356-65.
- 44. Desai NT, Sarkar R, Kang G. Cryptosporidiosis: An under-recognized public health problem. Trop Parasitol 2012; 2:91-8.
- 45. O'Donoghue PJ. *Cryptosporidium* and cryptosporidiosis in man and animals. Int J Parasitol 1995; 25:139-95.
- 46. Leav BA, Mackay M, Ward HD. Cryptosporidium species: new insights and old challenges. Clin Infect Dis 2003; 36:903-8.
- 47. Kumar VP, Frey KM, Wang Y, Jain HK, Gangjee A, Anderson KS. Substituted pyrrolo[2,3 d]pyrimidines as *Cryptosporidium hominis* thymidylate synthase inhibitors. Bioorg Med Chem Lett 2013; 23:5426-8.
- 48. Stals A, Mathijs E, Baert L, Botteldoorn N, Denayer S, Mauroy A, et al. Molecular detection and genotyping of noroviruses. Food Environ Virol 2012; 4:153-67.
- 49. Ruvoen N, Le Pendu J. [Genetic susceptibility to norovirus infection]. Pathol Biol (Paris) 2013; 61:28-35.
- 50. Zhang XF, Tan M, Chhabra M, Dai YC, Meller J, Jiang X. Inhibition of histo-blood group antigen binding as a novel strategy to block norovirus infections. PLoS One 2013; 8:e69379.
- 51. Wu Q, Xue L, Zhang J. [Norovirus epidemic strain GII. 4 evolution--a review]. Wei Sheng Wu Xue Bao 2012; 52:1431-8.
- 52. Patel MM, Widdowson MA, Glass RI, Akazawa K, Vinje J, Parashar UD. Systematic literature review of role of noroviruses in sporadic gastroenteritis. Emerg Infect Dis 2008; 14:1224-31.
- 53. Tamang MD, Sharma N, Makaju RK, Sarma AN, Koju R, Nepali N, et al. An outbreak of El Tor cholera in Kavre district, Nepal. Kathmandu Univ Med J (KUMJ) 2005; 3:138-42.
- 54. Lopez-Gigosos RM, Plaza E, Diez-Diaz RM, Calvo MJ. Vaccination strategies to combat an infectious globe: oral cholera vaccines. J Glob Infect Dis 2011; 3:56-62.
- 55. WHO. Cholera vaccines: WHO position paper. Wkly Epidemiol Rec 2010; 85:117-28.
- 56. Ferreccio C, Prado V, Ojeda A, Cayyazo M, Abrego P, Guers L, et al. Epidemiologic patterns of acute diarrhea and endemic *Shigella* infections in children in a poor periurban setting in Santiago, Chile. Am J Epidemiol 1991; 134:614-27.
- 57. Schroeder GN, Hilbi H. Molecular pathogenesis of *Shigella* spp.: controlling host cell signaling, invasion, and death by type III secretion. Clin Microbiol Rev 2008; 21:134-56.
- 58. Peirano G, Souza FS, Rodrigues DP. Frequency of serovars and antimicrobial resistance in Shigella spp. from Brazil. Mem Inst Oswaldo Cruz 2006; 101:245-50.
- 59. Lengerh A, Moges F, Unakal C, Anagaw B. Prevalence, associated risk factors and antimicrobial susceptibility pattern of Campylobacter species among under five diarrheic children at Gondar University Hospital, Northwest Ethiopia. BMC Pediatr 2013; 13:82.
- 60. Su CC, Radhakrishnan A, Kumar N, Long F, Bolla JR, Lei HT, et al. Crystal structure of the Campylobacter jejuni CmeC outer membrane channel. Protein Sci 2014.
- 61. Minnaganti VR, Patel PJ, Iancu D, Schoch PE, Cunha BA. Necrotizing fasciitis caused by Aeromonas hydrophila. Heart Lung 2000; 29:306-8.
- 62. Janda JM, Abbott SL. The genus Aeromonas: taxonomy, pathogenicity, and infection. Clin Microbiol Rev 2010; 23:35-73.
- 63. O'Hara CM, Brenner FW, Miller JM. Classification, identification, and clinical significance of Proteus, Providencia, and Morganella. Clin Microbiol Rev 2000; 13:534-46.
- 64. National Nosocomial Infections Surveillance (NNIS) report, data summary from October 1986- April 1996, issued May 1996. A report from the National Nosocomial Infections Surveillance (NNIS) System. Am J Infect Control 1996; 24:380-8.
- 65. Long C, Jones TF, Vugia DJ, Scheftel J, Strockbine N, Ryan P, et al. Yersinia pseudotuberculosis and Y. enterocolitica infections, FoodNet, 1996-2007. Emerg Infect Dis 2010; 16:566-7.
- 66. Gomez-Duarte OG, Romero-Herazo YC, Paez-Canro CZ, Eslava-Schmalbach JH, Arzuza O. Enterotoxigenic Escherichia coli associated with childhood diarrhoea in Colombia, South America. J Infect Dev Ctries 2013; 7:372-81.
- 67. Wenneras C, Erling V. Prevalence of enterotoxigenic Escherichia coli-associated diarrhoea and carrier state in the developing world. J Health Popul Nutr 2004; 22:370-82.
- 68. Das JK, Tripathi A, Ali A, Hassan A, Dojosoeandy C, Bhutta ZA. Vaccines for the prevention of diarrhea due to cholera, shigella, ETEC and rotavirus. BMC Public Health 2013; 13 Suppl 3:S11.
- 69. Harro C, Sack D, Bourgeois AL, Walker R, DeNearing B, Feller A, et al. A combination vaccine consisting of three live attenuated enterotoxigenic Escherichia coli strains expressing a range of colonization factors and heat-labile toxin subunit B is well tolerated and immunogenic in a placebo-controlled double-blind phase I trial in healthy adults. Clin Vaccine Immunol 2011; 18:2118-27.
- 70. Papatheodorou P, Hornuss D, Nolke T, Hemmasi S, Castonguay J, Picchianti M, et al. Clostridium difficile binary toxin CDT induces clustering of the lipolysis-stimulated lipoprotein receptor into lipid rafts. MBio 2013; 4:e00244-13.
- 71. McDonald LC LF, Sievert D, Wise M, Herrera R, Gould C, Malpiedi P, Dudeck M, Srinivasan A, Fridkin S, et al. Vital signs: preventing Clostridium difficile infections. . MMWR Morb Mortal Wkly Rep 2012; 61:157-62.
- 72. Hall AC CA, McDonald LC, Parashar UD, Lopman BA. The roles of norovirus and. Clostridium difficile among gastroenteritis deaths in the United States, 1997-2007. Presentation and the 49th annual Meeting of the Infectious Disease Society of America 2011.
- 73. Tian JH, Fuhrmann SR, Kluepfel-Stahl S, Carman RJ, Ellingsworth L, Flyer DC. A novel fusion protein containing the receptor binding domains of C. difficile toxin A and toxin B elicits protective immunity against lethal toxin and spore challenge in preclinical efficacy models. Vaccine 2012; 30:4249-58.
- 74. Serrano-Luna J, Pina-Vazquez C, Reyes-Lopez M, Ortiz-Estrada G, de la Garza M. Proteases from Entamoeba spp. and Pathogenic Free-Living Amoebae as Virulence Factors. J Trop Med 2013; 2013:890603.
- 75. Leippe M, Muller-Eberhard HJ. The pore-forming peptide of Entamoeba histolytica, the protozoan parasite causing human amoebiasis. Toxicology 1994; 87:5-18.
- 76. Petri WA, Jr., Haque R, Mann BJ. The bittersweet interface of parasite and host: lectincarbohydrate interactions during human invasion by the parasite Entamoeba histolytica. Annu Rev Microbiol 2002; 56:39-64.
- 77. Muller N, von Allmen N. Recent insights into the mucosal reactions associated with Giardia lamblia infections. Int J Parasitol 2005; 35:1339-47.
- 78. Fayer R, Morgan U, Upton SJ. Epidemiology of Cryptosporidium: transmission, detection and identification. Int J Parasitol 2000; 30:1305-22.
- 79. Tzipori S, Ward H. Cryptosporidiosis: biology, pathogenesis and disease. Microbes Infect 2002; 4:1047-58.
- 80. WHO WHO. WHO/PAHO informal consultation on intestinal protozoal infections. Tech Rep WHO/CDS/IPI/922, World Health Organisation, Geneva, Switzerland,, 1992.
- 81. O'Connor RM, Burns PB, Ha-Ngoc T, Scarpato K, Khan W, Kang G, et al. Polymorphic mucin antigens CpMuc4 and CpMuc5 are integral to *Cryptosporidium parvum* infection in vitro. Eukaryot Cell 2009; 8:461-9.
- 82. Rotavirus vaccines WHO position paper: January 2013 Recommendations. Vaccine 2013; 31:6170-1.
- 83. Ko EA, Jin BJ, Namkung W, Ma T, Thiagarajah JR, Verkman AS. Chloride channel inhibition by a red wine extract and a synthetic small molecule prevents rotaviral secretory diarrhoea in neonatal mice. Gut 2013.
- 84. Lorrot M, Vasseur M. [Physiopathology of Rotavirus diarrhea]. Arch Pediatr 2007; 14 Suppl 3:S145-51.
- 85. Tate JE, Burton AH, Boschi-Pinto C, Steele AD, Duque J, Parashar UD. 2008 estimate of worldwide rotavirus-associated mortality in children younger than 5 years before the introduction of universal rotavirus vaccination programmes: a systematic review and metaanalysis. Lancet Infect Dis 2012; 12:136-41.
- 86. National Center for Immunization and Respiratory Diseases (NCIRD) DoVD. Rotavirus Clinical Information. 2011.
- 87. Lee PI, Chen PY, Huang YC, Lee CY, Lu CY, Chang MH, et al. Recommendations for rotavirus vaccine. Pediatr Neonatol 2013; 54:355-9.
- 88. Ravn V, Dabelsteen E. Tissue distribution of histo-blood group antigens. APMIS 2000; 108:1- 28.
- 89. Tan M, Jiang X. Norovirus and its histo-blood group antigen receptors: an answer to a historical puzzle. Trends Microbiol 2005; 13:285-93.