SUPPLEMENTAL DATA

Molecular determinants of the N-terminal acetyltransferase Naa60 anchoring to the Golgi membrane

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Table S1. All mutations performed on Naa60, purpose of the mutation and consequence observed by microscopy and carbonate wash assay when applied. Abbreviations: <1/2, less than half of the proteins extracted; 1/2, half of the proteins extracted; >1/2, more than half of the proteins extracted; ϕ , hydrophobic amino acid; SF, by subcellular fractionation. Numbers in [] are internal plasmid numbers.

		Localization by microscopy		Org fraction extractable by high pH?	
Mutation	Concept tested	Cyto	Org		
Naa60-(1-242)-eGFP [134]	Ctrl, full-length		х	some (<1/2)	
C-truncations (Fig 3B/E)					
Naa60-(1-182)-eGFP [439]	Lacking last 61 aa	x			
Naa60-(1-216)-eGFP [207]	Lacking last 26 aa	x	х		
Naa60-(1-225)-eGFP [440]	Lacking last 17 aa	some	х	yes (1/2)	
Naa60-(1-230)-eGFP [449]	Lacking last 12 aa	some	х		
Naa60-(1-236)-eGFP [450]	Lacking last 6 aa	?	х		
N-truncations (eGFP targeting assay)) (Fig 3C/E)				
eGFP-Naa60-(182-242) [131]	eGFP targeting using the last 61 aa of Naa60		х		
eGFP-Naa60-(192-242) [349]	eGFP targeting using the last 51 aa of Naa60		х		
eGFP-Naa60-(202-242) [363]	eGFP targeting using the last 41 aa of Naa60	x	х		
eGFP-Naa60-(217-242) [132]	eGFP targeting using the last 26 aa of Naa60	х			
Deletione Devid A sold A sold A					
Deletions Pred-al and extended (Fig	3U/E)				
Delta 191-200 [364]	Most of Pred-a1	x	x	yes (>1/2)	
Delta 191-209 [409]	Pred-a1 extended	x			
Delta 203-210 [457]	Between Pred-α-Helices	x (more ER)			
Deletions Pred-α2 and extended (Fig	3D/E)				
Delta 213-222 [365]	Most of Pred-a2	x	x	yes (~	
Dolto 212 220 [450]	Dred - 2 outonded	~		completely)	
Della 213-230 [458]	Pred-az extended	x	X		
Delta 213-236 [459]	Pred-a2 further extended	x	x		
Delta 211-216 [421]	1^{st} half of Pred- $\alpha 2$	х	x		
Mutations in Pred- α 1 (Fig 4 and S1)					
I190A/L191A/Y193A/I194A [370]	ϕ in Pred- $\alpha 1 \rightarrow A$	x	х	yes (>1/2)	
I190E/L191E/Y193E/I194E [389]	ϕ in Pred- α 1 \rightarrow E	x x		yes (>1/2)	
L197A/L201A/L204A/I209A [428]	ϕ in Pred- α 1 \rightarrow A	x	some	yes (~ completely)	
L197E/L201E/L204E/I209E [429]	ϕ in Pred- α 1 \rightarrow E	х			
L197/L201/L204A [435]	∳ in Pred-α1 → A	x	x	yes (>1/2)	
L197/L201/L204E [436]	204/209E effective. Skew it Nt dir. E mut	x	some		
L201/L204/I209A [433]	ϕ in Pred- α -Helix1 \rightarrow A	x	х		
L201/L204/I209E [434]	204/209E effective. Extend it Nt dir. E mut	x			
I190/L191A [451]	ϕ in Pred- α -Helix1 \rightarrow A		х		
Y193/I194A [454]	ϕ in Pred- α 1 \rightarrow A		х		
Y194/L197A [455]	ϕ in Pred- α 1 \rightarrow A		х		
L197A/L201A [456]	ϕ in Pred- α 1 \rightarrow A		х		
L201/L204A [462]	ϕ in Pred- α 1 \rightarrow A		х		
L204/I209A [430]	ϕ in between Pred- α 1 and 2 \rightarrow A		x		
L204E/I209E [398]	ϕ in between Pred- α 1 and 2 \rightarrow E	x	x	yes(>1/2)	
L204E [431]	Effective when comb w 209E. Alone?		х		
I209E [432]	Effective when comb w 204E. Alone?	x	х		
I190E/L191E/Y193E/I194E + L204E/I209E [416]	Mut ϕ in extended Pred- α 1 to E	x			

D192A/H196A [366]	Electrostatic interactions?		x	some (like WT)
D192A [468]	Alone?		х	
H185/H196A [464]	Electrostatic interactions?		х	
Mutations in Pred-α2 (Fig 5 and S2)			•	
I209/V213/Y214A [424]	Mut ϕ in Pred- α 2+209 to A	some	х	
I209/V213/Y214E [425]	Mut ϕ in Pred- α 2+209 to E	х	х	
V213/Y214A [422]	Mut ϕ in Pred- α 2 to A	?	х	
V213/Y214E [423]	Mut ϕ in Pred- α 2 to E	some	х	
V213E [443]	Mut ϕ in Pred- α 2 to E	some	х	
L220/221E [396]	Mut ϕ in Pred- α 2 to E	?	х	
L220/L221/F224/L225A [460]	Mut ϕ in Pred- α 2 to A	some	х	yes (1/2)
F224/L225/W227/I230A [426]	Mut ϕ in Pred- α 2 to A	some	х	
F224/L225/W227/I230E [427]	Mut φ in Pred-α2 to E	some	х	yes (1/2)
F224A/L225A [441]	Mut ϕ in Pred- α 2 to A		х	
F224E/L225E [442]	Mut ϕ in Pred- α 2 to E	?	х	
H211A/R212A/R215A/H218A [367]	Electrostatic interactions?	SF	x	some (like WT)
H211A/R212A [465]	Electrostatic interactions?		х	
R212A/R215A [466]	Electrostatic interactions?		х	
R215A/H218A [467]	Electrostatic interactions?		х	
			•	
Combined mutations in Pred-α1 + Pred	l-α2 (Fig 5 and S2)	Cyto	Org	
Delta 191-200 + H211A/R212A/R215A/H218A [392]	Delete most of Pred- α 1 + Mut charge in Pred- α 2	x	x	
Delta 213-222 + D192A/H196A [394]	Delete most of Pred- α 2 + Mut charge in Pred- α 1	x	x	
l190E/L191E/Y193E/l194E + H211A/R212A/R215A/H218A [411]	Mut ϕ in Pred- $\alpha 1$ to E + Mut charge in Pred- $\alpha 2$	x	x	
I190E/L191E/Y193E/I194E + V213E [414]	Mut ϕ in Pred- $\alpha 1$ to E + Mut ϕ in Pred- $\alpha 2$ to E	x	some	
I190E/L191E/Y193E/I194E + L220/221E [415]	Mut ϕ in Pred- $\alpha 1$ to E + Mut ϕ in Pred- $\alpha 2$ to E	x		
I190E/L191E/Y193E/I194E + C222S [412]	Mut ϕ in Pred- $\alpha 1$ to E + Mut put palmitoylation site	x	x	
I190E/L191E/Y193E/I194E + F224E/L225E [413]	Mut ϕ in Pred- $\alpha 1$ to E + Mut ϕ in/near Pred- $\alpha 2$ to E	x		
D192A/H196A + H211A/R212A/R215A/H218A [419]	Mut charge in Pred- α 1 and Pred- α 2 to A	?	x	
L197/L201/L204A + H211A/R212A/R215A/H218A [445]	Mut ϕ in Pred- $\alpha 1$ to A + Mut charge in Pred- $\alpha 2$	x	x	
1 107/L 201 + 1200//213/V21/A				
[463]	Mut ϕ in Pred- $\alpha 1$ to A + Mut ϕ in Pred- $\alpha 2$ to A	x	weak	
[463] L197/201/204A + F224/L225/W227/I230A [448]	Mut ϕ in Pred- α 1 to A + Mut ϕ in Pred- α 2 to A Mut ϕ in Pred- α 1 to A + Mut ϕ in Pred- α 2 to A	x x	weak	
[463] L197/201/204A + F224/L225/W227/I230A [448] H211A/R212A/R215A/H218A + K233A/R240A [446]	Mut ϕ in Pred- α 1 to A + Mut ϕ in Pred- α 2 to A Mut ϕ in Pred- α 1 to A + Mut ϕ in Pred- α 2 to A Mut charge in Pred- α 1 + mut charge in C-term	x x	weak	
[463] L197/201/204A + F224/L225/W227/I230A [448] H211A/R212A/R215A/H218A + K233A/R240A [446] L204E/I209E + L220/221E [418]	Mut ϕ in Pred- α 1 to A + Mut ϕ in Pred- α 2 to A Mut ϕ in Pred- α 1 to A + Mut ϕ in Pred- α 2 to A Mut charge in Pred- α 1 + mut charge in C-term Mut ϕ in Pred- α 2 + 204/209 to E	x x x	weak x	

SUPPLEMENTAL FIGURE LEGENDS

Supplemental figure 1. Mutated constructs of Naa60-eGFP demonstrated hydrophobic amino acids within Pred- α 1 important for membrane association *in cellulo*. HeLa cells were transfected with the indicated Naa60-eGFP constructs (A) and imaged live (B) or subjected to subcellular fractionation followed by high pH/sodium carbonate mediated protein extraction from membranes (C). A. The sequence of the Naa60 C-terminal tail as in Figure 1. Arrows indicate the positions of mutations shown in B and C, and are color coded accordingly. B. Subcellular localization of Naa60eGFP mutated constructs in the area of Pred- α 1 as indicated with corresponding colors in A. Scale bar is 10 µm and is representative for all microscopic images. C. Cells were transfected with the indicated constructs and subjected to high pH/sodium carbonate mediated protein extraction from membranes following subcellular fractionation and immunoblotting. The peripheral membrane protein GM130 and transmembrane RCAS1 were used as controls for extractable and un-extractable modes of membrane binding. Supplements data in Figure 4B-C.

Supplemental figure 2. Mutated constructs of Naa60-eGFP demonstrated amino acids within Pred- α 2 important for membrane association *in cellulo*. HeLa cells were transfected with the indicated Naa60-eGFP constructs (A) and imaged live (B) or subjected to subcellular fractionation followed by high pH/sodium carbonate mediated protein extraction from membranes (C). A. The sequence of the Naa60 C-terminal tail as in Figure 1. Arrows indicate the positions of mutations shown in B and C, and are color coded accordingly. B. Subcellular localization of Naa60-eGFP mutated constructs in the area of Pred- α 1 as indicated with corresponding colors in A. C. Cells were transfected with the indicated constructs and subjected to high pH/sodium carbonate mediated protein extraction from membranes following subcellular fractionation and immunoblotting. The peripheral membrane protein GM130 and transmembrane RCAS1 were used as controls for extractable and unextractable modes of membrane binding. Supplements data in Figure 4D-E.

Supplemental figure 3. Anti-Naa60₁₉₂₋₂₄₂ antibody validation for detection of Naa60₁₈₉₋₂₄₂ peptide. The specificity of the anti-Naa60₁₉₂₋₂₄₂ (Abcam 103800) antibody used to detect the Naa60₁₈₉₋₂₄₂ peptide in pull down experiments was determined by a dot blot. Ratios from 0.1 to 10 pmol Naa60₁₈₉₋₂₄₂ peptide were spotted on a nitrocellulose membrane, in addition to 10 pmol of the negative control peptide SESS (SESSSKS RWGRPVGRRRPVRVYP). The dot blots were incubated with either anti-Naa60₁₉₂₋₂₄₂ or custom-made anti-Naa10 antibody before detection. Supplements data in Figure 7A.

Supplemental figure 4. Far UV-CD experiments supporting Naa60₁₈₉₋₂₄₂ folding most optimal in the presence of PI4P over all other PI. PI4P liposomes induce the most favorable Naa60₁₈₉₋₂₄₂ helical spectra, when compared to all other PI-liposomes. Far UV-CD spectra of 20 μ M Naa60₁₈₉₋₂₄₂ in the presence of 360 μ M liposomes containing 92 mol % PC and 8 mol % of either PI3P, PI4P, PI5P, PI(3,4)P2, PI(3,5)P2, PI(4,5)P2 or PIP3. n = 2 for all experiments. Related to data in Figure 7D.



RCAS1

Naa60-(I190E/L191E/Y193E/I194E)

Naa60-(D192A/H196A)



В WT	213/214A	209/213/214E	213/214E
-725	A.	4	
220/221E	224/225/227/230E	224/225A	224/225E
*		12	

С			рН	7.4	pH ′	11.5	
	P1	S1	P2	S2	P2	S2	
	1		-		•	-] GM130
	1		-		-		RCAS1
	1		1		1		Naa60-(F224E/L225E/W227E/I230E)

S-6



Naa60-(189-242) + 3' phosphorylated Pls



Naa60-(189-242) + 4' phosphorylated Pls



Naa60-(189-242) + 5' phosphorylated Pls

