

Supplementary Figure 1, LOX controls surface EGFR retention and AKT activation.

(a, b) Western blots of pY1068 EGFR, surface EGFR, total EGFR and GAPDH in EGF-stimulated control (shCtl) or LOX depleted (shLOX A) MDA-MB-231 and U87 cells. (c) Western blots of pY1068 EGFR, surface EGFR, total EGFR and GAPDH in control (shCtl) or LOX depleted (shLOX A,B) MDA-MB-231 and U87 cells cultured under EGF only condition for 24 hours. (d) Quantification of EGFR activation, and (e) surface EGFR level in MDA-MB-231 and U87 cells from c. (f,g) AKT activation in MDA-MB-231 cells and in U87 cells under EGF only condition for 24 hours. All data are represented as Mean \pm SD from 3 independent experiments. **p<0.01, Student's t-test.



Supplementary Figure 2, LOX RNA interference

(a) Western blot showing LOX and GAPDH in control (siCtl) or LOX depleted (siLOX #1, siLOX #2) MDA-MB-231 cells. (b) Time course (days) showing LOX and GAPDH in LOX depleted (siLOX #1) MDA-MB-231 cells. (c) Western blot showing expression of transfected secreted GFP (sGFP), LOX-GFP (LOX), LOX^{K314A}-GFP (K314A), LOX^{Δ BMP-1}-GFP (Δ BMP-1), endogenous LOX and tubulin in control (siCtl) or LOX depleted (siLOX #1) MDA-MB-231 cells. MOCK: transfection reagents only.



Supplementary Figure 3, LOX regulates MATN2 and HTRA1

(a) Schematic showing 10 EGF-like domains in matrilin2 (MATN2). (b)
Western blot showing MATN2 binding to LOX-GFP (LOX), LOX^{K314A}-GFP (K314A). MATN2/GFP ratio indicates binding strength. (c) Western blots showing MATN2 and GAPDH in control (shCtl) or LOX depleted (shLOX A, B) MDA-MB-231 and U87 cells. (d) Confocal photomicrographs of

extracellular MATN2 in control (shCtl) or LOX depleted (shLOX A) MDA-MB-231 and U87 cells in collagen gels. Scale bar: 40µm. (e) Quantification of extracellular MATN2 in d. All data are represented as Mean ± SD from 15 fields of views per condition. **p<0.01, Student's t-test. (f) Western blots showing LOX, MATN2 and GAPDH in control (shCtl) or LOX depleted (shLOX C) BJ fibroblasts. (g,h) Quantification of LOX and MATN2 protein expression from experiments in f. All data are represented as Mean ± SD from 3 independent experiments. **p<0.01, Student's t-test. (i) Confocal photomicrographs of extracellular MATN2 in cell derived matrix (CDM) of control (shCtl) or LOX depleted (shLOX C) BJ fibroblasts. Scale bar: 40µm. (j) Quantification of extracellular MATN2 in i. Data are represented as Mean \pm SD from 15 fields of views per condition. **p<0.01, Student's t-test. (k) Western blot showing MATN2 in DMSO or LY364947 treated control (shCtl) or LOX depleted (shLOX A) MDA-MB-231 cells. (I) Western blots showing pSMAD2, SMAD2/3 and GAPDH in control (shCtl) or LOX depleted (shLOX A) MDA-MB-231 and U87 cells. (m) Confocal photomicrographs of nuclear phospho-SMAD2 (pSMAD2) in control (shCtl) or LOX depleted (shLOX A) MDA-MB-231 and U87 cells. Scale bar: 10µm. (n) Western blot for MATN2, pSMAD2, SMAD2/3, and GAPDH in control (siCtl) or LOX depleted (siLOX #1) MDA-MB-231 cells expressing secreted GFP (sGFP), LOX-GFP (LOX), LOX^{K314A}-GFP (K314A) or LOX^{Δ BMP-1}-GFP (Δ BMP-1). (o) Western blot showing HTRA1 binding to LOX-GFP (LOX). HTRA1/GFP ratio indicates binding strength. (p) NativePAGE showing extracellular HTRA1 multimer and trimer in control (shCtl) or LOX depleted (shLOX A) MDA-MB-231 cell culture medium. (q) Western blot showing pSMAD2, SMAD2/3, GAPDH, MATN2 and Tubulin in PBS or rhHTRA1 treated control (shCtl) or LOX depleted (shLOX A) MDA-MB-231 cells.



Supplementary Figure 4, MATN2 upregulates cell surface EGFR.

(a) Confocal photomicrographs of rhMATN2 (Red) binding to the cell surface of a GFP expressing MDA-MB-231 cell (Green). Scale bar: 10μm. (b) Western blots of surface EGFR, total EGFR and GAPDH in rhMATN2 treated (500ng/ml) MDA-MB-231 and U87 cells. (c) Western blots showing MATN2 and GAPDH in control (shCtl) or MATN2 depleted (siMATN2) MDA-MB-231 cells. (d) Western blots showing surface EGFR, total EGFR and GAPDH in PBS or EGF treated (3 min) control (siCtl) or MATN2 depleted (siMATN2) MDA-MB-231 cells. (e) Western blots showing surface EGFR in rhMATN2 treated control (shCtl) or LOX depleted (shLOX A) MDA-MB-231 cells under 3min EGF stimulation. (f) Western blots showing surface EGFR, total EGFR and GAPDH in PBS or rhMATN2 treated control (siCtl) or MATN2 depleted (siMATN2) MDA-MB-231 cells. (g) Western blots showing pY1068EGFR, total EGFR and GAPDH in EGF (50ng/ml) or EGF(50ng/ml) and rhMATN2 (500ng/ml) treated MDA-MB-231 cells at indicated time points.



Supplementary Figure 5, LOX chemical inhibition controls surface EGFR via MATN2

(a) Schematic illustrating the LOX biosensor (LOX-roGFP2) mechanism used in this study. SP, signal peptide; PP, pro-peptide; CD, catalytic domain; LOX, wild type enzyme; K314A, catalytically inactive LOX mutant; Δ BMP-1; BMP-1 cleavage site deleted LOX mutant. (b,c) Western blots showing surface EGFR, total EGFR and GAPDH in EGF-stimulated DMSO, CCT365623, or BAPN treated MDA-MB-231 cells. (d) Western blots for pY1068 EGFR, total EGFR, pAKT, total AKT and GAPDH in EGF (3 min) stimulated, DMSO, CCT365623 or BAPN treated MDA-MD-231 cells. (e) Western blot from Native PAGE showing reduced HTRA1 multimer in

DMSO, CCT365623, or BAPN treated MDA-MB-213 cell culture media. (f) Confocal photomicrographs showing nuclear phospho-SMAD2 (pSMAD2) in DMSO, CCT365623 or BAPN treated MDA-MB-231 cells. Scale bar, 10µm. (g) Western blots showing pSMAD2, SMAD2/3, and GAPDH in DMSO, CCT365623, or BAPN treated MDA-MB-231 cells. (h) Western blots showing MATN2 and tubulin in DMSO, CCT365623, or BAPN treated MDA-MB-231 cells.



Supplementary Figure 6, (a) Mouse body weight in Vehicle or CCT365623 treated MMTV-PyMT mice. Data are represented as Mean \pm SD from 6 mice in Vehicle group, and 6 mice in CCT365623 group. (b) Tumour growth in Vehicle or BAPN treated MMTV-PyMT mice. Data are represented as Mean \pm SD from 6 mice in Vehicle group, and 4 mice in BAPN group.









Supplementary Figure 7, Full scans of blots in indicated Figures and Supplementary Figures.

		Proportion of lung	
	Mice	MDA-MB-231 Deposition	Score
	1	50%-75%	3
	2	>75%	4
	3	1-25%	1
shCtl	4	25-50%	2
	5	>75%	4
	6	>75%	4
	7	1-25%	1
	8	25-50%	2
	9	1-25%	1
	10	1-25%	1
shLOX A	11	0	0
	12	25-50%	2
	13	25-50%	2
	14	<1%, Micromets	0
		<1%	0
		1%-25%	1
Scoring sy	stem	25-50%	2
		50%-75%	3
		>75%	4

Supplementary Table 1, LOX affects MDA-MB-231 tumour growth in the lungs. Quantification of lung deposits in mice following tail vein injection with control (shCtl) or LOX depleted (shLOX A) MDA-MB-231 cells.

	LOX IC ₅₀	DAO IC ₅₀	MAO-A IC ₅₀	MAO-B IC ₅₀	hERG IC ₅₀	MLM
CCT365623	0.89µM	>1mM	333µM	>1mM	>20µM	65%
BAPN	14.7µM	204µM	>1mM	>1mM	>20µM	70%

Supplementary Table 2, Characterisation of CCT365623 and BAPN. The table shows the IC_{50} for CCT365623 and BAPN against LOX, diamine oxidase (DAO), monoamine oxidase A and B (MAO-A, MAO-B) and the potassium ion channel hERG. It also shows CCT365623 and BAPN stability in mouse liver microsomes (MLM) *in vitro* (% remaining after 30 minutes incubation).

		PK parameters								Therapy parameters		
	MW	PK Dose	PK Molar dose	C _{max} IV	C _{max} PO	F%	T _{1/2} PO	CI IV	AUClast PO PK	Therapy dose (PO, qd)	Therapy molar dose (PO, qd)	AUClast PO extrapolated at therapy dose
	(g/mol)	(mg/kg)	(mmol/kg)	(µM)	(µM)		(h)	(mL/min/kg)	(µM.h)	mg/kg	mmol/kg	(µM.h)
CCT365623	407	PO - 50 IV - 10	PO - 0.12 IV - 0.024	17	17	45	0.6	49	15	70	0.17	21
BAPN	128	PO - 50 IV - 10	PO - 0.39 IV - 0.078	132	192	41	0.6	25	195	1460	11.4	5694

Supplementary Table 3. *In Vivo* PK and Therapy Parameters for CCT365623 and BAPN. The table shows the PK parameters for CCT365623 and BAPN following a single PO dose at 50 mg/kg compared to a single IV injection at 10mg/kg respectively. Cmax: maximum concentration. F%: oral bioavailability. T_{12} : elimination half life. CI: clearance from plasma. AUClast.: area under the concentration time curve; and the therapy parameters for CCT365623 and BAPN following a single PO dose at 70 mg/kg per day for CCT365623 and an estimated daily dose of 1460 mg/kg for BAPN extrapolated from consumption of compound administered in the drinking water (1% w/v).

		0			Total
	MICe	Small	Mealum	Large	Score
	1	1	1	0	3
	2	12	12	12	72
	3	3	3	1	12
Water	4	0	1	0	2
	5	0	1	0	2
	6	10	1	2	18
	7	1	0	0	1
	8	0	0	0	0
	9	0	0	0	0
	10	1	0	0	1
BAPN	11	1	0	0	1
	12	0	0	0	0
	13	1	0	0	1
	14	0	0	0	0
	1	7	0	5	22
	2	2	1	4	16
	3	7	4	1	18
venicie	4	1	2	9	32
	5	0	4	15	53
	6	1	4	0	9
	7	0	0	0	0
	8	0	0	0	0
CCT365623	9	0	1	0	2
	10	0	1	0	2
	11	0	1	0	2
	12	1	1	0	3
		1		Size	Score
			None	0	0
Scorin	ig system	l	Small	<100µm	1
	-		Medium	100~200µm	2
			Large	>200µm	3

Number of Metastasis

Supplementary Table 4. Quantification of lung metastases in MMTV-PyMT mice. The table shows the quantification of lung metastases in water or BAPN treated, and vehicle or CCT365623 treated MMTV-PyMT mice. The score was calculated using the following formula (number of small metastasis x 1)+(number of medium metastasis x 2) + (number of large metastasis x 3) for each individual animal.

Supplementary Methods

Synthesis of CCT365623

CCT365623 is compound **2** in Fig.6a. It is referred as **2** in the following methods.

Commercial building blocks 3 and 7, reagents and solvents for reactions were reagent grade and used as purchased. Chromatography solvents were HPLC grade and were used without further purification. Thin layer chromatography (TLC) analysis was performed using silica gel 60 F-254 thin layer plates. Flash column chromatography was performed using columns pre-packed with 40-63 µm silica. LCMS and HRMS analyses were performed on a HPLC system with diode array detector operating at 254 nm, fitted with a reverse-phase 50×4.6mm column at a temperature of 22 °C, connected to a Time of Flight (ToF) mass spectrometer (ESI). ¹H and ¹³C NMR spectra were recorded on a Bruker Avance 500 MHz spectrometer using an internal deuterium lock. NMR data is given as follows: chemical shift (δ) in ppm, multiplicity, coupling constants (*J*) given in Hz and integration.

(5-((5-(Methylsulfonyl)-[1,1'-biphenyl]-3-yl)sulfonyl)thiophen-2yl)methanamine (2)

Synthesis route for 3-bromo-5-(methylthio)benzenethiol intermediate 6



a) NaSMe (21% aq.; 50.8 mL, 152 mmol) was added to a solution of 1,3,5-tribromobenzene **3** (40.0 g, 127 mmol) in DMF (508 mL), and the

reaction was stirred at 100 °C for 16 h. After cooling to rt, EtOAc (1.5 L) was added. The organic phase was washed with H_2O (3 × 1.0 L), brine (1.0 L), dried over MgSO₄, filtered and the solvent was removed under reduced pressure. The crude sulfide was used in the subsequent transformation immediately.

b) The intermediate sulfide was dissolved in DCM (400 mL) and cooled to 0 °C. A solution of *m*-CPBA (34.2 g, 152 mmol) in DCM (234 mL) was slowly added and the mixture was stirred at 0 °C for 1 h before allowing to warm to rt over 1 h. EtOAc (800 mL) was added. The organic phase was washed with sat. NaHCO₃ (3 × 500 mL), dried over MgSO₄, filtered and the solvent was removed under reduced pressure. The crude was recrystallised from DCM/cyclohexane to give 22.3 g of beige crystals containing a 4.5:1 mixture of the desired 1,3-dibromo-5-(methylsulfinyl)benzene **4** and the by-product (5-bromo-1,3-phenylene)bis(methylsulfane). Data for pure **4** - mp: 121–124 °C. ¹H **NMR** (500 MHz, Chloroform-*d*) δ 7.75 (t, *J* = 1.7 Hz, 1H), 7.67 (d, *J* = 1.7 Hz, 2H), 2.73 (s, 3H). ¹³C **NMR** (126 MHz, Chloroform-*d*) δ 149.76, 136.51, 125.23, 124.02, 44.14. **LCMS (ESI)** *m*/z 297/299/301 [M+H]⁺.

c) NaSMe (21% aq.; 27.5 mL, 82.3 mmol) was added to a solution of sulfoxide 4 (22.3 g, 74.8 mmol) in DMF (374 mL), and the reaction was stirred at 40 °C for 6 h. After cooling to rt, EtOAc (400 mL) was added. The organic phase was washed with 1:1 $H_2O/brine$ (3 × 400 mL), dried over MgSO₄, filtered and the solvent was removed under reduced pressure. The crude was purified by chromatography (EtOAc/cyclohexane 5→80%) afford (3-bromo-5to (methylsulfinyl)phenyl)(methyl)sulfane 5 as an orange oil (6.26 g, 19% from **3**). ¹**H NMR** (500 MHz, Chloroform-*d*) δ 7.46 – 7.41 (m, 3H), 2.75 (s, 3H), 2.53 (s, 3H). ¹³C NMR (126 MHz, Chloroform-d) δ 148.35, 143.41, 130.46, 123.61, 122.31, 118.88, 44.06, 15.34. LCMS (ESI) *m/z* 265/267 [M+H]⁺

d) TFAA (6.30 mL, 43.3 mmol) was added to a solution of (3-bromo-5-(methylsulfinyl)phenyl)(methyl)sulfane **5** (4.0 g, 15.1 mmol) and 2,6-

lutidine (7.04 mL, 60.4 mmol) in MeCN (101 mL). The reaction was stirred at rt for 0.5 h and all volatiles were removed under reduced pressure. 1:1 Et₃N/MeOH (100 mL) was then added. The solution was stirred at rt for 0.5 h and was subsequently acidified to pH <4 with 2M HCI. The aqueous phase was extracted with EtOAc (400 mL). The organic phase was washed with 0.5 M HCI (200 mL) and brine (200 mL), dried over MgSO₄, filtered and the solvent was removed under reduced pressure. The crude was purified by chromatography (EtOAc/cyclohexane $0 \rightarrow 30\%$) to afford 3-bromo-5-(methylthio)benzenethiol 6 as an orange oil (3.42 g, 96%). ¹H NMR (500 MHz, Chloroform-d) δ 7.18 (m, 1H), 7.14 (m, 1H), 7.04 (m, 1H), 3.49 (s, 1H), 2.47 (s, 3H). ¹³C NMR (126 MHz, Chloroform-d) δ 141.86, 133.65, 127.99, 125.72, 124.83, 123.03, 15.49. LCMS - did not ionise.

Synthetic route for compound 2



e) A mixture of 5-bromothiophene-2-carbonitrile 7 (5.20 g, 27.6 mmol),
3-bromo-5-(methylthio)benzenethiol 6 (5.41 g, 23.0 mmol), K₂CO₃

(4.77 g, 34.5 mmol) and DMF (115 mL) was stirred at 40 °C for 16 h. After cooling to rt, the mixture was diluted with EtOAc. The organic phase was washed with 1:1 H₂O/brine (3 ×), dried over MgSO₄ and filtered. The solvent was removed under reduced pressure. The crude was purified by chromatography (EtOAc/cyclohexane 0→20%) to afford 5-((3-bromo-5-(methylthio)phenyl)thio)thiophene-2-carbonitrile **8** as a yellow oil (7.53 g, 96%). ¹H NMR (500 MHz, Chloroform-*d*) δ 7.57 (d, *J* = 3.9 Hz, 1H), 7.25 (t, *J* = 1.7 Hz, 1H), 7.21 (d, *J* = 3.8 Hz, 1H), 7.16 (t, *J* = 1.6 Hz, 1H), 7.09 (t, *J* = 1.6 Hz, 1H), 2.47 (s, 3H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 142.81, 141.05, 138.03, 137.77, 134.07, 128.10, 127.78, 125.10, 123.63, 113.71, 113.35, 15.52. LCMS (ESI) *m/z* 342/344 [M+H]⁺.

f) and **g**) BH₃.THF (1.0 M in THF; 66.1 mL, 66.1 mmol) was added to a solution of 5-((3-bromo-5-(methylthio)phenyl)thio)thiophene-2-carbonitrile **8** (7.53g, 22.0 mmol) in THF (66 mL) and the mixture was stirred at rt for 1 h. EtOH (132 mL) was carefully added to quench the reaction. Subsequently, the solution was heated at 70 °C for 1 h to aid decomplexation. The solvent was removed under reduced pressure. The crude was dissolved in DCM (110 mL). Et₃N (9.21 mL, 66.1 mmol), followed by Boc₂O (7.60 mL, 33.0 mmol) were added and the mixture was stirred at rt for 16 h. When complete conversion was achieved, more DCM was added (110 mL). The organic phase was washed with H₂O (220 mL) and brine (220 mL), dried over MgSO₄ and filtered. The solvent was removed under reduced pressure. The crude was used in the subsequent transformation immediately.

h) *m*-CPBA (50-55%; 32.6 g, ~99.1 mmol) was added in small portions to a solution of *tert*-butyl ((5-((3-bromo-5-(methylthio)phenyl)thio)thiophen-2-yl)methyl)carbamate (crude of step **f** and **g**) in DCM (148 mL) at 0 °C and the mixture was stirred at rt for 3 h. When complete conversion was achieved, EtOAc was added (250 mL). The organic phase was washed with sat. NaHCO₃ (4 × 200 mL) and sat. Na₂S₂O₃ (200 mL), dried over MgSO₄ and filtered. The solvent was removed under reduced pressure. The crude was purified by column chromatography (EtOAc/cyclohexane $0\rightarrow60\%$) to afford *tert*-butyl ((5-((3-bromo-5-(methylsulfonyl)phenyl)sulfonyl)thiophen-2yl)methyl)carbamate as a yellow foam **9** (3.83 g, 34%). ¹**H NMR** (500 MHz, Chloroform-*d*) δ 8.41 (m, 1H), 8.32 (m, 1H), 8.24 (m, 1H), 7.63 (d, *J* = 3.6 Hz, 1H), 6.97 (d, *J* = 3.9 Hz, 1H), 5.15 (br, 1H), 4.47 (m, 2H), 3.12 (s, 3H), 1.47 (s, 9H). ¹³**C NMR** (126 MHz, Chloroform-*d*) δ 155.07, 145.73, 143.77, 138.80, 135.21, 135.07, 134.81, 126.06, 124.89, 124.59, 80.68, 44.48, 39.91, 28.42. **LCMS** (ESI) *m/z* 532/534 [M+Na]⁺, 393/395 [M-BocNH]⁺.

i) А mixture of *tert*-butyl ((5-((3-bromo-5-(methylsulfonyl)phenyl)sulfonyl)thiophen-2-yl)methyl)carbamate 9 (2.00 g, 3.92 mmol), Pd(PPh₃)₄ (453 mg, 10%), benzeneboronic acid (574 mg, 4.71 mmol), Cs₂CO₃ (1.53 g, 4.71 mmol) and 1,4-dioxane (13 mL) was degassed with argon and then stirred at 100 °C for 16 h. After cooling to rt. The suspension was filtered through celite, washed with DCM and the solvent was removed under reduced pressure. The crude was purified by chromatography (EtOAc/cyclohexane $0 \rightarrow 60\%$) to afford *tert*-butyl ((5-((5-(methylsulfonyl)-[1,1'-biphenyl]-3yl)sulfonyl)thiophen-2-yl)methyl)carbamate 10 as a brown foam (1.59 g, 80%). ¹H NMR (500 MHz, Chloroform-d) δ 8.47 (m, 1H), 8.42 (m, 1H), 8.33 (m, 1H), 7.70 – 7.62 (m, 3H), 7.58 – 7.43 (m, 3H), 6.96 (d, J = 3.8 Hz, 1H), 5.15 (br, 1H), 4.53 – 4.42 (m, 2H), 3.15 (s, 3H), 1.43 (s, 9H). ¹³C NMR (126 MHz, Chloroform-d) δ 154.41, 144.61, 144.58, 142.79, 139.48, 137.18, 134.67, 130.35, 130.09, 129.44, 127.34, 125.88, 124.56, 124.53, 80.38, 44.44, 39.77, 28.32. LCMS (ESI) m/z 530 [M+Na]⁺, 391 [M-BocNH]⁺.

j) 4M HCl in dioxane (10.5 mL) was added to a solution of *tert*-butyl ((5-((5-(methylsulfonyl)-[1,1'-biphenyl]-3-yl)sulfonyl)thiophen-2yl)methyl)carbamate **10** (1.59 g, 3.14 mmol) in 1,4-dioxane (10.5 mL) and the mixture was stirred at rt 1or 16 h. Cyclohexane was added to precipitate the solid product. The solids were filtered, washed with EtOAC/cyclohexane (1:1) and then treated with 2N NH₃ in MeOH. The solvent was removed under reduced pressure to afford compound **2** as a light brown solid (1.12 g, 88%). mp: 98-101 °C. ¹H **NMR** (500 MHz, Chloroform-*d*) δ 8.46 (t, *J* = 1.7 Hz, 1H), 8.44 (t, *J* = 1.7 Hz, 1H), 8.32 (t, *J* = 1.7 Hz, 1H), 7.67 (d, *J* = 3.9 Hz, 1H), 7.65 – 7.61 (m, 2H), 7.55 – 7.45 (m, 3H), 6.93 (m, 1H), 4.09 (s, 2H), 3.14 (s, 3H), 1.68 (br, 2H). ¹³C **NMR** (126 MHz, Chloroform-*d*) δ 145.00, 144.60, 142.82, 138.62, 137.33, 134.96, 130.41, 130.02, 129.54, 127.42, 124.66, 124.18, 44.56, 41.68. **HRMS** (ESI) for C₁₈H₁₈NO₄S₃ ([M+H]⁺): Calculated 408.0392; Observed 408.0401.









































