## SUPPLEMENTAL MATERIAL

## Substrate recognition and modification by a pathogen-associated aminoglycoside-resistance 16S rRNA methyltransferase

Kellie Vinal<sup>a,b</sup> and Graeme L. Conn<sup>a,\*</sup>

Department of Biochemistry, Emory University School of Medicine, Atlanta, Georgia<sup>a</sup>; Graduate Program in Microbiology and Molecular Genetics, Graduate Division of Biological and Biomedical Sciences, Emory University<sup>b</sup>

\*Address correspondence to Graeme L. Conn, gconn@emory.edu

## SUPPLEMENTAL FIGURES S1-S5

- Fig S1 NpmA-E184C\* binds only 30S subunits with unmodified A1408 and is displaced by addition of SAM and/or wild-type NpmA.
- Fig S2 Solution optimization and effect of ionic strength on 30S-NpmA interaction.
- Fig S3 Measurement of NpmA  $\beta$ 2/3 linker variant binding affinity for cosubstrate SAM and reaction byproduct SAH.
- Fig S4 Linker deletion variants are folded and monomeric.
- Fig S5 ITC analysis of SAM/SAH interaction with  $\beta$ 5/6 and  $\beta$ 6/7 linker deletion and single residue substitution NpmA variants.



**FIG S1** NpmA-E184C\* binds only 30S subunits with unmodified A1408 and is displaced by addition of SAM and/or wild-type NpmA. NpmA-E184C\* alone (black) and pre-methylated 30S with NpmA-E184C\* (open black squares) controls and binding reactions containing 30S and NpmA-E184C\* (green open squares) subject to addition of SAM (green closed squares), addition of wild-type NpmA (orange closed squares) or no treatment (green open squares) at the point indicated by the arrowhead and dotted vertical line.



**FIG S2** Solution optimization and effect of ionic strength on 30S-NpmA interaction. Fluorescence polarization comparison of free NpmA-E184C\* (dark blue/ green) with 30S-bound (light blue/ green) NpmA-E184C\* with increasing concentrations of (A) KCI and (B) Mg<sup>2+</sup>. In each panel, error bars indicate standard deviation of averaged measurements and the arrow indicates the concentration of each component used in the final competition binding assays with wild-type and variant NpmA proteins.



**FIG S3** Measurement of NpmA  $\beta$ 2/3 linker variant binding affinity for cosubstrate SAM and reaction by-product SAH. ITC titrations for each protein-ligand pair tested: (A),(B) wild-type NpmA; (C),(D) NpmA-K66E/K67E; (E),(F) NpmA-K70E/K71E; and, (G),(H) NpmA-K66E/K67E/K70E/K71E (4x K $\rightarrow$ E) with SAM and SAH, respectively. Values for binding affinity (K<sub>d</sub>) are those derived from the individual titration shown.



**FIG S4** Linker deletion variants are folded and monomeric. Gel filtration chromatograms showing the essentially identical elution profile of wild-type NpmA and the NpmA- $\Delta\beta$ 5/6 and NpmA- $\Delta\beta$ 6/7 linker deletion variants during protein purification.



**FIG S5** ITC analysis of SAM/SAH interaction with  $\beta$ 5/6 and  $\beta$ 6/7 linker deletion and single residue substitution NpmA variants. ITC titrations for each protein-ligand pair tested: (A),(B) wild-type NpmA (same data as shown in FIG S3); (C),(D)  $\Delta\beta$ 5/6 linker; (E),(F)  $\Delta\beta$ 6/7 linker; (G),(H) NpmA-R207E; (I),(J) NpmA-V190P; (K),(L) NpmA-L201P; (M),(N) NpmA-L196G; and, (O),(P) NpmA-L196 $\Delta$  with SAM and SAH, respectively. Values for binding affinity (K<sub>d</sub>) are those derived from the individual titration shown.