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# Main Figures: 8

# Supplementary Figures: 6

# Supplementary Tables: 1

# Supplementary Videos: N/A

# Reporting Checklist for Nature Neuroscience

This checklist is used to ensure good reporting standards and to improve the reproducibility of published results. For more information, please read Reporting Life Sciences Research.

**Please note that in the event of publication, it is mandatory that authors include all relevant methodological and statistical information in the manuscript.**

## Statistics reporting, by figure

- Please specify the following information for each panel reporting quantitative data, and where each item is reported (section, e.g. Results, & paragraph number).
- Each figure legend should ideally contain an exact sample size (n) for each experimental group/condition, where n is an exact number and not a range, a clear definition of how n is defined (for example x cells from x slices from x animals from x litters, collected over x days), a description of the statistical test used, the results of the tests, any descriptive statistics and clearly defined error bars if applicable.
- For any experiments using custom statistics, please indicate the test used and stats obtained for each experiment.
- Each figure legend should include a statement of how many times the experiment shown was replicated in the lab; the details of sample collection should be sufficiently clear so that the replicability of the experiment is obvious to the reader.
- For experiments reported in the text but not in the figures, please use the paragraph number instead of the figure number.

Note: Mean and standard deviation are not appropriate on small samples, and plotting independent data points is usually more informative. When technical replicates are reported, error and significance measures reflect the experimental variability and not the variability of the biological process; it is misleading not to state this clearly.

| | | TEST USED | | n | | | DESCRIPTIVE STATS (AVERAGE, VARIANCE) | | P VALUE | | DEGREES OF FREEDOM & F/t/z/R/ETC VALUE | |
|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | FIGURE NUMBER | WHICH TEST? | SECTION & PARAGRAPH # | EXACT VALUE | DEFINED? | SECTION & PARAGRAPH # | REPORTED? | SECTION & PARAGRAPH # | EXACT VALUE | SECTION & PARAGRAPH # | VALUE | SECTION & PARAGRAPH # |
| example | 1a | one-way ANOVA | Fig. legend | 9, 9, 10, 15 | mice from at least 3 litters/group | Methods para 8 | error bars are mean +/- SEM | Fig. legend | p = 0.044 | Fig. legend | F(3, 36) = 2.97 | Fig. legend |
| example | results, para 6 | unpaired t-test | Results para 6 | 15 | slices from 10 mice | Results para 6 | error bars are mean +/- SEM | Results para 6 | p = 0.0006 | Results para 6 | t(28) = 2.808 | Results para 6 |
| + | | | | | | | | | | | | |
| - | | | | | | | | | | | | |

| | | TEST USED | | n | | | DESCRIPTIVE STATS (AVERAGE, VARIANCE) | | P VALUE | | DEGREES OF FREEDOM & F/t/z/R/ETC VALUE | |
|---|---|---|---|---|---|---|---|---|---|---|---|---|
| | FIGURE NUMBER | WHICH TEST? | SECTION & PARAGRAPH # | EXACT VALUE | DEFINED? | SECTION & PARAGRAPH # | REPORTED? | SECTION & PARAGRAPH # | EXACT VALUE | SECTION & PARAGRAPH # | VALUE | SECTION & PARAGRAPH # |
| + - | Figure 1b | One-way ANOVA with Tukey–Kramer test | Methods and figure legend | 6, 6, and 6 | Yes. Restraint(-):IR(-), Restraint(-):IR(+), and Restraint(+):IR(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | < 0.0001 | Figure legend | F (2, 15) = 35.64 | Figure legend |
| + - | Figure 1c | Kruskal-Wallis test with Steel-Dwass test | Methods and figure legend | 6, 6, and 6 | Yes. Restraint(-):IR(-), Restraint(-):IR(+), and Restraint(+):IR(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | < 0.0001 | Figure legend | Kruskal-Wallis statistic (H) = 14.36 | Figure legend |
| + - | Figure 1d | Kruskal-Wallis test with Steel-Dwass test | Methods and figure legend | 6, 6, and 6 | Yes. Restraint(-):IR(-), Restraint(-):IR(+), and Restraint(+):IR(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | < 0.0001 | Figure legend | Kruskal-Wallis statistic (H) = 14.36 | Figure legend |
| + - | Figure 1e | Two-way ANOVA with Tukey–Kramer test | Methods and figure legend | 6, 6, 6, and 6 | Yes. α7WT:Restraint(-), α7WT:Restraint(+), α7KO:Restraint(-), and α7KO:Restraint(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P < 0.0001 | Figure legend | F (1, 18) = 37.03 | Figure legend |
| + - | Figure 2a2 | Two-way ANOVA with Tukey–Kramer test | Methods and figure legend | 5, 8, 5, 8, 6 and 6 | Yes. 1*10^6:Restraint(-):IR(-), 1*10^6:Restraint(+):IR(+), 5*10^6:Restraint(-):IR(+), 5*10^6:Restraint(+):IR(+), 1*10^7:Restraint(-):IR(+), and 1*10^7:Restraint(+):IR(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P = 0.0173 | Figure legend | F (1, 32) = 6.299 | Figure legend |
| + - | Figure 2b2 | Unpaired t test | Methods and figure legend | 7 and 6 | Yes. Vehicle:IR(+) and Noradrenaline:IR(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P < 0.0001 | Figure legend | t=7.657 df=11 | Figure legend |
| + - | Figure 3b2 | Kruskal-Wallis test with Steel-Dwass test | Methods and figure legend | 6, 6, 6, and 6 | Yes. ChR2:Laser(-):IR(-), ChR2:Laser(-):IR(+), ChR2:Laser(+):IR(+), and mCherry:Laser(+):IR(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P = 0.0003 | Figure legend | Kruskal-Wallis statistic (H) = 18.62 | Figure legend |
| + - | Figure 3b3 | Kruskal-Wallis test with Steel-Dwass test | Methods and figure legend | 6, 6, and 6 | Yes. ChR2:Laser(-):IR(-), ChR2:Laser(-):IR(+), ChR2:Laser(+):IR(+), and mCherry:Laser(+):IR(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P = 0.0002 | Figure legend | Kruskal-Wallis statistic (H) = 19.22 | Figure legend |

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|---|---|---|---|---|---|---|---|---|---|---|---|---|
| + - | Figure 3b4 | Kruskal-Wallis test with Steel-Dwass test | Methods and figure legend | 6, 6, and 6 | Yes. ChR2:Laser(-):IR(-), ChR2:Laser(-):IR(+), ChR2:Laser(+):IR(+), and mCherry:Laser(+):IR(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P = 0.0002 | Figure legend | Kruskal-Wallis statistic (H) = 19.77 | Figure legend |
| + - | Figure 4d | Repeated two-way ANOVA with Tukey–Kramer test | Methods and figure legend | 10 and 7 | Yes. Group: Control and Caspase; Distance: 611, 620, 629, 638, 647, 656, 665, 674, 683, 692, 701, 710, and 719 | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P < 0.0001 | Figure legend | F (12, 180) = 25.99 | Figure legend |
| + - | Figure 4e | One-way ANOVA with Tukey–Kramer test | Methods and figure legend | 6, 6, 6, 6, 6, and 6 | N/A:N/A:Restraint(-):IR(+), N/A:N/A:Restraint(+):IR(+), DREADD:Vehicle:Restraint(+):IR(+), DREADD:CNO:Restraint(+):IR(+), Caspase:N/A:Restraint(-):IR(+), and Caspase:N/A:Restraint(+):IR(+) | Methods and figure legend | Yes. Mean ± SEM are shown. | In Figure | P < 0.0001 | Figure legend | F (5, 30) = 14.11 | Figure legend |
| + - | Figure 5a | Unpaired t test | Methods and figure legend | 6 and 6 | Yes. DBH:Restraint(-) and DBH:Restraint(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P = 0.0005 | Figure legend | t=4.991 df=10 | Figure legend |
| + - | Figure 5b | One-way ANOVA with Tukey–Kramer test | Methods and figure legend | 9, 6, and 9 | Yes. ChR2:Laser(-), mCherry:Laser(+), and ChR2:Laser(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P = 0.0048 | Figure legend | F (2, 21) = 6.959 | Figure legend |
| + - | Figure 5c | Unpaired t test | Methods and figure legend | 6 and 6 | Yes. Vehicle:Laser(+):IR(+) and Mifepristone:Laser(+):IR(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P = 0.6313 | Figure legend | t=0.4950 df=10 | Figure legend |
| + - | Figure 6a | Repeated one-way ANOVA with Tukey–Kramer test | Methods and figure legend | 6, 6, 6, 6, and 6 | Yes. Dummy, 5Hz, 10Hz, 15Hz, and 20Hz | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P < 0.0001 | Figure legend | F (4, 25) = 112.9 | Figure legend |
| + - | Figure 6b | Repeated one-way ANOVA with Tukey–Kramer test | Methods and figure legend | 6, 6, 6, 6, and 6 | Yes. Dummy, 5Hz, 10Hz, 15Hz, and 20Hz | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P < 0.0001 | Figure legend | F (4, 25) = 14.4 | Figure legend |
| + - | Figure 6d | Repeated one-way ANOVA with Tukey–Kramer test | Methods and figure legend | 6, 6, 6, 6, and 6 | Yes. Dummy, 5Hz, 10Hz, 15Hz, and 20Hz | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P = 0.0061 | Figure legend | F (4, 25) = 4.651 | Figure legend |
| + - | Figure 7a | One-way ANOVA with Tukey–Kramer test | Methods and figure legend | 9, 9, 6, and 6 | Yes. Veh:Laser(-):IR(+), Veh:Laser(+):IR(+), But:Laser(+):IR(+), and Hex:Laser(+):IR(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P < 0.0001 | Figure legend | F (3, 26) = 22.17 | Figure legend |
| + - | Figure 7b | Unpaired t test | Methods and figure legend | 6 and 6 | Yes. Splenectomy:Laser(-):IR(+) and Splenectomy:Laser(+):IR(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P = 0.6331 | Figure legend | t=0.4924 df=10 | Figure legend |

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| + - | Figure 7c | One-way ANOVA with Tukey–Kramer test | Methods and figure legend | 5, 5, and 6 | Yes. Sham:Laser(-):IR(+), Sham:Laser(+):IR(+), and sVNX:Laser(+):IR(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P < 0.0001 | Figure legend | F (2, 13) = 44.75 | Figure legend |
| + - | Figure 8b | Repeated one-way ANOVA with Tukey–Kramer test | Methods and figure legend | 4, 4, and 4 | Yes. Pre, Restraint, and Post | Methods and figure legend | Yes. Mean ± SEM is shown. | In Figure | P = 0.0001 | Figure legend | F (2, 9) = 29.11 | Figure legend |
| + - | Figure 8c | Repeated one-way ANOVA with Tukey–Kramer test | Methods and figure legend | 4, 4, and 4 | Yes. Pre, Restraint, and Post | Methods and figure legend | Yes. Mean ± SEM is shown. | In Figure | P = 0.0003 | Figure legend | F (2, 9) = 21.93 | Figure legend |
| + - | Figure 8e | Repeated one-way ANOVA with Tukey–Kramer test | Methods and figure legend | 4, 4, and 4 | Yes. Pre, C1 stimulation, and Post | Methods and figure legend | Yes. Mean ± SEM is shown. | In Figure | P = 0.0024 | Figure legend | F (2, 9) = 12.7 | Figure legend |
| + - | Figure 8f | Repeated one-way ANOVA with Tukey–Kramer test | Methods and figure legend | 4, 4, and 4 | Yes. Pre, C1 stimulation, and Post | Methods and figure legend | Yes. Mean ± SEM is shown. | In Figure | P = 0.0066 | Figure legend | F (2, 9) = 9.249 | Figure legend |
| + - | Figure S2d | Repeated one-way ANOVA with Tukey–Kramer test | Methods and figure legend | 27, 27, 27, and 27 | Yes. Dummy, 5Hz, 10Hz, 15Hz, and 20Hz | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P < 0.0001 | Figure legend | F (3, 104) = 128.1 | Figure legend |
| + - | Figure S2e | Repeated one-way ANOVA with Tukey–Kramer test | Methods and figure legend | 27, 27, 27, and 27 | Yes. Dummy, 5Hz, 10Hz, 15Hz, and 20Hz | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P < 0.0001 | Figure legend | F (3, 104) = 12.01 | Figure legend |
| + - | Figure S2f | Repeated one-way ANOVA with Tukey–Kramer test | Methods and figure legend | 27, 27, 27, and 27 | Yes. Dummy, 5Hz, 10Hz, 15Hz, and 20Hz | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P < 0.0001 | Figure legend | F (3, 104) = 87.49 | Figure legend |
| + - | Figure S3d | Unpaired t test | Methods and figure legend | 6 and 6 | Yes. Laser(-) and Laser(+) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P = 0.0067 | Figure legend | t=3.41 df=10 | Figure legend |
| + - | Figure S7 | One-way ANOVA with Tukey–Kramer test | Methods and figure legend | 5, 5, and 5 | Yes. IR(-)Laser(-), IR(-)Laser(+), and IR(+)Laser(-) | Methods and figure legend | Yes. Dot plots and mean ± SEM are shown. | In Figure | P = 0.0186 | Figure legend | F (2, 12) = 5.653 | Figure legend |
| + - | Supplemental Table 1 (fR) | One-way ANOVA with Tukey–Kramer test | Methods and figure legend | 27, 27, 27, and 27 | Yes. Dummy, 5Hz, 10Hz, 15Hz, and 20Hz | Methods and figure legend | Yes. mean ± SEM is shown. | In Table | P = 0.5613 | Table legend | F (2.284, 59.38) = 0.6214 | Table legend |
| + - | Supplemental Table 1 (Vt) | One-way ANOVA with Tukey–Kramer test | Methods and figure legend | 27, 27, 27, and 27 | Yes. Dummy, 5Hz, 10Hz, 15Hz, and 20Hz | Methods and figure legend | Yes. mean ± SEM is shown. | In Table | P = 0.1246 | Table legend | F (2.335, 60.7) = 2.092 | Table legend |
| + - | Supplemental Table 1 (Ve) | One-way ANOVA with Tukey–Kramer test | Methods and figure legend | 27, 27, 27, and 27 | Yes. Dummy, 5Hz, 10Hz, 15Hz, and 20Hz | Methods and figure legend | Yes. mean ± SEM is shown. | In Table | P = 0.5350 | Table legend | F (2.378, 61.83) = 0.6795 | Table legend |

## Representative figures

1. Are any representative images shown (including Western blots and immunohistochemistry/staining) in the paper?

   If so, what figure(s)?

Yes.

Representative images of Immunohistochemistry
Figure 3b1, 4a, 4c, S1, S4, and S5

Representative physiological data
Figure 6c, 8a, 8d, S2c, and S3a

2. For each representative image, is there a clear statement of how many times this experiment was successfully repeated and a discussion of any limitations in repeatability?

   If so, where is this reported (section, paragraph #)?

Yes.
For each representative image (Figure 3b1, 4a, 4c, S1, S4, and S5) and data (Figure 6c, 8a, 8d, S2c, and S3a), summarized data are shown in the Figures.

## Statistics and general methods

1. Is there a justification of the sample size?

   If so, how was it justified?

   Where (section, paragraph #)?

   Even if no sample size calculation was performed, authors should report why the sample size is adequate to measure their effect size.

Yes.

From our previous study (Inoue et al., JCI, 2016), we determined that the value of creatinine in IRI with no treatment is 1.47 (mA), IRI with treatment is 0.45 (mB), standard deviation is 0.34 (sd), maximum number of pairwise comparisons is 21 (tau), type I error is 0.05 (alpha), and power is 0.8 (power). We measured sample size using R as follows.

mA=1.47
mB=0.45
sd=0.34
tau=21
alpha=0.05
power=0.80
(n=2*(sd*(qnorm(1-alpha/(2*tau))+qnorm(power))/(mA-mB))^2)
ceiling(n)

This examination shows that the sample size is 5, so we used 5 to 6 mice for each group in IRI experiment.

This is described in the subsection "Statistical analyses" of the Methods section.

2. Are statistical tests justified as appropriate for every figure?

   Where (section, paragraph #)?

Yes.
Statistical tests used for all figures are appropriate as described in the subsection "Statistical analyses" of the Methods section.

   a. If there is a section summarizing the statistical methods in the methods, is the statistical test for each experiment clearly defined?

Yes.

   b. Do the data meet the assumptions of the specific statistical test you chose (e.g. normality for a parametric test)?

      Where is this described (section, paragraph #)?

Yes.
The normality test was conducted (D'Agostino & Pearson omnibus normality test or Kolmogorov-Smirnov test). This is described in the subsection "Statistical analyses" of the Methods section.

c. Is there any estimate of variance within each group of data?

Is the variance similar between groups that are being statistically compared?

Where is this described (section, paragraph #)?

Yes.

We used Brown-Forsythe test to check a similar valiance. If p value of Brown-Forsythe test is more than 0.05, we did one- or two-way ANOVA test followed by Tukey–Kramer test. If p value of Brown-Forsythe test is less than 0.05, we did Kruskal-Wallis test followed by Steel-Dwass test.

This is described in the subsection "Statistical analyses" of the Methods section.

d. Are tests specified as one- or two-sided?

Two-sided.

e. Are there adjustments for multiple comparisons?

Yes, all post-hoc tests are adjusted for multiple comparisons.

3. Are criteria for excluding data points reported?

Was this criterion established prior to data collection?

Where is this described (section, paragraph #)?

N/A

4. Define the method of randomization used to assign subjects (or samples) to the experimental groups and to collect and process data.

If no randomization was used, state so.

Where does this appear (section, paragraph #)?

"Injections of viral vector into the rostral ventrolateral medulla (RVLM)" in Methods

5. Is a statement of the extent to which investigator knew the group allocation during the experiment and in assessing outcome included?

If no blinding was done, state so.

Where (section, paragraph #)?

Yes.
"Optogenetic activation of C1 cells and renal ischemia-reperfusion injury (IRI)" and "Renal histology" in Methods

6. For experiments in live vertebrates, is a statement of compliance with ethical guidelines/regulations included?

Where (section, paragraph #)?

Yes.
"Animals" in Methods

7. Is the species of the animals used reported?

Where (section, paragraph #)?

Yes.
"Animals" in Methods

8. Is the strain of the animals (including background strains of KO/ transgenic animals used) reported?

Where (section, paragraph #)?

Yes.
"Animals" in Methods

9. Is the sex of the animals/subjects used reported?

Where (section, paragraph #)?

Yes.
"Animals" in Methods

10. Is the age of the animals/subjects reported?

Where (section, paragraph #)?

Yes.
"Animals" in Methods

11. For animals housed in a vivarium, is the light/dark cycle reported?

    Where (section, paragraph #)?

Yes.
"Injections of viral vector into the rostral ventrolateral medulla (RVLM)" in Methods

12. For animals housed in a vivarium, is the housing group (i.e. number of animals per cage) reported?

    Where (section, paragraph #)?

Yes.
"Injections of viral vector into the rostral ventrolateral medulla (RVLM)" in Methods

13. For behavioral experiments, is the time of day reported (e.g. light or dark cycle)?

    Where (section, paragraph #)?

Yes.
Methods, Whole-body plethysmography and Corticosterone measurement

14. Is the previous history of the animals/subjects (e.g. prior drug administration, surgery, behavioral testing) reported?

    Where (section, paragraph #)?

N/A

    a. If multiple behavioral tests were conducted in the same group of animals, is this reported?

       Where (section, paragraph #)?

N/A

15. If any animals/subjects were excluded from analysis, is this reported?

    Where (section, paragraph #)?

N/A

    a. How were the criteria for exclusion defined?

       Where is this described (section, paragraph #)?

N/A

    b. Specify reasons for any discrepancy between the number of animals at the beginning and end of the study.

       Where is this described (section, paragraph #)?

N/A

## Reagents

1. Have antibodies been validated for use in the system under study (assay and species)?

Yes.

    a. Is antibody catalog number given?

       Where does this appear (section, paragraph #)?

Catalog numbers are given of commercial sources.
"Brain histology" in Methods

    b. Where were the validation data reported (citation, supplementary information, Antibodypedia)?

       Where does this appear (section, paragraph #)?

Abbott et al., J Eur J Neurosci, 2014
"Brain histology" in Methods

2. Cell line identity

   a. Are any cell lines used in this paper listed in the database of commonly misidentified cell lines maintained by ICLAC and NCBI Biosample?

   Where (section, paragraph #)?

N/A

   b. If yes, include in the Methods section a scientific justification of their use--indicate here in which section and paragraph the justification can be found.

N/A

   c. For each cell line, include in the Methods section a statement that specifies:
   - the source of the cell lines
   - have the cell lines been authenticated? If so, by which method?
   - have the cell lines been tested for mycoplasma contamination?

   Where (section, paragraph #)?

N/A

## Data deposition

Data deposition in a public repository is mandatory for:

a. Protein, DNA and RNA sequences
b. Macromolecular structures
c. Crystallographic data for small molecules
d. Microarray data

Deposition is strongly recommended for many other datasets for which structured public repositories exist; more details on our data policy are available here. We encourage the provision of other source data in supplementary information or in unstructured repositories such as Figshare and Dryad.

We encourage publication of Data Descriptors (see Scientific Data) to maximize data reuse.

1. Are accession codes for deposit dates provided?

   Where (section, paragraph #)?

N/A

## Computer code/software

Any custom algorithm/software that is central to the methods must be supplied by the authors in a usable and readable form for readers at the time of publication. However, referees may ask for this information at any time during the review process.

1. Identify all custom software or scripts that were required to conduct the study and where in the procedures each was used.

Software: LabView
Use: Poincaré plot (Supplementary Figure 2)

2. If computer code was used to generate results that are central to the paper's conclusions, include a statement in the Methods section under "**Code availability**" to indicate whether and how the code can be accessed. Include version information as necessary and any restrictions on availability.

We do not use computer code in LabView (we make a program using icons). We explain how to calculate two standard deviation of Poincaré plot in Figure legend (Supplementary Figure 2).

## Human subjects

1. Which IRB approved the protocol?

   Where is this stated (section, paragraph #)?

   N/A

2. Is demographic information on all subjects provided?

   Where (section, paragraph #)?

   N/A

3. Is the number of human subjects, their age and sex clearly defined?

   Where (section, paragraph #)?

   N/A

4. Are the inclusion and exclusion criteria (if any) clearly specified?

   Where (section, paragraph #)?

   N/A

5. How well were the groups matched?

   Where is this information described (section, paragraph #)?

   N/A

6. Is a statement included confirming that informed consent was obtained from all subjects?

   Where (section, paragraph #)?

   N/A

7. For publication of patient photos, is a statement included confirming that consent to publish was obtained?

   Where (section, paragraph #)?

   N/A

## fMRI studies

For papers reporting functional imaging (fMRI) results please ensure that these minimal reporting guidelines are met and that all this information is clearly provided in the methods:

1. Were any subjects scanned but then rejected for the analysis after the data was collected?

   N/A

   a. If yes, is the number rejected and reasons for rejection described?

      Where (section, paragraph #)?

      N/A

2. Is the number of blocks, trials or experimental units per session and/or subjects specified?

   Where (section, paragraph #)?

   N/A

3. Is the length of each trial and interval between trials specified?

   N/A

| | |
|---|---|
| 4. Is a blocked, event-related, or mixed design being used? If applicable, please specify the block length or how the event-related or mixed design was optimized. | N/A |
| 5. Is the task design clearly described?<br>Where (section, paragraph #)? | N/A |
| 6. How was behavioral performance measured? | N/A |
| 7. Is an ANOVA or factorial design being used? | N/A |
| 8. For data acquisition, is a whole brain scan used?<br>If not, state area of acquisition. | N/A |
| a. How was this region determined? | N/A |
| 9. Is the field strength (in Tesla) of the MRI system stated? | N/A |
| a. Is the pulse sequence type (gradient/spin echo, EPI/spiral) stated? | N/A |
| b. Are the field-of-view, matrix size, slice thickness, and TE/TR/ flip angle clearly stated? | N/A |
| 10. Are the software and specific parameters (model/functions, smoothing kernel size if applicable, etc.) used for data processing and pre-processing clearly stated? | N/A |
| 11. Is the coordinate space for the anatomical/functional imaging data clearly defined as subject/native space or standardized stereotaxic space, e.g., original Talairach, MNI305, ICBM152, etc? Where (section, paragraph #)? | N/A |
| 12. If there was data normalization/standardization to a specific space template, are the type of transformation (linear vs. nonlinear) used and image types being transformed clearly described? Where (section, paragraph #)? | N/A |
| 13. How were anatomical locations determined, e.g., via an automated labeling algorithm (AAL), standardized coordinate database (Talairach daemon), probabilistic atlases, etc.? | N/A |
| 14. Were any additional regressors (behavioral covariates, motion etc) used? | N/A |
| 15. Is the contrast construction clearly defined? | N/A |
| 16. Is a mixed/random effects or fixed inference used? | N/A |

| | |
|---|---|
| a. If fixed effects inference used, is this justified? | N/A |
| 17. Were repeated measures used (multiple measurements per subject)? | N/A |
| a. If so, are the method to account for within subject correlation and the assumptions made about variance clearly stated? | N/A |
| 18. If the threshold used for inference and visualization in figures varies, is this clearly stated? | N/A |
| 19. Are statistical inferences corrected for multiple comparisons? | N/A |
| a. If not, is this labeled as uncorrected? | N/A |
| 20. Are the results based on an ROI (region of interest) analysis? | N/A |
| a. If so, is the rationale clearly described? | N/A |
| b. How were the ROI's defined (functional vs anatomical localization)? | N/A |
| 21. Is there correction for multiple comparisons within each voxel? | N/A |
| 22. For cluster-wise significance, is the cluster-defining threshold and the corrected significance level defined? | N/A |

## Additional comments

| | |
|---|---|
| Additional Comments | N/A |