

Corresponding Author:	<u>John K. Park/Dorian B. McGavern</u>	# Main Figures:	<u>8</u>
Manuscript Number:	<u>NN-T57125A</u>	# Supplementary Figures:	<u>4</u>
Manuscript Type:	<u>Technical Report</u>	# Supplementary Tables:	<u>1</u>
		# Supplementary Videos:	<u>0</u>

Reporting Checklist for Nature Neuroscience

This checklist is used to ensure good reporting standards and to improve the reproducibility of published results. For more information, please read [Reporting Life Sciences Research](#).

Please note that in the event of publication, it is mandatory that authors include all relevant methodological and statistical information in the manuscript.

□ Statistics reporting, by figure

- Please specify the following information for each panel reporting quantitative data, and where each item is reported (section, e.g. Results, & paragraph number).
- Each figure legend should ideally contain an exact sample size (n) for each experimental group/condition, where n is an exact number and not a range, a clear definition of how n is defined (for example x cells from x slices from x animals from x litters, collected over x days), a description of the statistical test used, the results of the tests, any descriptive statistics and clearly defined error bars if applicable.
- For any experiments using custom statistics, please indicate the test used and stats obtained for each experiment.
- Each figure legend should include a statement of how many times the experiment shown was replicated in the lab; the details of sample collection should be sufficiently clear so that the replicability of the experiment is obvious to the reader.
- For experiments reported in the text but not in the figures, please use the paragraph number instead of the figure number.

Note: Mean and standard deviation are not appropriate on small samples, and plotting independent data points is usually more informative. When technical replicates are reported, error and significance measures reflect the experimental variability and not the variability of the biological process; it is misleading not to state this clearly.

FIGURE NUMBER	TEST USED		n			DESCRIPTIVE STATS (AVERAGE, VARIANCE)		P VALUE		DEGREES OF FREEDOM & F/t/z/R/ETC VALUE	
	WHICH TEST?	SECTION & PARAGRAPH #	EXACT VALUE	DEFINED?	SECTION & PARAGRAPH #	REPORTED?	SECTION & PARAGRAPH #	EXACT VALUE	SECTION & PARAGRAPH #	VALUE	SECTION & PARAGRAPH #
example 1a	one-way ANOVA	Fig. legend	9, 9, 10, 15	mice from at least 3 litters/group	Methods para 8	error bars are mean +/- SEM	Fig. legend	p = 0.044	Fig. legend	F(3, 36) = 2.97	Fig. legend
example results, para 6	unpaired t-test	Results para 6	15	slices from 10 mice	Results para 6	error bars are mean +/- SEM	Results para 6	p = 0.0006	Results para 6	t(28) = 2.808	Results para 6

FIGURE NUMBER	TEST USED		n			DESCRIPTIVE STATS (AVERAGE, VARIANCE)		P VALUE		DEGREES OF FREEDOM & F/t/z/R/ETC VALUE	
	WHICH TEST?	SECTION & PARAGRAPH #	EXACT VALUE	DEFINED?	SECTION & PARAGRAPH #	REPORTED?	SECTION & PARAGRAPH #	EXACT VALUE	SECTION & PARAGRAPH #	VALUE	SECTION & PARAGRAPH #
+ - 1b-j	NA		6	Cells from three different differentiation experiments. Staining was done in replicates.	Fig legend						
+ - 2a	NA		3	Three different cell culture samples	Fig. legend						
+ - 2b	one-way ANOVA FDR MCC followed by Tukey HSD	Methods Pg 18, 19 para # 4 and 1	3	RNA from cells from three different differentiation experiments.	Fig. legend			ANOVA corrected P<0.05 post hoc P <0.05	Methods Pg 18, 19 para # 4 and 1		
+ - 2c	one-way ANOVA FDR MCC followed by Tukey HSD	Methods Pg 18, 19 para # 4 and 1	3	RNA from cells from three different differentiation experiments.	Fig. legend			ANOVA corrected P<0.05 post hoc P <0.05	Methods Pg 18, 19 para # 4 and 1		
+ - 3	NA		3	Three different cell culture samples done in replicates.	Fig. legend						
+ - 4	NA		6	Cells from three different differentiation experiments. Staining was done in replicates.	Fig legend						
+ - 5a	NA		4	Two technical replicates in two different experiments	Fig. legend						
+ - 5b	one-way ANOVA FDR MCC followed by Tukey HSD	Methods Pg 18, 19 para # 4 and 1	3	RNA from cells from three different differentiation experiments.	Fig. legend			ANOVA corrected P<0.05 post hoc P <0.05	Methods Pg 18, 19 para # 4 and 1		
+ - 5c	one-way ANOVA FDR MCC followed by Tukey HSD	Methods Pg 18, 19 para # 4 and 1	3	RNA from cells from three different differentiation experiments.	Fig. legend			ANOVA corrected P<0.05 post hoc P <0.05	Methods Pg 18, 19 para # 4 and 1		

+ -	6a,b	NA		6	Cells from three different differentiation experiments, done in replicates.	Fig. legend						
+ -	6c	NA		4	Cells from two different implantation experiments. Staining was carried out for at least two sections.	Fig. legend						
+ -	7	log-rank test (Mantel-Cox test)	Fig leg	10	10 animals for each treatment condition	Fig legend Results, Pg. 7			<0.0001	Fig legend	df=1	Results, Pg. 7
+ -	Table 1	NA		7	At least two technical replicates in three different experiments	Table 1 legend	Standard Error of Mean (SEM)	Table 1				
	Supplementary Fig 1	NA		2	From two differentiation experiments	Fig. leg						
	Supplementary Fig 2	one-way ANOVA FDR MCC followed by Tukey HSD	Methods Pg 18, 19 para # 4 and 1	3	cells from three different differentiation experiments.	Fig. leg			ANOVA corrected P<0.05 post hoc P <0.05	Methods Pg 18, 19 para # 4 and 1		
	Supplementary Fig 3	NA		3	Cells from three different differentiation experiments	Fig. leg.						
	Supplementary Fig 4	NA		3, 4	Three different cell-culture for the alkaline phosphatase and immunocytochemistry experiments and 4 mice for the chimera experiments.	Fig. leg.						
	Supplementary Table 1	NA					NA					

□ Representative figures

1. Are any representative images shown (including Western blots and immunohistochemistry/staining) in the paper?

If so, what figure(s)?

Yes, figures 1,3, 4 and 6

2. For each representative image, is there a clear statement of how many times this experiment was successfully repeated and a discussion of any limitations in repeatability?

If so, where is this reported (section, paragraph #)?

Figure legends

□ Statistics and general methods

1. Is there a justification of the sample size?

If so, how was it justified?

Where (section, paragraph #)?

Even if no sample size calculation was performed, authors should report why the sample size is adequate to measure their effect size.

Yes, Methods, Page 20

2. Are statistical tests justified as appropriate for every figure?

Where(section, paragraph#)?

Yes, Methods, Pg. 18, 19 (Microarray data analysis), Fig. legend 7, Results- Last paragraph, Pg.6

- a. If there is a section summarizing the statistical methods in the methods, is the statistical test for each experiment clearly defined?

Yes, Methods, Page 21.

- b. Do the data meet the assumptions of the specific statistical test you chose (e.g. normality for a parametric test)?

Where is this described (section, paragraph #)?

Yes, Methods, Pg. 18, 19 (Microarray data analysis).

- c. Is there any estimate of variance within each group of data?

Is the variance similar between groups that are being statistically compared?

Where is this described (section, paragraph #)?

Yes, Methods, Pg. 18,19 (Microarray data analysis). Data has equal variance across groups after log₂ transformation and RMA normalization.

- d. Are tests specified as one- or two-sided?

Used one-way ANOVA under FDR MCC (BH) condition followed by Tukey HSD post hoc test.

- e. Are there adjustments for multiple comparisons?

Yes, FDR MCC was used.

3. To promote transparency, *Nature Neuroscience* has stopped allowing bar graphs to report statistics in the papers it publishes. If you have bar graphs in your paper, please make sure to switch them to dot-plots (with central and dispersion statistics displayed) or to box-and-whisker plots to show data distributions.

4. Are criteria for excluding data points reported?
Was this criterion established prior to data collection?
Where is this described (section, paragraph #)?
- No data points were removed
5. Define the method of randomization used to assign subjects (or samples) to the experimental groups and to collect and process data.
If no randomization was used, state so.
Where does this appear (section, paragraph #)?
- Block randomization for tumor sizes was used for animal studies.
Methods, Pg. 20. Section on Tumor implantation and treatment delivery of mice with neonatal brain-isolated or mouse iPSC microglia.
6. Is a statement of the extent to which investigator knew the group allocation during the experiment and in assessing outcome included?
If no blinding was done, state so.
Where (section, paragraph #)?
- Block randomization for tumor sizes was used for animal studies.
7. For experiments in live vertebrates, is a statement of compliance with ethical guidelines/regulations included?
Where (section, paragraph #)?
- Yes
Methods, Pg. 20, Para # 1 and Page 21, Para # 1.
8. Is the species of the animals used reported?
Where (section, paragraph #)?
- Yes, Methods, Pg. 14 (Mouse strains) and Pg. 20, Para # 1 and Para # 3
9. Is the strain of the animals (including background strains of KO/transgenic animals used) reported?
Where (section, paragraph #)?
- Yes, Methods, Pg. 14 (Mouse strains).
10. Is the sex of the animals/subjects used reported?
Where (section, paragraph #)?
- Yes, Methods, Pg. 20, Para # 1 and Para # 3
11. Is the age of the animals/subjects reported?
Where (section, paragraph #)?
- Yes, Methods, Pg. 20, Para # 1 and Para # 3
12. For animals housed in a vivarium, is the light/dark cycle reported?
Where (section, paragraph #)?
- Yes, Methods, Pg. 20, Para # 1 and Page 21, Para # 1.
13. For animals housed in a vivarium, is the housing group (i.e. number of animals per cage) reported?
Where (section, paragraph #)?
- Yes, Methods, Pg. 20, Para # 1 and Page 21, Para # 1.
14. For behavioral experiments, is the time of day reported (e.g. light or dark cycle)?
Where (section, paragraph #)?
- NA

15. Is the previous history of the animals/subjects (e.g. prior drug administration, surgery, behavioral testing) reported?

NA

Where (section, paragraph #)?

a. If multiple behavioral tests were conducted in the same group of animals, is this reported?

Where (section, paragraph #)?

16. If any animals/subjects were excluded from analysis, is this reported?

No animals were excluded from the analysis. Methods, Page 21, (Tumor implantation and treatment delivery of mice with mouse neonatal brain-isolated or mouse iPSC microglia.)

Where (section, paragraph #)?

a. How were the criteria for exclusion defined?

NA

Where is this described (section, paragraph #)?

b. Specify reasons for any discrepancy between the number of animals at the beginning and end of the study.

NA

Where is this described (section, paragraph #)?

□ Reagents

1. Have antibodies been validated for use in the system under study (assay and species)?

Yes

a. Is antibody catalog number given?

Where does this appear (section, paragraph #)?

Yes, Methods- Immunofluorescence, FACS analyses and miPSC-engraftment studies sections.

b. Where were the validation data reported (citation, supplementary information, Antibodypedia)?

Where does this appear (section, paragraph #)?

Yes, Methods - Immunofluorescence, FACS analyses and miPSC-engraftment studies sections.

2. Cell line identity

No

a. Are any cell lines used in this paper listed in the database of commonly misidentified cell lines maintained by [ICLAC](#) and [NCBI Biosample](#)?

Where (section, paragraph #)?

b. If yes, include in the Methods section a scientific justification of their use—indicate here in which section and paragraph the justification can be found.

NA

- c. For each cell line, include in the Methods section a statement that specifies:
- the source of the cell lines
 - have the cell lines been authenticated? If so, by which method?
 - have the cell lines been tested for mycoplasma contamination?

Where (section, paragraph #)?

Methods - Creation of Cx3cr1gfp/+ miPSC and human iPSC cell lines and Cell culture section. Pages 14, 15.

□ Data availability

Provide a Data availability statement in the Methods section under "Data availability", which should include, where applicable:

- Accession codes for deposited data
- Other unique identifiers (such as DOIs and hyperlinks for any other datasets)
- At a minimum, a statement confirming that all relevant data are available from the authors
- Formal citations of datasets that are assigned DOIs
- A statement regarding data available in the manuscript as source data
- A statement regarding data available with restrictions

See our [data availability and data citations policy page](#) for more information.

Data deposition in a public repository is mandatory for:

- a. Protein, DNA and RNA sequences
- b. Macromolecular structures
- c. Crystallographic data for small molecules
- d. Microarray data

Deposition is strongly recommended for many other datasets for which structured public repositories exist; more details on our data policy are available [here](#). We encourage the provision of other source data in supplementary information or in unstructured repositories such as [Figshare](#) and [Dryad](#).

We encourage publication of Data Descriptors (see [Scientific Data](#)) to maximize data reuse.

Where is the Data Availability statement provided (section, paragraph #)?

Microarray data for both the mouse and human samples are available for download from the NCBI Gene Expression Omnibus (<http://www.ncbi.nlm.nih.gov/geo/>); see SuperSeries GSE78116.

Accession codes, Pg.

Microarray data for both the mouse and human samples are available for download from the NCBI Gene Expression Omnibus (<http://www.ncbi.nlm.nih.gov/geo/>); see SuperSeries GSE78116.

Accession codes, Pg. 8.

□ Computer code/software

Any custom algorithm/software that is central to the methods must be supplied by the authors in a usable and readable form for readers at the time of publication. However, referees may ask for this information at any time during the review process.

2. If computer code was used to generate results that are central to the paper's conclusions, include a statement in the Methods section under "**Code availability**" to indicate whether and how the code can be accessed. Include version information as necessary and any restrictions on availability.

□ Human subjects

1. Which IRB approved the protocol?

Where is this stated (section, paragraph #)?

Lonza, NIAID Institutional Review Board (IRB). Human subjects were not directly involved in the current study. Cells from human patients were used which were obtained and characterized in a previous study (Ref. 37 and 38).

2. Is demographic information on all subjects provided?

Where (section, paragraph #)?

Please refer to References 37 and 38.

3. Is the number of human subjects, their age and sex clearly defined?

Where (section, paragraph #)?

Please refer to References 37 and 38.

4. Are the inclusion and exclusion criteria (if any) clearly specified?

Where (section, paragraph #)?

Please refer to References 37 and 38.

5. How well were the groups matched?

Where is this information described (section, paragraph #)?

Please refer to References 37 and 38.

6. Is a statement included confirming that informed consent was obtained from all subjects?

Where (section, paragraph #)?

We confirm that written signed informed consent was obtained from normal healthy donors.

7. For publication of patient photos, is a statement included confirming that consent to publish was obtained?

Where (section, paragraph #)?

NA

□ fMRI studies

For papers reporting functional imaging (fMRI) results please ensure that these minimal reporting guidelines are met and that all this information is clearly provided in the methods:

1. Were any subjects scanned but then rejected for the analysis after the data was collected?

- a. If yes, is the number rejected and reasons for rejection described?

Where (section, paragraph #)?

2. Is the number of blocks, trials or experimental units per session and/or subjects specified?

Where (section, paragraph #)?

3. Is the length of each trial and interval between trials specified?

4. Is a blocked, event-related, or mixed design being used? If applicable, please specify the block length or how the event-related or mixed design was optimized.

5. Is the task design clearly described?

Where (section, paragraph #)?

6. How was behavioral performance measured?

7. Is an ANOVA or factorial design being used?

8. For data acquisition, is a whole brain scan used?

If not, state area of acquisition.

- a. How was this region determined?

9. Is the field strength (in Tesla) of the MRI system stated?

- a. Is the pulse sequence type (gradient/spin echo, EPI/spiral) stated?

- b. Are the field-of-view, matrix size, slice thickness, and TE/TR/flip angle clearly stated?

10. Are the software and specific parameters (model/functions, smoothing kernel size if applicable, etc.) used for data processing and pre-processing clearly stated?

11. Is the coordinate space for the anatomical/functional imaging data clearly defined as subject/native space or standardized stereotaxic space, e.g., original Talairach, MNI305, ICBM152, etc? Where (section, paragraph #)?

12. If there was data normalization/standardization to a specific space template, are the type of transformation (linear vs. nonlinear) used and image types being transformed clearly described? Where (section, paragraph #)?

13. How were anatomical locations determined, e.g., via an automated labeling algorithm (AAL), standardized coordinate database (Talairach daemon), probabilistic atlases, etc.?

14. Were any additional regressors (behavioral covariates, motion etc) used?
15. Is the contrast construction clearly defined?
16. Is a mixed/random effects or fixed inference used?
- a. If fixed effects inference used, is this justified?
17. Were repeated measures used (multiple measurements per subject)?
- a. If so, are the method to account for within subject correlation and the assumptions made about variance clearly stated?
18. If the threshold used for inference and visualization in figures varies, is this clearly stated?
19. Are statistical inferences corrected for multiple comparisons?
- a. If not, is this labeled as uncorrected?
20. Are the results based on an ROI (region of interest) analysis?
- a. If so, is the rationale clearly described?
- b. How were the ROI's defined (functional vs anatomical localization)?
21. Is there correction for multiple comparisons within each voxel?
22. For cluster-wise significance, is the cluster-defining threshold and the corrected significance level defined?

□ Additional comments

Additional Comments