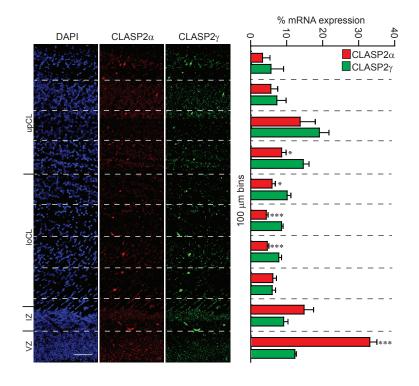
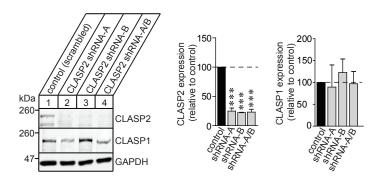
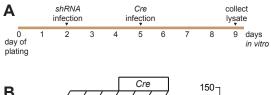
Suppl. Figure 1

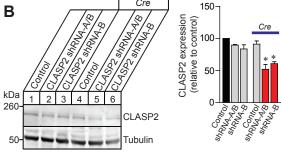


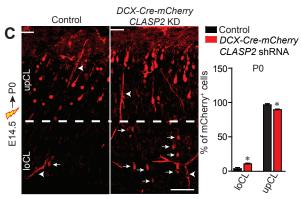
Suppl Figure 2



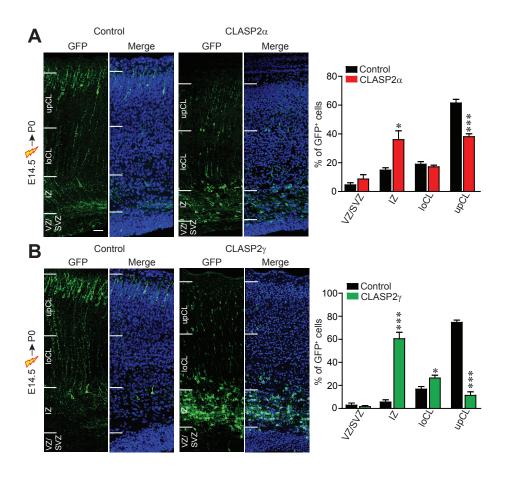
Suppl Figure 3



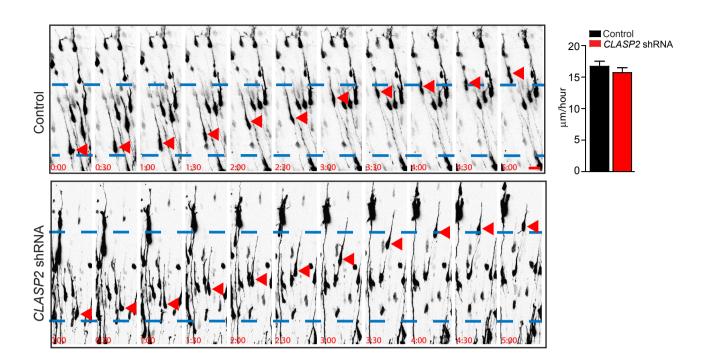




Suppl. Figure 4



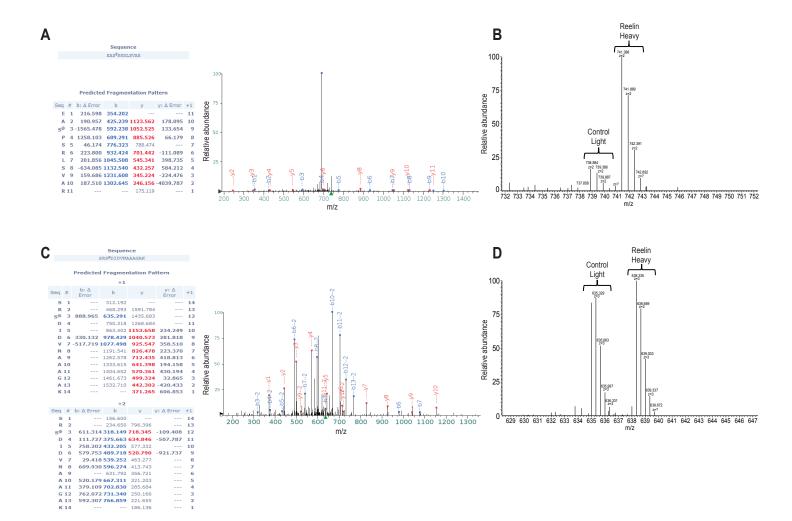
Suppl. Figure 5



Suppl Figure 6

Amino acids mutated in human CLASP2α	Equivalent amino acid sites in mouse CLASP2γ	Phosphosites regulated by Reelin in M/S
568	505	
572	509	
576	513	513
594	531	
598	535	
602	539	
606	543	543
610	547	
614	551	551

Suppl. Figure 7



SUPPLEMENTAL INFORMATION

Supplemental information includes seven figures and two movies.

Figure S1. CLASP2α and CLASP2γ mRNA Expression in the Mouse Cerebral Cortex at Postnatal Day 0. Representative images showed *in situ* hybridization of cortical sections for CLASP2α and CLASP2γ at postnatal day 0. Fluorescent expression were quantitated every 100 μm bins from the ventricle to the pial surface regions (VZ = ventricular zone, IZ = intermediate zone, loCL = lower cortical layer, upCL = upper cortical layer). CLASP2α expression was the highest in the first 100 μm which represents the proliferative ventricular zones (n = 6 brain sections for each group). Data are means \pm SEM and statistical significance was assessed using one-way ANOVA (*p < 0.05, ***p<0.0001). Scale bar represents 50 μm.

Figure S2. Efficiency of Individual *CLASP2* shRNAs in Knocking Down Expression of Endogenous CLASP2 in Primary Mouse Neurons. Lentiviral infection of primary cortical neurons of individual (shRNA-A, shRNA-B) and combinations (shRNA-A/B) of *CLASP2* shRNAs efficiently dowregulated CLASP2 but not CLASP1 protein expression (n = 2 independent experiments).

Figure S3. Neuron-Specific Knockdown of CLASP2 Affects Neuronal Migration.

- (A) Time line of lentiviral infection of conditional pSico-*CLASP2* shRNA and *cre* recombinase to test the levels of CLASP2 knockdown in primary cortical neurons.
- (B) Lysates of primary cortical neurons infected with individual conditional pSico-shRNA-B and combinations (shRNA-A/B) of *CLASP2* shRNAs efficiently downregulated CLASP2 protein expression in *cre*-dependent manner.

(C) Mouse embryonic brains were electroporated *in utero* with pSico-*CLASP2* shRNA-A/B or scrambled control and DCX-*cre*-mCherry at E14.5 and analyzed at P0 (control, n = 3 brains; *CLASP2* shRNA, n = 3 brains). Coronal sections of the cortex were visualized for transfected mCherry-positive neurons (red). Solid and dash white lines indicate the demarcations for pial surface and lower and upper cortical regions (loCL = lower cortical layer, upCL = upper cortical layer), respectively. Arrows indicates the number of mCherry positive neurons in the loCL. Arrowheads indicate blood vessels.

Data are means \pm SEM and statistical significance was assessed using one-way ANOVA (*p < 0.05). Scale bar represents 50 μ m.

Figure S4. CLASP2 Overexpression Alters Neuronal Migration.

(A-B) Mouse embryos were electroporated *in utero* with control GFP, GFP-CLASP2 α or GFP-CLASP2 γ full-length expression vectors at E14.5 and analyzed at P0 (control, n = 4 brains; CLASP2 α , n = 4 brains and CLASP2 γ , n = 4 brains). Coronal sections of the cortex were visualized for transfected GFP-positive neurons (green) and cell nuclei (Hoeschst 33342, blue). White lines indicate the demarcations for different cortical regions (VZ = ventricular zone, SVZ = subventricular zone, IZ = intermediate zone, loCL = lower cortical layer, upCL = upper cortical layer).

Data are means \pm SEM and statistical significance was assessed using one-way ANOVA (*p < 0.05, ***p<0.0001). Scale bar represents 50 μ m.

Figure S5. CLASP2 Knockdown Does Not Alter the Rate of Neuronal Migration.

Mouse embryos were electroporated *in utero* with GFP-tagged *CLASP2* shRNAs or scrambled control at E14.5 and brain slices were analyzed at E17.5. Time-lapse imaging of migrating neurons expressing GFP scrambled control or GFP-*CLASP2* shRNA (arrowhead) were captured every 30 seconds (see Supplementary Movie 1 and 2). Average rates of migration showed CLASP2 knockdown did not alter the rate of neuronal migration (control, n = 82 neurons from 4 brain slices; *CLASP2* shRNA, n = 71 neurons from 3 brain slices).

Data are means \pm SEM and statistical significance was assessed using unpaired t test. Scale bar represents 20 μ m.

Figure S6. Phosphosites in Murine CLASP2γ Regulated by Reelin.

The first column indicates amino acids mutated in human CLASP2α phosphomutants within the positively-charged S/A rich region. The second column represents the equivalent amino acid sites in mouse CLASP2γ. The third column indicates the three amino acid phosphosites in murine CLASP2γ regulated by Reelin in mouse primary neurons by tandem mass spectrometry.

Figure S7. Phosphorylation Sites to CLASP2 in Primary Mouse Neurons in Response to Reelin Stimulation. Representative tandem mass spectra showing the fragmentation pattern of two phosphopeptides (A and C). The peptide in panel A shows an approximately eight-fold increase in phosphorylation following Reelin treatment, while panel C shows a control peptide with no change following Reelin. Panels B and D depict the full mass scan showing the ratio of the light (control-treated neurons) and heavy (Reelin-treated neurons) peptides for the same peptides.

Movie S1. Movie of time-lapse imaging of GFP-positive migrating neurons electroporated with control GFP-scrambled plasmid.

Movie S2. Movie of time-lapse imaging of GFP-positive migrating neurons electroporated with GFP-CLASP2 shRNAs.