

Cell Line	A6 Mapped Reads	COII Mapped Reads	CYb Mapped Reads	MURF2 Mapped Reads	ND7 Mapped Reads	Total Number of Mapped Reads	Unmapped Reads Excluded from analysis	Total Reads in Sample	% of Mapped Reads
427wt	66,513	204,222	287,122	300,691	252,105	1,110,653	353,574	1,464,227	75.9
MGA	29,984	106,824	142,268	169,248	136,959	585,283	179,268	764,551	76.6
KREN2 null	220,567	631,677	863,295	1,076,317	692,339	3,484,195	859,029	4,343,224	80.2
KREN3 null	57,028	81,897	172,841	153,394	85,745	550,905	247,288	798,193	69.0
KREN1 only	136,160	228,734	299,137	418,238	289,855	1,372,124	296,590	1,668,714	82.2
KREN2 only	133,800	417,694	468,452	527,111	455,485	2,002,542	737,825	2,740,367	73.1
KREN3 only	141,482	262,820	414,425	401,130	308,994	1,528,851	420,566	1,949,417	78.4
Triple null	77,114	132,076	170,174	188,131	142,762	710,257	199,634	909,891	78.1
sum:	862,648	2,065,944	2,817,714	3,234,260	2,364,244	11,344,810	3,293,774	14,638,584	77.5

Table S1. Read counts mapped to target mitochondrial transcripts by cell line. Total number of reads identified for target amplicons analysed by Illumina sequencing.

Cell Line	A6 Unique Reads	COII Unique Reads	CYb Unique Reads	MURF2 Unique Reads	ND7 Unique Reads
427wt	167	47	1546	1929	1166
MGA	95	37	1592	2108	1012
KREN2 null	419	48	701	5480	3288
KREN3 null	118	21	586	4426	729
KREN1 only	103	21	103	573	335
KREN2 only	1397	34	1605	3392	7775
KREN3 only	1238	34	732	3678	3521
Triple null	49	19	85	90	141

Table S2. Unique read sequence counts mapped to target mitochondrial transcripts by cell line. Numbers of unique read sequences identified for target amplicons analysed by Illumina sequencing.

	A6 (ED)	A6 (Partial)	A6 (PRE)	COII (ED)	COII (Partial)	COII (PRE)	CYb (ED)	CYb (Partial)	CYb (PRE)	MURF2 (ED)	MURF2 (Partial)	MURF2 (PRE)	ND7 (ED)	ND7 (Partial)	ND7 (PRE)	Total Mapped
427wt	-	1,955	64,558	1,567	956	201,699	45,099	42,903	199,120	132,879	89,365	78,447	58,582	50,031	143,492	1,110,653
MGA	-	808	29,176	496	323	106,005	1	22,198	120,069	52,265	61,339	55,644	26,880	27,756	82,323	585,283
KREN2 null	9	67,091	153,467	1,055	1,211	629,411	2	32,788	830,505	9,184	934,361	132,772	128,889	237,778	325,672	3,484,195
KREN3 null	-	1,397	55,631	1	144	81,752	2,510	6,020	164,311	23,153	81,580	48,661	26,261	13,575	45,909	550,905
KREN1 only	-	43,005	93,155	-	442	228,292	1	2,595	296,541	6	311,566	106,666	6	131,654	158,195	1,372,124
KREN2 only	-	19,291	114,509	-	797	416,897	8,472	42,217	417,763	450	391,937	134,724	30	339,395	116,060	2,002,542
KREN3 only	-	22,555	118,927	249	588	261,983	-	14,576	399,849	22,778	315,805	62,547	22	173,702	135,270	1,528,851
Triple Null	-	7,454	69,660	-	248	131,828	-	1,587	168,587	1	1,843	186,287	4	16,470	126,288	710,257
sum:	9	163,556	699,083	3,368	4,709	2,057,867	56,085	164,884	2,596,745	240,716	2,187,796	805,748	240,674	990,361	1,133,209	11,344,810

Table S3. Read counts identified as pre-edited, partially edited, or fully edited by PARERS for each mRNA target and cell line. Numbers of reads identified for each mRNA from each cell line after classification as pre-edited (PRE), partially edited (Partial), or fully edited (ED).

	INSERTION								DELETION					
	1U	2U	3U	4U	5U	6U	7U	8U	1U	2U	3U	4U	5U	
427wt	4740	2371	174	3	78	16	7	0	427wt	790	334	128	1	0
MGA	6392	2454	222	25	102	20	1	1	MGA	1125	386	183	0	0
KREN2 null	3904	1429	159	29	228	42	9	2	KREN2 null	1478	949	429	0	0
KREN3 null	4677	1220	39	4	105	1	6	0	KREN3 null	2404	895	194	4	0
KREN1 only	192	7	1	0	46	0	2	0	KREN1 only	6050	76896	12657	154	0
KREN2 only	5540	12236	1003	98	108	38	45	0	KREN2 only	599	15	4	0	0
KREN3 only	4149	18049	4115	266	354	165	57	29	KREN3 only	331	23	2	0	0
Triple null	763	59	2	2	51	5	4	0	Triple null	24612	1354	191	9	0
	30358	37825	5715	426	1072	288	132	33		37389	80851	13789	168	0

Table S4. Normalized read counts (reads per million reads) identified within reads that contained a single edited site. Data for Figure 5 in tabular form, showing read numbers normalized to the total number of reads for each cell line (Column 6 in Table S1). Columns show the number of Us either deleted or inserted at a single editing site.

Oligo Name	Oligo Sequence
<i>Oligos for knockout constructs</i>	
KREN1 FOR long A	GAAACGGTTACCAGCGACG
KREN1 REV long H	TTGCTGAAGCACGCAAATTTTC
KREN1 FOR nested A	CTCACAAGGAATTGGATA
KREN1 FOR nested H	CGCAAAGATAAGTGACATATG
KREN1 REV B	ATACGAAGTTATAAGCTTATCAATGCTTTTTACTCCTTTTC
KREN1 FOR C	ACCATGGTGTGCTTAATTAC
KREN1 REV D	ATACGAAGTTATAAGCTTATCCTACCTTTAGCCACTTAGCAA
KREN1 FOR E	TTATGGATCCTAAGTGGGTCTAATTATTTGGATGATGAGGT
KREN1 REV F	TGTACATGAGTGGAGGACATTG
KREN1 FOR G	TTATGGATCCTAAGTGGGTCCAAGGATGGGAATATGCAT
KREN2 FOR long A	TCTGGTGGAAATAGAGAAG
KREN2 REV long H	TTGTAGAGCTGAGACGCT
KREN2 FOR nested A	TCCTCTTCTTCTTACTCAC
KREN2 FOR nested H	AATACCAAATTTAAGAAACG
KREN2 REV B	ATACGAAGTTATAAGCTTATCGAAGTTCGTGGTTCCTTGC
KREN2 FOR C	AGTGTTCACTTACCTCCCA
KREN2 REV D	ATACGAAGTTATAAGCTTATCATGGTAATGCCTTACTTCAA
KREN2 FOR E	TTATGGATCCTAAGTGGGTCTGATTACATTCCTACTTTC
KREN2 REV F	CACATGAACATAAAGAAGAA
KREN2 FOR G	TTATGGATCCTAAGTGGGTCCACCGGGTGTGCATCCAT
KREN3 FOR long A	GGTAGTAGCAAACAATTGTGC
KREN3 REV long H	ATACGACAAGAAGTCTTGCTA
KREN3 FOR nested A	TTGTGGTCTTTTTCCCATCT
KREN3 FOR nested H	CATGATAGGAAAAGTAACAA
KREN3 REV B	ATACGAAGTTATAAGCTTATCGAGGTATAAAGCAACTAGGGAG
KREN3 FOR C	CCTGCGGCACTACGACTAC
KREN3 REV D	ATACGAAGTTATAAGCTTATCTTGGCGAATTTACAAAGCT
KREN3 FOR E	TTATGGATCCTAAGTGGGTCTCGTCATTGGAGGTTTGC
KREN3 REV F	TGACAAAACGTGCGGAGC
KREN3 FOR G	TTATGGATCCTAAGTGGGTCCGCATAGGTGGTTATGGCG
<i>Oligos for First Round RT-PCR for RNAseq analysis</i>	
11209 ND7-5' FOR	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGATGACTACATGATAAGTA
11210 ND7-5' REV	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGACTTTTCTGTACCACGATGC
11215 MURF2 FOR	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGATAGAAAGGTATATAATCTATAATG
11216 MURF2 REV	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGCATAACAAATCAAAAACACGAC
11217 A6 FOR	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGAGAGCAGGAAAGGTTAG
11218 A6 REV	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGTTTGTATCTTATTCTATAACTCC
11211 CYb FOR	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGGTTAAGAATAATGGTTATAAATT
11212 CYb REV	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGCAACCTGACATTAAGAC
11213 COII FOR	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGATTACAGTGAACCATGTATTGACATT

Oligos for Second Round PCR for RNAseq
analysis

P5-TAGATCGC-F1 (427wt)	AATGATACGGCGACCACCGAGATCTACACTAGATCGCTCGTCGGCAGCGTC
P5-CTCTCTAT-F2 (MGA)	AATGATACGGCGACCACCGAGATCTACACCTCTCTATTCGTCGGCAGCGTC
P5-TATCCTCT-F3 (N2null)	AATGATACGGCGACCACCGAGATCTACACTATCCTCTTCGTCGGCAGCGTC
P5-AGAGTAGA-F4 (N3null)	AATGATACGGCGACCACCGAGATCTACACAGAGTAGATCGTCGGCAGCGTC
P5-GTAAGGAG-F5 (N1only)	AATGATACGGCGACCACCGAGATCTACACGTAAGGAGTCGTCGGCAGCGTC
P5-ACTGCATA-F6 (N2only)	AATGATACGGCGACCACCGAGATCTACACACTGCATATCGTCGGCAGCGTC
P5-AAGGAGTA-F7 (N3only)	AATGATACGGCGACCACCGAGATCTACACAAGGAGTATCGTCGGCAGCGTC
P5-CTAAGCCT-F8 (Triple null)	AATGATACGGCGACCACCGAGATCTACACCTAAGCCTTCGTCGGCAGCGTC
P7-TCGCCTTA-R1 (427wt)	CAAGCAGAAGACGGCATAACGAGATTCGCCTTAGTCTCGTGGGCTCGG
P7-CTAGTACG-R2 (MGA)	CAAGCAGAAGACGGCATAACGAGATCTAGTACGGTCTCGTGGGCTCGG
P7-TTCTGCCT-R3 (N2null)	CAAGCAGAAGACGGCATAACGAGATTTCTGCCTGTCTCGTGGGCTCGG
P7-GCTCAGGA-R4 (N3null)	CAAGCAGAAGACGGCATAACGAGATGCTCAGGAGTCTCGTGGGCTCGG
P7-AGGAGTCC-R5 (N1only)	CAAGCAGAAGACGGCATAACGAGATAGGAGTCCGTCTCGTGGGCTCGG
P7-CATGCCTA-R6 (N2only)	CAAGCAGAAGACGGCATAACGAGATCATGCCTAGTCTCGTGGGCTCGG
P7-GTAGAGAG-R7 (N3only)	CAAGCAGAAGACGGCATAACGAGATGTAGAGAGTCTCGTGGGCTCGG
P7-CCTCTCTG-R8 (Triple null)	CAAGCAGAAGACGGCATAACGAGATCCTCTCTGGTCTCGTGGGCTCGG

Table S7. Oligo sequences used in this study.

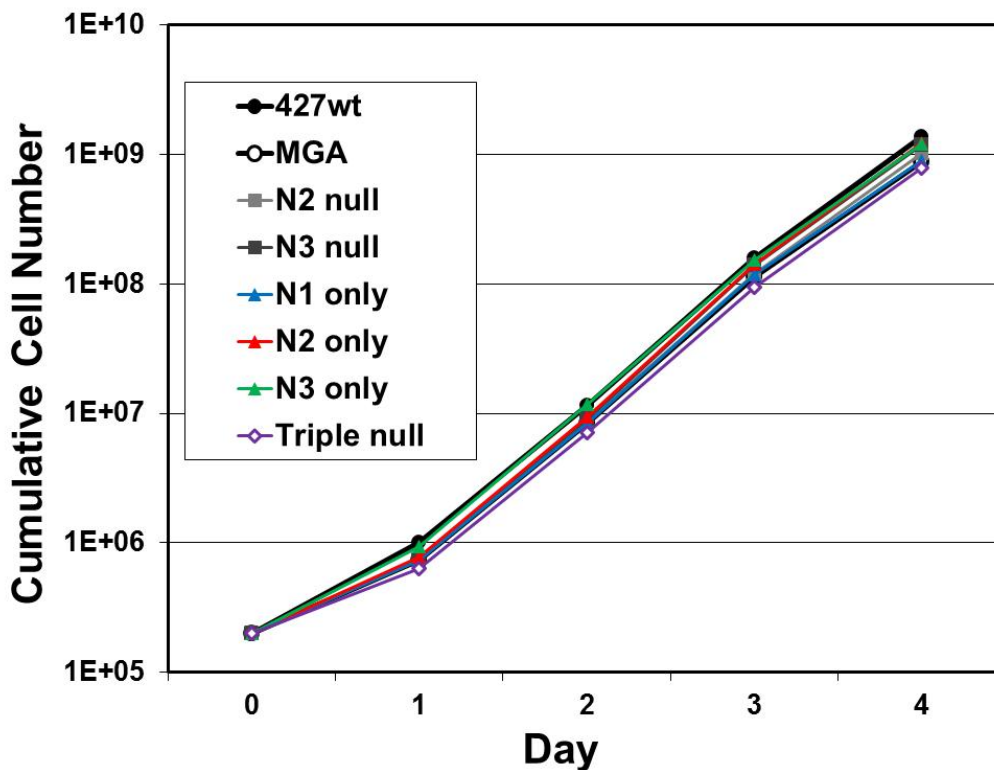
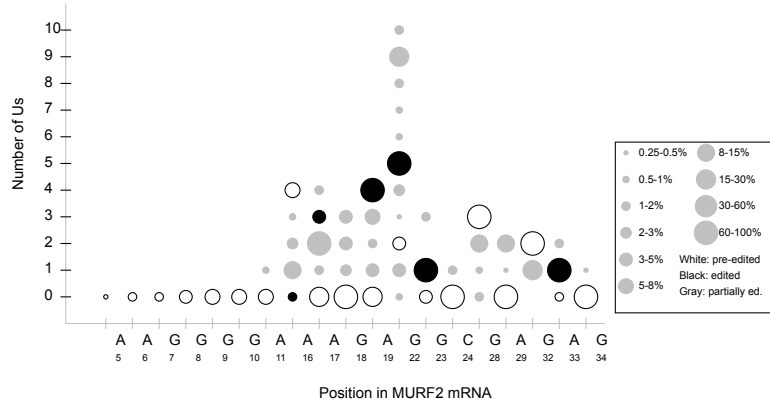


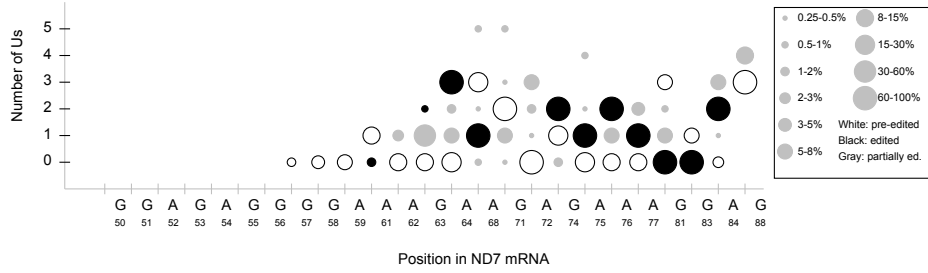
Figure S1. Cumulative growth curves of cell lines used in this study. No notable differences in growth among cell lines were observed.

427wt

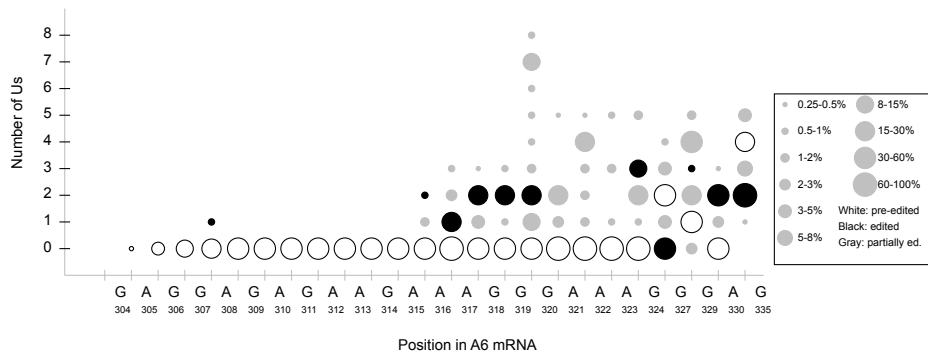
MURF2



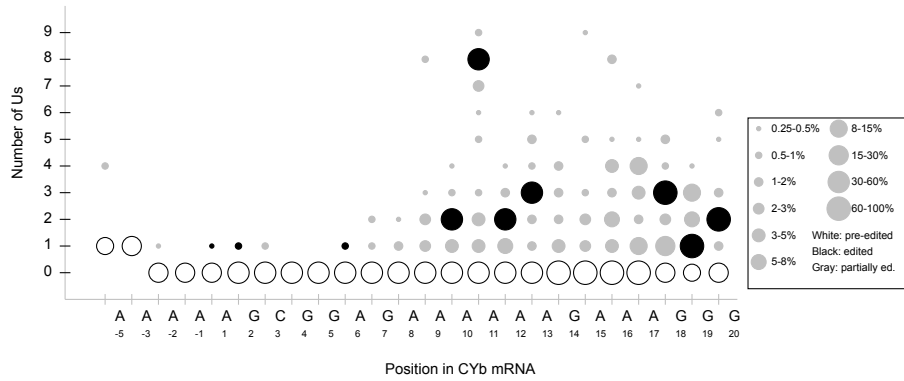
ND7



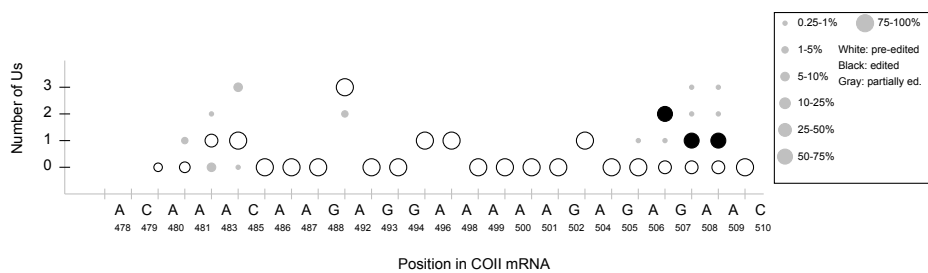
A6



CYb

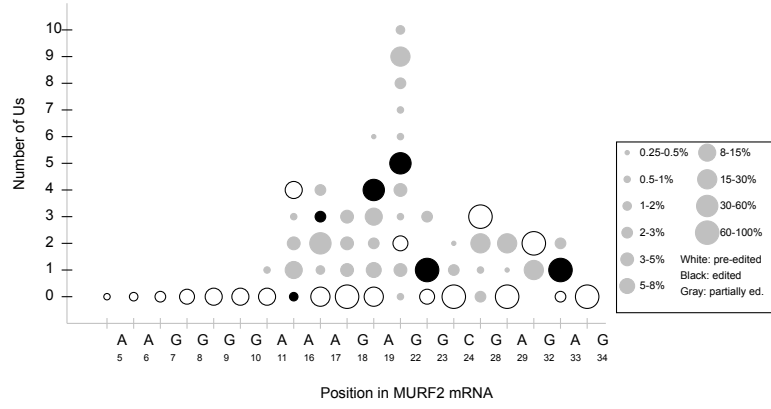


COII

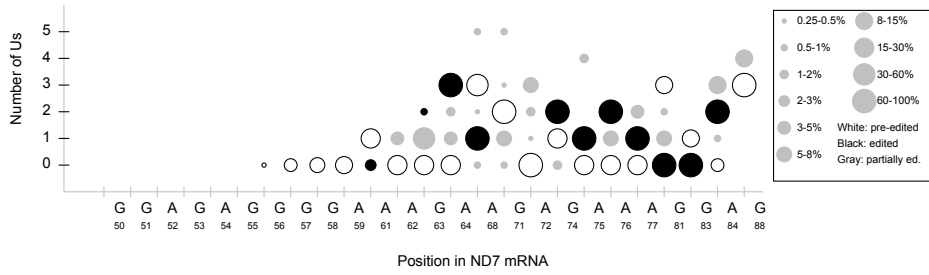


MGA

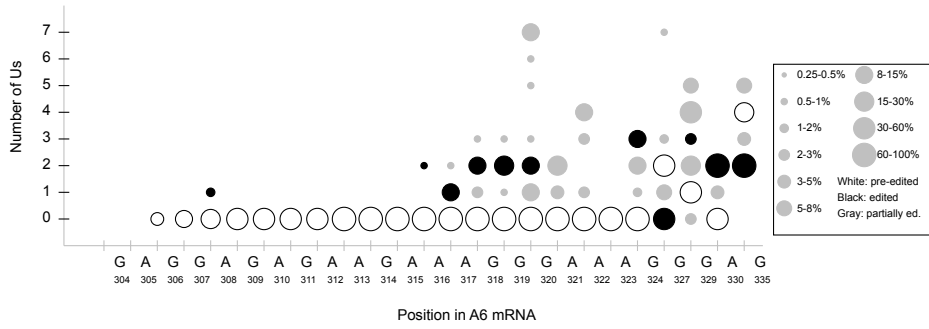
MURF2



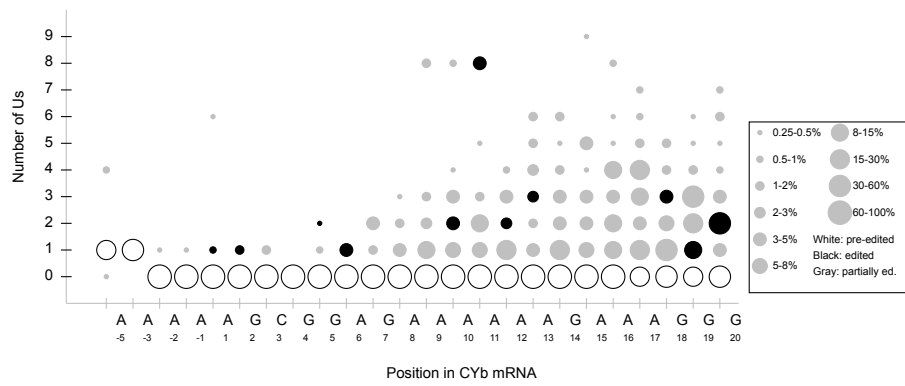
ND7



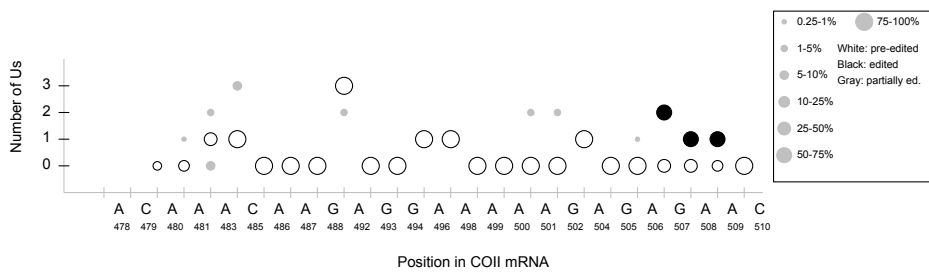
A6



CYb

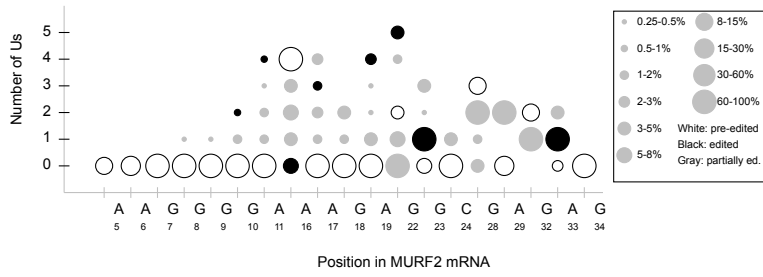


COII

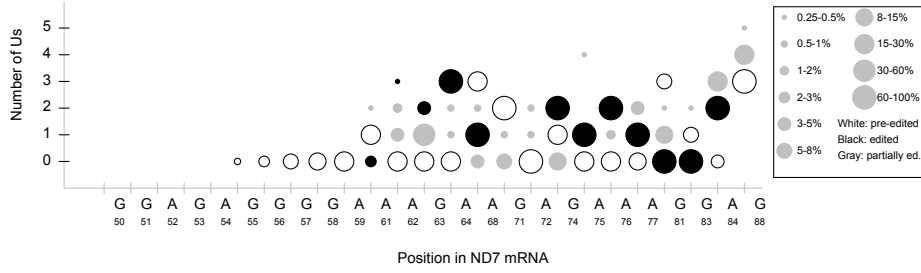


KREN2 null

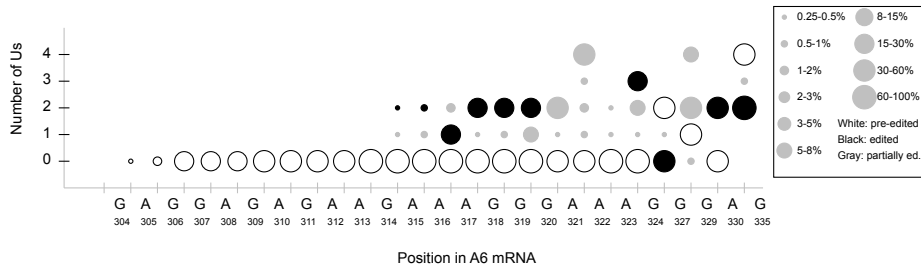
MURF2



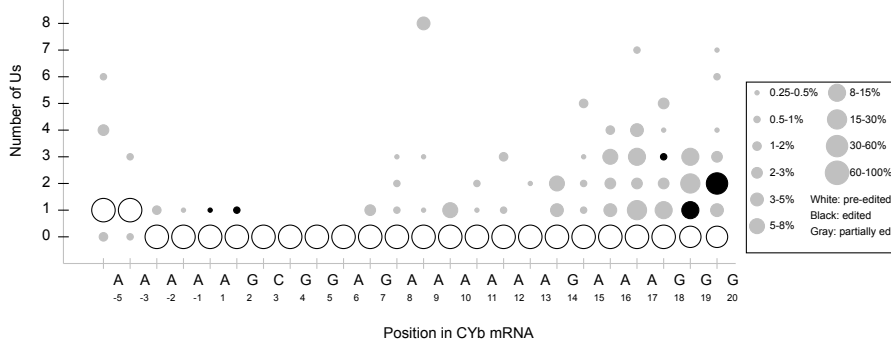
ND7



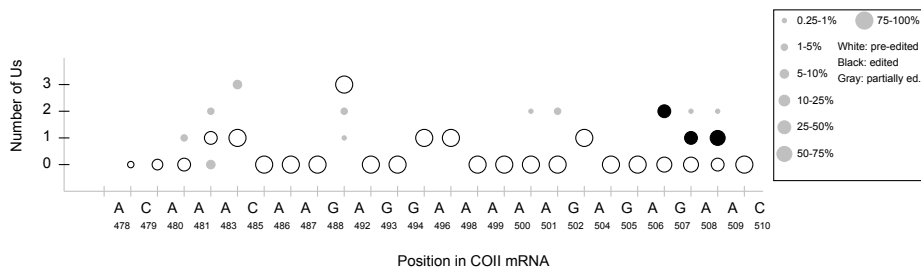
A6



CYb

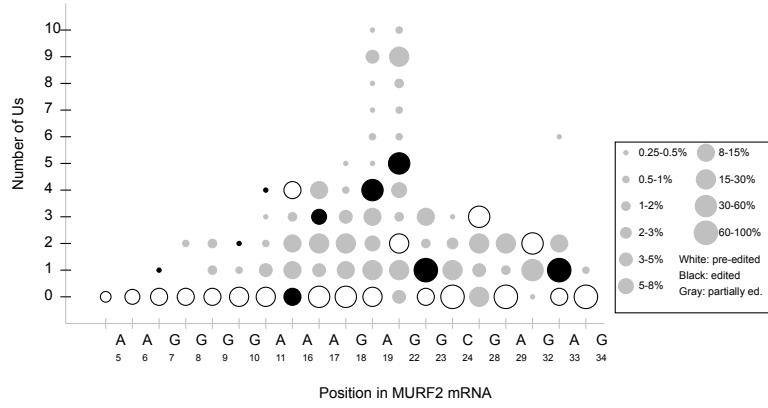


COII

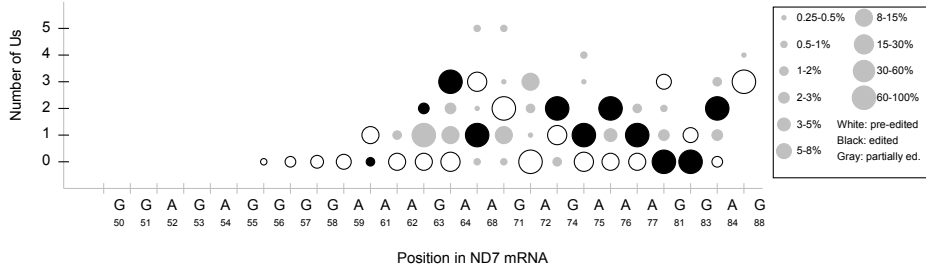


KREN3 null

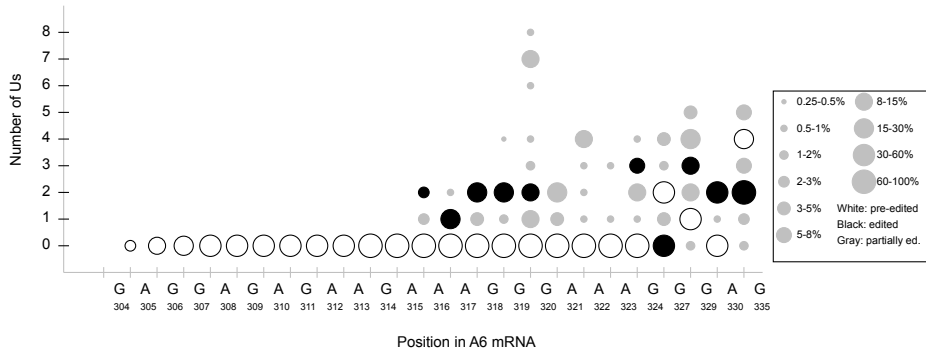
MURF2



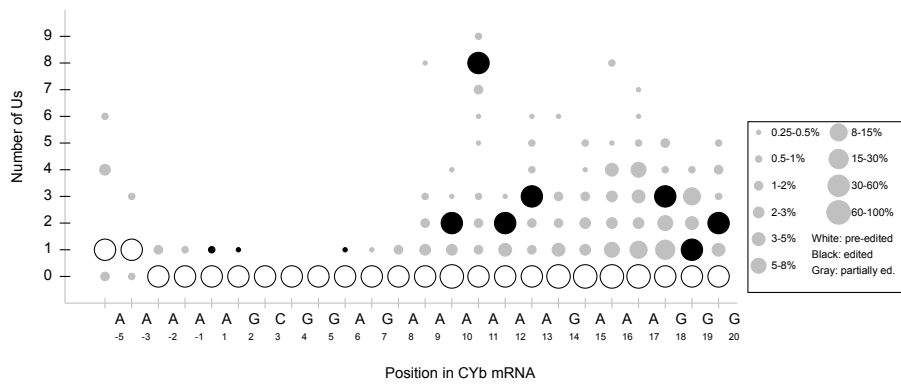
ND7



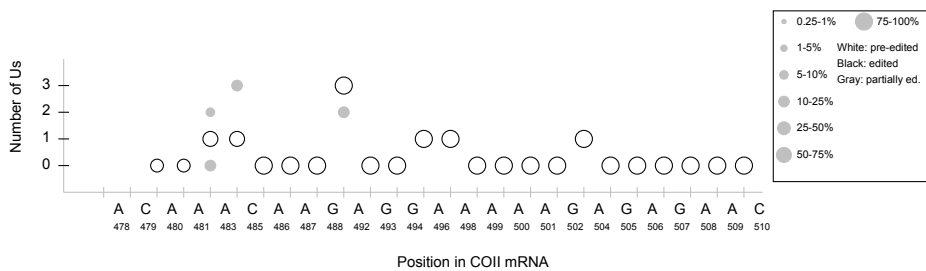
A6



CYb

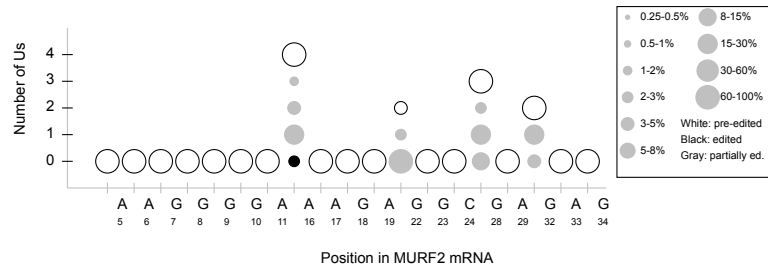


COII

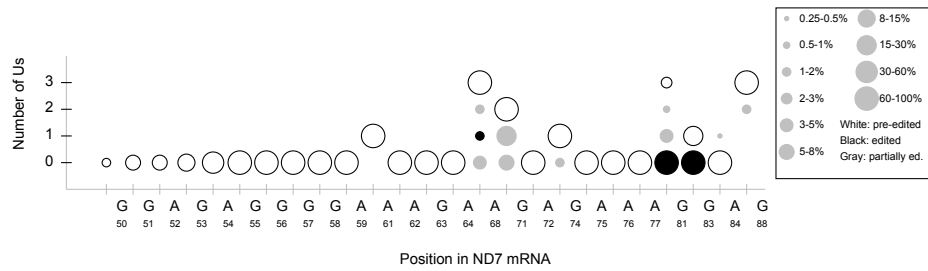


KREN1 only

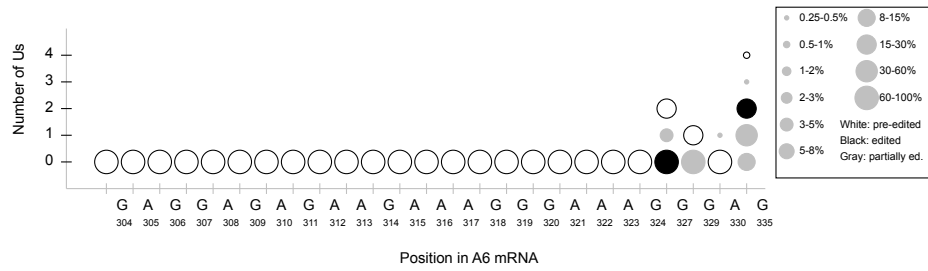
MURF2



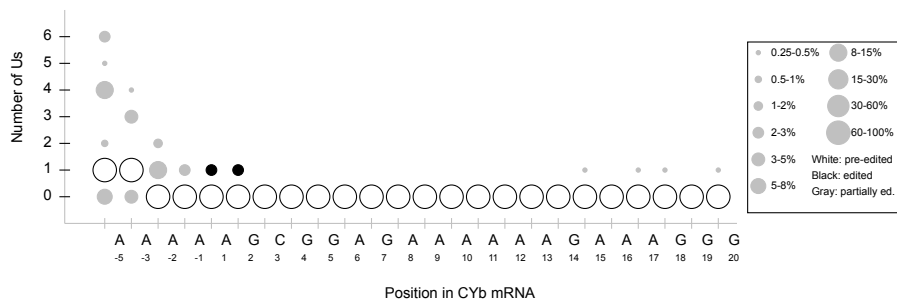
ND7



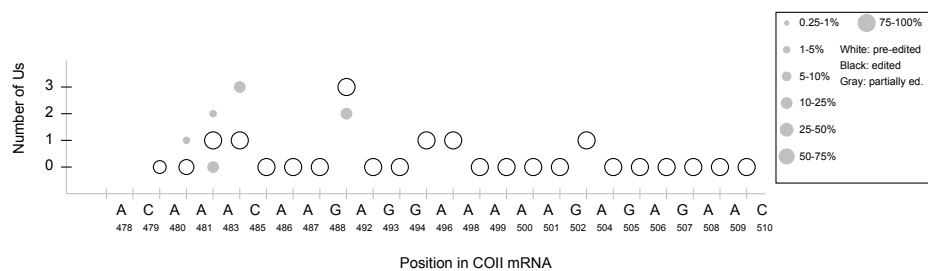
A6



CYb

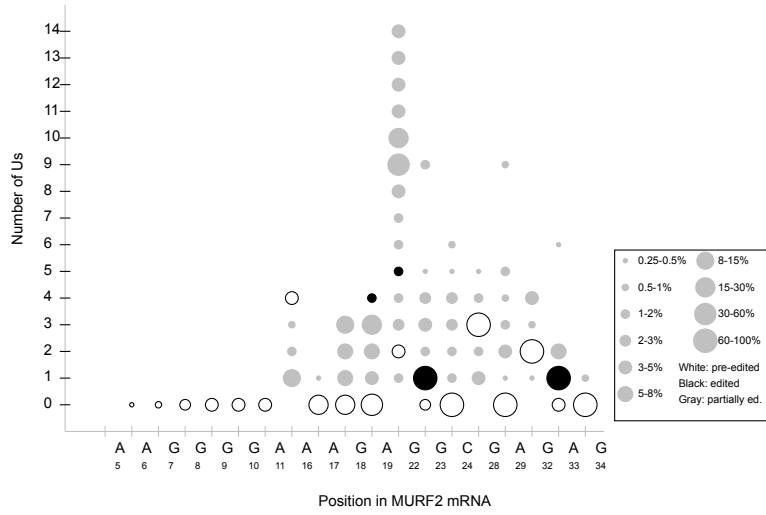


COII

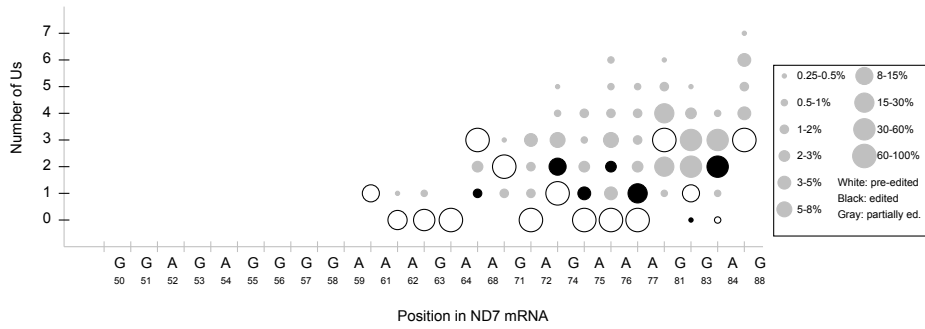


KREN2 only

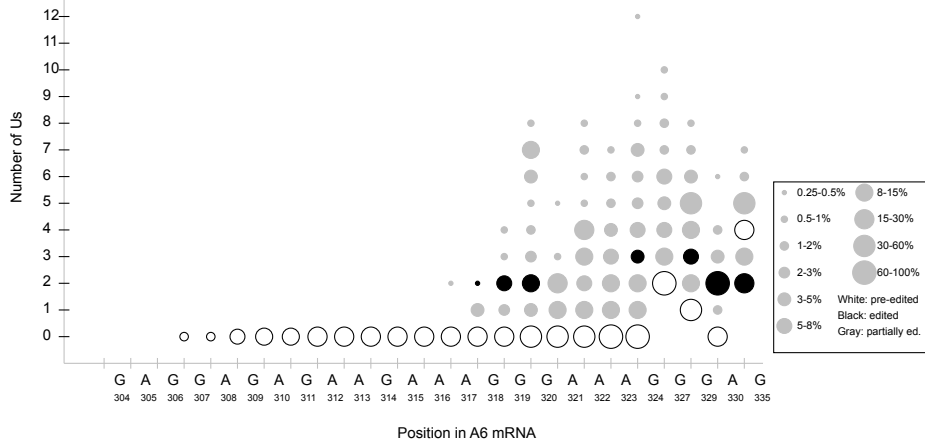
MURF2



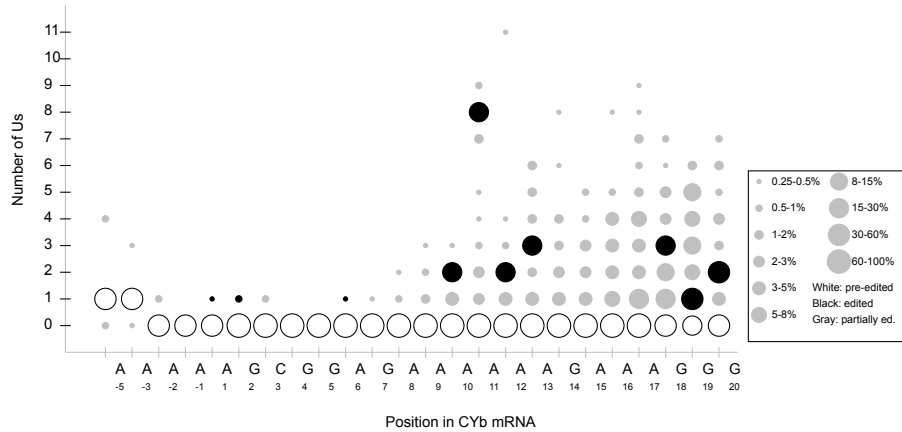
ND7



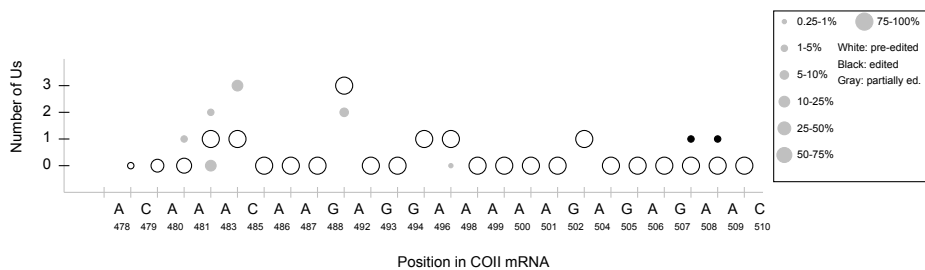
A6



CYb

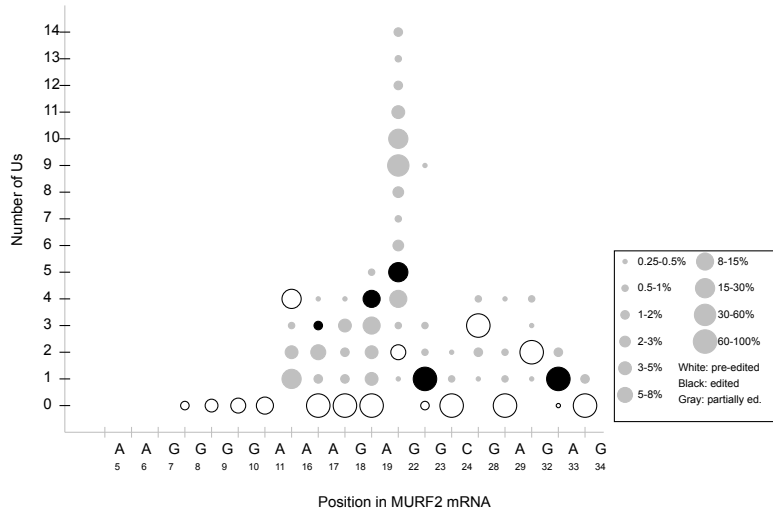


COII

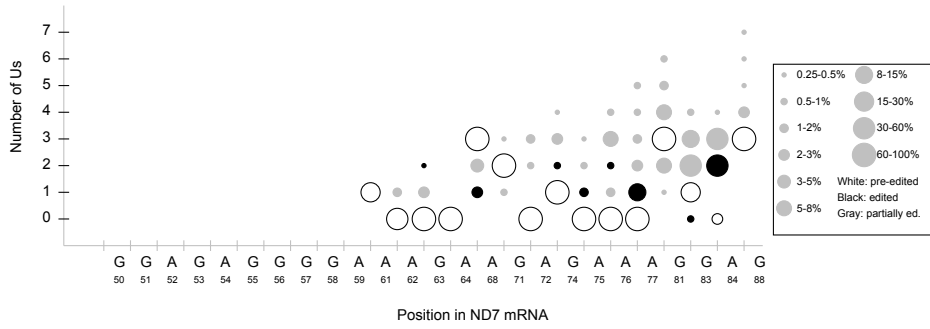


KREN3 only

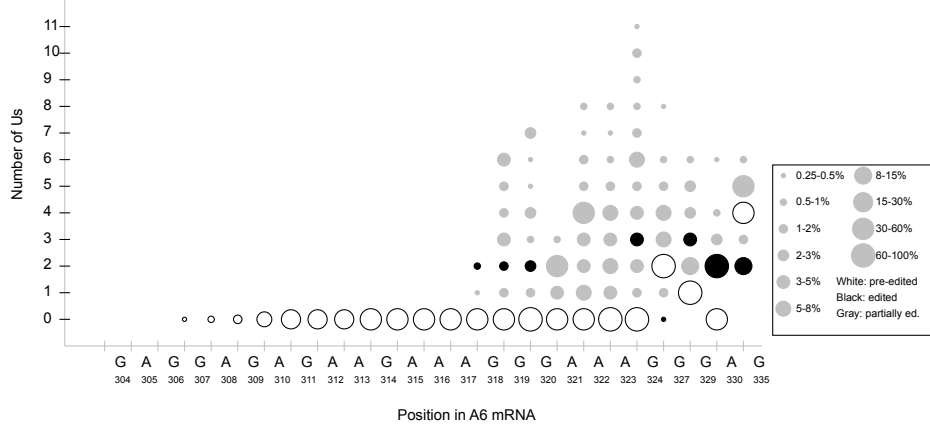
MURF2



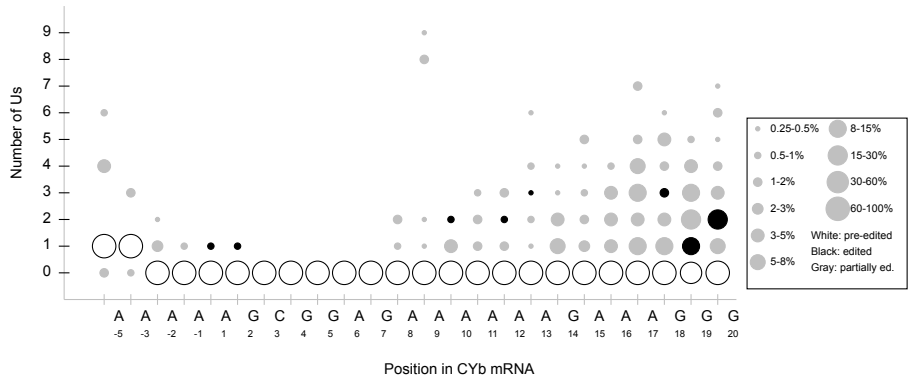
ND7



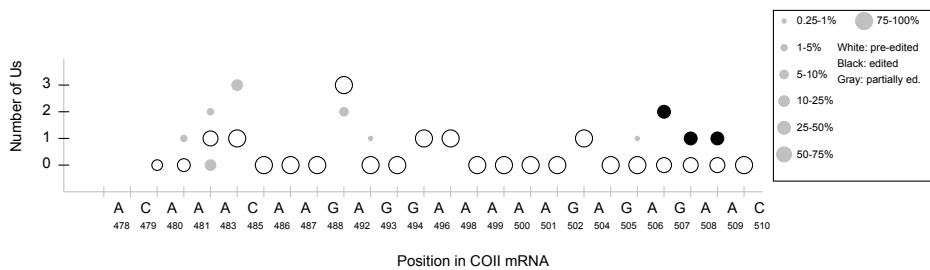
A6



CYb

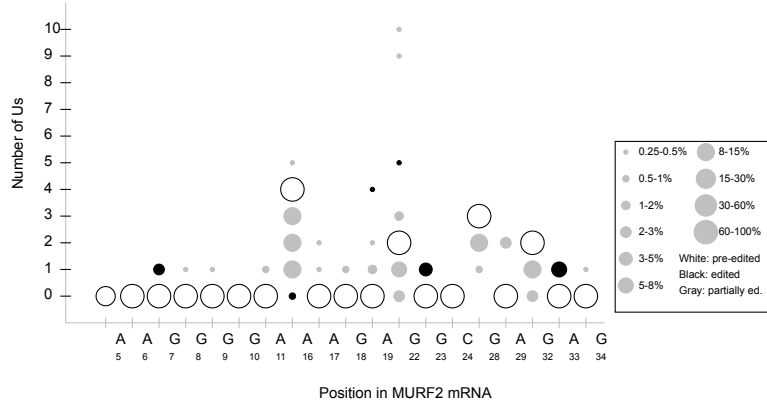


COII

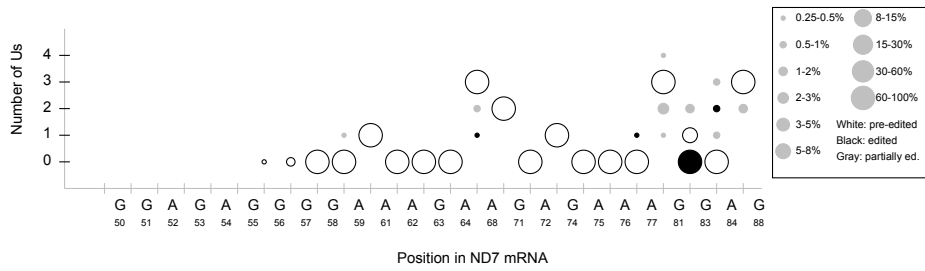


Triple null

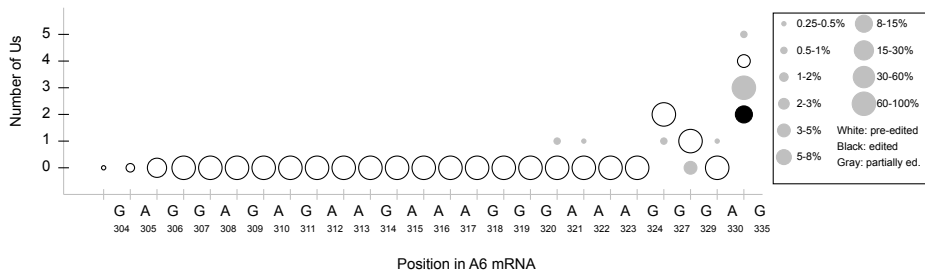
MURF2



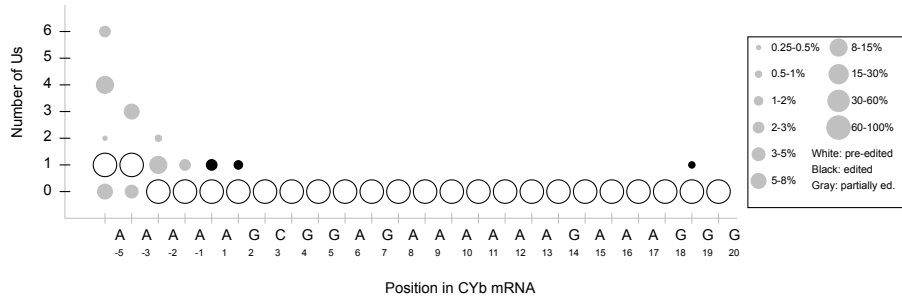
ND7



A6



CYb



COII

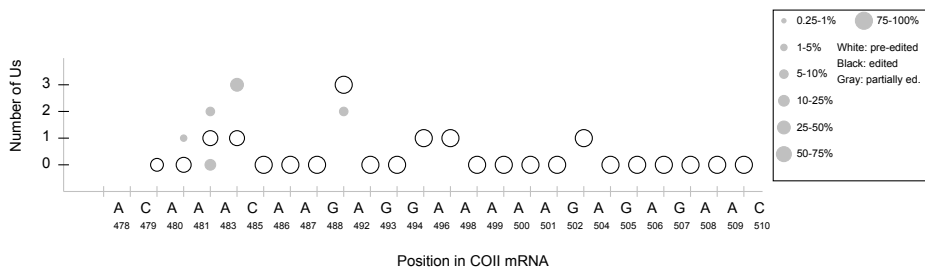


Figure S2. Bubble plot analyses of edited read sequences show the frequency of site-specific insertion and deletion activities. Bubble plots show number of Us inserted at potential editing sites at the 3' end of indicated transcripts for each cell line. In comparison to Figures 7-11, these plots include all of the sites in the dataset, not just the first 10 sites at the 3' end of the sequence. All mapped reads for each transcript are analyzed to determine the amount (number of Us, y-axis) and position (location within 3' end of edited region, x-axis) of editing events within each cell line. The x-axis shows the mRNA position using the non-U nucleotide sequence, with position numbering starting at the first base of the start codon in the pre-edited sequence. When the number of Us at a position matches the pre-edited number of Us, the bubble is white; when the number of Us is altered by editing, the number of Us matching a fully edited mRNA is colored black. When the number of Us at a position matches neither pre-edited nor fully edited sequence, bubbles are colored grey to denote partial editing. The size of the bubble correlates with the proportion of reads that have that number of Us at each position, with the legend giving the percent range for each size bubble. To decrease noise in these plots, data points that represent less than 0.25% of reads or fewer than 5 reads total are not shown.